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(54) **EQUINE PARASITE DETECTION**

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USPC ..... **435/7.22**; 435/7.92; 435/7.95

(58) **Field of Classification Search**

None  
See application file for complete search history.

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(74) *Attorney, Agent, or Firm* — Barnes & Thornburg LLP

(57) **ABSTRACT**

The present invention provides a method of diagnosing a *cyathostomin* infection, said method comprising the step of identifying a level of anti-*cyathostomin* larval antigen antibodies in a sample, wherein a level of anti-*cyathostomin* larval antigen antibodies is indicative of a *cyathostomin* infection.

**12 Claims, 11 Drawing Sheets**

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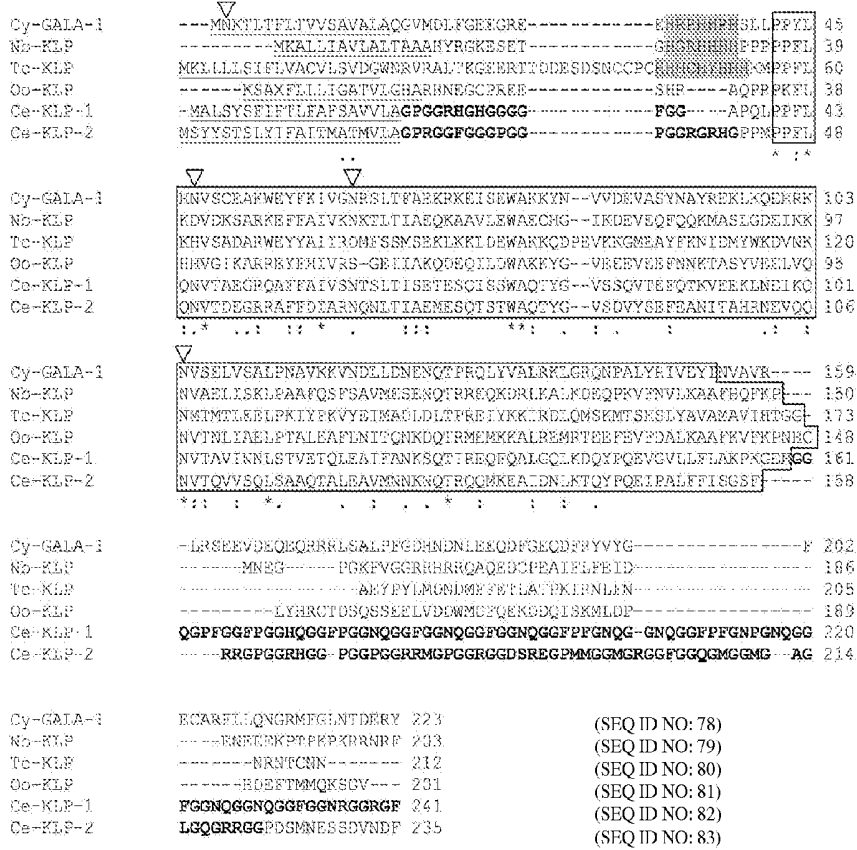


Figure 1

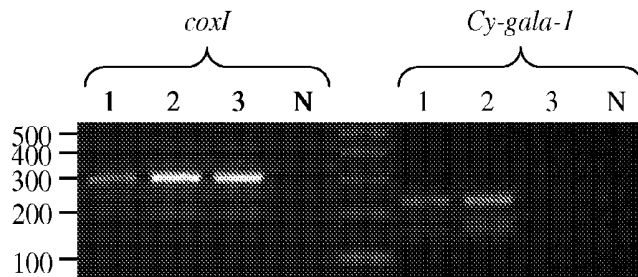


Figure 2

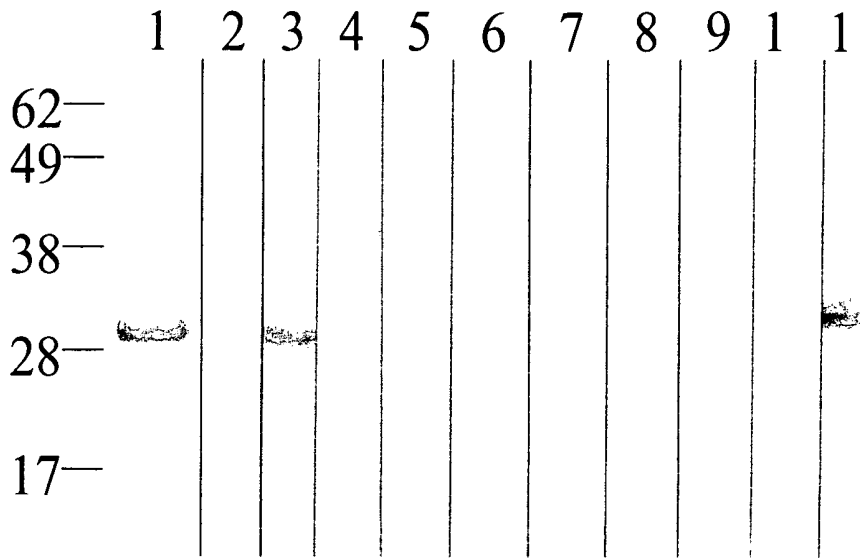


Figure 3A

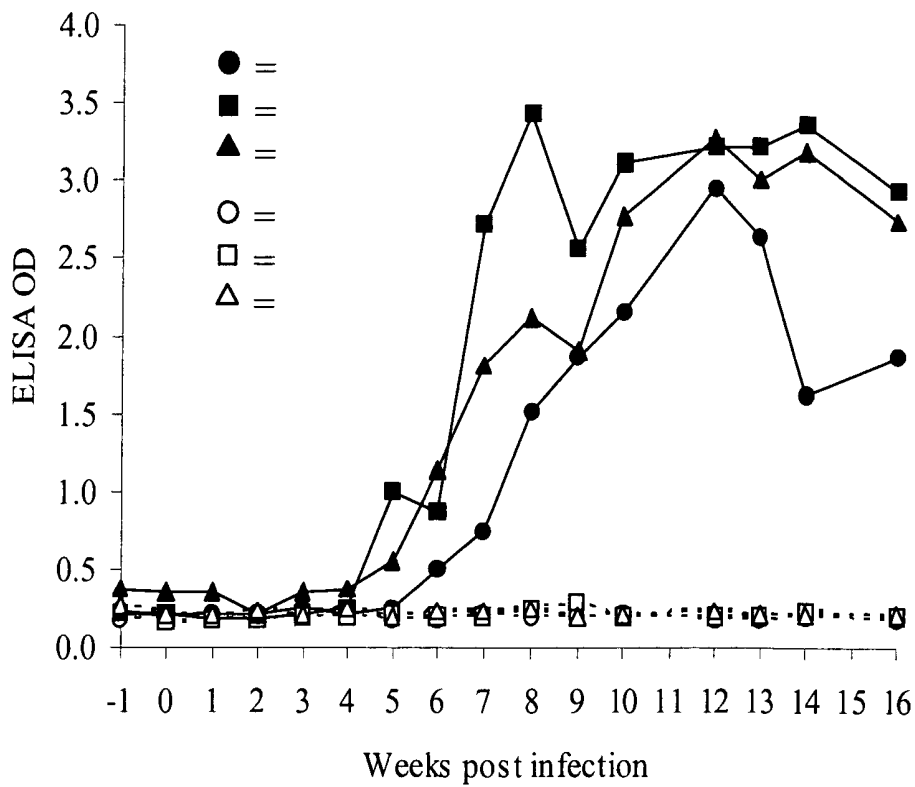


Figure 3B

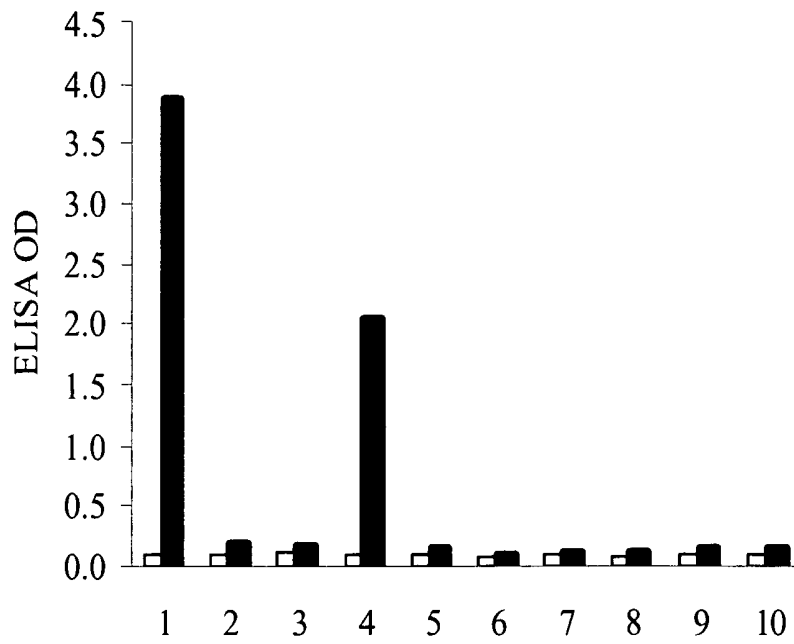


Figure 4A

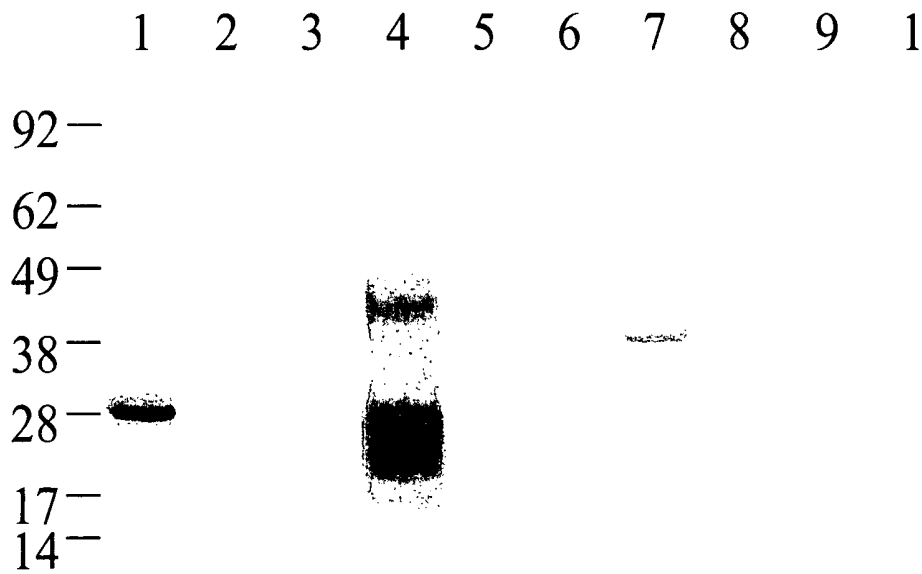


Figure 4B

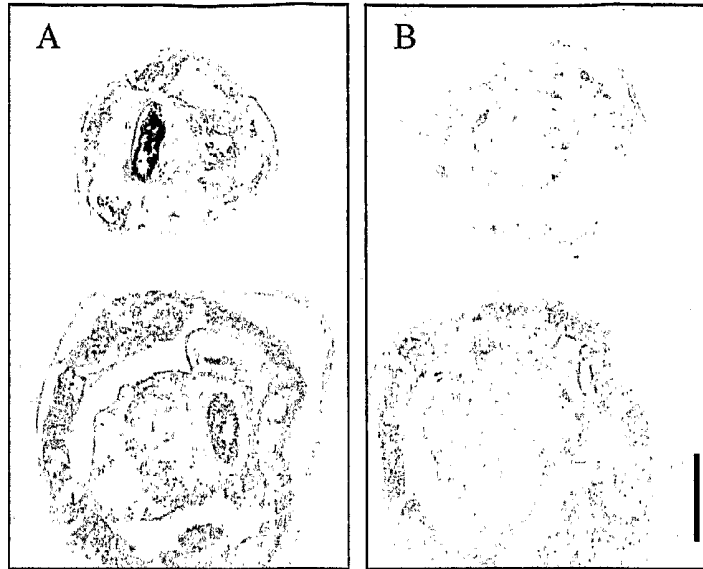


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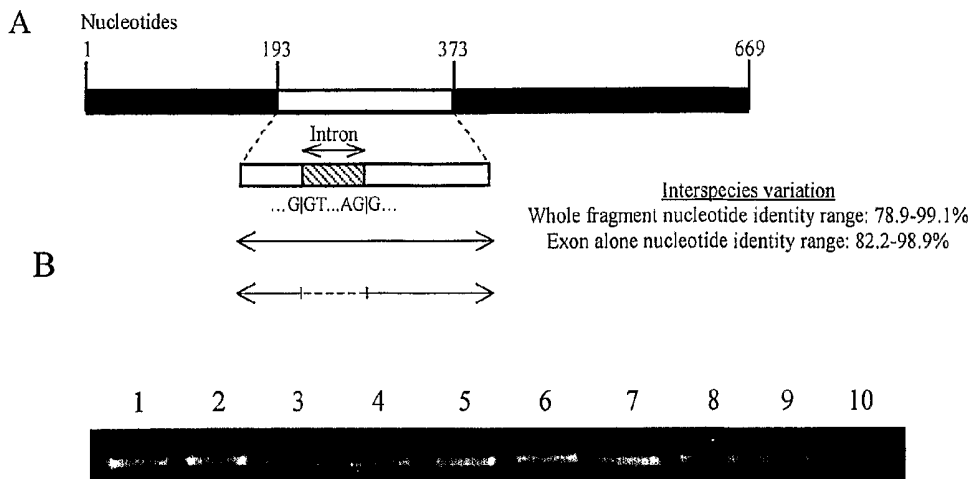
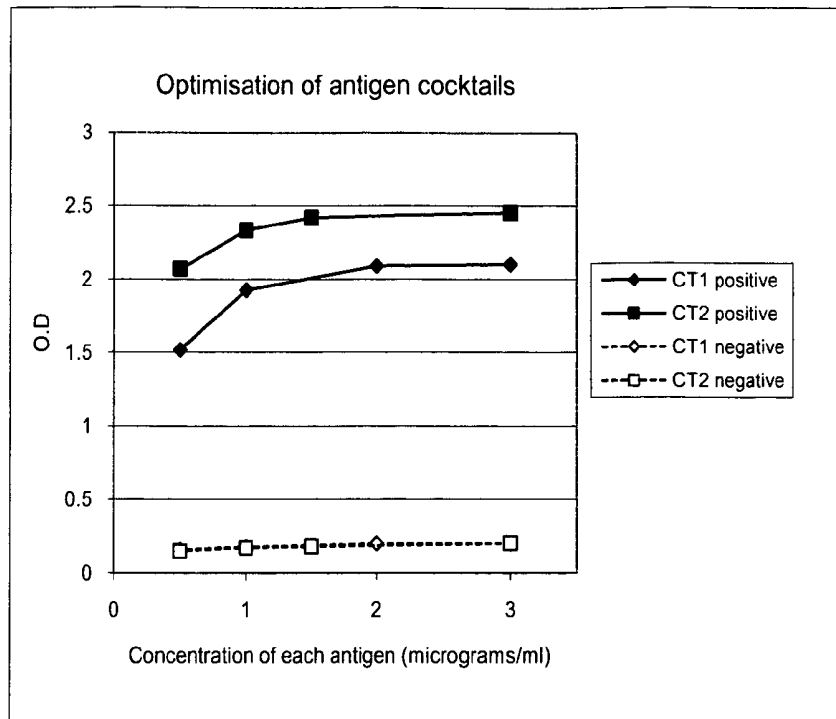
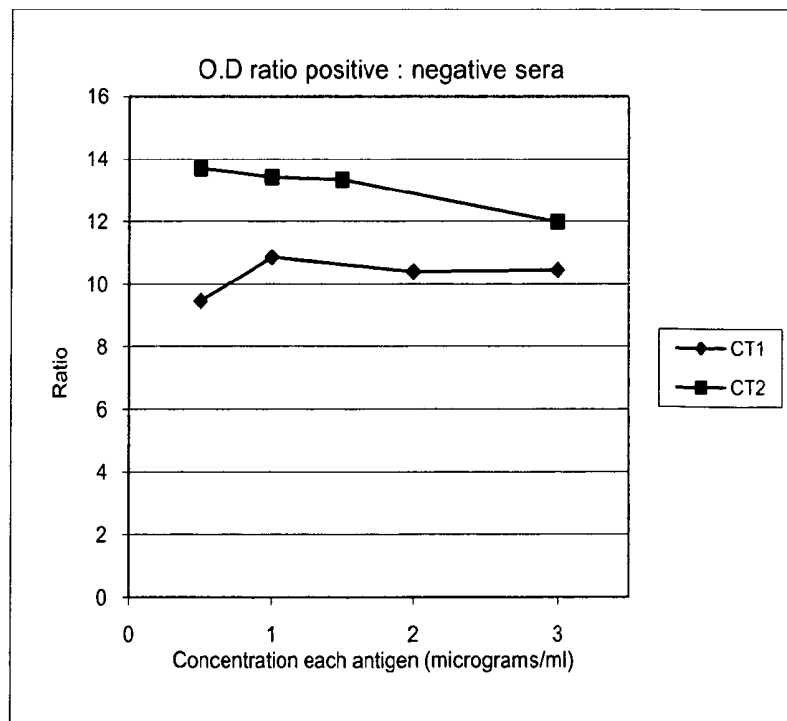


Figure 6



A



B

Figure 7 A & B

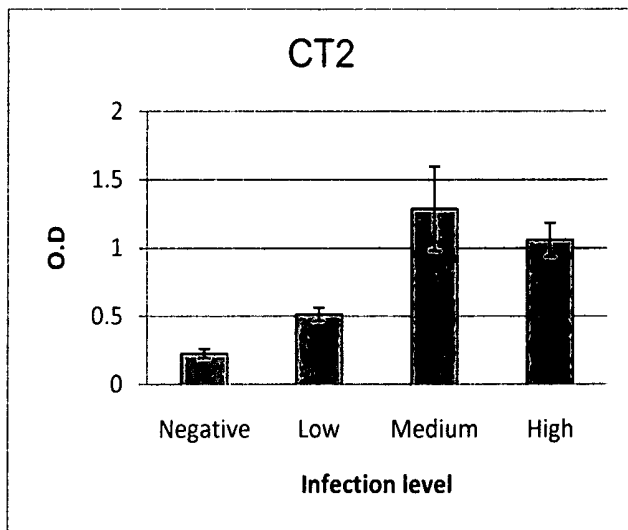
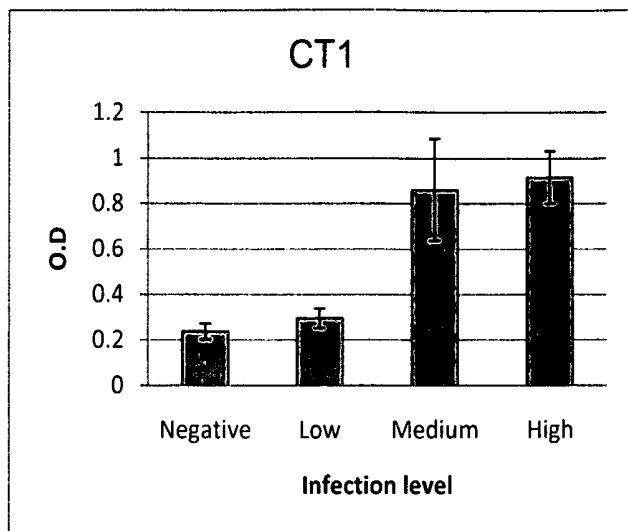


Figure 7 C and D

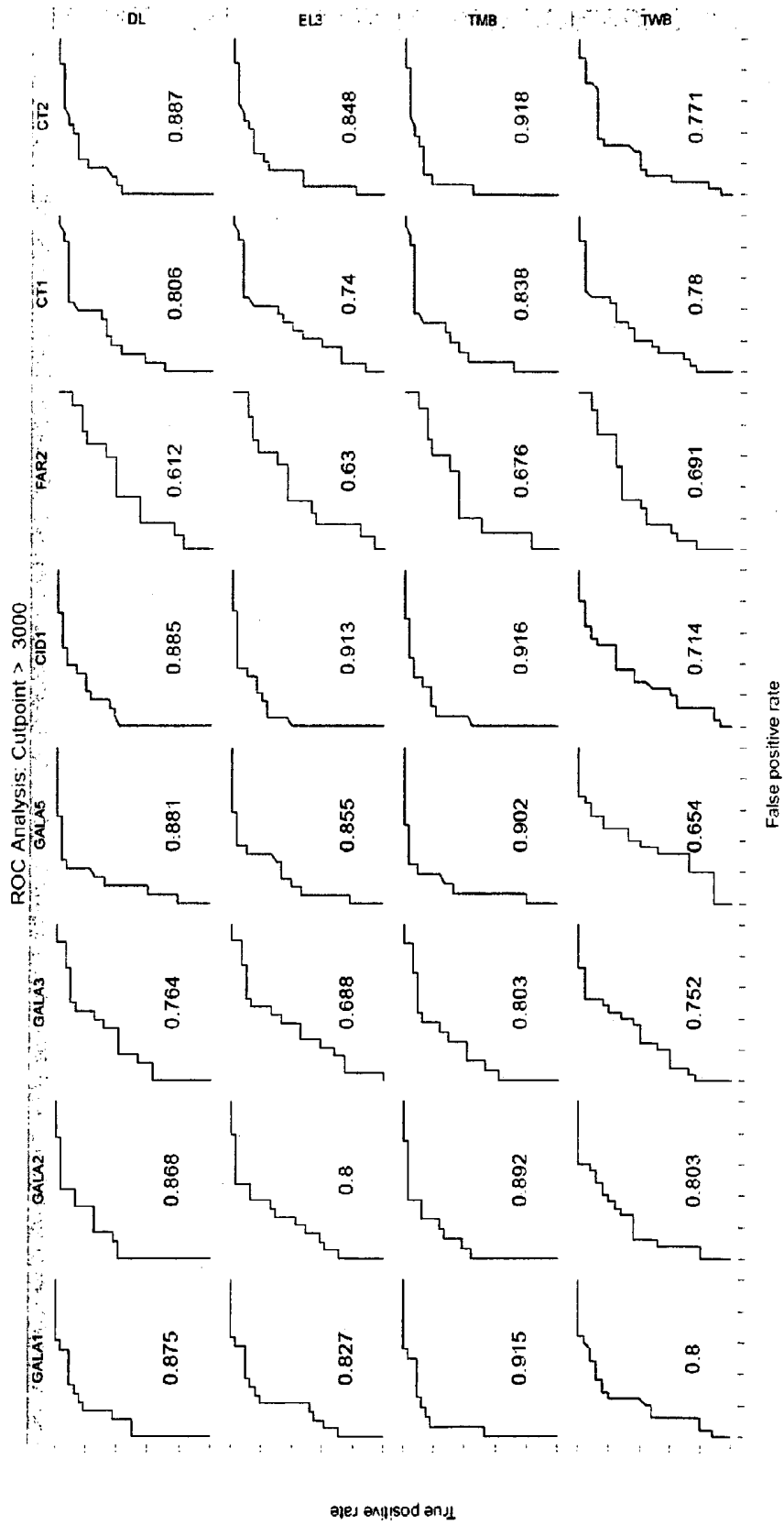


Figure 8A

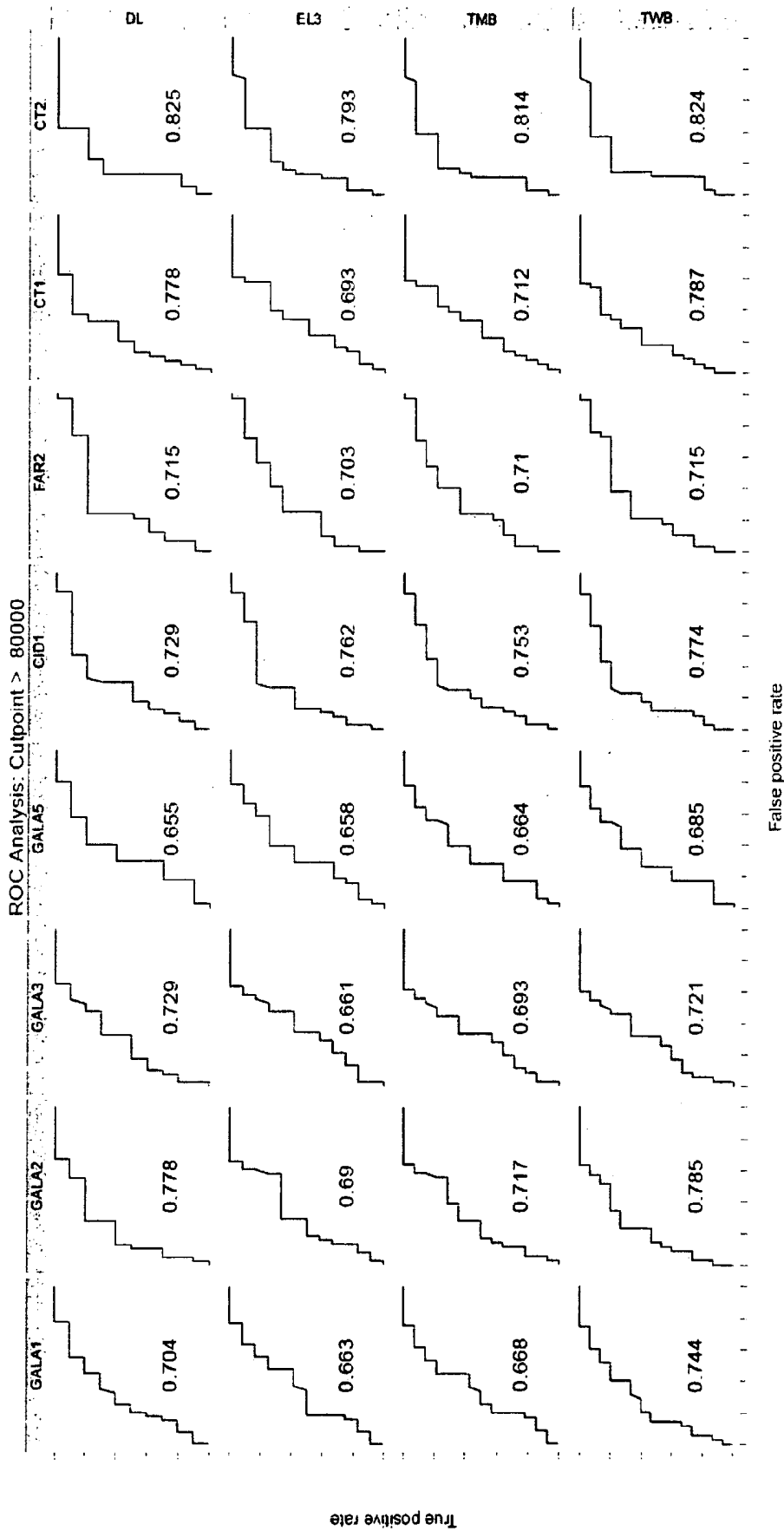


Figure 8B

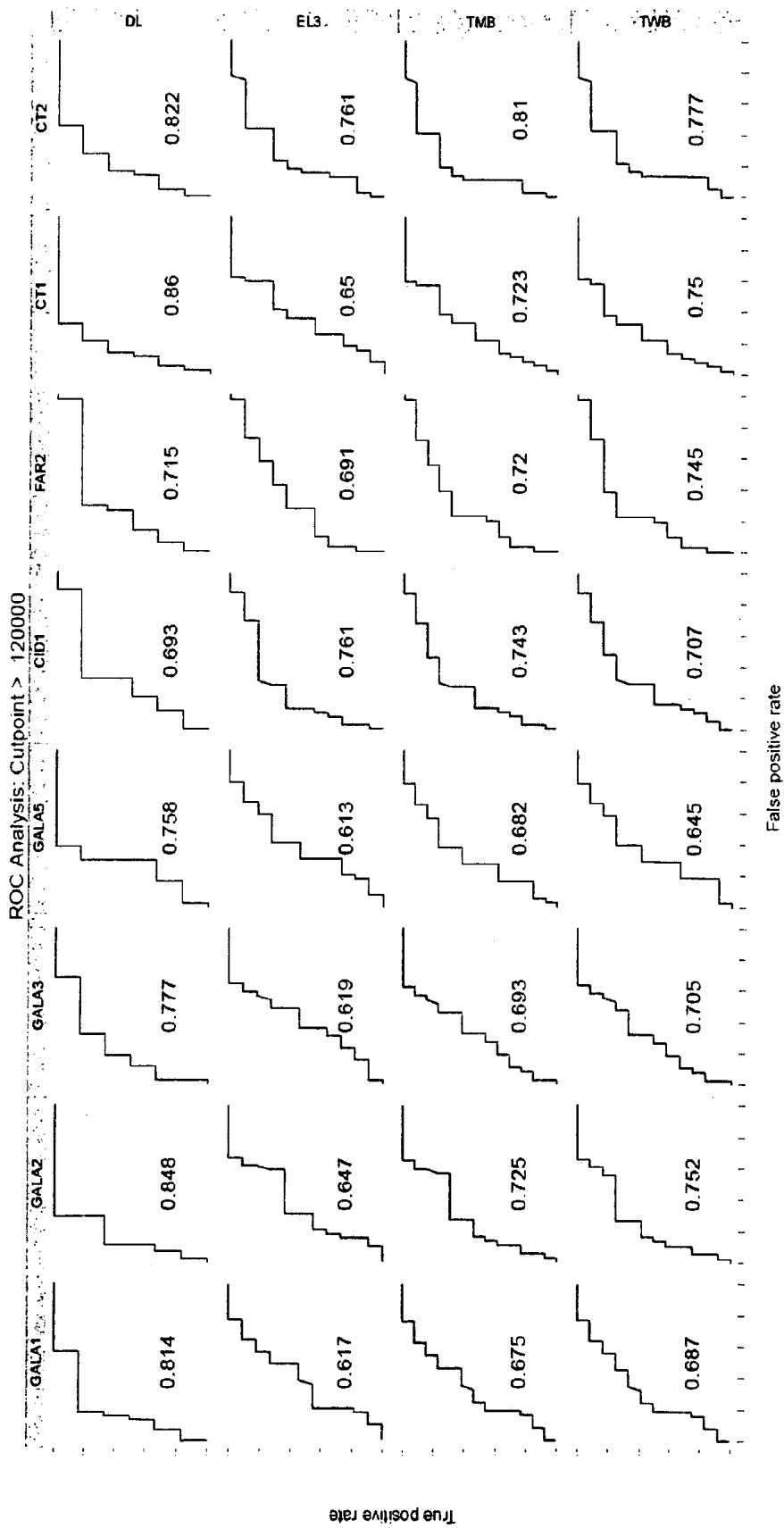


Figure 8C

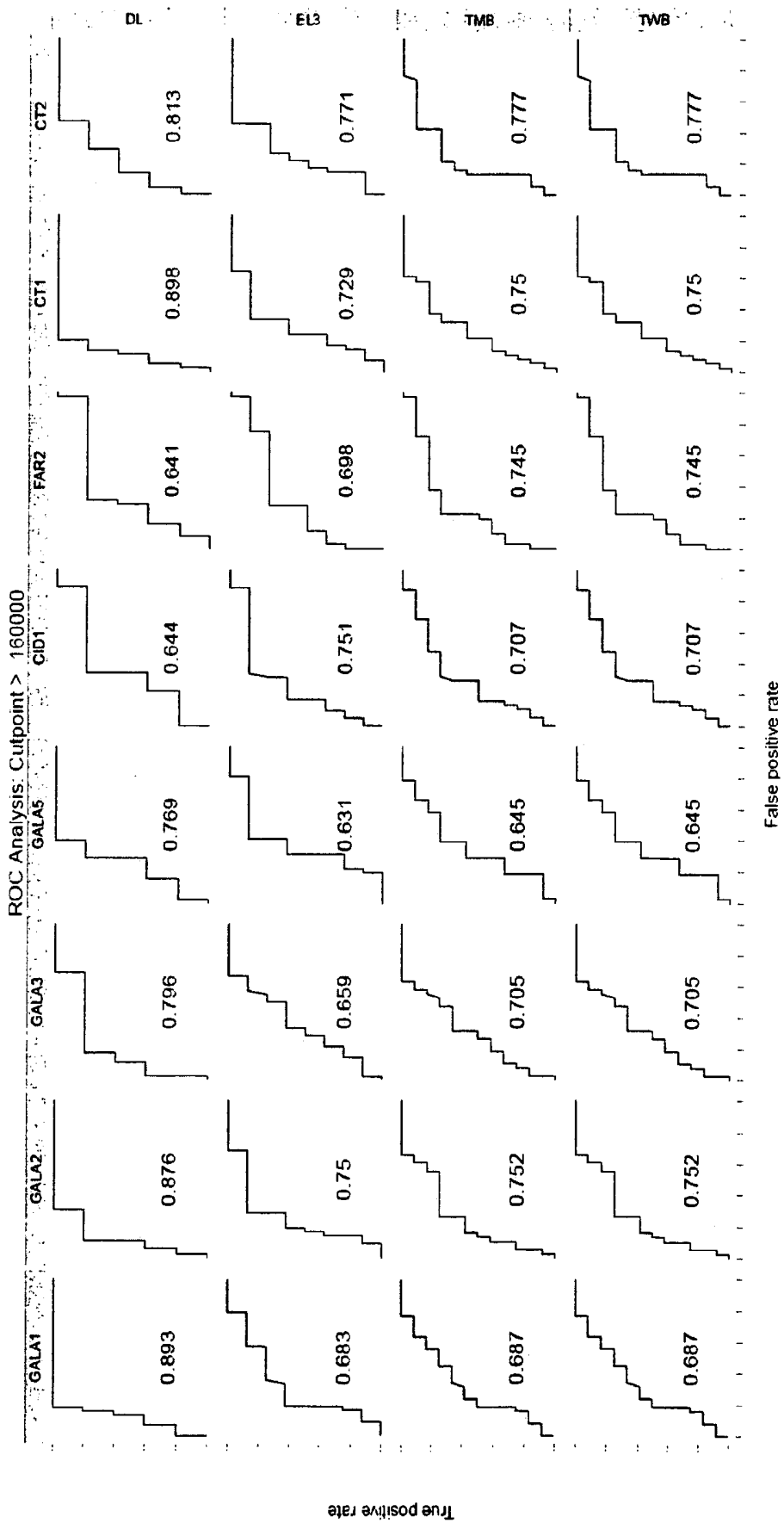


Figure 8D

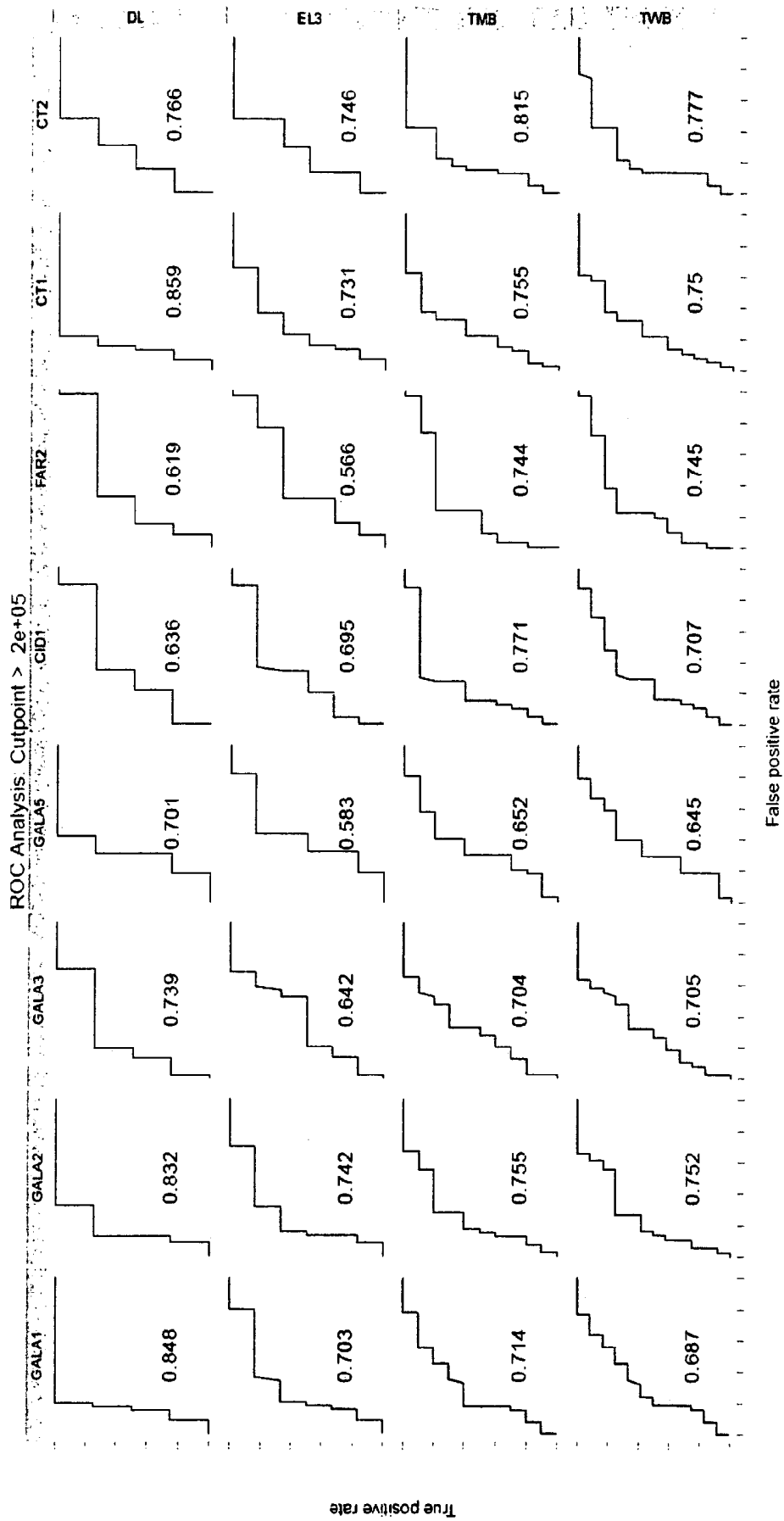


Figure 8E

## EQUINE PARASITE DETECTION

## CROSS-REFERENCE TO RELATED APPLICATIONS

This application is a U.S. national counterpart application of PCT international Application Serial No. PCT/GB2010/000616, filed Mar. 31, 2010, which claims priority to United Kingdom Patent Application Serial Number 0905511.2, filed Mar. 31, 2009, the disclosures of both which are hereby incorporated herein by reference.

## FIELD OF THE INVENTION

The present invention provides a means of detecting infections caused by parasitic nematodes belonging to the Cyathostominae group in Equine subjects and in particular in horses.

## BACKGROUND OF THE INVENTION

Members of the Cyathostominae group of nematodes infect almost all grazing horses. Most horses have burdens to the order of tens of thousands of cyathostomins and usually do not exhibit clinical disease, however, in some animals, infection leads to a severe inflammatory enteropathy [15]. This disease occurs following accumulation of *cyathostomin* larvae that encyst and undergo inhibited development as early third larvae (EL3) in the large intestinal wall. Vast numbers of encysted larvae can accumulate and these can reactivate simultaneously to cause an inflammatory enteropathy known as larval cyathostominosis. The principal effect of this syndrome is weight loss, but horses can exhibit other signs including diarrhoea, colic, subcutaneous oedema and/or pyrexia [25]. Up to 50% of animals with larval cyathostominosis die as a result of the condition [15]. This disease most commonly occurs in younger horses, however horses have a life-long susceptibility to infection and disease may occur at any age [15, 35]. Encysted larvae can persist for prolonged periods (up to two years in some cases) and it has been proposed that encystment is favoured by a variety of factors including; negative feedback from mature worms in the large intestinal lumen, a large larval challenge or a 'trickle' infection [29]. *cyathostomin* EL3 have limited susceptibility to several currently available anthelmintics [12, 19] and drug resistance is common, particularly with regard to benzimidazole and pyrantel compounds [17]. Moxidectin is now only drug available that has high efficacy against EL3, but for which resistance is not yet widespread. It is therefore important that the high efficacy of this anthelmintic be maintained for as long as possible.

To reduce the spread of anthelmintic resistance, it is important that only animals with moderate to high *cyathostomin* burdens are targeted strategically for treatment [32]. Targeted treatments can be undertaken on the basis of faecal egg counts however the latter have no value in estimating burdens of mucosal larvae. Indeed, horses with high mucosal burdens often have low or negative faecal egg counts [31] and there is no specific, non-invasive method to diagnose pre-patent *cyathostomin* infection. A diagnostic test for mucosal larvae would allow veterinarians to identify horses that require larvicidal anthelmintic treatments. Recently, we identified two larval antigen complexes (observed to migrate at 20 and 25 kDa by 1-dimensional SDS PAGE) that have diagnostic potential [9-11]. Significant increases in serum IgG(T) specific to these antigen complexes were observed as early as 6 weeks post infection (PI) in experimentally-infected ponies [11]. Antigens present in both complexes appeared to be specific for mucosal larval cyathostomins, indicating their utility as markers of pre-patent infection [11]. When serum IgG(T) levels were compared amongst groups of naturally- and experimentally-infected horses, there was a strong sig-

nificant correlation of anti-25 kDa serum IgG(T) responses with total mucosal burden, particularly EL3 burden [10]. In naturally infected horses, IgG(T) responses to both larval complexes were significantly greater than those in uninfected individuals [10] and IgG(T) levels to both complexes were significantly higher in larval cyathostominosis clinical cases than in helminth-naïve ponies and parasite-negative horses from an abattoir [10]. These results indicate that an immunoassay based on antigens present in these complexes could ultimately be used to differentially diagnose larval cyathostominosis, or used to target horses with high mucosal burdens for treatment. The native mucosal larval preparations are extremely time-consuming to prepare and rely on a continuous source of infected mucosa. Therefore, it would be advantageous if genes encoding proteins present in these complexes were isolated and cloned and the associated proteins expressed in recombinant form.

## SUMMARY OF THE INVENTION

The present invention is based upon the finding that parasitic nematodes belonging to the Cyathostominae group express proteins which can be used to diagnose, detect or identify incidences of *cyathostomin* infection in animals, particularly horses. Although *cyathostomin* infections are treatable, the range of effective drugs is rapidly diminishing and at present only moxidectin exhibits a high efficacy against the encysted *cyathostomin* parasite.

In order to ensure that the development of resistance to moxidectin can be delayed for as long as possible, it is essential that only animals with moderate to high *cyathostomin* burdens are targeted for treatment. However, the encysted larval stages of this parasite can remain undetected for months or even years eventually emerging from the intestinal wall to cause severe pathology (including symptoms of diarrhoea, weight loss, colic, oedema and pyrexia); as such, it is often difficult to know whether or not a particular animal should be treated.

The inventors have identified a number of proteins that are expressed predominantly during the mucosal larval stages (i.e. the early third larval (EL3) and late third (LL3)/developing fourth (DL4) stages). These proteins are highly immunogenic and exhibit low cross-reactivity to proteins present in other helminth species.

Accordingly, a first aspect of this invention provides a method of diagnosing a *cyathostomin* infection, said method comprising the step of identifying a level of anti-*cyathostomin* larval antigen antibodies in a sample, wherein a level of anti-*cyathostomin* larval antigen antibodies is indicative of a *cyathostomin* infection.

Animals positively diagnosed as having a "*cyathostomin* infection" by the method provided by the first aspect of this invention may harbour high numbers of encysted *cyathostomin* in the gut mucosa, particularly the large intestinal wall, as such they may generate a significant immune response to *cyathostomin* antigens including any antigens produced by the EL3, LL3 and DL4 stages. Animals with infections of this type may otherwise be referred to as having high mucosal burdens. In other instances, positive diagnoses may indicate animals with larval cyathostominosis, an inflammatory enteropathy manifesting with symptoms of weight loss, diarrhoea, colic, subcutaneous oedema and/or pyrexia. Conditions of this type are often fatal if untreated.

In one embodiment, "a level" of anti-*cyathostomin* larval antigen antibodies may be evaluated relative to the "a level" of anti-*cyathostomin* larval antigen antibodies present in reference or control samples derived from healthy animals or animals not having high mucosal burdens of *cyathostomin* parasites or larval cyathostominosis. In this way levels and, in particular high levels, of anti-*cyathostomin* larval antigen antibodies, may easily be detected. Accordingly, the term "a level" may be taken to include levels of anti-*cyathostomin*

3

larval antigen antibodies which are less or greater than levels of anti-*cyathostomin* larval antigen antibodies identified in reference or control samples.

It should be understood that in addition to providing methods in which levels of anti-*cyathostomin* larval antigen antibodies are detected in samples, the present invention might also be adapted to provide methods in which levels of *cyathostomin* larval antigens are detected in samples. Methods of this type, rather than “indirectly” diagnosing *cyathostomin* infections via immune responses, may provide a more direct means of diagnosing *cyathostomin* infections. As above, “a

4

level” of *cyathostomin* larval antigens may be taken to include levels of anti-*cyathostomin* larval antigen which are less or greater than levels of anti-*cyathostomin* larval antigen identified in reference or control samples.

5 The *cyathostomin* larval antigens described/mentioned herein may be derived from larval antigen complexes having a molecular weight of about 20 to about 25 kDa. An exemplary larval antigen is obtained from the parasite *cyathostomin pateratum* and comprises or consists of the following amino acid sequence (designated SEQ ID NO: 1):

SEQ ID NO: 1

MNKTLTFLTVVSAVALAQGVMDLFGEEGEEHRRHRSLLPPYLHNVSCEAKWEYF  
 KIVGNRSLTFAEKRKEISEWAKKYNVDEVASYNAYREKLKQEHRKNVSELVSALPN  
 AVKKVNDLLDNENQTPRQLYVALRKLGRQNPALYRIVEYINVAVRLRSEEVDEQEQR  
 RRLSALPFGDHDNLEEQDFGEQDFRYVYGFECARFLLQNGRMFGLNTDERY

20 The nucleic acid sequence encoding the protein provided by SEQ ID NO: 1 has also been determined and is given as SEQ ID NO: 2 below.

SEQ ID NO: 2

Atgaacaaaacgttaacatttctcacagtcgtagtgccgtagctctggcccaaggt  
 gtcattggaccttttgggtgaagagggctcgtgaagaacatcgtcgtcaccatcgtcat  
 tcacttttaccaccatctccacaatgtgagctgtgaggctaaatgggagtagcttcc  
 aaaattgtgggaacaggagttgaccttctgctgagaaaagaagaaattagcggag  
 tgggcaaaaaatacaatggtggtgagtgcaagctacaatgcttacagggaa  
 aaactcaagcaggagcacagaaaaacgtagcgaactgtttctgctcttccaaac  
 gcagtgaaagaagtcaatgatctcttagacaatgaaaatcagactcctaggcaactt  
 tacgtagcccttagaaaacttggtagacaaaatccggcactttaccgtagttgtagag  
 tacattaatgtggctgaagactaagaagtgaagaagtgagtagcaagaacaacga  
 agaaggctgtcagctctacctttggcgaccataacgataaatttggaaagagcaggac  
 ttcggtgaacaagacttctgctatgtctatggcttgagtggtgcaagatttctcctt  
 caaaatggaagaatggttgacttaacacagatgaaagatat

45 One of skill in the art will appreciate that while SEQ ID NO: 1 represents the entire coding sequence of an exemplary *cyathostomin* larval antigen, after removal of the signal peptide the mature antigen may comprise 206 amino acids yielding a protein having a molecular weight of approximately 25.6 kDa.

50 In addition, the inventors have isolated homologous antigens from other *cyathostomin* spp., and the amino acid sequences of these are provided below as SEQ ID NOS: 3, 5 and 7 respectively. In addition, the nucleic acid sequences encoding each of the proteins encoded by SEQ ID NOS: 3, 5  
 55 and 7 have been designated SEQ ID NOS: 4, 6 and 8 respectively and each is detailed below.

SEQ ID NO: 3

HEELRHHRSLLPPYLHNVSCEAKWEYFKIVGNRSLTFAEKKKGSSEWAKKYNVVD  
 EVASYNAYREKLKQEHRKNVSELVSLPGAVKKVNEILLDNENQTPRQLYVALRKLKGLK  
 QNPVLYRVVEFVNLVVRFRREDSDEQEQLSTLPSFENNEEQDLGEQDFQYIYGF  
 ECARFIFQNGRMFGLNTDRRY

The antigen encoded by SEQ ID NO: 3 was isolated from *Cylicocyclus nassatus*.

SEQ ID NO: 4

Catgaagaacttcgctcgtcaccatcgctcattcacttttaccaccctatctccacaat  
 gtgagctgtgaagcacaatgggaatacttcaagattgtggggaacaggagcttgact  
 tttgctgaaaagaagggaagtagcgagtgaggcaaaaaatacaatggttgatg  
 gaagttgcaagttacaatgcctatagagaaaaacttaagcaggagcacaggaaaaac  
 gttagcgaacttgtttctggtcttcccggtgctgtgaagaaagtaacgaactcttg  
 gataatgagaatcagactcctaggcaactttacgttgctctaagaaagcttggtaaa  
 caaaatccagtaactctaccgtgtgtcgagttgtcaatttggttgagatttaga  
 cgtgaagattcggatgagcaagaacaacgagaaatgctgtcaactttacctttcagc  
 gaaaaaatgaagagcaggaccttggtgaacaagacttccagtacatctatggtttt  
 gaatgtgcaagattcatctttcaaatgggagaatggttgactcaacacggataga  
 agat at

SEQ ID NO: 5

SCVAKWEYFKIVINRSLTFAQRKEEISKWAKKYKVEDEVASYNAYREKLKQOHRKNV  
 SELVSNLPGAVERVNKLLDNENQTPKQLYLALRELKQNPALYHVVEYVNVVRLKR  
 EELDQQDQRRALSGSLFGENNDNLEEQDFGEEDFRVYVGFECARFILQNGRMFGLNM  
 DRNY

30

The antigen encoded by SEQ ID NO: 5 was isolated from *Coronocyclus coronatus*.

SEQ ID NO: 6

Agctgtgtggctaagtgggagtaacttcaagatcgtgatgaacaggagctcagcgttt  
 gctcaaaagaaggaagaaattagcaagtgggcgaaaaatacaaaagttgaggatgaa  
 gttgcaagctacaatgcttatagagaaaaactcaagcagcagcacaggaaaaacgtt  
 agcgaacttgtttctagtcttcccggtgcaatggaaagagtgaacaaaactttggac  
 aatgaaaaaccagacccttaagcaactttaccttgccctacgagaacttggaacaaa  
 aatccggcactttaccatgtgtcgagatgtcaatgtggttgtagacttaaacga  
 gaagaattggatgaacaagatcaatgaagagcgtgtcgggttcactttttggcgag  
 aataacgacaatctagaagagcaggactttggtgaagaagacttctogctatgtctat  
 gggtttgaatgtgcaagattcatccttcaaatggaagaatggttggtctaaacatg  
 gataggaattat

SEQ ID NO: 7

GEEDREEHRRHRHSLPPLYLHNVCVAKWEYFRIVGNRSLTFAEKKKEISEWAKKY  
 NVLDEVASYNAYREKLKQHRKNVSELVSDLPKAVKKNVNDLLDNENQTPRQLYVALR  
 ELGRQNPPLYRIVEYINVAVRRRSEELDEQEQRRLSALPFGDNDNLEEQDFGEQD  
 FRYVYGFECARFLQNGRMFGLNTERD

The antigen encoded by SEQ ID NO: 7 was isolated from *Cyathostomum catinatum*.

SEQ ID NO: 8

Gaggatcgtgaagaacatcgccgtcaccatcgctcattcactcttgccaccatctc  
 cacaacgtgagctgtgtggcacaatgggaatactttagaattgtggggaacaggagt

- continued

ttaacgctttagcgagaaaaagaagaatagcgagtgaggcaaaaaatacaatgtt  
 ctggatgaagtagcaagctacaatgcttatagggaaaaactcaagcaggagcacaga  
 aaaaaagcttagcgaactgtttctgatcttccaaggcagtaagaaagctcaacgat  
 cttctagacaatgaaaatcagactcctaggcaactttatggtgaccttagagagctt  
 ggtagacaaaaatccgacactttaccgtattgtcgagtacatcaatgtggctgaagg  
 cgaagaagtgaagaactggatgagcaagaacaaggaagaaggctgtcagctttacct  
 ttcggcgacaacaacgataaattggaagagcaggacttcggtgaacaagactttcgc  
 tatgtctacggctttgagtgcaagatttctccttcaaaatggaagaatgttcgga  
 ctcaacacagatgaaagagat

SEQ ID NOS 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33,  
 and 35 have been translated into amino acid sequences by  
 removing the non-coding regions (introns) identified by com-  
 parison with the coding sequence of Gala-1. <sup>20</sup>

The antigen encoded by SEQ ID NO: 9 was isolated from  
*Cylicocyclus ashworthi*.

SEQ ID NO: 9

ATGAACAAAACGTTAACATTTCTCACAGTCGTTAGTGCCGTAGTTCTGGCCCAAGGT  
 GTCATGGACCTTTTTGGTGAAGAGGGTCGTGAAGAACATCGCCGTCACCATCGTCA  
 TCACTCTTACCACCATATCTCCACAACGTGAGCTGTGTGGCTAAATGGGAGTACTTC  
 AAAATTGTAGGGAACAGGAGTTTAAAGTTGCTGAGAAAAAAGAAATAGCCAG  
 TGGGCAAAAAATACAATGTTGTGGTAAGCTTTTCTGAATTAATGTAATACACTCG  
 CATGCTGGCCTTTTTAGGATGAAGTTGCAAGCTACAATGCTTACAGGGAGAACTCA  
 AGCAGGAGCACAGAAAAACGTTAGCGAAGCTGTTTCTGCTCTTCCAAACGCAGTAA  
 AGAAAGTCAACAATCTTCTAGACAATGAAAATCAGACTCTTAGGCAACTTTACGTTG  
 CCCTTAGAGAACTTGGTAGACAAAATCCGGCAGTAAGTAGAAGAGCTGCACTCCTG  
 GGCTTAATAAAAACAAATTATTTAAGCTTTACCGTATTGTGAGTACATCAATGTGGC  
 TGTAAGACGAAGAAGTGAAGGACTGGATGAGCAAGAACAACGAAGAAGCTATCAGC  
 TTTACCTTTCGGCGACAACAACGATAAATATGGAAGAGCAGGACTTCGGTGAACAAGA  
 CTTTCGCTATGTCTACGGCTTTGAGTGTGCAAGATTTCTCCTTCAAAATGGAAGAAT  
 GTTTGGGCTCAACACAGATGAAAGAGATTAGCAAAGAATCAATTGTAGTTCAAAGCG  
 GTAGAGTTTGAGCTGCAAACTCAGCATGCCATCATCACCTCCT

SEQ ID NO: 10 (i.e., SEQ ID NO: 9 translated)

MNKLTFLTLVVSAVVLAQGVMDLFGEEGREHRRHRSLLPPYLHNVS CVAKWEYF  
 KIVGNRSLTFAEKKEEISQWAKKYNVVDEVASYNAYREKLKQEHRKNVSELVSALPN  
 AVKKVNNLLDNENQTLRQLYVALRELGRQNPALYRIVEYINAVRRRSEGLDEQEQR  
 RKLALPFGDNDNMEEQDFGEQDFRYVYGFECARFLQNGRMFGLNTERD

The antigen encoded by SEQ ID NO: 11 was isolated from  
*Cyathostomum catinatum*.

SEQ ID NO: 11

ATGAACAAAACGTTAACATTTCTCACAGTCGTTAGTGCCGTAGTCTGGCT  
 CAAGGTGTCATGGACCTTTTTGGTGAAGAAGCCGTGAAGAACATCGCCG  
 TCACCGTCGTCATCACTCTTGCCACCATATCTCCACAACGTGAGCTGTGT

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GGCTAAATGGGAATACTTCAGAATTGTGGGAAACAGGAGTTTGACGTTTG  
 CTGAGAAAAAGGAAGAGATTAGCGAGTGGGCAAAAAAGTACAATGTTGT  
 GGTAAGCTTTTCTGAATTGATGTAATACTCGCATGCTGGCCTTTTGTAG  
 GATGAAGTTGCAAGCTACAATGCTTACAGGGAAAACTCAAGCAGGAGC  
 ACAGAAAAACGTTAGCGAACTTGTCTCTTCCAAACGCAGTAAAG  
 AAAGTCAACGATCTTCTAGACAATGAAAAATCAGACTCCTAGGCAACTTA  
 CGTTGCCCTTAGAGAACTTGGTAGACAAAATCCGGCAGTAAGTCGAAAGA  
 GCTGCACTCTTGGGCATAAGTAAAAAAGTATTTTAGCTTTACCGTATTG  
 TGGAGTACATCAATGTGGCTGTAAGACTAAGAAGTGAAGAAGTGGATGA  
 GCAAGAACAACGAAGAAGGCTATCAGCTTTACCTTTTGGTGACCATAACG  
 ATAATATGGAAGAGCAGGACTTTGGTGATCAAGACTTTTCGCTATGCTAC  
 GGCTTTGAGTGTCAAGATTTCTCCTTCAAATGGAAGAATGTTTGGACTT  
 AACACAGATGAAAGATATTAGTAAAAATTAAGTGTAGCTCAAAGCGGTAG  
 AGTTTGAGCTGCAAACTCAGCATGCCATCATCACCTCCT

SEQ ID NO: 12 (i.e., SEQ ID NO: 11 translated)

MNKTLLTFLTVVSAVLAQGVMDLFGEEGREEHRHRRHSLPPLYLHNVSCVAKWEYF  
 RIVGNRSLTFAEKKEI SEWAKKYNVDEVASYNAYREKLKQEHKKNVSELVSLPN  
 AVKKVNDLLDENQTPRQLYVALRELGRQNPALYRIVEYINVAVRLRSEEVDEQEQR  
 RRLSALPFGDNDNMEEQDFGDQDFRYVYGFECARFLLQNGRMFGLNTERY

The antigen encoded by SEQ ID NO: 13 was isolated from  
*Cylicostephanus goldi*.

SEQ ID NO: 13

ATGAACAAAACGTTAACATTTCTCACAGTCGTTAGTGCCGTAGTCCTGGCTCAAGGT  
 GTCGTGGACCTTTTGGTGAAGAGGGTCGTGAAGAACATCGCCGTCACCATCGTCAT  
 TCACTCTTACCACCATATCTCCACAACGTCAGCTGTGTGGCTAAATGGGAATACTTC  
 AAAATGTGGGGAATAGGAGTTTGACATTTGCTGAGAAAAAGAAATAGCGAG  
 TGGGCTAAAAATACAATGTAGTGGTAAGCTTTTTGACTTGATGTAATGCACTCG  
 TATGCCGCCCTTTTAGGATGAAGTTGCAAGGTACAATGCTTATAGAGAAAACTTA  
 AGCAGGAACACAGGAAAAACGTCAGCGAACTTGTCTGATCTTCCAAACGCAGTAA  
 AGAAGTGAATGATCTCCTGGACAATGAGAATCAAACCTTAGGCAACTTTACATTG  
 CCCTCAGAGAACTTGGTAGACAAAATCCAGAAGTAAAGTTGAAAGTGTGCAATTTTA  
 GGCTTAGATAAAAACAGTTGTTAAGCTTTACCGTGTGTCGAGTTTATCAATGTGGC  
 TGTAAAGAATAAGACGTGAAGATTGGATGAGCAAGAACAACGAACAAGGCTGTCAAC  
 TTTACCTTTTGGCGACAACAACGACAATTTGGAAGAGCAAGACTTCGGTGAACAAGA  
 CTTTCGCTATGTCTATGGCTTTGAGTGTGCAAGATTTCTCCTTCAAATGGAAGAAT  
 GTTTGACTTAACACGGATAGAAGATAC

SEQ ID NO: 14 (i.e., SEQ ID NO: 13 translated)

MNKTLLTFLTVVSAVLAQGVVDLFGEEGREEHRHRRHSLPPLYLHNVSCVAKWEYF  
 KIVGNRSLTFAEKKEI SEWAKKYNVDEVARYNAYREKLKQEHKKNVSELVSLPN  
 AVKKVNDLLDENQTPRQLYIALRELGRQNPPELYRVVEFINVAVIRREDLDEQEQR  
 TRLSTLPGDNDNDFEEQDFGEQDFRYVYGFECARFLLQNGRMFGLNTERY

The antigen encoded by SEQ ID NO: 15 was isolated from *Cylicostephanus goldi*

SEQ ID NO: 15
ATGAACAAAACGTTAACATTTCTCACAGTCGTTAGTGCCCGTAGTCC
TGGCCCAAGGTGTCATGGACCTTTCTGATGAAGAGGCTCGTGGAGA
GCATCGCCGTCACCATCGTCATTCACTCTTACCACCATATCTCCAC
AACGTGAGCTGTGTGGCTAAATGGGAATACTTCAAATTTGTGGGGA
ACAGGAGTTTGACGTTTGTCTGAGAAAAAGAAAGAAATTAGCGAGTG
GGCAAAAAATACAACGTTGTGGTAAGCTTTTGTGACTCGATGTAG
ATACCCAGATATTCTAGATACCCATGCTGGCCTTTTGTAGGATGAA
GTTGCAAGCTACAATGCTTATAGAGAAAACTCAAGCAGGAACACA
GGAAAAACGTTAGCGAATTTGATCTGATCTTCCCAATGCGAGTGAA
GAAAGTGAATGATCTCTGGACAATGAGAATCAAACCTCTAGGCAA
CTTTACGTTGGCCCTCAGAGAATTTGGTAGACAAAATCCAGCAGTAA
GTTGAAAGTGTGCAATTTCCAGGCTTAGATAAAACAGTTGTTTAAAG
CTTTACCGTGTGTGCGAGCTCATCAATGTGGCTGTAAGATTAAGAC
GTGAAGATTTGGATGAGCAAGAAACAACGAAACAGGCTGTCAACCTT
ACCTTTTGGCGACAACAACAATTTGATGAGCAGGACTTCGGT
GAACAAGACTTTCGCTATGTCTATGGCTTTGAGTGTGCAAGATTTT
TCCCTCAAATGGAAGATGTTTGGACTTAAACACGGATAGAAGATA
CTAGTAAGAGTCAACTGTAGCTCAAAGTGGTTCGAGCTACGAACAG
CATGCCATCATCACCTCT

SEQ ID NO: 16 (i.e., SEQ ID NO: 15 translated)
MNKTLTFLTVVAVVLAQGVMDLLEDEARGEHRHRHRLSLPPYLH
NVSCVAKWEYFKIVGNRSLTFAEKKKEISEWAKKYNVVEVASYNA
YREKLKQEHKKNVSELVSDLPNAVVKVNDLLDNENQTPRQLYVALR
ELGRQNPALYRVVEYINVLVRLRREDLDEQEQRTLSTLPLFGDNNN
NFBEQDFGEQDFRYIYGFECARFILQNGRMFGLNTRDRY

The antigen encoded by SEQ ID NO: 17 was isolated from *Cylicostephanus goldi*

SEQ ID NO: 17
ATGAACAAAACGTTAACATTTCTCACAGTCGTTAGTGCCCGTTGTC
TGGCCCAAGGTGTCATGGACCTTTTGGTGAAGAGAGTCTGGAAGA
ACACCCGCGTCACCATCGTCATTCACTCTTACCACCATATCTCCAC
AACGTGAGCTGTGTGGCTAAATGGGAGTACTTCAAATTTGTGGGGA
ACAGGAGTTTGACGTTTGTCTGAGAAAAAGAAAGAAATCAGCGAGTG
GGCTAAAAATACAATGTTGTGGTAAGCTTTTGTGACTGATGTA
ATGCACTCGCATGCCGCTTTATAGGATGAAGTTGCAAGCTACAA
TGCTTATAGAGAAAACTCAAGCAGGAACACAGGAAAAACGTTAGC
GAACTGTTTCTGATCTTCCCAACGCGATAAAGAAAGTCAAGGATC
TTTTGGACAACGAAATCAGACTTCTAGGCAACTTTATGTTGCACT
CAGAGAACTTGGTAGACAAAATCCGGCAGTAAGTTGAAGAGGCTCC
AATTTTGGCTCAAGCAAAAATAATATTTTAGCTATACCGTGTG
TCGAGTATATCAATGTGGCTGTGAGATTAAGACGAAAGAACAGGA
TGAACAAGAACGCAAGGAAACGCTGTGAGCTCTACCTTTTGGCGAG
AATAACGACAATTTGGAAGAGCAGGACTTTGGTGAAACAAGACTTC
GCTATGTCTATGGCTTTGAGTGTGCAAGATTTCTCCTCAAATGG
AAGAAATGTTGACTCAACACGGATAGAAGATACAGTAAGAGTCA
ACTGTAGCTCAAAGTGGGTTTGGACTACGAACAGCATGCCATCATC
ACCTCT

SEQ ID NO: 18 (i.e., SEQ ID NO: 17 translated)
MNKTLTFLTVVAVVLAQGVMDLLEDEARGEHRHRHRLSLPPYLH
NVSCVAKWEYFKIVGNRSLTFAEKKKEISEWAKKYNVVEVASYNA
YREKLKQEHKKNVSELVSDLPNAVVKVNDLLDNENQTPRQLYVALR
ELGRQNPALYRVVEYINVLVRLRREDLDEQEQRTLSTLPLFGDNNN
NLEBQDFGEQDFRYIYGFECARFILQNGRMFGLNTRDRY

The antigen encoded by SEQ ID NO: 19 was isolated from *Cylicostephanus longibursatus*

SEQ ID NO: 19
ATGAACAAAACGTTAACATTTCTCACCGTCGTTAGTGCCCGTAGTCC
TGGCCCAAGGTGTCATGGACCTTTTGGTGAAGAGGCTCGTGAAGA
ACATCGCCGTCACCATCGTCATTCACTCTTACCACCATATCTCCAC
AATGTGAGCTGTGTGGCTAAATGGGAATACTTCAAATTTGTGGGGA
ACAGGAGTTTGACGTTTGTCTGAGAAAAAGGAAGAAATAGCAAGTG
GGCAAAAAATACAATGTTGTGGTACGCTTTTGTAAACCCCGTATA
TATACTCTCGATACTGGCCCTTTCCAGGATGAAGTTGCAAGCTACA
GTGCTTGCAGGAAAAAGCTTAAAGCAGGAACACAGGAAAAACGTTAG
CGAAATGTTTCTAATCTTCCCAATGCAAGTGAAGAAAGTAAACGAT
CTTTTGGACAATGAAATCAGACCCAGGCAACTTTACGTTGGCT
TCAGAAAATTTGGTAAACAAAATCCGGCAGTAAGTTGAAAGAGCTG

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CAATTTTGGGTTTGGAGGAAAAAACTATTTAGCTTTATCGTGT
GTCGAGTATATCAATGTGCTTGTGAGACTAAGACGTGAAGAATTTG
ATGAAGATCAGCGAAGATCGCTGTCAGCTTTACCTTTTGGCGACAA
TAACGACGATTTGGAAGAGCAGGACTTTGGTGAACAGGACTTTGCG
TATATCTATGGCTTTGAGTGTGCAAGATTTATCCTTCAAATGGAA
GAATGTTCCGACTCAACACGGATAGAAGATATAGTAAGAGTCAAC
GTAGCTCGAGGTTTGGACTACGAACGTCATGCCATCATCACCTC
CT

SEQ ID NO: 20 (i.e., SEQ ID NO: 19 translated)
MNKTLTFLTVVAVVLAQGVMDLFGEEGREHRRHRLSLPPYLH
NVSCVAKWEYFKIVGNRSLTFAEKKKEISEWAKKYNVVEVASYSA
CREKLNQEHKKNVSEIVSNLPNAVVKVNDLLDNENQTPRQLYVAFR
KLGKQNPALYRVVEYINVLVRLRREDLDEQEQRTLSTLPLFGDNNN
LEEQDFGEQDFRYIYGFECARFILQNGRMFGLNTRDRY

The antigen encoded by SEQ ID NO: 21 was isolated from *Cylicocylus insigne*.

SEQ ID NO: 21
ATGAACAAAACGTTAACATTTCTCACCGTCGTTAGTGCCCGTAGTCC
TGGCCCAAGGTGTCATGGACCTTTTGGTGAAGAAAGTCTGGAAGA
ACATCGCCGTCACCATCGTCATTCACTCTTACCACCATATCTCCAC
AATGTGAGCTGTGTGGCTAAATGGGAATACTTCAAATTTGTGGGGA
ACAGAAGTTTGGCTTGTCTGAGAAAAAGGAAAAATCAGCGAGTG
GGCAAAAAGTACAATGTTGTGGTACGCTTTTGTAACTCCGTATAA
TATACCTCGCATGCTGGCCGTTTCCAGGATGAAGTTGCAAGCTACA
ATGCTTGCAGGAAAAAGCTTAAAGCAGGAACACAGGAAAAACGTTAG
CGAAATGTTTCTAATCTTCCCAATGCAAGTAAAGAAAGTAAACGAT
CTTTTGGACAATGAAATCAGACTCCAGGCAACTTTACGTTGGCC
TCAGAAAATCGGTAAACAAAATCCGGCAGTAAGTTGAAAGACTG
AACTTTGGGTTTAAAGGAAAAAACTATTTTAGCTTTACCGCGTTG
TCGAGTATATCAATGTGGTGTGAGACTAAGACGTGAAGAACTGTA
TGAAGAACAACGAAAGACGCTGTGAGCTTTACCTTTTGGCGACAAT
AACGCAACTTGGAAAGACGAAGCTTTGGTGAAGAAAGCTTTGCT
ATATTTATGGCTTTGAGTGTGCAAGATTTATCTTCAAATGGGAG
AATGTTCCGACTCAACACGGATAGAAGATATCAGTAAGAGTCAACT
GTAGCTTAAAAGTTTGGACTACGAACAGCATGCCATCATCACCTCC
T

SEQ ID NO: 22 (i.e., SEQ ID NO: 21 translated)
MNKTLTFLTVVAVVLAQGVMDLFGEEGREHRRHRLSLPPYLH
NVSCVAKWEYFKILGNRSLTFAEKKKEISEWAKKYNVVEVASYNA
CREKLNQEHKKNVSEIVSNLPNAVVKVNDLLDNENQTPRQLYVALR
KLGKQNPALYRVVEYINVLVRLRREESDEEQRRLLSALPLFGDNNN
LEEQDFGEQDFRYIYGFECARFILQNGRMFGLNTRDRY

The antigen encoded by SEQ ID NO: 23 was isolated from *Cylicostephanus longibursatus*.

SEQ ID NO: 23
ATGAACAAAACGTTAACATTTCTCACCGTCGTTAGTGCCCGTAGTCC
TGGCCCAAGGTGTCATGGACCTTTTGGTGAAGAGGCTCTGGAAGA
ACATCGCCGTCACCATCGTCATTCACTCTTACCACCATATCTCCAC
AATGTGAGCTGTGTGGCTAAATGGGAATACTTCAAATTTGTGGGGA
ACAGGAGTTTGACGTTTGTCTGAGAAAAAGGAAAAATCAGCGAGTG
GGCAAAAAGTACAATGTTGTGGTACGCTTTTGTAACTCAGTATAA
TATATCTCGCATACTGGCCGTTTCCAGGATGAAGTTGCAAGCTACA
ATGCTTGCAGGAAAAAGCTTAAAGCAGGAACACAGGAAAAACGTTAG
CGAAATGTTTCTAATCTTCCCAATGCAAGTAAAGAAAGTAAACGAT
CTTTTGGACAATGAAATCAGACCCAGGCAACTTTACGTTGGCC
TCAGAAAATTTGGTAAACAAAATCCGGCAGTAAGTTGAAAGAGCTG
CAATTTTGGGTTTGGAGAAAAAACTATTTTAGCTTTATCGTGT
GTCGAGTATATCAATGTGCTTGTGAGACTAAGACGTGAAGAATTTG
ATGAAGATCAGCGAAGATCGCTGTCAGCTTTACCTTTTGGCGACAA
TAACGACGATTTGGAAGAGCAGGACTTTGGTGAACAGGACTTTGCG
TATATCTATGGCTTTGAGTGTGCAAGATTTATCCTTCAAATGGAA
GAATGTTCCGACTCAACACGGATAGAAGATATAGTAAGAGTCAAC
GTAGCTCAAGGTTTGGACTACGAACGTCATGCCATCATCACCTC
CT

SEQ ID NO: 24 (i.e., SEQ ID NO: 23 translated)
MNKTLTFLTVVAVVLAQGVMDLFGEEGLEHRRHRLSLPPYLH
NVSCVAKWEYFKILGNRSLTFAEKKKEISEWAKKYNVVEVASYNA
CREKLNQEHKKNVSEIVSNLPNAVVKVNDLLDNENQTPRQLYVALR
KLGKQNPALYRVVEYINVLVRLRREDLDEQEQRTLSTLPLFGDNNN
LEEQDFGEQDFRYIYGFECARFILQNGRMFGLNTRDRY

The antigen encoded by SEQ ID NO: 25 was isolated from *Cylicocycclus nassatus*.

SEQ ID NO: 25
ATGAACAAAACGTTAACATTTCTCATCGTCGTTAGTGCCGTAGTCC
TGACCCAAAGTGTATGGACTTTTTTCGATGAAGACGGTCTGTAAGA
ACATCGCCGTCATCATCGTCATTTCCCTTTTACCACCGTATCTCCAC
AATATGAGCTCGCTGGCCAAATGGGAATACTTCGAGATTGTGGGGG
ACAGGAGTCTGACGTTTGGCTGAAAAGAAGGAAAAAATCGGCGAGTG
GGCTAAAAATAACAATGTTGTGGTAAGATTTTGTAACTCTATGTAA
AGATACCCCGTACGTGCGCCCTGTTTAGGATGAAGTTGCAAGCTAC
AATGCTTATAGAGAAAAAATAAGCAGGAGCACAGGAAAAACGTTA
CGGAGCTTGTCTCTGGTCTTCCCAATGCTGTGAAGAAAAATAACGA
ACTTTTAGACAATGAAAATCAGACTGTTAGGCAACTTTATGTTGCT
TTAAGAGAACTTGGTAAACAAAATCCAGCAGTAAGTTAAAGAAGT
GCAATTTTGGGCTTAACCTAATGAGACAATTTAGCTCTACCGTGT
GTCGAGTATATCAATGTGGTGTGAGACTTAGACGTGAAGATTGG
ATGAGCAGGAACAACAGAGAAGCTGTCAACCCACCTTTTCGGCGA
GAATAACGAAGCAAGACTTTGGTGAACAAGACTTCACTATATC
TATGGTTTTGAGTGTCCAGATTATCCTTCAAAATGGAAGAATGT
TTGGACTTAACACCGATAGAGAATATTAGTAAGAGTTAACTGCAGC
TCAATGTGATAGAGATTGAGCCACAACCAACATGCCATCATCACC
TCCCT

SEQ ID NO: 26 (i.e., SEQ ID NO: 25 translated)
MNKTLTFLIVSAVVLVTSQVMDFFDEEDGREHRRHRRHSLPPYLHN
NMSCVAKWEYFEIVGDRSLTFAEKKEKIGEWAKKYNVVDEVASYN
YREKLKQEHKRNVSSELVSLPNAVKKINELLDNENQTVRQLYVALR
ELGKQNPALYRVVEYINVVRLRREDSDEQEQRRRLSTSPFGENNE
EQDFGEQDFHYIYGFECARFILQNGRMFGLNTDRRY

The antigen encoded by SEQ ID NO: 27 was isolated from *Cylicocycclus nassatus*.

SEQ ID NO: 27
ATGAACAAAACGTTAACATTTCTCATCGTCGTTAGTGCCATAGTCC
TGGCCCAAAGTGTATGGACTTTTTTCGATGAAGAAGGTCTGAGGG
ACATCGCCGTCATCATCGTCATTTCACTTTTACCACCATATCTCCAC
AATATGAGCTCGCTGGCCAAATGGGAATACTTCGAGATTGTGGGGG
ACAGGAGTCTGACGTTTGGCTGAAAAGAAGGAAAAAATCGGCGAGTG
GGCTAAAAATAACAATGTTGTGGTAAGATTTTGTAACTCCATGTTA
GGATACCTCCGCACGTGCGCCCTGTTTAGGATGAAGTTGCAAGCTAC
AATGCTTATAGAGAAAAAATAAGCAGGAGCACAGGAAAAACGTTA
CGGAGCTTGTCTCTGGTCTTCCCAATGCTGTGAAGAAAGTAAACGA
ACTTTTAGACAATGAAAATCAGACTGTTAGGCAACTTTATGTTGCT
TTAAGAGAACTTGGTAAACAAAATCCAGCAGTAAGTTAAAGAAGT
ACAATTTTGGGCTTAACCTAATGAGACAATTTAGCTCTACCGTGT
GTCGAGTATATCAATGTGGTGTGAGACTTAGACGTGAAGATTCCG
ATGAGCAGGAACAACGAAGAAGCTGTCAACCTCACCTTTTCGGCGA
GAATAACGAAGCAAGACTTTGGTGAACAAGATTTTCACTATATC
TATGGTTTTGAGTGTGCAAGATTATCCTTCAAAATGGAAGAATGT
TTGGACTCAATACGGATAGAAGATAT

SEQ ID NO: 28 (i.e., SEQ ID NO: 27 translated)
MNKTLTFLIVSAIVLAQVMDFFDEEGREHRRHRRHSLPPYLHN
NMSCVAKWEYFEIVGDRSLTFAEKKEKIGEWAKKYNVVDEVASYN
YREKLKQEHKRNVSSELVSLPNAVKKINELLDNENQTVRQLYVALR
ELGKQNPALYRVVEYINVVRLRREDSDEQEQRRRLSTSPFGENNE
EQDFGEQDFHYIYGFECARFILQNGRMFGLNTDRRY

The antigen encoded by SEQ ID NO: 29 was isolated from *Cylicocycclus nassatus*.

SEQ ID NO: 29
ATGAACAAAACGTTAACATTTCTCATCGCCGTTAGTGCCATAGTCC
TGGCCCAAAGTATGGACTTTTTTCGATGAAGACGGTCTGTAAGAACA
TCGCCGTCATCATCGTCATTTCACTTTTACCACCATATCTCCACAAT
ATGAGCTCGCGGGCAAATGGGAATACTTCGAGATTGTAGGGGACA
GGAGTCTGACGTTTGGCTGAAAAGAAGGAAAAAATCGGCGAGTGGC
TAAAAATAACAATGTTGTGGTAAGATTTTGTAACTCCATGTAAGA
TACCCCTCCATGTCGTCCTCCGTTTAGGATGAAGTTGCAAGCTACAAT
GCTTGCAGAGAAAAAATGAAGCAAGAGCACAGGAAAAACGTCAGCG
AGCTTGTCTCTGGTCTTCCCAATGCTGTGAAGAAAGTAAACGAAC
TTTAGACAATGAAAATCAGACTGTTAGGCAACTTTATGTTGCTTTA
AGAGAACTTGTGTAACAAAATCCAGCAGTAAGTTGAAGAAGTGCA
TTTTGGGCTTAACCTAACGAGACAATTTAGCTCTACCGTGTGTCG
AGTATATCAATGTGGCTGTGAGACTTAGACGTGAAGATTCCGATGA

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GCAGGAAAAACGAAGAACGCTGTCAACCTCACCTTTCGGCGAGAA
AACGAAGAGCAGGACCTTGGTGAACAAGATTTTCACTATATCTATG
GCTTTGAGTGTGCAAGATTATCCTTCAAAATGGGAAGATGTTTGG
ACTTAAACACGGATAGAAGATATTAGTAAAATTTGACTGCAGCTCAA
AGTGGTAGAGATTGAGCTACCAACCCAACATGCCATCATCACCTCC
T

SEQ ID NO: 30 (i.e., SEQ ID NO: 29 translated)
MNKTLTFLIIVSAIVLAQVMDFFDEEDGREHRRHRRHSLPPYLHN
MSCAKWEYFEIVGDRSLTFAEKKEKIGEWAKKYNVVDEVASYNAC
REKLKQEHKRNVSSELVSLPNAVKKINELLDNENQTVRQLYVALRE
LGKQNPALYRVVEYINVVRLRREDSDEQEQRRRLSTSPFGENNE
QDLGEQDFHYIYGFECARFILQNGRMFGLNTDRRY

The antigen encoded by SEQ ID NO: 31 was isolated from *Cyathostomum pateratum*.

SEQ ID NO: 31
ATGAACAAAACGTTAACATTTCTCACAGTCTGTTAGTGCCGTAGTCT
GGCCCAAAGGTGTATGGACTTTTTTGGTGAAGAGGGTCTGTAAGAAC
ATCGTCGTCAACATCGTCATTTCACTTTTACCACCATATCTCCACAAT
GTGAGCTGTGAGGCTAAATGGGAGTACTTCAAAATTTGGGGAACAG
GAGTTTACGCTTTGCTGAGAAAAAGGAGAAAAATAGCCGAGTGGCCAA
AAAAATACAATGTTGTGGTAAGCTTTTTTGAATTGATGTAATTCAC
TCGCATGCTGGCCTTTTTTAGGATGAAGTTGCAAGCTACAATGCTTAC
AGGGAAAAACTCAAGCAGGAGCACAGAAAAAAGCTTAGCGAAGCTTGT
TTCTGCTCTTCAAAACGAGTAAGAAAGTCAACGATCTTCTAGACA
ATGAAAATCAGACTCTTAGGCAACTTACGTTGCCCTTAGAAAACCTT
GGTAGACAAAATCCGGCAGTAAGTCGAAAGAGCTGCGCTCTGGACT
TAAGCGGAAAAAATATTTTACGCTTTACCGTATTGTCGAGTACATTA
TGTGGCTGTAAGACTAAGAGTGAAGAGTGGATGAGCAAGAACAA
GAAGAAGGCTGTCAGCTCTACCTTTTGGCGACCATAACGATAATTTG
GAAGCAGGACTTCCGTGAACAAGACTTTCGCTATGCTATGCTATGGCT
TGAGTGTGCAAGATTTCTCCTTCAAAATGGAAGAATGTTCCGACTCA
ACACGGATGGAAGATATTAGTAAGAAAAAGTGTAGCTCAAGTGGT
AGAGTTTGAGTACGAACCTCAACATGCCATCATCACCTCTCT

SEQ ID NO: 32 (i.e., SEQ ID NO: 31 translated)
MNKTLTFLIVSAVVLVLAQVMDLFGEEEGREHRRHRRHSLPPYLHN
VSCAKWEYFKIVGNRSLTFAEKKEKISEWAKKYNVVDEVASYNAYR
EKLKQEHKRNVSSELVSLPNAVKKINELLDNENQTVRQLYVALRKL
GQNPALYRIVEYINVVRLRREDSDEQEQRRRLSALPFGDHNLDLEE
QDFGEQDFRYVYGFECARFILQNGRMFGLNTDGRY

The following sequences (SEQ ID NOS: 33 and 35) represent *Cyathostomin* GALA sequences obtained from cDNA clones.

The antigen encoded by SEQ ID NO: 33 was isolated from *Cylicostephanus goldi*.

SEQ ID NO: 33
ATGAACAAAACGTTAACATTTCTCACAGTCTGTTAGTGCCGTTGTCCT
GGCCCAAAGGTGTATGGCCCTATTTGGTGAAGAGAGTCTGTAAGAAC
ACCGCCGTCACCATCGTCATTTCACTTTTACCACCATATCTCCACAAT
GTGAGCTGTGAGGCTAAATGGGAGTACTTCAAAATTTGGGGAACAG
GAGTTTACGCTTTGCTGAGAAAAAGAAGAAATCAGCGAGTGGGCTA
AAAAATACAATGTTGTGGATGAAGTTGCAAGCTACAATGCTTATAGA
GAAAAACTCAAGCAGGAACACAGGAAAAAAGCTTAGCGAAGCTGTTTC
TGATCTTCCCAACGAGTAAGAAAGTCAACGATCTTTTGGACAACG
AAAATCAGACTTCTAGGCAACTTATGTTGCACTCAGAGAAGCTTGTG
AGACAAAATCCGGCACTATACCGTGTGCTGAGTATATCAATGTGGC
TGTGAGATTAAGACGAAAAAGAACAGGATGAACAAGAACGACAGGAA
CGCTGTGAGCTCTACCTTTTGGCGAGAAATACGACAAATTTGGAGAG
CAGGACTTTGGTGAACAAGACTTTCCGCTATGCTATGGCTTTGAGTG
TGCAAGATTTCTCCTTCAAAATGGAAGAATGTTTGGACTCAACACGG
ATAGAAGATACAGTAAGAGTCAACTGTAGCTCAAAGTGGGTTTGGAG
CTACGAACAGCATGCCATCATCACCTCTCT

SEQ ID NO: 34 (i.e., SEQ ID NO: 33 translated)
MNKTLTFLIVSAVVLVLAQVMDLFGEEESREHRRHRRHSLPPYLHN
VSCVAKWEYFKIVGNRSLTFAEKKEKISEWAKKYNVVDEVASYNAYR
EKLKQEHKRNVSSELVSLPNAVKKINELLDNENQTVRQLYVALREL
GQNPALYRVVEYINVVRLRREKQEQERQGLSALPFGENNDNLEE
QDFGEQDFRYVYGFECARFILQNGRMFGLNTDRRY

The antigen encoded by SEQ ID NO: 35 was isolated from *Cylicostephanus longibursatus*.

C. lon91-GALA  
 ATGAACAAAACGTTAACATTTCTCACCGTCGTCTATGCCGTAGTCTCT  
 GGCCCAAGGTGTCATGGACCTTTTGGTGAAGAGGGTCTGGAAGAAC  
 ATCGCCGTCACCATCGTCATTCACTCTTACCACCATATCTCCACAAT  
 GTGAGCTGTGTGGCTAAATGGGAATACTTCAAATTTCTGGGAACAG  
 GAGTTTGACGTTTGGCTGAGAAAAAGGAAAAATCAGCGAGTGGGCAA  
 AAGAACTACAATGTTGTGGAAGAAGTTCGAAGCTATAATGCTTGCAGG  
 GAAAAGCTTAAGCAGGAACACAGGAAAAACGTTAGCGAAATGTTTC  
 TAATCTTCCCAATGCAGTGAAGAAAGTAAACGATCTTTGGACAATG  
 AAAATCAGACCCCGCAGCACTTTACGTTGCCCTCAGAAAACTTGGT  
 AAACAAAATCCGGCACTTTATCGTGTGTGCGAGTATATCAATGTGCT  
 TGTGAGACTAAGACGTGAAGAATTTGATGAAGATCAACGAAGATCGC  
 TGTGAGCTTTACCTTTTGGCGACAATAACGACGATTTGGAAGAGCAG  
 GACTTTGGTGAACAGGACTTTCCCTATATCTATGGCTTTGAGTGTGC  
 AAGATTTATCTTCAAATGGAAGAAATGTTCCGAATCAACACGGATA  
 GAAGATATTAGTAAAGTCAACTGTAGCTCAAGGGTTTGGAGTACGA  
 ACTGCATGCCATCATCACCTCTCT

SEQ ID NO: 36 (i.e., SEQ ID NO: 35 translated)  
 MNKTLTFLTVVYAVVLAQGVMDLFGEEGREHRRHRHSLPPYLHN  
 VSCVAKWEYFKILGNRSLTFAEKKEKISEWAKKYNVVDVAVSYNACR  
 EKLKQEHKRVSEIVSNLNPVAVKVVNDLNDNENQTPRQLYVALRKLK  
 KQNPALYRVVEYINVLVRLRREFEDEQRRSLSALPFGDNDLLEEQ  
 DFGEDFRYIYGFECARFILLQNGRMFGINTDRRY

Each of the proteins provided by SEQ ID NO: 1, 3, 5, 7, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, and 36 (or encoded by the nucleic acid sequences of SEQ ID NOS: 2, 4, 6, 8, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, and 35) may be classified as a member of the "keratin-like" proteins although, because they lack the glycine-rich domains characteristic of other KLP proteins and are localised to the gut of larval *cyathostomin*, the inventors have chosen to designate these proteins *cyathostomin* gut-associated larval antigens (Cy-GALA).

Using any of the Cy-GALA sequences described herein, one of skill in the art could readily identify related or homologous sequences in other species, such as, for example, other *cyathostomin* spp. etc. For example, the nucleic acid sequence encoding these proteins could be used to probe for homologous sequences in other *cyathostomin* species.

Other potentially useful *cyathostomin* larval antigens include those encoded by the following sequences, designated SEQ ID NOS: 37-58. (SEQ ID NOS: 37, 43, and 45 are amino acid sequences and SEQ ID NOS: 38-42, 44, 46-58 are nucleic acid sequences, SEQ ID NOS: 38, 44, and 46 encoding SEQ ID NOS: 37, 43, and 45 respectively). It should be understood that the invention further encompasses proteins, peptides and amino acids having sequences encoded by SEQ ID NOS: 39-42 and 46-58.

SEQ ID NO: 37 (CID-1):  
 REKARI IQDEYTKRMQVTPQAQEFBLAKWEKTFWTFNVQYSGDKKAF  
 FKQMI ELI PQLMEEVHGFS EETWKS LEEQFP EQTA AWKDNEDRLKQF  
 YEFIKSLPKQDLAEDPEAFRKF AHLGLQLKLLPIEALRA

SEQ ID NO: 38 (nucleic acid sequence encoding CID-1)  
 AGGGAGAAGGCTAGAATTATTCAAGACGAATACTAAACGATGCA  
 GCAGGTACACCAAGCTCAGGAATCTCTGGCAAATGGGAGAAGA  
 CATGGTTCACGAATGTGACGAATA TAGCGGAGATAAGAAAGCTTTC  
 TTCAAGCAGATGATGAGCTAAATCCCTCAACTAATGGAGGAGTTC  
 TGGGTTCTCGGAAGAGACTTGAAGAGCCTTGGAGGACAAATCCCG  
 AGCAGACAGCCGATGGAAAGATAATGAGGATCGCCTAAAGCAATTT  
 TATGAGTTTATCAAGAGCCTACCAAGCAGGACTTAGCTGAGGATCC  
 GGAAGCATTGAGAAAGTTCGCTCACCTCGGACTCCAGAAACTTCTTC  
 CAATTGAAGCTCTCAGAGCT

CID antigens from other *Cyathostomin* organisms may include those encoded by the genomic DNA sequences provided as SEQ ID NOS: 39-42 provided below.

SEQ ID NO: 39

C. cat01-CID  
 TGGTCACACCACAAGCTCAGGAGTTCCTGGCCAAGGTAAGCTATTAC  
 CTTACCAGGGTGAGGGGAAGAAGTTGGCAGCGGTCGGAACCCGGT  
 AATCTACTGACTTTACCAATATTTTTCAGTGGGGAAGACATGGTTTC  
 ACGAATATACAGCAATACAGTGGAGACAAGCAAGCCCTCTTTAAGCA  
 GATGATTGAACTAATCCTCAACTTATGGAGGAGGTTTCAGGTAAGTT  
 AGCCGCAAAAATTTTAAACCAATGGTTGAGCTCGACATTTTTCAGG  
 GATTACAGAGGAGACTTGGAAATAGCCTGAGGGAGCAATTCGCCGAG  
 CAGACAGCCGATGGAAGGATCGTGAGTATCTTTCATAATTACTGTA  
 CTTGGAATTATACTTTACAATCATAATCCTACTCTTAGACGAGGATC  
 GCCTGAAGCAATCTATGAGTTCATTAAAGAGCCTACCCAAACAACAA  
 TTAGCTGAGGTGATTTTCATTGATTTTTTCGAAAAATATATTTTGT  
 ACATCTTTTTTCAGGATCCGGAAGCTTTCAGAAAGTTTCGCTCACCTC  
 G

SEQ ID NO: 40

C. cat02-CID  
 TTGT CACACCACAAGCTCAGGAGTTCCTGGCTAAGGTAAGCTATTAC  
 CTTACCAGGGTGAGGGGAAGAAGTTGGGAGCGGTCGGAACCCGGT  
 AATCTACTGACTTTACCAATATTTTTCAGTGGGAGAGGACATGGTTTC  
 ACGAATATACAGCAATACAGTGGAGACAAGCAAGCCCTCTTTAAGCA  
 GATGATTGAACTAATCCTCAACTTATGGAGGAGGTTTCAGGTAAGTT  
 GGCCGCAAAAATTTTAAACCAATGGTTGAGCTCGACATTTTTCAGG  
 GATTACAGAGGAGACTTGGAAATAGCCTGAGGGAGCAATTTCCCGGAG  
 CAGACAGCCGATGGAAGGATCGTAAGTATCTTTCATAATTACTGTA  
 CTTGGAATTATACTTTACAATCATAATCCTACTCTTAGACGAGGATC  
 GCCTGAAGCAATCTATGAGTTCATTAAAGAGCCTACCCAAACAACAA  
 TTAGCTGAGGTGATTTTCATTGATTTTTTCGTACGAAAAATATATTTT  
 TGATACATCTTTTTTCAGGATCCGGAAGCTTTCAGAAAGTTTCGCTCA  
 CCTCG

SEQ ID NO: 41

C. lon91-CID  
 AGGTCACACCACAAGCTCAGGAATTCCTGGCAAAGGTAAGCTATCAC  
 CTTACCAGGGTGAGGGGTAGAAGTTAGGAGCGAGGGAACCCGGTGT  
 CTCTTATACCATTACTTTCAGTGGGAGAAGATATGGTTCCAGGAATGT  
 ACAGCAATATAGTGGAGACAAGCAAGCCTTCTTCAAGCAGATGATTG  
 AACTAATTCCTCAACTTATGGAGGAGTACAGGTAAGTCAGCTAAG  
 TGATTTAAGAAAAAATAAGCCGTGATTTTCTTTCAGGGATTCTCA  
 GAGGAGACTTGGAAATAGCCTTAAAGGAGCAATTCCTGAGGAGACAGC  
 CGCATGGAAGGATAGTGAATTTTTTCATAATTACTGTAAGTTCGGAAT  
 TATACTTTACAAATCATAATCCTACCCCTCAGACGAGGAGCGCCTGAAG  
 CAATTCATGAGTTCATTAAAGAGCCTACCCAAACAACAATAGCTGA  
 GGTGATTTTCATTGATTTTTTCGTACGAAAAAGTATATTTTAAATCAT  
 TCTTTTGCAGGATCCGGAAGCCTTCAGAAAGTTTCGCTCACCTCG

SEQ ID NO: 42

C. nas07-CID  
 AGGTCACACCACAAGCTCAGGAATTCCTGGCAAAGGTAAGCTACCAT  
 ATTTTCAGGGGGAGGGCAATTTTGGAGCGAGGGAGGAGGAAGAGG  
 AGAGAAACACTGGTTGGGATCACTAATCTACCCGCACTTCCAGTG  
 GGAGAAGACATGGTTTACGAAATGTGACAGCAATATAGCGGAGATAAGA  
 AAGCCTTTTTCAAACAGATGATTGAGCTAATCCCTCAACTAATGGAA  
 GAGGTTTCATGTAAGTCAACCAAGTGGCTTTTAAAGCGGAGATTAAC  
 TCGAATTTTTCTCAGGGTTCCTCGGAGGAGACTTGAAGAGCCTTG  
 AGGAGCAATTCAGAGCAGACAGCCGATGGAAGGATAGTAAAGCAT  
 TCTTACATAGCTCCCGCTTATCATTATCTTACCAGTAACTTCTT  
 ATTTTATGATGAGGATCGCCTGAAGCAATTTTATGAGTTCATCAAGA  
 GCCTACCAAGCAGGACTTAGCTGAGGTAACCTTTCATGTTTTTTTCC  
 TGAGCTGTAATAATGCTTGAACATAACACTTTTCTAGGATCCGGAA  
 GCTTTCAGAAAGTTTCGCTCACCTCG

SEQ ID NO: 43 (FAR-2):  
 KKESQGFPSI PVDNLRASPFLLQYIKEYIPDYKNAMEK FEDIPKQYR  
 DLIP EVATHLKAITAE EKAVLKEVMKYAKYKDEEFLKALKEKSE  
 GLHEKASKLHNF IKGKVDALGDEBAKFVKKVI AAAREVHAKLLAGDK  
 PSLEDI KKKAKEHMGEP EKLSDDAKEDL KKNFP I LTVSWTNEKRAL  
 IDKYVEN

SEQ ID NO: 44 (nucleic acid sequence encoding (FAR-2))  
 ATGCTTCGAATAACTTTCTTCTTCTGCTCTTTTGGTGTCTACACTTT  
 TTCTGCACCTCTGGACCCGCTGAAGAGAAGATAGATGTGAAAAAAA  
 TGGAAAAATTTGAAGATATTCCAAAGCAATATCGAGACCTTATCTCG  
 GAAGAGGTAGCTACACACCTCAAAGCCATCACCCTGAAGAGAAAGC  
 TGTTCTAAAAGAGGTAATGAAGAAATATGCAAAAGTACAAGACAGG  
 AGGAGTTTTTGAAGCGTTGAAAGAAAAATCAGAGAGTTCATGATGAG  
 AAAGCCAGCAAACTTCAAAATTTTATCAAAGGGAAGGTTGACGCACT  
 TGGAGATGAAGCAAGGCAATTTGTGAAGAAAGTTATCGCAGCTGCTC  
 GAGAAGTGATGCCAAACTTCTTCCCGGGGACAAACCATCGCTGAA

-continued

GATATCAAGAAGAAAGCCAAGGAGCATATGGCTGAATTCGAGAAACT  
AAGCGATGATGCCAAGGAGGATCTCAAAAAGAATTTCCAATCCTTA  
CTTCCGCTGGACAAATGAGAAAACAAGAGCGTTGATTGACAAATAT  
GTGGAGAAC

SEQ ID NO: 45 (UNK-50a):

GKMSDLWTAISETNKVRLFNTLSLGIAGVLCITTAIPVPEVQVCAV  
LITLLQGVIGFNSAGYNKAAVIVARQHAHLLLTFCFLIVTFVPLVQP  
FIVQLVAPDHSWDQWFLVFGHGLVLVIANLRFCLTIEAKPAAFTQK  
TDS5

SEQ ID NO: 46 (nucleic acid sequence encoding  
UNK-50a)

GGTAAAATGTCAGATTTATGGACGGCAATAAGCGAAACAAATAAAGT  
CCGCTTGTTCACACCTTGTGCTGGGAATGCTGGCGTACTGTGTA  
TAACACTGCTTTCATCTCTGTGAAAATCAGGTTGTTTTCGCTGTT  
TTAATCACGTTATTGCAAGGAGTTATCGGATTCATTCAGCTGGATA  
TAACAAAGCTGCAGTCATTGTTGCTAGGCAGCATGCTCATCTTCTGT  
TGACCTGCTTTGGGCTCATTGTCACTTTTGTCCCTTGTGTCAGCCA  
TTCATAGTTCAACTTGTGGCCCTGACCATAGCTGGGCAAAATGGTT  
TTATCTGTTTGTGGGCATGGTCTCGTACTTGTATAGCGAATTTAT  
TCTTTTGTCTCACTATCGAGGCGAAACCGGCAGCTTACACAGAAA  
ACTGATTCATCA

The following sequences represent nucleic acid sequences encoding potentially useful EL3 antigens (or fragments or portions thereof). As above, it should be understood that the in addition to these nucleic acid sequences, the present invention relates to amino acid sequences comprising sequences encoded by SEQ ID NOS: 47-58 or derivatives, variants or homologues thereof.

SEQ ID NO: 47

EL3sequence1  
GGTTTAATTACCCAAGTTTGGAGTACTTTCTAAATCTGACCCGATCAA  
CTGATTGTGGTCTGATTAATTTTGAATACTCTCCCTGAATAGGGAG  
AGTACAAGAGTGATATCCAAAAAATAAAAAAAAAAAAAAAAAAAAA  
ACATGTCGGCCGCTCGGCCTCTAGAATA

SEQ ID NO: 48

EL3sequence2  
GGTTTAATTACCCAAGTTTGGAGTGTATGAAGCTTGCCTGAAAAAGCA  
GAGAAACCAAGAGGAGATAGTTTACAGTTCGCCAGACAGGAAATGCG  
TGCCAAGATGTTTTCGGAAGAGGAGAAACGTCGTTCACTTAGAATGAG  
AAGCATTGATTTCTGTTTGTGTTGAGATATTTAAAAATCTTTTCAG  
AAAACTTTTCAATCATAAAGTCGAAAGCCAAAAAATAAAAAAAAA  
AAAAAAAAAAAAAAAAACATGTCGGCCGCTCGGCCTCTAGAATA

SEQ ID NO: 49

EL3sequence3 (Cy-Ins-1)  
GGTTTAATTACCCAAGTTTGGAGTCTTCAACAGTAGGTTTGAATGA  
CATCGCGGATATGGCGCGCACCCAGAGCCCTCCATTAATGCTACTCTG  
TTGTTGATCAGTCTACCAGTAGCTGAGTGTAGTATTCGACTATGTGGAGT  
GCGACTAACACGAACTCTTATGGCTATCTGCAGGAATCAATATGCGGTT  
ATTCGCAAGTAAAGATCTGCTATGTGGGAAGAGCCCTGACTGGAAC  
GTGCACCTAACCAATGAAACGATCAGGGATCGCCACCGAATGCTGCGAGAA  
TCGTTGCTCATTAGCTACTTAAAGACATACTGCTGCAGCACTTAGCCTT  
GGCATCTTAAGCCGCTTTTATCTCTCTCCATGATCTCTCTCGTTATCT  
GTATAACCGAATATAGTCAATTCGGAATGCGGATGCTTAGGCCAATTTG  
TTGACGTTTGGCGCATGAATCATTGCTGTTGCTCATTATCTCACAGCG  
TGTAAGATCTCTTTTATGAAAGTCTATTTGTTTGGAGTGCACCAT  
AAACCGTTTCAAAAAAATAAAAAAAAAAAAAAAAAAATGTCGGC  
CGCTCGGCCTCTAGAATA

SEQ ID NO: 50

EL3sequence4  
GGTTTAATTACCCAAGTTTGGAGTACTTTCTAGATCTGACCCGATCAACT  
GATTGTGGTCTGATTAATTTTGGAAATCTCTCCTGAACAGGAGGAGTA  
CAAGAGTGTATATGAAAAAATAAAAAAAAAAAAAAAAAAATGTC  
GGCCGCTCGGCCTCTAGAATA

SEQ ID NO: 51

EL3sequence5  
GGTTTAATTACCCAAGTTTGGAGTACTTTCTAGCTGTTTCAAGCTCGTTCTC  
TTCGTACTTCTCACAGCTTGTGTGCTAACAGATCCAAGAGTGTAAATCCG  
AGAAAAGCGAATGGACTGGAGAGCTTACTATAGCAGATGGGTCGCGGAA  
GCTCTAATTTGGGAAACCGCGGAGTACCTTCGCGGACGAAAAATGGAGT  
TACCCGACTTTTGGACAATGGGACATTAACATCTGATGATGAAAAAGAT  
CTAATGAAATAAAGCTTCGAAAAAATAAAAAAAAAAAAAAAAAAATGTC  
ATGTCGGCCGCTCGGCCTCTAGAATA

-continued

SEQ ID NO: 52

EL3sequence6 (Cy-Cbg-1)  
GGTTTAATTACCCAAGTTTGGAGATGTTTCGAAAAATTCCTTCTGCTACTG  
5 ATCGTTGTGATCGCCCTCATTCTTTGGCGTCTGCAGATTTTTTCATGCTT  
CTTCCGCTGATACCATCTGCAAGAGCATTACATGCAGGGGCTGCACCGTGC  
CCACTTGCCTTAATGGAGACTGTATGTCACACTATGTAACACTGATGATCT  
TCACATGTCGATTAACATTTGTAACAAATACATTTTCTCTGTTTCATAA  
TAAATTTTCTACTCAAAAAAATAAAAAAAAAAAAAAAAAAATGTC  
GGCCGCTCGGCCTCTAGAATA

10

SEQ ID NO: 53

EL3sequence7  
GGCCGCGGGATTTTCTAGAGCCGAGGCGGGTTTTAGGTTGTTCTCATAA  
CTTGGGTAATTAACCACGAGGCCGAGGCGGGTTTTAGGTTGTTCTCATAA  
CTTGGGTAATTAACCACGATGGCGAGGCGGGTTTTAGGTTGTTCTCATAA  
15 CTTGGGTAATTAACCACGATGGCGAGGCGGGTTTTAGGTTGTTCTCATAA  
ACTTGGGTAATTAACCACGAGGCCGAGGCGGGTTTTAGGTTGTTCTCATAA  
AACTTGGGTAATTAACCACGATGGCGAGGCGGGTTTTAGGTTGTTCTCATAA  
AACTTGGGTAATTAACCACACTAGT

SEQ ID NO: 54

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EL3sequence8  
GGCCGCGGGATTTTCTAGAGCCGAGGCGAGTGGTATCAACGAGAGTGG  
CCATTACGGCCGGGAGAGGAAAAGTTTCTTTCTCTCGATACCCATG  
TCGGCCGCTCGGCCTCTAGAATA

SEQ ID NO: 55

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EL3sequence9  
GGCCGCGGGATTTTCTAGAGCCGAGGCGGCTTACTTGGTGGCTCAATA  
25 CTGAAAGCTTAGAATTCATTAACCTTAACCACAGGGGTTATTTGACA  
TGCTTACTTGAATGATGCTCTCTGCTTGTAGTGTGTTTATTATGCT  
AGCTGTAAGTATACTCTGGTAGACAGAACATCAATGTGCTAGTTGAATG  
TATCATGTTATCACTTTGTCACTCTATACGAATCTAGGTTGGCAGGC  
CACACCCCTCTCCTGACCTGTTTCCACATCAATAGCTTTAGCTGTTAT  
TTAATAACATCACACTGATGCAAAAAAATAAAAAAAAAAAAAAAAAA  
AACATGTCGGCCGCTCGGCCTCTAAAAATCACTAGT

SEQ ID NO: 56

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EL3sequence10  
GGCCGCGGGATTTTCTAGAGCCGAGGCGAGTGGTATCAACGAGAGTGG  
CCATTACGGCCGAGCAGTGGTATCAACGAGAGTGGCCATTACGCGCGG  
GTGGTACACGGGTGACGGGAATTAGGTTGATTCGCGAGAGGAGAC  
CTGAGAAACGGCTACCACATCAAGGAAGCAGCAGGCGCGCAAAATACC  
35 CACTCCGACCCGGGAGGTAGTGACGAAAAAATAAAAAAAAAAAAA  
AAAAAAAAACATGTCGGCCGCTCGGCCTCTAGAATAACTACTAGT

SEQ ID NO: 57

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EL3sequence11  
GGCCGCGGGATTTTCTAGAGCCGAGGCGGGTTTTAGCTCAAACCTGGGT  
AATTAACCGGTAGGATGGCGAGGCGGGTTCTCAAACCTGGGTAATTA  
ACCAAGTGGATGGCGAGGCGGGTTCTCAAACCTGGGTAATTAACCGGT  
AGGAGGCCGAGGCGGGTCTCAAACCTGGGTAATTAACCAATCACTAGT

SEQ ID NO: 58

45

EL3sequence12  
CAAGTTTGGAGTACTTTCTAGATCTGACCCGATCAACTGATTGTGGTCTG  
ATTAATTTTGGAAATCTTCTCCTGAACAGGAGAGTACAAGAGTGTATA  
TTAAGAAAAAATAAAAAAAAAAAAAAAAAAATGTCGGCCGCTC  
GGCTCTAGAATAACTACTAGT

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As such, the present invention relates to the proteins encoded by the sequences designated as SEQ ID NOS: 1, 3, 5, 7, 37, 43 and 45, the corresponding gene sequences (such as, for example, those given as SEQ ID NOS: 2, 4, 6, 8, 38, 44, 46) and proteins, peptides and/or amino acids comprising sequences encoded by SEQ ID NOS 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 39-42, and 47-58, as well as any fragments, portions, mutants, variants, derivatives, analogues and/or homologues/orthologues thereof. Furthermore, the methods described herein may provide means for detecting levels of antibodies which bind to proteins comprising (or encode by) any of SEQ ID NOS: 1-58 (or fragments, portions, mutants, derivatives, analogues or variants thereof).

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Typically the fragments, portions, mutants, variants, derivatives, analogues and/or homologues/orthologues mentioned in this invention are immunogenic or encode

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immunogenic *cyathostomin* larval antigens—that is, they are capable of generating immune, preferably humoral, responses.

The term “mutants” may encompass naturally occurring mutants or those artificially created by the introduction of one or more amino acid/nucleic acid additions, deletions, substitutions or inversions.

One of skill in this field will readily understand that proteins or nucleic acids homologous to the proteins encoded by SEQ ID NOS: 1, 3, 5, 7, 37, 43 and 45 or nucleic acid sequences of SEQ ID NOS: 2, 4, 6, 8, 38, 39-42, 44, 46, and 47-58, may exhibit as little as 20 or 30% sequence homology or identity thereto (or to a portion thereof). In other instances however, homologous proteins or nucleic acid sequences may exhibit at least 40, 50, 60, 65, 70, 75, 80, 85, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99% homology or identity the whole or part of SEQ ID NOS: 1-9, 11, 13, 15, 17, and 19 detailed above. As such, proteins or nucleic acids homologous to (or partially identical with) the proteins and/or nucleic acid sequences provided by SEQ ID NO: 1-9, 11, 13, 15, 17, and 19 are also included within the scope of this invention.

It should also be understood that natural variations due to, for example, polymorphisms, may exist between related (or homologous) proteins/genes from any given *cyathostomin* species. These variants may manifest as proteins/genes which exhibit one or more amino/nucleic acid substitutions, additions, deletions and/or inversions relative to a reference sequence (for example any of the sequences provided by SEQ ID NOS: 1-58 described above). All such variants, especially those which are functional and/or are immunogenic (or encode functional/immunogenic proteins or peptides) are to be included within the scope of this invention.

Additionally, or alternatively, analogues of the various peptides described herein may be made by introducing one or more amino acid substitutions into the primary sequence. In certain embodiments, one or more of these substitutions may represent a “conservative substitution”. One of skill in this field will understand that the term “conservative substitution” is intended to embrace the act of replacing one or more amino acids of a protein or peptide sequence with an alternate amino acid with similar properties and which does not substantially alter the physico-chemical properties and/or structure or function of the native (or wild type) protein.

As is well known in the art, the degeneracy of the genetic code permits substitution of one or more bases in a codon without changing the primary amino acid sequence. Consequently, although the nucleic acid sequences described in this application are known to encode potentially useful *cyathostomin* larval antigens, the degeneracy of the code may be exploited to yield variant nucleic acid sequences which encode the same primary amino acid sequences.

Also encompassed by this invention are splice variants of the primary gene transcripts encoded by any of the gene sequences described herein, as well as and the translated *Cyathostomin* larval antigen splice variant proteins which are encoded thereby. By way of example, splice variants of the Cy-GALA proteins described herein, including, for example, variants encoded by transcripts having 115 bp segment deletions, are within the scope of this invention. Furthermore, one of skill in this field will readily appreciate that polyadenylation variants and start codon variants, including cDNA sequences encoding the same, may also be included within the scope of this invention.

As stated, this invention finds particular application in the identification or diagnosis of *cyathostomin* infections in horses but may be more generally be used to diagnose or

identify *cyathostomin* infections present in other species of the Equidae family including, for example, donkeys and zebra

The term “sample” should be understood as including any samples comprising antibodies and/or *cyathostomin* larval antigens. For example, suitable samples may include fluids such as whole blood, plasma, serum, saliva, sweat and/or semen. In other instances “samples” such as tissue biopsies and/or scrapings may be used. In particular biopsies or scrapings from the gut may be used. In addition, a sample may comprise a tissue or gland secretion and washing protocols may be used to obtain samples of fluid secreted into, for example, the gut. In other embodiments, faecal samples may be used. One of skill will understand that in order to prepare a faecal sample for use, it may be necessary to add buffers and various protease inhibitors and subject the sample to procedures such as centrifugation, to remove particulate material. As such, “faecal samples” may represent suitable samples for use in the methods provided by this invention. As stated, a “reference” or “control” sample may be derived from healthy animals or from animals not having high mucosal burdens of *cyathostomin* parasites or larval cyathostominosis.

In order to identify a level of anti-*cyathostomin* larval antigen antibodies present in a sample, the sample may be contacted with one or more *cyathostomin* larval antigen(s) (such as those provided, comprising or encoded by SEQ ID NOS: 1-58) under conditions which permit binding between any anti-*cyathostomin* larval antigen antibodies present in the sample and the *cyathostomin* larval antigen(s). Anti-*cyathostomin* larval antigen antibodies bound to *cyathostomin* larval antigen may easily be detected with the use of agents capable of binding anti-*cyathostomin* larval antigen antibodies. In one embodiment, the agents capable of binding anti-*cyathostomin* larval antigen antibodies may be conjugated or linked to a detectable moiety.

One of skill will appreciate that while the methods provided by this invention may provide a means of detecting antibodies having affinity for, or specificity/selectivity to a single *cyathostomin* antigen (such as any described herein), in certain embodiments, the methods may exploit the use of one or more of the *cyathostomin* antigens. Since, for example, horses tend to be infected with one or more different *Cyathostomin* species, assays/methods which utilise cocktails of *cyathostomin* antigens provide a means of increasing the likelihood of a positive diagnosis. Accordingly, it should be understood that the methods described herein may use one or more of the *cyathostomin* antigens described herein.

In one embodiment, the methods provided by this invention may utilise substrates to which one or more *cyathostomin* larval antigens have been bound, conjugated or immobilised. One of skill in that art will appreciate that in addition to techniques which allow antigens to be bound, conjugated or immobilised “directly” on to the surface of substrates, other techniques may involve the use of substrates which have been coated with agents capable of binding *cyathostomin* larval antigens.

It is to be understood that the term “agents capable of binding *cyathostomin* larval antigens” may include, for example, antibodies such as monoclonal or polyclonal antibodies and/or other types of peptide or small molecule capable of binding to *cyathostomin* larval antigens. It should be noted that this definition applies to all types of binding agent mentioned herein. Furthermore, references to “antibodies” herein are intended to encompass “anti-*cyathostomin* larval antigen antibodies”.

The techniques used to generate antibodies (either monoclonal or polyclonal) are well known to one of skill and may

involve the use of *cyathostomin* antigens (or fragments or portions thereof) either isolated or purified from *cyathostomin* parasites or recombinantly generated as described herein.

Suitable substrates may include, for example, glass, nitrocellulose, paper, agarose and/or plastics. A substrate such as, for example, a plastic material, may take the form of a microtitre plate.

In order to detect a level of antibody present in a sample, immunological detection techniques such as, for example, enzyme-linked immunosorbent assays (ELISA) may be used. One of skill in this field will appreciate that ELISAs may use substrates to which *cyathostomin* larval antigens have been "captured" or bound by binding agents (capable of binding *cyathostomin* larval antigens) bound or immobilised to the substrate. Alternatively, substrates may comprise *cyathostomin* larval antigens, which have been directly bound or immobilised to the substrate.

An ELISA may involve contacting the sample to be tested with a substrate under conditions which permit binding between any antibodies present in the sample and the *cyathostomin* larval antigens bound or immobilised to the substrate as described above. One familiar with these techniques will appreciate that prior to contacting the sample to be tested with the substrate, a blocking step may be introduced to reduce incidences of non-specific binding.

An ELISA may comprise the further step of contacting the substrate with a further binding agent capable of binding one or more of the antibodies present in the sample. Such agents may otherwise be known as "secondary antibodies" and may take the form of rodent or ruminant antibodies specific to particular forms of equine antibody.

Secondary antibodies useful in the present invention may be conjugated to moieties which permit them to be detected (referred to hereinafter as "detectable moieties"). For example, the secondary antibodies may be conjugated to an enzyme capable of reporting a level via a colourimetric chemiluminescent reaction. Such conjugated enzymes may include but are not limited to Horse Radish Peroxidase (HRP) and Alkaline Phosphatase (AlkP). Additionally, or alternatively, the secondary antibodies may be conjugated to a fluorescent molecule such as, for example a fluorophore, such as FITC, rhodamine or Texas Red. Other types of molecule which may be conjugated to binding agents include radiolabelled moieties.

The amount of secondary antibody (identifiable by means of the detectable moiety) bound to the anti-*cyathostomin* larval antibodies, may be representative of the anti-*cyathostomin* larval antibodies present in the sample tested.

Alternatively, in order to identify a level of *cyathostomin* larval antigen present in a sample, a substrate or substrate comprising one or more agents capable of binding one or more *cyathostomin* larval antigens, may first be contacted with a sample to be tested. Any *cyathostomin* larval antigen bound to the substrate or to the agents capable of binding the *cyathostomin* larval antigen, may be detected with the use of a further agent capable of binding the *cyathostomin* larval antigen (referred to hereinafter as the "primary binding agent"). Additionally, or alternatively, the primary binding agents may have affinity for, or bind to *cyathostomin* larval antigen: substrate complexes or complexes comprising *cyathostomin* larval antigen and the abovementioned agents capable of binding the *cyathostomin* larval antigen. In one embodiment, the primary binding agent may be an antibody conjugated to a detectable moiety as described above.

Alternatively, any *cyathostomin* larval antigen bound to the substrate or agents capable of binding the *cyathostomin* larval

antigen, may be detected by means of a yet further binding agent having affinity for the primary binding agents. In certain embodiments, the further binding agents may be conjugated to detectable moieties.

In one embodiment, the methods for identifying a level of *cyathostomin* larval antigen or a level of anti-*cyathostomin* larval antigen antibodies, may take the form of "dip-stick" test, wherein a substrate (or portion thereof) is contacted with a sample to be tested under conditions which permit the binding of any *cyathostomin* larval antigen or anti-*cyathostomin* larval antigen antibodies present in the sample, to the substrate or a binding agent bound or immobilised thereto.

Other techniques which exploit the use of agents capable of binding the *cyathostomin* larval antigen or antibodies which bind thereto include, for example, techniques such as western blot or dot blot. A western blot may involve subjecting a sample to electrophoresis so as to separate or resolve the components, for example the proteinaceous components, of the sample. In other embodiments, electrophoresis techniques may be used to separate proteins purified from *cyathostomin* parasites and/or proteins generated in a recombinant form. The resolved components/proteins may then be transferred to a substrate, such as nitrocellulose.

In order to identify any *cyathostomin* larval antigen present in a sample, the substrate (for example nitrocellulose substrate) to which the resolved components and/or proteins have been transferred, may be contacted with a binding agent capable of binding *cyathostomin* larval antigens under conditions which permit binding between any *cyathostomin* larval antigen in the sample (or transferred to the substrate) and the agents capable of binding the *cyathostomin* larval antigen.

Advantageously, the agents capable of binding the *cyathostomin* larval antigen may be conjugated to a detectable moiety.

Additionally, the substrate may be contacted with a further binding agent having affinity for the binding agent(s) capable of binding the *cyathostomin* larval antigen. Advantageously, the further binding agent may be conjugated to a detectable moiety.

Similar techniques may also be used to detect levels of anti-*cyathostomin* larval antigen antibodies present in samples. Techniques of this type may be known as "immunoblots" or "dotblots" or "dipsticks" where *cyathostomin* antigen(s) is/are immobilised onto suitable substrates (for example a nitrocellulose substrate) and contacted with agents capable of binding *cyathostomin* antigen(s). In certain embodiments any of the samples described above may be used a source of *cyathostomin* antigen. Additionally or alternatively, the *cyathostomin* larval antigen may be isolated or purified from the parasite, or produced in recombinant form.

Other immunological techniques which may be used to identify a level of *cyathostomin* larval antigen in a sample include, for example, immunohistochemistry wherein binding agents, such as antibodies capable of binding *cyathostomin* larval antigens, are contacted with a sample such as those described above, under conditions which permit binding between any *cyathostomin* larval antigen present in the sample and the *cyathostomin* larval antigen binding agent. Typically, prior to contacting the sample with the binding agent, the sample is treated with, for example a detergent such as Triton X100. Such a technique may be referred to as "direct" immunohistochemical staining.

Alternatively, the sample to be tested may be subjected to an indirect immunohistochemical staining protocol wherein, after the sample has been contacted with a *cyathostomin* larval antigen binding agent, a further binding agent (a secondary binding agent) which is specific for, has affinity for, or

is capable of binding the *cyathostomin* larval antigen binding agent, is used to detect *cyathostomin* larval antigen/binding agent complexes.

The skilled person will understand that in both direct and indirect immunohistochemical techniques, the binding agent or secondary binding agent may be conjugated to a detectable moiety. Preferably, the binding agent or secondary binding agent is conjugated to a moiety capable of reporting a level of bound binding agent or secondary binding agent, via a colour-metric chemiluminescent reaction.

In order to identify the levels of *cyathostomin* larval antigen present in the sample, one may compare the results of an immunohistochemical stain with the results of an immunohistochemical stain conducted on a reference sample. By way of example, a sample revealing more bound *cyathostomin* larval antigen binding agent (or secondary binding agent) than in a reference sample, may have been provided by a subject with a *cyathostomin* infection.

In addition to the methods and techniques described above, the present invention also contemplates the use of a range of PCR based techniques which may be used to detect levels of *cyathostomin* antigen gene expression or gene quantity in a given sample. Useful techniques may include, for example, polymerase chain reaction (PCR) using genomic DNA as template or reverse transcriptase (RT)-PCR (see below) based techniques in combination with real-time PCR (otherwise known as quantitative PCR). In the present case, real time-PCR may be used to determine the level of expression of the genes encoding any of the *cyathostomin* larval antigens described herein. Typically, and in order to quantify the level of expression of a particular nucleic acid sequence, RT-PCR may be used to reverse transcribe the relevant mRNA to complementary DNA (cDNA). Preferably, the reverse transcriptase protocol may use primers designed to specifically amplify an mRNA sequence of interest (in this case a *cyathostomin* mRNA encoding a *cyathostomin* larval antigen). Thereafter, PCR may be used to amplify the cDNA generated by reverse transcription. Typically, the cDNA is amplified using primers designed to specifically hybridise with a certain sequence and the nucleotides used for PCR may be labelled with fluorescent or radiolabelled compounds.

One of skill in the art will be familiar with the technique of using labelled nucleotides to allow quantification of the amount of DNA produced during a PCR. Briefly, and by way of example, the amount of labelled amplified nucleic acid may be determined by monitoring the amount of incorporated labelled nucleotide during the cycling of the PCR.

Further information regarding the PCR based techniques described herein may be found in, for example, PCR Primer: A Laboratory Manual, Second Edition Edited by Carl W. Dieffenbach & Gabriela S. Dveksler: Cold Spring Harbour Laboratory Press and Molecular Cloning: A Laboratory Manual by Joseph Sambrook & David Russell: Cold Spring Harbour Laboratory Press.

Other techniques that may be used to determine the level of *cyathostomin* larval antigen gene expression in a sample, include, for example, northern and/or Southern blot techniques. A northern blot may be used to determine the amount of a particular mRNA present in a sample and as such, could be used to determine the amount of *cyathostomin* larval antigen gene expression. Briefly, total or messenger (m)RNA may be extracted from any of the samples described above using techniques known to the skilled artisan. The extracted RNA may then be subjected to electrophoresis. A nucleic acid probe, designed to hybridise (i.e. complementary to) an RNA sequence of interest—in this case the mRNA encoding a

*cyathostomin* larval antigen, may then be used to detect and quantify the amount of a particular mRNA present in a sample.

Additionally, or alternatively, a level of *cyathostomin* larval antigen gene expression may be identified by way of microarray analysis. Such a method would involve the use of a DNA micro-array which comprises nucleic acid derived from *cyathostomin* larval antigen genes. To identify a level of *cyathostomin* larval antigen gene expression, one of skill in the art may extract the nucleic acid, preferably the mRNA, from a sample and subject it to an amplification protocol such as, RT-PCR to generate cDNA. Preferably, primers specific for a certain mRNA sequence—in this case sequences encoding *cyathostomin* larval antigen genes may be used.

The amplified *cyathostomin* larval antigen cDNA may be subjected to a further amplification step, optionally in the presence of labelled nucleotides (as described above). Thereafter, the optionally labelled amplified cDNA may be contacted with the microarray under conditions which permit binding with the DNA of the microarray. In this way, it may be possible to identify a level of *cyathostomin* larval antigen gene expression.

In addition, other techniques such as deep sequencing and/or pyrosequencing may be used to detect *cyathostomin* larval antigen sequences in any of the samples described above, particularly faecal matter extracts. Further information on these techniques may be found in “Applications of next-generation sequencing technologies in functional genomics”, Olena Morozovaa and Marco A. Marra, Genomics Volume 92, Issue 5, November 2008, Pages 255-264 and “Pyrosequencing sheds light on DNA sequencing”, Ronaghi, Genome Research, Vol. 11, 2001, pages 3-11.

The present invention also extends to kits comprising reagents and compositions suitable for diagnosing *cyathostomin* infections. For example, depending on whether or not the kits are intended to be used to identify levels of *cyathostomin* larval antigen or antibodies thereto in samples, the kits may comprise substrates having *cyathostomin* larval antigens or agents capable of binding *cyathostomin* larval antigens, bound thereto. In addition, the kits may comprise agents capable of binding *cyathostomin* larval antigens—particularly where the kit is to be used to identify levels of *cyathostomin* larval antigens in samples. In other embodiments, the kit may comprise agents capable of binding the *cyathostomin* larval antigens, for example specifically raised polyclonal antibodies or monoclonal antibodies. Where the kits are intended to diagnose equine *cyathostomin* larval infections, these binding agents may take the form of antibodies capable of binding equine antibodies. The antibodies may be conjugated to detectable moieties. Kits for use in detecting the expression of genes encoding *cyathostomin* larval antigen gene may comprise one or more oligonucleotides/primers for detecting/amplifying/probing *cyathostomin* larval antigen encoding sequences. The kits may also comprise other reagents to facilitate, for example, sequencing, PCR and/or RFLP analysis. All kits described herein may further comprise instructions for use.

It will be appreciated that the uses, medicaments and methods of treatment described herein may require the generation of recombinant *cyathostomin* larval antigens (or genes encoding the same) and as such, the present invention further contemplates methods of generating and/or expressing recombinant *cyathostomin* larval antigen genes and/or proteins (such as for example those described above as SEQ ID NOS: 1-58). One of skill in this field will appreciate that PCR techniques may be exploited to selectively obtain *cyathostomin* larval antigen gene sequences from a variety of sources including,

for example, equine gut tissue, faecal matter or extracts prepared from *cyathostomin* nematodes. In one embodiment, molecular cloned *cyathostomin* larval antigen gene sequences may be introduced into a vector (such as a plasmid or expression cassette). In one embodiment, the vector may further comprise a nucleotide sequence of a tag or label to assist in protein purification procedures.

A host cell may be transformed with the vector and maintained under conditions suitable to induce expression of the *cyathostomin* larval antigen gene sequence and production of recombinant *cyathostomin* larval antigen. Techniques used to purify recombinant proteins generated in this way are known and, where the recombinant protein is tagged or labelled, these may include the use of, for example, affinity chromatography techniques.

In view of the above, further aspects of this invention provide an expression vector comprising a *cyathostomin* larval antigen gene sequence and a host cell transformed therewith, respectively.

In a further aspect, the present invention provides a method for determining whether or not an equine subject should be treated with anthelmintic drug, said method comprising the step of detecting a level of anti-*cyathostomin* larval antigen antibodies in a sample as per the first aspect of this invention and/or a level of *cyathostomin* larval antigen in a sample, wherein a level of anti-*cyathostomin* larval antigen antibodies and/or antigen, is indicative of an equine subject that should be administered an anthelmintic drug. In one embodiment, the anthelmintic drug may be Moxidectin.

#### DETAILED DESCRIPTION

The present invention will now be described in detail and with reference to the following Figures which show:

FIG. 1. ClustalW alignment of Cy-GALA-1 with its orthologues in other nematode species. *Cyathostomin* (Cy) GALA-1 is compared to *N. brasiliensis* keratin-like protein (Nb-KLP) (accession number: BAB68205); *T. circumcincta* (Tc) (AAM45145); *O. ostertagi* (Oo) (CAD22110); *C. elegans* (Ce) KLP-1 (NP\_502026) and Ce-KLP-2 (NP\_501448). The signal peptide for each sequence is underlined and the domain of unknown function (DUF148) is boxed. The histidine-rich region is highlighted in grey and the glycine-rich regions of the *C. elegans* sequences are shown in bold.

FIG. 2: Development transcription pattern of Cy-gala-1. RT-PCR was performed using gene-specific primers for Cy-gala-1 and the housekeeping gene cytochrome oxidase c subunit I (coxI), from mixed-species pools of EL3 (lane 1), DL (lane 2) and LP (lane 3) cDNA. For each reaction no-template controls were performed (N). Sizes in base pairs (bp) are labelled on the left-hand side.

FIG. 3. Immunoreactivity of rCy-GALA-1. IgG(T) reactivity to rCy-GALA-1 in horses infected with cyathostomins or other helminths as assessed by (A) immunoblot and (B) ELISA. FIG. 3A. Lane 1: Coomassie blue. Lanes 2-11: IgG (T) reactivity of specific equine sera: HF (2); CI (3); a pool of sera from *cyathostomin*-free horses (n=5) from an abattoir (4); a pool of sera from *cyathostomin*-infected horses which harboured total mucosal larval burdens of >100,000 (n=6) from an abattoir (5); horses mono-specifically infected with *P. equorum* (6), *S. edentatus* (7), *S. westeri* (8) or *S. vulgaris* (9). Also shown is IgG reactivity in sera from a rabbit before (lane 10) and after two immunisations (lane 11) with a 20 kDa complex purified from EL3/DL somatic extracts [11]. FIG. 3B. ELISA indicating IgG(T) reactivity to rCy-GALA-1 antigen in equine sera over an experimental infection [29].

Responses in the CI group are depicted by the solid lines and black shapes and in the HF group by dashed lines and white shapes.

FIG. 4: Reactivity of anti-rCy-GALA-1 antiserum to cyathostomins and other equine helminths. IgG(T) responses were assessed by (A) ELISA and (B) immunoblot. ELISA results depict binding of anti-rCy-KLP-1 anti-sera (black) and pre-immunisation serum (white). For both assays, the antigens were as follows: 1=rCy-GALA-1; 2=*cyathostomin* IL3; 3=*cyathostomin* EL3; 4=*cyathostomin* DL; 5=*cyathostomin* LP; 6=adult *A. perfoliata*; 7=adult *P. equorum*; 8=adult *S. edentatus*; 9=adult *S. vulgaris*; 10=adult *S. equinus*.

FIG. 5: Immunolocalisation of Cy-GALA. Transverse sections of DL cyathostomins were probed with anti-rCy-GALA-1 antiserum (A) and pre-immunization serum (B). Specific binding of antiserum in the parasite gut is indicated by the black arrows. The vertical bar represents 40  $\mu$ m.

FIG. 6: Schematic representation of Cy-gala-1 and the 220 bp fragment of the gene amplified from 10 *cyathostomin* species. Cy-gala-1 cDNA sequence is represented by black boxes (A). The 220 bp region PCR amplified from genomic DNA samples from 10 *cyathostomin* species is represented by the white box. The latter is expanded to indicate the position of the intron (hatched box). The range in interspecies variation for the whole gene fragment (and also without the intron sequence) are depicted. A representative PCR product of Cy-gala-1 is shown for each species (B): *C. catinatum* (1); *C. nassatus* (2); *C. goldi* (3); *C. longibursatus* (4); *C. coronatum* (5); *C. pateratum* (6); *C. ashworthi* (7); *C. leptosomum* (8); *C. minutus* (9) and *C. labiatus* (10).

FIG. 7A: Optimisation of antigen cocktails. The antibody response of encysted *cyathostomin* infected (positive) and non-infected (negative) animals is shown for varying concentrations of antigen and two different cocktails of antigen (CT1 and CT2). CT1 contains Gala 1, Gala 2, Gala 3. CT2 contains Gala 1, Gala 2, Gala 3 and CID 1. Individual antigen concentration is shown on the x axis and optical density (O.D) on the y axis. 7B: Ratio of signal for encysted *cyathostomin* infected (positive) to uninfected (negative) animals in an ELISA. Individual antigen concentration is on the x axis and ratio of positive to negative optical density on the y axis. C: shows mean serum antibody response to cocktail 1 (CT1) in groups of horses with varying infection levels. CT1 contains Gala 1, Gala 2, Gala 3. Horses were grouped as follows according to total mucosal parasite burden (TMB). Neg; uninfected horses TMB=0 (n=5), Low; TMB=0-20000, (n=8), Medium; TMB=20000-100000, (n=7), High; TMB=>100000 (n=26). Error bars show +/- standard error of the mean. O.D=optical density. D: shows mean serum antibody response to cocktail 2 (CT2) in groups of horses with varying infection levels. CT2 contains Gala 1, Gala 2, Gala 3 and CID 1. Horses were grouped as follows according to total mucosal parasite burden (TMB). Negative; uninfected horses TMB=0 (n=5), Low; TMB=0-20000, (n=8), Medium; TMB=20000-100000, (n=7), High; TMB=>100000 (n=26). Error bars show +/- standard error of the mean. O.D=optical density.

FIG. 8A-E: ROC analysis of ELISA data derived from cocktail (CT) 1 (which includes GALA-1, 2 and -3) and CT2 (which includes GALA-1, -2, -3 and CID-1). The Areas Under the Curve (AUC) are shown on each graph for each CT at the specified *cyathostomin* burden cut-off value indicated on each set of charts. The results indicate that CT1 and CT2 allow clear discrimination at different levels of *cyathostomin* mucosal burden, especially developing larval (DL) burdens above 120,000; however, it is likely that the AUC values could be improved by developing the assay to take into account *cyathostomin* species complexity and by including proteins

that specifically relate to EL3. These additional proteins have been identified and will be added systematically to the cock-tails to test their effect on AUC in the ROC analysis.†

## MATERIALS AND METHODS

### Parasite Material

Cyathostomins were collected from equine large intestinal tissue as described previously [9]. Briefly, caecum and ventral colon samples were removed at an abattoir and luminal parasites (LP), consisting of fifth stage larvae and adults, were collected from intestinal washings using sieves. Mucosal larval stages were recovered by pepsin-HCl digestion [9]. The mucosal parasites were separated into two populations based on size following previous recommendations [13]: (i) EL3 and (ii) late third stage (LL3)/developing fourth stage (DL4), collectively termed developing larvae (DL). Nematode samples for RNA extraction were placed into RNAlater (Ambion) at 4° C., while those for protein extraction and genomic DNA isolation were snap frozen in liquid nitrogen and stored at -80° C. For immunolocalisation experiments, DL were fixed in 10% formal saline. Infective third-stage larvae (IL3) were collected from horse faeces as described previously [8]. Individual adult cyathostomins were identified to species according to published recommendations [16]. Adult stage large strongyles, *Anoplocephala perfoliata* and *Parascaris equorum*, were also obtained and stored at -80° C.

### Construction of a Complementary (c)DNA Library and Immunoscreening

*Cyathostomin* RNA was extracted from DL populations by homogenisation in a mortar and pestle under liquid nitrogen, then using TRIzol (Invitrogen) according to the manufacturer's instructions. Integrity of RNA samples was assessed using a 2100 Bioanalyser (Agilent Technologies) and RNA stored in RNase-free water at -80° C. A mixed-species DL cDNA library was constructed using a SMART cDNA Library Construction Kit (Clontech Laboratories, Inc) using long distance PCR according to manufacturer's instructions. Briefly, the cDNA was synthesised by reverse transcriptase (RT)-PCR using 1 µg total RNA pooled from 11 separate DL RNA samples collected over a 6-month period from a range of intestinal sites. This was done to maximise *cyathostomin* species representation within the cDNA library. After ligation into the λ-Triplex2 vector, the cDNA was packaged into Gigapack Gold III packaging extract (Stratagene) and amplified in *Escherichia coli* XL1-Blue strain, (Stratagene). Library quality was assessed by analysing insert size in 40 plaques chosen at random. Length and identity of the inserts were determined by PCR and sequencing; the majority of plaques contained an insert with an average size of 500 base pairs (bp).

An EL3 cDNA library was constructed using the same method as for the construction of the DL cDNA library with the exception being the use of a SL1 primer to amplify nematode specific DNA prior to ligation into the Triplex2 vector. (Martin, et al, 1995). Briefly, the cDNA was synthesised by reverse transcriptase (RT)-PCR using 1 µg total RNA pooled from EL RNA samples from EL3 larvae collected from a range of intestinal sites from 6 individual horses. This cDNA was then used in a PCR with SL1 forward primer sequence: GGTTTAATTACCCAAGTTTGAG (SEQ ID NO: 59) and reverse primer sequences: ATTCTAGAGGCCGAGGC (SEQ ID NO: 60) and TTCTAGAGGCCGAGGCG (SEQ ID NO: 61). Products of this PCR were then used for packaging into the Triplex2 vector as described for generation of the DL cDNA library.

Immunoscreening was performed according to the manufacturer's protocol. For immunoscreening, two types of sera

were used: *cyathostomin*-infected (CI) and helminth-free (HF) sera [29]. Ponies in the CI group (n=3) had been trickle infected with a total of 3.9 million *cyathostomin* IL3 over a period of 9 weeks, while the HF control group (n=3) were maintained helminth-free. Serum was obtained weekly from both groups. For immunoscreening, a pool of CI sera was prepared by combining samples obtained from the three ponies at 12, 13, 14 and 16 weeks PI. The pool of HF sera was made by combining samples obtained from the three ponies at 2, 3, 4 and 6 weeks before the start of the infection period. To reduce background reactivity, both pools of sera were pre-absorbed with *E. coli* lysate by incubating equal volumes of each and rocking for 4 h at room temperature [37]. After centrifugation at 18,000×g for 10 min, the supernatant was retained for probing library filter lifts. The primary immunoscreen consisted of approximately 108,000 cDNA clones in *E. coli* XL1-Blue strain. Plaque lifts were made onto nitrocellulose filters (Hybond-C Extra, GE Healthcare). The membranes were washed [five×10 min in Tris-buffered saline (10 mM Tris, 150 mM NaCl, pH 7.4) containing 0.05% Tween-20 (TBST)1, then blocked for 1 h with 1% gelatin/TBST. In the first screen, the serum pool from the CI ponies was used at 1:200 in TBST and incubated with the membranes overnight at 4° C. The secondary antibody (goat anti-equine IgG(T), Serotec) and tertiary antibody (rabbit anti-goat[IgG]:HRP, Sigma), were incubated at 1:200 and 1:500 respectively, for 1 h each, with washing (as above) between steps. Filters were developed using SIGMAFAST DAB with Metal Enhancer (Sigma). Positive clones were isolated by taking agar plugs from the corresponding plate. Plaques that reacted non-specifically with equine sera (false positives) were identified by performing a second screen. Here, clones selected in the first round were screened as described above, except that filters were cut in half and one half probed with the CI serum pool and the other with the HF serum pool. Only plaques that reacted with the CI serum pool and not the HF serum pool were selected for sequence analysis. Vector-specific primers were used to amplify selected phage inserts and the PCR products purified using a QIAquick PCR Purification Kit (Qiagen). Each purified PCR product was sequenced using a commercial service (MWG Biotech). The resultant sequences were translated and searched against the GenBank 'non-redundant protein' database using BLASTp, and then against the 'non-human, non-mouse' EST database using tBLASTn, from the National Centre for Biotechnology Information [3]. Sequence alignments were performed using ClustalW2 [21] from the European Bioinformatics Institute (<http://www.ebi.ac.uk/Tools/clustalw2/>) and analysis for signal peptides performed using SignalP 3.0 [5]. Sequence identities were calculated using MegAlign 8.0.2 (DNASTAR) based on ClustalW alignments. Molecular mass estimations were made using an online tool from the Sequence Manipulation Suite ([http://www.bioinformatics.org/sms2/protein\\_mw.html](http://www.bioinformatics.org/sms2/protein_mw.html)) and glycosylation sites identified using ExPASy Pro site (<http://ca.expasy.org/prosite/>).

RT-PCR to Determine Temporal Transcription Pattern of the mRNA Encoding the *Cyathostomin* Gut-Associated Larval Antigen-1 (Cy-GALA-1)

Stage-specific cDNA was synthesised from 1 µg each of EL3, DL and LP total RNA using a SMART cDNA Library Construction Kit (Clontech Laboratories, Inc.). Briefly, first-strand cDNA was synthesised and amplified using the long-distance PCR method (22 cycles). Double-stranded cDNA was purified using a QIAquick PCR Purification Kit (Qiagen), eluted in 50 µl dH<sub>2</sub>O and stored at -20° C. until required. Integrity and loading of each cDNA population was assessed by amplifying a portion of the cytochrome oxidase c

subunit I (cox1) gene using primers designed to conserved sequences among cyathostomins (sense: 5'-AAAAAGGAG-GTGTGGTTC-3' (SEQ ID NO: 62); antisense: 5'-CT-TGAATTTGATAAACTACACC-3' (SEQ ID NO: 63)). PCR conditions were as follows: 0.3  $\mu$ M primers, 0.25  $\mu$ M dNTPs and 1.5 mM MgCl<sub>2</sub> with the following cycling: 94° C. for 5 min, 40 cycles at 94° C. for 30 sec, 60° C. for 30 sec and 72° C. for 30 sec, with a final extension at 72° C. for 7 min. PCR was performed using Platinum Taq (Invitrogen) with 1  $\mu$ l cDNA from each developmental stage. Primers were designed for the most abundant immunoreactive clone identified in Section 2.2 and designated *cyathostomin* gut-associated larval antigen-1 (Cy-GALA-1). The primer sequences were as follows: sense, 5'-AATTGTGGGAACAGGAG-3' (SEQ ID NO: 64); antisense, 5'-AATGAAAATCAGACTC-TAGG-3' (SEQ ID NO: 65). PCR conditions were as above, but using 35 cycles. This experiment was repeated twice and the PCR products were analysed on 2% w/v agarose gels using TrackIt 100 bp DNA Ladder (Invitrogen) for size determination. The gels were stained with 1xGelRed (Biotium). Expression of Recombinant Cy-GALA-1

The Cy-GALA-1 clone from the library immunoscreen that contained the largest insert was chosen for expression of recombinant protein. This clone incorporated the full-length coding sequence of Cy-gala-1 including the putative initiating methionine, signal peptide and poly-A tail. Primers were designed to amplify the coding sequence of Cy-gala-1 (minus the sequence that encoded the signal peptide) for sub-cloning into pET-22b(+) vector (Novagen). Appropriate sequences encoding flanking restriction enzyme sites were incorporated for uni-directional cloning. The primer sequences were as follows (NB: BamH1 and HindIII sites underlined): sense 5'-AATTCGGATCCGCAAGGTGTCATGGACCTTTTGTG-3' (SEQ ID NO: 66); antisense, 5'-CCGCAAGCT-TAATCTTTCACTGTGTGAGTCCAAAC-3'

(SEQ ID NO: 67). The PCR step was performed as described above except that the annealing temperature was 58° C. and 30 cycles were used. The PCR product was purified as described above. The pET-22b(+) vector and PCR product were digested with BamH1 and HindIII and ligation, using a 1:1 ratio of vector to PCR product, performed according to Novagen's protocol. Plasmids were transformed into *E. coli* JM109 Competent Cells (Promega) following manufacturer's instructions and selected on ampicillin-agar. A selection of colonies was subjected to colony PCR to ensure the presence of the cDNA encoding Cy-GALA-1. A colony which contained an insert of the correct estimated size was subjected to plasmid purification using a Wizard Plus SV Miniprep kit (Promega) and the purified plasmid was both sequenced and transformed into *E. coli* BL21-CodonPlus (DE3)-RIL competent cells (Stratagene) for expression of recombinant protein (rCy-GALA-1). Following induction with 1 mM isopropyl-beta-D-thiogalactopyranoside (Bioline), soluble rCy-GALA-1, present in the bacterial lysate supernatant, was purified on a His-trap HP column (GE Healthcare), following manufacturer's instructions. The purified protein was dialysed into phosphate buffered saline, pH 7.4 (PBS), using cellulose dialysis tubing (Sigma) and stored at -20° C. until required. Purified rCy-GALA-1 (0.5  $\mu$ g) was separated by SDS-PAGE, and a band at the expected size excised and subjected to matrix-assisted laser desorption/ionization time-of-flight (MALDI-ToF-ToF) mass spectrometry using an Ultraflex II MALDI-ToF-ToF mass spectrometer (Bruker Daltonics). The identity of the protein was confirmed by comparing the peptide mass fingerprint (PMF) generated to the theoretical peptide mass fingerprint (PMF) of Cy-GALA-1.

#### Preparation of rCy-GALA-1 Antiserum in Rabbits

Anti-rCy-GALA-1 antiserum was generated by injecting a rabbit with 50  $\mu$ g of rCy-GALA-1, in 0.5 mg/ml QuilA/PBS (1 ml total injection). A secondary injection was administered three weeks later, after which a test bleed indicated a specific antibody response to the recombinant antigen. This experiment was performed under the legislation of a UK Home Office Licence.

#### Immunoblotting

Soluble somatic antigen extracts were prepared from *cyathostomin* stages (IL3, EL3, DL and LP) and adult worms of other helminth species (*A. perfoliata*, *Strongylus equinus*, *Strongylus edentatus*, *Strongylus vulgaris* and *P. equorum*) as described previously [9]. However, IL3 were disrupted using a Ribolyser Fast Prep FP120 (Thermo Scientific) instead of a glass homogeniser. Proteins were separated on 4-12% polyacrylamide Bis-Tris gels (NuPAGE MES system, Invitrogen) according to the manufacturer's protocol. For immunoblotting, proteins were transferred to nitrocellulose membranes. To assess cross-reactivity, 0.1  $\mu$ g rCy-GALA-1 was loaded onto lanes of a 15-well 12% NuPAGE gel using SeeBlue Plus2 protein standards (Invitrogen) for molecular weight estimations. In one lane, 0.4  $\mu$ g was loaded and after electrophoresis was cut from the gel and stained with Coomassie blue. After transfer, the blot was sliced into separate lanes and blocked in TNTT (10 mM Tris, 0.5M NaCl, 0.05% Tween-20, 0.01% thimerosal, pH 7.4). Each of the following sera was used, diluted 1:200 in TNTT: CI and HF sera pools (described above); a pool of 5 horses found to be *cyathostomin*-free (CF) from a local abattoir and a pool of 12 horses (from the same abattoir) with mucosal *cyathostomin* burdens of >100,000 (endemic infected—EI); horses mono-specifically infected with *S. edentatus* or *S. vulgaris* [20], *P. equorum* or *Strongylus westeri* [11]. Also tested was rabbit antiserum (and pre-immunisation samples) generated to the native 20 kDa *cyathostomin* complex [11]. Sera were incubated at room temperature for 1.5 h. Washing consisted of three, 5 min incubations in TNTT. The secondary and tertiary steps were as described for the immunoscreening (above), with the exception that the anti-20 kDa antiserum blots were incubated with goat anti-rabbit Ig:HRP (Dako) at 1:500. The blots were developed as for the library screen.

For detection of Cy-GALA-1 protein in somatic extracts of cyathostomins and other helminth species, somatic extracts (9  $\mu$ g each antigen) were loaded onto 10-well, 4-12% NuPAGE gels, using 10 ng rCy-GALA-1 for comparison. After transfer to nitrocellulose, periodate treatment of the blots was performed as described previously [9]. The blots were probed with pre-immunisation rabbit serum and anti-rCy-GALA-1 serum at 1:300 in TNTT, followed by goat anti-rabbit(Ig):HRP (Dako) at 1:500, and developed as described above. Three 5 min washes were applied between steps.

#### Enzyme-Linked Immunosorbant Assay (ELISA)

To test reactivity of experimentally infected pony sera (CI) to rCy-GALA-1 over the course of infection, the following conditions were used. Each well of a Microton High Binding plate (Greiner Bio-One) was coated with 100  $\mu$ l of rCy-GALA-1 (1  $\mu$ ml<sup>-1</sup> in bicarbonate coating buffer, 0.1 M, pH 9.6) overnight at 4° C. Plates were washed with 0.05% Tween-20 in PBS (PBST), six times. Block solution (2% soya infant powder (w/v) in PBST) was added, 200  $\mu$ l per well, and incubated for 1 h at 37° C. Plates were washed six times and Cland HF sera (1:200 in block solution), from weekly time points 2 weeks before infection to 16 weeks PI, added and incubated for 2 h at 37° C. After washing, 100  $\mu$ l goat anti-equine IgG(T) were added, diluted 1:200 in blocking solu-

tion. After 1 h at 37° C. and washing, 100 µl rabbit anti-goat (Ig):HRP were added, diluted in block at 1:500, and incubated for 1 h at 37° C. To develop the reaction, Sigma-FAST OPD tablets (Sigma) were dissolved in H<sub>2</sub>O according to the manufacturer's instructions and 100 µl added to each well and incubated for 15 min. Fifty µl of 2.5 M H<sub>2</sub>SO<sub>4</sub> were added to stop the reaction and the absorbance read at 490 nm. The same conditions were used to measure the anti-rCy-GALA-1 antiserum response to somatic extracts of *cyathostomin* stages and other adult helminth species extracts, except that these were coated at 2 µgml<sup>-1</sup>. The antiserum and goat anti-rabbit:HRP were used at 1:500.

#### Immunolocalisation

*Cyathostomin* DL were fixed in 10% formal saline and immobilised in a solidified gelatin plug by mixing with molten 5% gelatin/PBS (<30° C.) and allowing to set. The plugs were then dehydrated with alcohol and xylene and embedded in paraffin wax. Sections were cut at 3 µm using a microtome and the slides stored at 4° C. Immunolocalisation was performed using an EnVision+ System-HRP for rabbit primary antibodies (DakoCytomation) in a Sequenza Slide Rack (Thermo Scientific) at room temperature. After de-waxing, the slides were incubated in 0.5% Tween-80/PBS (PBST80) with 0.3% H<sub>2</sub>O<sub>2</sub> for 20 min, to inactivate endogenous peroxidases. Blocking was performed using 100 µl 25% normal goat serum (NGS) in PBST80 for 1 h. Rabbit antisera obtained prior to and after two immunisations with rCy-GALA-1 were diluted 1:100 in 10% NGS/PBST80, and 100 µl incubated on the slides for 1 h. After two washes in PBS at room temperature, 100 µl of HRP-labelled polymer conjugated to goat anti-rabbit Ig, was incubated (neat) for 30 min. The reactions were developed in neat 3-amino-9-ethylcarbazole substrate chromogen for 7.5 min. Slides were washed in H<sub>2</sub>O and counterstained using haematoxylin.

#### Single Worm PCR to Identify the Gene Encoding GALA in Different *Cyathostomin* Species

Genomic DNA was isolated from 54 individually identified adult cyathostomins using the DNeasy Blood and Tissue kit (Qiagen) according to their protocol, but with the addition of a homogenisation step before the proteinase K digestion step; each individual was disrupted briefly using a 1.5 ml microfuge tube homogeniser in 50 µl ATL buffer supplied with the kit. The following 10 species were examined (NB: numbers of worms used for each species is shown in parenthesis): *Cyathostomum catinatum* (10), *Cylicostephanus goldi* (8), *Coronocyclus coronatus* (6), *Cyathostomum pateratum* (6), *Cylicocyclus nassatus* (6), *Cylicostephanus longibursatus* (5), *Cylicocyclus ashworthi* (4), *Cylicocyclus leptostomum* (3), *Coronocyclus labiatus* (1) and *Cylicostephanus minutus* (1). The same primers used in Section 2.3 for RT-PCR were used to amplify a conserved fragment of Cy-gala in each species. The cycling conditions were: 2 min at 94° C., followed by 40 cycles of 15 sec at 94° C., 30 sec at 58° C. and 60 sec at 72° C., and a final extension at 72° C. for 7 min. PCR products were analysed on agarose gels as described above and PCR products from each of the 54 individuals cloned into pGEMT-Easy (Promega) according to manufacturer's instructions. Each clone was sequenced in forward and reverse directions with vector-specific primers using the commercial sequence facility described above.

#### Results

##### Immunoscreening of the *Cyathostomin* DL cDNA Library and Sequence Analysis of Cy-Gala-1

The primary immunoscreening yielded 33 positive clones; five of which were excluded as false positives on the basis of the secondary screen using HF sera. The remaining 28 clones contained inserts ranging in size from approximately 500 to

1500 bp. Sequence analysis indicated that 15 of these showed high identity to one another (73-100% at the amino acid [aa] level). One of these (Cy-gala-1) represented a full-length coding sequence: i.e. it contained a putative initiation codon, signal peptide and termination codon upstream of a poly-A tail. The entire coding sequence was 223 aa which, after cleavage of the signal peptide, would result in a 206 aa mature protein estimated at 25.6 kDa. Cy-GALA-1 contains a highly conserved domain as revealed by a domain search via BLASTp analysis [28]. The function of this domain is unknown and in *Caenorhabditis elegans* is designated Domain of Unknown Function 148 (DUF148). The Cy-GALA-1 sequence displayed highest aa identity to a sequence from *Nippostrongylus brasiliensis* (accession number: BAB68205; 35% identity over 128 residues), also identified via immunoscreening [38]. Two predicted proteins from *C. elegans* showed 34% identity over 105 residues to Cy-GALA-1. These proteins were 44.5% identical to each other. Also identified, were two trichostrongyloid ESTs: one from *Teladorsagia circumcincta* L3 (accession number: AAM45145), which displayed 32% identity to Cy-GALA-1 over 102 aa, and one from *Ostertagia osteragi* adult worms (accession number: CAD22110) with showed 32% identity over 140 aa. The two *C. elegans* orthologues (referred to here as Ce-KLP-1 (NP\_502026) and Ce-KLP-2 (NP\_501448)) contain glycine-rich domains which gives them homology to keratin sequences and hence their designation as 'keratin-like' proteins (KLP). All the parasitic nematode sequences described here lack this glycine rich sequence, despite some being previously designated as 'KLP-like' proteins [38]. Rather than classifying Cy-GALA-1 as a KLP, it was instead named to reflect its localisation to the gut (see below). An alignment of Cy-GALA-1 with its orthologous sequences in *N. brasiliensis* and *C. elegans* is depicted in FIG. 1. In all the parasitic nematode sequences, except that of *T. circumcincta*, a histidine-rich motif precedes DUF148 (FIG. 1); its function is unknown. In addition, four potential N-linked glycosylation sites were identified. Searching Cy-GALA-1 at Nembase gave additional significant hits. All of these EST sequences contained regions with high identity to DUF148 and some had glycine-rich regions. The closest matches were to sequences identified in adult *Ancylostoma ceylanicum* (accession numbers: CB176510, CB190303 and CB339159), with 45-46% aa identity to Cy-GALA-1 over 110 residues.

##### Temporal Transcription Pattern of Cy-Gala-1

Cy-gala-1 transcript was detected in DL and EL3 cDNA and not in cDNA from LP parasites (FIG. 2). After 40 cycles, similar levels of coxI PCR product were observed in DL and LP cDNA. However, a coxI PCR product from EL3 was less intense, indicating low quality of EL3 cDNA. This was due to degradation of EL3 RNA caused by the extensive digestion method required to harvest these larvae. These results indicate the apparent specificity of this transcript for mucosal stages; hence the gene was selected for expression of recombinant protein for assessment as a diagnostic marker.

##### Expression of rCy-GALA-1 and its Immunoreactivity

rCy-GALA-1 was obtained from the soluble fraction of the *E. coli* lysate; the purified protein was approximately 28 kDa (FIG. 3). The identity of this protein as rCy-GALA-1 was confirmed by MALDI-ToF-ToF (data not shown). Its molecular weight was slightly higher than the expected size of native Cy-GALA, calculated to be 25.6 kDa, and was due to addition of the His-tag and *E. coli* signal peptide. Anti-rCy-GALA-1 antiserum predominantly recognised the expected size band in somatic DL extracts (Section 3.4 and FIG. 4). The immunoreactivity of the recombinant antigen is shown in FIG. 3. Only IgG(T) in CI and EI sera equine sera bound rCy-GALA-

1, indicating that both experimentally and naturally infected horses recognise this antigen. Sera from horses harbouring other parasitic helminths did not contain IgG(T) that bound Cy-GALA-1. The rabbit antiserum to the *cyathostomin* larval anti-20 kDa complex generated previously [11], showed strong reactivity to rCy-GALA-1.

Levels of rCy-GALA-1-specific IgG(T) in sera from infected vs. non-infected ponies [29] were measured by ELISA (FIG. 3). Increases in rCy-GALA-1-specific IgG(T) levels were observed in all infected ponies by 6 weeks PI. A more rapid increase was observed in pony 104. Antigen-specific IgG(T) levels plateaued at 8 weeks PI for 104 and 12 weeks PI for 101 and 105; these levels remained elevated until the end of the measurement period at 16 weeks PI. No significant increases in rCy-GALA-1-specific IgG(T) levels were observed in any of the HF ponies throughout the experiment. Murphy and Love (1997) [29] described clinical signs in the infected animals from 4-6 weeks PI. While all showed a slower increase in percentage weight gain than the control group, pony 104 showed a drop in weight gain over weeks 4-8 PI. These signs may indicate a higher level of infection in 104. Anti-rCy-GALA-1 antiserum reactivity was tested against somatic extracts from *A. perfoliata*, *P. equorum*, *S. edentatus*, *S. vulgaris* and *S. equorum* [FIG. 4]. No reactivity was observed except to a band at 38 kDa in the *P. equorum* extract. Binding to this band was less than that seen in the *cyathostomin* DL lane (FIG. 4).

#### Detection of Cy-GALA-1 in Different *Cyathostomin* Stages

Antiserum raised to rCy-GALA-1 was used to investigate the presence of the native protein in different *cyathostomin* stages (FIG. 4). This antiserum bound the 28 kDa recombinant antigen (FIG. 4, lane 1): an additional band at 53 kDa was bound and may represent a dimeric form of Cy-GALA-1. The anti-rCy-GALA-1 antisera showed reactivity to EL3 and DL somatic extracts but not to adult extract (FIG. 4). Immunoreactivity to antigens in EL3 and DL stages was primarily directed at molecules of approximately 26 kDa, corresponding to the calculated molecular mass of Cy-GALA-1. Two other EL3 and DL antigens were bound by IgG in anti-Cy-GALA-1 antisera, one at approximately 45 kDa and the other at 55 kDa. The ELISA results indicated high reactivity to DL, however no binding was observed in EL3 or adult extract.

#### Immunolocalisation of Cy-GALA-1

DL were subjected to immunolocalisation studies (FIG. 5). Reactivity was detected in the gut of individual worms, where considerable staining was observed on the gut epithelium and in the gut lumen. No reactivity was detected to any other structures in the nematodes.

#### Single Worm PCR to Identify the Gene Encoding Cy-GALA in Different *Cyathostomin* Species

Single worm PCR experiments were performed using primers to amplify a 220 bp fragment of Cy-gala-1 from 50 morphologically-identified adult worms encompassing 10 species. A PCR product was obtained from all nematodes tested and sequencing confirmed that PCR products representative of each species encoded Cy-gala sequence. FIG. 6 shows a schematic representation of this fragment and PCR products from each species. There was variation in size of the PCR product obtained from different species; from 267 bp (for all *C. coronatus* individuals) to 284 bp (for one *C. goldi* individual). This variation was due to a difference in intron size at this site amongst the species. The precise location of the intron was conserved as indicated by splice site analysis (FIG. 6). Nucleotide identities between individuals from different species ranged from 78.9-99.1% for the whole fragment. Higher nucleotide identities were observed in the coding region; interspecies variation ranged from 82.2-98.9% over 180 nt, while the amino acid identities were 80-100% over 60 residues. At the aa level, intra-species variation was as follows: *C. catinatum* 93.3-100%; *Cs. goldi* 90.0-100.0%; *Co. coronatus* 96.7-100.0%; *C. pateratum* 93.3-100%; *Cc. nassatus* 88.3-98.3%; *Cs. longibursatus* 91.7-100%; *Cc. ash-*

*worthi* 90.0-100.0%; *Cc. leptostomum* 88.3-100.0%. In an attempt to assign a species for the library clone, Cy-gala-1, the coding sequence from each individual was compared against Cy-gala-1 in this 220 bp fragment. The highest identity was found to a *C. pateratum* individual (97.8% nt identity and 98.3% aa identity). Therefore, with the available sequence data for each species, we have provisionally identified Cy-gala-1 as belonging to *C. pateratum*.

#### Optimisation of Antigen Cocktails.

The optimum concentration of antigen to use in an ELISA using a cocktail of antigens was evaluated using sera from *cyathostomin* infected (positive) and non-infected (negative) animals. FIG. 7A shows the serum antibody response to varying concentrations of antigen in two different cocktails of antigen (CT1 and CT2). CT1 contains GALA-1, -Gala 2 and -3. CT2 contains these three antigens plus CID1. Individual antigen concentration is shown on the x-axis and optical density (O.D) on the y-axis. FIG. 7B shows the ratio of the OD signal obtained on *cyathostomin* infected (positive) vs. uninfected (negative) animals in an ELISA. Individual antigen concentration is on the x-axis and ratio of positive to negative optical density on the y-axis.

#### Evaluation of Antigen Cocktail for Discriminating Different Levels of Infection.

Two different cocktails of antigen were tested in an ELISA to assess their potential for discriminating different levels of mucosal infection. FIGS. 7 C and D shows mean serum antibody response to cocktail 1 (CT1) and cocktail 2 (CT2) respectively in groups of horses with varying infection levels. CT1 and CT2 were as described above. Horses were grouped as follows according to total mucosal parasite burden (TMB). Neg; uninfected horses TMB=0 (n=5), Low; TMB=0-20000, (n=8), Medium; TMB=20000-100000, (n=7), High; TMB=>100000 (n=26). Error bars show +/- standard error of the mean. O.D=optical density.

#### Discussion

Identification of the *cyathostomin* GALA sequence is an advance in the development of an ELISA for the diagnosis of larval cyathostomiasis. Three important criteria were met by this protein: 1) it appeared to be specific to larval stages; 2) there was no cross reactivity with the other equine helminth species assessed here and 3) the gene encoding the protein was isolated from all *cyathostomin* species examined with a relatively low level of sequence variation amongst the species. Furthermore, serum IgG(T) responses to rCy-GALA-1 increased within 5 weeks of the administration of an experimental infection and the protein was also the target of IgG(T) responses in naturally infected horses.

The RT-PCR, immunoblot and ELISA results indicated that Cy-GALA-1 is restricted to parasitic larval stages, particularly DL stages. This is a vital feature for a diagnostic marker that specifically indicates mucosal larval burden. Despite numerous attempts, RNA extracted from EL3 was of relatively poor quality so it was difficult to judge precise levels of transcription in these stages. EL3 require extensive digestion in pepsin/HCl at 37° C. to remove them in sufficient quantity from the intestinal mucosa and submucosa and so it is technically difficult to obtain sufficient high quality RNA. The EL3 somatic protein extracts also contained a small amount of contaminating host protein (it is impossible to totally separate every single worm from its host capsule), and this may have resulted in the lower levels of reactivity of EL3 extracts to Cy-GALA-1 antiserum as indicated by the ELISA results. Immunolocalisation was also attempted in EL3, but degradation resulted in a lack of distinct morphology and no specific binding was observed (data not shown). Therefore it remains to be fully elucidated if Cy-GALA is a significant immunogen of EL3, or is predominantly an antigen of the later larval stages. Immunolocalisation studies of diseased equine mucosa are planned, to provide EL3 embedded in their mucosal cysts. Serum IgG(T) responses to rCy-GALA-1 over the time course of an experimental infection showed that the

antigen is a reasonably early indicator of infection and these responses were identified whilst the infections were not patent [29]. Indeed, in these ponies, the infections never progressed to patency even though the experiment was continued until 60 and 62 weeks PI in two of the animals. Substantial increases in reactivity were observed at 5 weeks PI in one animal (pony 104) and by 6 weeks in all ponies. *cyathostomin* larval-specific serum IgG(T) responses were analysed previously in these animals and similar dynamics of responses were observed to the 20 and 25 kDa complexes purified from EL3/DL mixtures [11]. Furthermore, serum IgG(T) reactivity to crude larval antigen was also observed to increase only after 6 weeks PI in these ponies [9], suggesting that only by this time point do larvae stimulate a detectable serum IgG(T) response. Pony 104 had the most pronounced increase in IgG(T) to rCy-GALA-1 and this is similar to its response to crude larval antigen and the purified 20- and 25-kDa antigen complexes [9, 11]. The clinical signs observed in this pony (reduced weight gain, lowest plasma fructosamine) indicate that it may have had a greater burden of mucosal larvae [29]. Indeed, when this animal was euthanized at 20 weeks PI it was found to have a high *cyathostomin* burden. Unfortunately the other two ponies in the group were necropsied at 60 and 62 weeks PI so their burdens cannot be directly compared with pony 104. Nevertheless, the data provides preliminary evidence that this recombinant antigen may be able to distinguish varying degrees of disease.

As mentioned above, there is similarity of the IgG(T) response to rCy-GALA-1 and to the two larval antigen complexes purified and shown to have diagnostic potential previously [10, 11]. The molecular mass of Cy-GALA, estimated at 25.6 kDa, means that it could feasibly be a component of the 25 kDa antigen complex, an observation supported by the results using anti-rCy-GALA-1 against EL and DL somatic extracts in western blots. Antiserum generated to the 20 kDa complex in rabbits also bound rCy-GALA-1 indicating its presence in this complex also. This is not altogether surprising as these complexes were excised rather crudely from SDS-polyacrylamide gels [10, 11].

Specificity of Cy-GALA-1 in the cyathostominae was confirmed by probing the recombinant protein with sera from horses infected mono-specifically with heterologous helminth species. While experimentally infected (CI) and naturally infected (EI) horses recognised rCy-GALA-1, IgG(T) in serum from horses with large strongyle infections (*S. edentatus*, *S. westeri* or *S. vulgaris*) and *P. equorum* infection, did not bind the antigen. Cross-reactivity was further explored by probing somatic extracts of other equine parasites with anti-rCy-GALA-1 serum: extracts from *A. perfoliata*, *P. equorum*, *S. edentatus*, *S. vulgaris* and *S. equorum* were analysed. In the ELISA no binding above background levels was observed in any of the five other parasite extracts. In the immunoblot, there was a degree of binding to a band of approximately 38 kDa in the *P. equorum* extract, but this was of far less intensity than binding observed in the *cyathostomin* DL samples. Furthermore, there was no cross reactivity to *P. equorum* antigens when the samples were assessed using the ELISA.

The presence of sequences encoding GALA-like proteins was confirmed in 10 *cyathostomin* species, indicating ubiquity of this gene in the group. There are currently 50 recognised *cyathostomin* species [23], and while a large number of species are often found in infected individuals [6,7], the bulk of the burden is consistently found to comprise 5-10 species [26, 27, 36]. Nine of the species explored in this study belong to the 10 most common cyathostomins as identified by Reinemeyer et al. (1984) [36], Ogbourne (1976) [30] and Lichtenfels et al 2001 [22]. The presence of Cy-GALA in these species indicates it is likely to be present in most, if not all, cyathostomins. An analysis of the sequence of Cy-gala-1 amongst the cyathostomins indicated a low level of sequence diversity across the selected 220 bp region. It is possible that greater diversity exists outside this region and the full-length cDNA

sequences of Cy-gala are currently being isolated from a number of species to investigate this further. Promisingly, for development of a specific immunoassay, the levels of sequence diversity identified thus far are substantially lower among cyathostomins than they are when the Cy-gala sequences are compared to orthologous sequences in other nematode species, i.e. 80-100% vs. 25-35% identity. The nematodes that were present in the CI pony group unfortunately had not been identified, so it is difficult to compare levels of rCy-GALA-1 IgG(T) with the species present.

A factor that must be considered in the development of any helminth immunodiagnostic assay is the length of time that circulating specific immunoglobulin levels take to return to normal values after anthelmintic treatment. Since the ponies used in the experimental infection were not treated with anthelmintic before necropsy, this could not be assessed here. Studies on a commercially-available serological ELISA for *A. perfoliata* [33, 34], which is based on the specific binding of IgG(T) to a purified 12/13 kDa antigen complex, indicated that post-treatment IgG(T) levels can take months to reduce to 'non-infection' levels [2,4]. Also, Kjaer et al. (2007) [18] found that two thirds of horses which had no visible signs of tapeworm infection at necropsy had ELISA ODs higher than the current accepted cut-off for infection (0.2). Despite this, the *A. perfoliata* 12/13 kDa antigen ELISA is still regarded as the most useful diagnostic tool for infection [1, 18]. These observations suggest that circulating IgG(T) levels may remain high for a time after treatment and this will be considered when designing how a *cyathostomin* diagnostic assay, based on IgG(T), could be used in future.

No function has been ascribed to orthologues of Cy-GALA in other nematode species and only Nb-KLP has been characterised in any detail [38]. It was speculated that Nb-KLP may be a cuticular protein, based on its identity to Ce-KLPs, which are described as 'keratin-like'. However the authors did not explore this further. Ce-KLP-1 and -2 encode hypothetical proteins, and some information regarding these is available in WormBase (www.wormbase.org). Both are predicted to be alpha-helical proteins, and Ce-KLP-1 has been confirmed by transcript evidence, while Ce-KLP-2 has been partially confirmed. Ce-KLP-1 shows no RNAi phenotype, while Ce-KLP-2 displays 'embryonic lethal', indicating that it may play a role in development. An anatomic expression plan is available for Ce-KLP-2, showing expression in pharyngeal muscles and tail neurons which is different to what was observed here with localisation of Cy-GALA to the worm intestinal lumen. The function of this molecule remains to be elucidated.

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Additional immunoreactive clones: The following table lists additional sequences encoding immunoreactive cyathostomin antigens which were identified from the larval cDNA library, from two screenings which revealed distinct clones. The first immunoscreen (A) used serum from experimentally cyathostomin-infected ponies (from a previous study<sup>1</sup>), and the second immunoscreen (B) used a pool of sera from naturally infected horses; both groups had high parasite burdens. The antigens which have been checked for immunogenicity and cross specificity by recombinant bacterial expression are also indicated.

Antigen	Number of clones per screen		Closest homologues (Accession numbers in brackets)	Amino acid identity	Transcription pattern			Immuno-genicity	Cross speci-ficity
	A	B			EL3	DL	LP		
Gut-associated larval antigen (GALA)	15	1	Keratin-like protein, <i>Nippostrongylus brasiliensis</i> , (BAB68205). Keratin-like proteins, <i>Caenorhabditis elegans</i> (NP_502026 and NP_501448)	35% over 128 a.a. 34% over 104 a.a.	+	++	-	+	+
Glutathione-S-transferase (GST)	1	0	Cytosolic GST from <i>Oesophagostomum dentatum</i> (ACA30415)	85% over 209 a.a.	-	+	+		
Galectin-1 (GAL-1)	1	0	Galectin family member, <i>C. elegans</i> (NP_495163)	83% over 279 a.a.	-	++	+		
Galectin-2 (GAL-2)	0	1	Galectin family member, <i>Haemonchus contortus</i> (AAF63406)	91% over 259 a.a.	+	++	++		
Nematode polyprotein allergen/antigen (NPA)	4	0	NPA from <i>Dictyocaulus viviparus</i> (Q24702)	42% over 314 a.a.	-	++	++		
Cyathostomin immunodominant antigen-1 (CID-1)	3	0	EST from larval-stage <i>Necator americanus</i> (BG467549). Function of this is unknown.	59% over 61 a.a.	-	++	++	+	
Surface associated antigen (SAA)	4	8	SAA-2, <i>N. americanus</i> (ACE79378)	71% over 146 a.a.	+	++	++		
Fatty acid/retinol binding protein -1 (FAR-1)	0	1	Putative ES protein with FAR binding domain, <i>Ostertagia ostertagi</i> (CAD20464)	45% over 100 a.a.	+	++	++		
Fatty acid/retinol binding protein -2 (FAR-2)	0	1	FAR binding protein, <i>Ancylostoma ceylanicum</i> (ACC76809)	72% over 160 a.a.	-	++	+		
Globin (GLO)	0	15	Cuticle globin, from <i>Syngamus trachea</i> (AAL56426)	54% over 161 a.a.	+	++	++		
Clone of unknown function -20a (Unk-20a)	0	1	No homology found	NA	-	++	++		
Unk-46a	0	1	Third-stage larval EST, <i>N. brasiliensis</i> (EH359049)	33% over 124 a.a.	-	++	++		
Unk-50a	0	1	Hypothetical protein, <i>C. elegans</i> (NP_490737)	33% over 140 a.a.	-	++	+		

<sup>1</sup>Murphy D., Love S., The pathogenic effects of experimental cyathostome infections in ponies, Vet. Parasitol. (1997) 70: 99-110.

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Homologues of cyathostomin gut-associated larval antigen (Cy-GALA)			Homologues of cyathostomin gut-associated larval antigen (Cy-GALA)		
Homologue	Amino acid identity to Cy-GALA-1	Putative species	Homologue	Amino acid identity to Cy-GALA-1	Putative species
Cy-GALA-1	—	<i>C. patemtum</i>	Cy-GALA-3	77.7 %	<i>C. coronatus</i>
Cy-GALA-2	83.3 %	<i>C. nassatus</i>	Cy-GALA-4	93.0 %	<i>C. catinatum</i>

Sequences of a conserved region of GALA from individual cyathostomin species

Species	Sequence	Number of individuals	Intraspecies aa identity range
<i>C. ashworthi</i>	LTFAEKKGKISEWAKKYNVVDEVASYNAYREKLKQEHKKNVS (E/V) LVSGLP (G/D) AVKKVN (E/V) LLD (SEQ ID NO: 68)	4	90.0-100%
<i>C. catinatum</i>	LTFAEKK (E/K) EISEWAKKYNVVDEVASYNAYREKLKQEHKKNVSE LVSALPNAVKKVNDLLD (SEQ ID NO: 69)	10	91.7-100%
<i>C. coronatus</i>	LTFAEKKKISEWAKKYKVEDEVASYNAYREKLKQEHKKNVSELVSA LPGAVKKVNELLD (SEQ ID NO: 70)	6	96.7-100%

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Sequences of a conserved region of GALA from individual cyathostomin species			
Species	Sequence	Number of individuals	Intraspecies aa identity range
<i>C. goldi</i>	LTFAEKKKEISEWAKKYNVVDEVASYNAYREKLKQEHRKNVSELVSD LPSAVKKVNDLLD (SEQ ID NO: 71)	8	90.0-100%
<i>C. labiatus</i>	LTFAEKKKEKISEWAKKYNVVDEVARYNAYREKLKQEYRKNVSELVSG LPNAVKKVNDLLD (SEQ ID NO: 72)	1	—
<i>C. leptostomum</i>	LTFAEKKKGISEWAKKYNVVDEVASYNAYREKLKQEHRKNVSELVSG LPGAVKKVNELLD (SEQ ID NO: 73)	3	88.3-100%
<i>C. longibursatus</i>	LTFAEKKKEISKWAKKYNVVDEVASYNAYREKLKQEHRKNVSEIVSD LPNAVKKVNDLLD (SEQ ID NO: 74)	5	91.7-100%
<i>C. minutus</i>	LTFAEKKKEKISEWAKKYNVVDEVASYNAYREKLKQEHRKNVSQLVSA LPNAVKKVNDLLD (SEQ ID NO: 75)	1	—
<i>C. nassatus</i>	LTFAEKKKEKIGEWAKKYNVVDEVAXYNAYREKLKQEHRKNVSELVSG LPNAVKKVNELLD (SEQ ID NO: 76)	6	88.3-100%
<i>C. pateratum</i>	LTFAEKK(K/E)EISEWAKKYNVVDEVASYNAYREKLKQEHRKNVSE LVSALPNAVKKVNDLLD (SEQ ID NO: 77)	6	93.3-100%

## SEQUENCE LISTING

&lt;160&gt; NUMBER OF SEQ ID NOS: 83

&lt;210&gt; SEQ ID NO 1

&lt;211&gt; LENGTH: 223

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: cyathostomum pateratum

&lt;400&gt; SEQUENCE: 1

Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Ser Ala Val Ala Leu  
1 5 10 15

Ala Gln Gly Val Met Asp Leu Phe Gly Glu Glu Gly Arg Glu Glu His  
20 25 30

Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val  
35 40 45

Ser Cys Glu Ala Lys Trp Glu Tyr Phe Lys Ile Val Gly Asn Arg Ser  
50 55 60

Leu Thr Phe Ala Glu Lys Arg Lys Glu Ile Ser Glu Trp Ala Lys Lys  
65 70 75 80

Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys  
85 90 95

Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Ala Leu  
100 105 110

Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln  
115 120 125

Thr Pro Arg Gln Leu Tyr Val Ala Leu Arg Lys Leu Gly Arg Gln Asn  
130 135 140

Pro Ala Leu Tyr Arg Ile Val Glu Tyr Ile Asn Val Ala Val Arg Leu

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145	150	155	160
Arg Ser Glu Glu Val Asp Glu Gln Glu Gln Arg Arg Arg Leu Ser Ala	165	170	175
Leu Pro Phe Gly Asp His Asn Asp Asn Leu Glu Glu Gln Asp Phe Gly	180	185	190
Glu Gln Asp Phe Arg Tyr Val Tyr Gly Phe Glu Cys Ala Arg Phe Leu	195	200	205
Leu Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Glu Arg Tyr	210	215	220

<210> SEQ ID NO 2  
 <211> LENGTH: 669  
 <212> TYPE: DNA  
 <213> ORGANISM: cyathostomum pateratum

<400> SEQUENCE: 2

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atgaacaaaa cgtaacatt ttcacagtc gttagtgcc tagctctggc ccaaggtgtc      60
atggaccttt ttggtgaaga gggtcgtgaa gaacatcgtc gtcaccatcg tcattcactt    120
ttaccacat atctccacaa tgtgagctgt gaggctaaat gggagtactt caaaattgtg     180
gggaacagga gtttgacctt tgctgagaaa agaaaggaaa ttacgagtg ggcaaaaaaa     240
tacaatggtg tggatgaagt tgcaagctac aatgcttaca gggaaaaact caagcaggag    300
cacagaaaaa acgtagcga acttgtttct gctcttccaa acgcagtgaa gaaagtcaat    360
gatcttctag acaatgaaa tcagactcct aggcaacttt acgttgccct tagaaaactt    420
ggtagacaaa atccggcact ttaccgtatt gtcgagtaca ttaatgtggc tgtaagacta    480
agaagtgaag aagtggatga gcaagaacaa cgaagaaggc tgtagctct accttttggc     540
gaccataacg ataatttga agagcaggac ttcggtgaac aagactttcg ctatgtctat    600
ggctttgagt gtgcaagatt tctccttcaa aatggaagaa tgtttgact taacacagat    660
gaaagatat                                     669
    
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<210> SEQ ID NO 3  
 <211> LENGTH: 192  
 <212> TYPE: PRT  
 <213> ORGANISM: Cylicocyclus nassatus

<400> SEQUENCE: 3

His Glu Glu Leu Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr	1	5	10	15
Leu His Asn Val Ser Cys Glu Ala Lys Trp Glu Tyr Phe Lys Ile Val	20	25	30	
Gly Asn Arg Ser Leu Thr Phe Ala Glu Lys Lys Gly Lys Ser Ser Glu	35	40	45	
Trp Ala Lys Lys Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala	50	55	60	
Tyr Arg Glu Lys Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu	65	70	75	80
Val Ser Gly Leu Pro Gly Ala Val Lys Lys Val Asn Glu Leu Leu Asp	85	90	95	
Asn Glu Asn Gln Thr Pro Arg Gln Leu Tyr Val Ala Leu Arg Lys Leu	100	105	110	
Gly Lys Gln Asn Pro Val Leu Tyr Arg Val Val Glu Phe Val Asn Leu	115	120	125	
Val Val Arg Phe Arg Arg Glu Asp Ser Asp Glu Gln Glu Gln Arg Glu	130	135	140	

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Met Leu Ser Thr Leu Pro Phe Ser Glu Asn Asn Glu Glu Gln Asp Leu  
145 150 155 160

Gly Glu Gln Asp Phe Gln Tyr Ile Tyr Gly Phe Glu Cys Ala Arg Phe  
165 170 175

Ile Phe Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr  
180 185 190

<210> SEQ ID NO 4  
<211> LENGTH: 576  
<212> TYPE: DNA  
<213> ORGANISM: *Cylicocyclus nassatus*

<400> SEQUENCE: 4

catgaagaac ttcgctgtca ccatcgatc tcaactttac cacctatct ccacaatgtg 60  
agctgtgaag ccaaatggga atacttcaag attgtgggga acaggagctt gacttttgc 120  
gaaaagaagg gaaaaagtag cgagtgggca aaaaaataca atgttgggga tgaagttgca 180  
agttacaatg cctatagaga aaaacttaag caggagcaca ggaaaaacgt tagcgaactt 240  
gtttctggtc ttcccgggtc tgtgaagaaa gtaaacgaac tcttgataa tgagaatcag 300  
actctaggc aactttactg tgcctcaaga aagcttggtg aacaaaatcc agtactctac 360  
cgtgtgtcag agtttgcata tttggtgtg agatttagac gtgaagattc ggatgagcaa 420  
gaacaacgag aaatgctgtc aactttacct ttcagcgaat ataatgaaga gcaggacctt 480  
ggtgaacaag acttccagta catctatggt tttgaatgtg caagattcat ctttcaaaat 540  
gggagaatgt ttggactcaa cacggataga agatat 576

<210> SEQ ID NO 5  
<211> LENGTH: 175  
<212> TYPE: PRT  
<213> ORGANISM: *Coronocyclus coronatus*

<400> SEQUENCE: 5

Ser Cys Val Ala Lys Trp Glu Tyr Phe Lys Ile Val Ile Asn Arg Ser  
1 5 10 15

Leu Thr Phe Ala Gln Arg Lys Glu Glu Ile Ser Lys Trp Ala Lys Lys  
20 25 30

Tyr Lys Val Glu Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys  
35 40 45

Leu Lys Gln Gln His Arg Lys Asn Val Ser Glu Leu Val Ser Asn Leu  
50 55 60

Pro Gly Ala Val Glu Arg Val Asn Lys Leu Leu Asp Asn Glu Asn Gln  
65 70 75 80

Thr Pro Lys Gln Leu Tyr Leu Ala Leu Arg Glu Leu Gly Lys Gln Asn  
85 90 95

Pro Ala Leu Tyr His Val Val Glu Tyr Val Asn Val Val Val Arg Leu  
100 105 110

Lys Arg Glu Glu Leu Asp Gln Gln Asp Gln Arg Arg Ala Leu Ser Gly  
115 120 125

Ser Leu Phe Gly Glu Asn Asn Asp Asn Leu Glu Glu Gln Asp Phe Gly  
130 135 140

Glu Glu Asp Phe Arg Tyr Val Tyr Gly Phe Glu Cys Ala Arg Phe Ile  
145 150 155 160

Leu Gln Asn Gly Arg Met Phe Gly Leu Asn Met Asp Arg Asn Tyr  
165 170 175

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<210> SEQ ID NO 6
<211> LENGTH: 525
<212> TYPE: DNA
<213> ORGANISM: Coronocyclus coronatus

<400> SEQUENCE: 6
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caaagaaagg aagaaattag caagtgggcg aaaaaataca aagttgagga tgaagttgca    120
agctacaatg cttatagaga aaaactcaag cagcagcaca ggaaaaacgt tagcgaactt    180
gtttctagtc ttcccgggtc aatggaaaga gtgaacaaac ttttgacaaa tgaaaaccag    240
accocctaagc aactttacct tgcocctacga gaacttgcca aacaaaatcc ggcactttac    300
catgtttgctg agtatgtcaa tgtggttgag agacttaaac gagaagaatt ggatgaacaa    360
gatcaatgaa gagcgctgtc gggttcactt tttggcgaga ataacgacaa tctagaagag    420
caggactttg gtgaagaaga ctttcgctat gtctatgggt ttgaatgtgc aagattcatc    480
cttcaaaatg gaagaatggt ttggtctaac atggatagga attat                    525

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<210> SEQ ID NO 7
<211> LENGTH: 199
<212> TYPE: PRT
<213> ORGANISM: Cyathostomum catinatum

<400> SEQUENCE: 7
Gly Glu Glu Asp Arg Glu Glu His Arg Arg His His Arg His Ser Leu
1          5          10          15
Leu Pro Pro Tyr Leu His Asn Val Ser Cys Val Ala Lys Trp Glu Tyr
20        25        30
Phe Arg Ile Val Gly Asn Arg Ser Leu Thr Phe Ala Glu Lys Lys Lys
35        40        45
Glu Ile Ser Glu Trp Ala Lys Lys Tyr Asn Val Leu Asp Glu Val Ala
50        55        60
Ser Tyr Asn Ala Tyr Arg Glu Lys Leu Lys Gln Glu His Arg Lys Asn
65        70        75        80
Val Ser Glu Leu Val Ser Asp Leu Pro Lys Ala Val Lys Lys Val Asn
85        90        95
Asp Leu Leu Asp Asn Glu Asn Gln Thr Pro Arg Gln Leu Tyr Val Ala
100       105       110
Leu Arg Glu Leu Gly Arg Gln Asn Pro Thr Leu Tyr Arg Ile Val Glu
115       120       125
Tyr Ile Asn Val Ala Val Arg Arg Arg Ser Glu Glu Leu Asp Glu Gln
130       135       140
Glu Gln Gly Arg Arg Leu Ser Ala Leu Pro Phe Gly Asp Asn Asn Asp
145       150       155       160
Asn Leu Glu Glu Gln Asp Phe Gly Glu Gln Asp Phe Arg Tyr Val Tyr
165       170       175
Gly Phe Glu Cys Ala Arg Phe Leu Leu Gln Asn Gly Arg Met Phe Gly
180       185       190
Leu Asn Thr Asp Glu Arg Asp
195

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<210> SEQ ID NO 8
<211> LENGTH: 591
<212> TYPE: DNA
<213> ORGANISM: Cyathostomum catinatum

<400> SEQUENCE: 8

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gaggatcgtg aagaacatcg ccgtcaccat cgtcattcac tcttgccacc atatctccac   60
aacgtgagct gtgtggccaa atgggaatac tttagaattg tggggaacag gagtttaacg   120
tttctgaga aaaagaaaga aattagcgag tgggcaaaaa aatacaatgt tctggatgaa   180
gtagcaagct acaatgctta tagggaaaaa ctcaagcagg agcacagaaa aaacgttagc   240
gaacttgttt ctgatcttcc caaggcagta aagaaagtca acgatcttct agacaatgaa   300
aatcagactc ctaggcaact ttatgttgcc cttagagagc ttggtagaca aaatccgaca   360
ctttaccgta ttgtcgagta catcaatgtg gctgtaaggc gaagaagtga agaactggat   420
gagcaagaac aaggaagaag gctgtcagct ttacctttcg gcgacaacaa cgataatttg   480
gaagagcagg acttcgggtga acaagacttt cgctatgtct acggctttga gtgtgcaaga   540
tttctccttc aaaatggaag aatgttcgga ctcaacacag atgaaagaga t           591

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<210> SEQ ID NO 9
<211> LENGTH: 841
<212> TYPE: DNA
<213> ORGANISM: Cylicocyclus ashworthi

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<400> SEQUENCE: 9

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atgaacaaaa cgtaacatt tctcacagtc gttagtgcgc tagttctggc ccaaggtgtc   60
atggaccttt ttggtgaaga gggctgtgaa gaacatcgcc gtcaccatcg tcattcactc   120
ttaccaccat atctccacaa cgtgagctgt gtggctaaat gggagtactt caaaattgta   180
gggaacagga gtttaacggt tgctgagaaa aaagaagaaa ttagccagtg ggcaaaaaaa   240
tacaatggtg tggtaaagctt ttctgaatta atgtaatac actcgcacatg tggccttttt   300
aggatgaagt tgcaagctac aatgcttaca gggagaaaact caagcaggag cacagaaaaa   360
acgttagcga acttgtttct gctcttccaa acgcagtaaa gaaagtcaac aatcttctag   420
acaatgaaaa tcgactcctt aggcaacttt acgttgccct tagagaactt ggtagacaaa   480
atccggcagt aagtagaaaag agctgcactc ctgggcttaa taaaacaaat tatttaagct   540
ttaccgtatt gtcagatata tcaatgtggc tgtaagacga agaagtgaag gactggatga   600
gcaagaacaa cgaagaagac tatcagcttt acctttcggc gacaacaacg ataatatgga   660
agagcaggac ttcggtgaac aagactttcg ctatgtctac ggctttgagt gtgcaagatt   720
tctccttcaa aatggaagaa tgtttgggt caacacagat gaaagagatt agcaaagaat   780
caattgtagt tcaaagcggg agagtttgag ctgcaaaact agcatgccat catcacctcc   840
t                                                                           841

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<210> SEQ ID NO 10
<211> LENGTH: 223
<212> TYPE: PRT
<213> ORGANISM: Cylicocyclus ashworthi

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<400> SEQUENCE: 10

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Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Ser Ala Val Val Leu
 1             5             10            15
Ala Gln Gly Val Met Asp Leu Phe Gly Glu Glu Gly Arg Glu Glu His
 20            25            30
Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val
 35            40            45
Ser Cys Val Ala Lys Trp Glu Tyr Phe Lys Ile Val Gly Asn Arg Ser
 50            55            60
Leu Thr Phe Ala Glu Lys Lys Glu Glu Ile Ser Gln Trp Ala Lys Lys
 65            70            75            80

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Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys  
 85 90 95  
 Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Ala Leu  
 100 105 110  
 Pro Asn Ala Val Lys Lys Val Asn Asn Leu Leu Asp Asn Glu Asn Gln  
 115 120 125  
 Thr Leu Arg Gln Leu Tyr Val Ala Leu Arg Glu Leu Gly Arg Gln Asn  
 130 135 140  
 Pro Ala Leu Tyr Arg Ile Val Glu Tyr Ile Asn Val Ala Val Arg Arg  
 145 150 155 160  
 Arg Ser Glu Gly Leu Asp Glu Gln Glu Gln Arg Arg Lys Leu Ser Ala  
 165 170 175  
 Leu Pro Phe Gly Asp Asn Asn Asp Asn Met Glu Glu Gln Asp Phe Gly  
 180 185 190  
 Glu Gln Asp Phe Arg Tyr Val Tyr Gly Phe Glu Cys Ala Arg Phe Leu  
 195 200 205  
 Leu Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Glu Arg Asp  
 210 215 220

<210> SEQ ID NO 11  
 <211> LENGTH: 841  
 <212> TYPE: DNA  
 <213> ORGANISM: Cyathostomum catinatum

<400> SEQUENCE: 11

atgaacaaaa cgtaaactt tctcacagtc gttagtgccg tagtctggc tcaaggtgtc 60  
 atggaccttt ttgggtaaga aggcctgtaa gaacatcgcc gtcaccgtcg tcattcactc 120  
 ttgccaccat atctccacaa cgtgagctgt gtggctaaat gggaaactt cagaattgtg 180  
 gggaaacagga gtttgacgtt tgctgagaaa aaggaagaga ttagcgagtg ggcaaaaaag 240  
 tacaatggtt tgtaagctt ttctgaattg atgtaaatac actcgcagtc tggcctttt 300  
 aggatgaagt tgcaagctac aatgcttaca gggaaaaact caagcaggag cacagaaaa 360  
 acgtagcga acttgtttct gctcttccaa acgcagtaaa gaaagtcaac gatcttctag 420  
 acaatgaaaa tcgactcct aggcaacttt acgttgccct tagagaactt ggtagacaaa 480  
 atccggcagt aagtcgaaag agctgcactc ttgggcataa gtaaaaaaaa gtatttttagc 540  
 tttaccgtat tgtggagtac atcaatgtgg ctgtaagact aagaagtga gaagtggatg 600  
 agcaagaaca acgaagaagg ctatcagctt taccttttgg tgaccataac gataaatatgg 660  
 aagagcagga ctttggtgat caagactttc gctatgtcta cggctttgag tgtgcaagat 720  
 ttctccttca aaatggaaga atgtttggac ttaacacaga tgaaagatat tagtaaaaa 780  
 taactgtagc tcaaagcggg agagtttgag ctgcaaaact agcatgcat catcacctcc 840  
 t 841

<210> SEQ ID NO 12  
 <211> LENGTH: 223  
 <212> TYPE: PRT  
 <213> ORGANISM: Cyathostomum catinatum

<400> SEQUENCE: 12

Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Ser Ala Val Val Leu  
 1 5 10 15  
 Ala Gln Gly Val Met Asp Leu Phe Gly Glu Glu Gly Arg Glu Glu His  
 20 25 30

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Arg Arg His Arg Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val  
 35 40 45

Ser Cys Val Ala Lys Trp Glu Tyr Phe Arg Ile Val Gly Asn Arg Ser  
 50 55 60

Leu Thr Phe Ala Glu Lys Lys Glu Glu Ile Ser Glu Trp Ala Lys Lys  
 65 70 75 80

Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys  
 85 90 95

Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Ala Leu  
 100 105 110

Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln  
 115 120 125

Thr Pro Arg Gln Leu Tyr Val Ala Leu Arg Glu Leu Gly Arg Gln Asn  
 130 135 140

Pro Ala Leu Tyr Arg Ile Val Glu Tyr Ile Asn Val Ala Val Arg Leu  
 145 150 155 160

Arg Ser Glu Glu Val Asp Glu Gln Glu Gln Arg Arg Arg Leu Ser Ala  
 165 170 175

Leu Pro Phe Gly Asp His Asn Asp Asn Met Glu Glu Gln Asp Phe Gly  
 180 185 190

Asp Gln Asp Phe Arg Tyr Val Tyr Gly Phe Glu Cys Ala Arg Phe Leu  
 195 200 205

Leu Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Glu Arg Tyr  
 210 215 220

<210> SEQ ID NO 13  
 <211> LENGTH: 769  
 <212> TYPE: DNA  
 <213> ORGANISM: *Cylicostephanus goldi*

<400> SEQUENCE: 13

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atgaacaaaa cgtaaaccatt tctcacagtc gttagtgcgc tagtctctggc tcaaggtgtc      60
gtggaccttt ttggtgaaga gggctcgtgaa gaacatcgcc gtcaccatcg tcattcactc      120
ttaccaccat atctccacaa cgctcagctgt gtggctaaat ggggaatactt caaaattgtg      180
gggaatagga gtttgacatt tgctgagaaa aagaaagaaa ttagcgcgagtg ggctaaaaaa      240
tacaatgtag tggtaagctt ttttgacttg atgtaaatgc actcgtatgc cggccctttt      300
aggatgaagt tgcaaggtac aatgcttata gagaaaaact taagcaggaa cacaggaaaa      360
acgtcagcga acttgtttct gatcttccca acgcagtaaa gaaagtgaat gatctcctgg      420
acaatgagaa tcaaaactct aggcaacttt acattgcctt cagagaactt ggtagacaaa      480
atccagaagt aagttgaaaag tgctgcaatt ttaggcttag ataaaacagt tgtttaagct      540
ttaccgtggt gtgcagttta tcaatgtggc tgtaagaata agacgtgaag atttgatga      600
gcaagaacaa cgaacaaggc tgtcaacttt accttttggc gacaacaacg acaatttcga      660
agagcaagac ttcggatgaa aagactttcg ctatgtctat ggctttgagt gtgcaagatt      720
tctccttcaa aatggaagaa tgtttggact taacacggat agaagatac      769
    
```

<210> SEQ ID NO 14  
 <211> LENGTH: 223  
 <212> TYPE: PRT  
 <213> ORGANISM: *Cylicostephanus goldi*

<400> SEQUENCE: 14

Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Ser Ala Val Val Leu  
 1 5 10 15

-continued

Ala Gln Gly Val Val Asp Leu Phe Gly Glu Glu Gly Arg Glu Glu His  
 20 25 30  
 Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val  
 35 40 45  
 Ser Cys Val Ala Lys Trp Glu Tyr Phe Lys Ile Val Gly Asn Arg Ser  
 50 55 60  
 Leu Thr Phe Ala Glu Lys Lys Lys Glu Ile Ser Glu Trp Ala Lys Lys  
 65 70 75 80  
 Tyr Asn Val Val Asp Glu Val Ala Arg Tyr Asn Ala Tyr Arg Glu Lys  
 85 90 95  
 Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Asp Leu  
 100 105 110  
 Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln  
 115 120 125  
 Thr Pro Arg Gln Leu Tyr Ile Ala Leu Arg Glu Leu Gly Arg Gln Asn  
 130 135 140  
 Pro Glu Leu Tyr Arg Val Val Glu Phe Ile Asn Val Ala Val Arg Ile  
 145 150 155 160  
 Arg Arg Glu Asp Leu Asp Glu Gln Glu Gln Arg Thr Arg Leu Ser Thr  
 165 170 175  
 Leu Pro Phe Gly Asp Asn Asn Asp Asn Phe Glu Glu Gln Asp Phe Gly  
 180 185 190  
 Glu Gln Asp Phe Arg Tyr Val Tyr Gly Phe Glu Cys Ala Arg Phe Leu  
 195 200 205  
 Leu Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr  
 210 215 220

<210> SEQ ID NO 15  
 <211> LENGTH: 847  
 <212> TYPE: DNA  
 <213> ORGANISM: Cylicostephanus goldi

<400> SEQUENCE: 15

atgaacaaaa cgtaacatt tctcacagtc gttagtgcg tagtctggc ccaaggtgtc 60  
 atggaccttc ttgatgaaga ggctcgtgga gagcatcgcc gtcaccatcg tcattcactc 120  
 ttaccaccat atctccacaa cgtgagctgt gtggctaaat ggaataactt caaaattgtg 180  
 gggaacagga gtttgacgtt tgctgagaaa aagaaagaaa ttagcgagtg ggcaaaaaaa 240  
 tacaacgttg tgtaagctt ttgtgactcg atgtagatac cccagatatt ctagataccc 300  
 atgctggcct ttttaggatg aagttgcaag ctacaatgct tatagagaaa aactcaagca 360  
 ggaacacagg aaaaacgta gcgaacttgt atctgatctt cccaatgcag tgaagaaagt 420  
 gaatgatctc ctggacaatg agaatcaaac tctaggcaa ctttactgtg cctcagaga 480  
 acttggtaga caaaatccag cagtaagttg aaagtgtgac aatttcaggc ttagataaaa 540  
 cagttgttta agctttaccg tgtgtgctgag ctcatcaatg tggtgtgtaag attaagactg 600  
 gaagatttgg atgagcaaga acaacgaaca aggctgtcaa ccttacctt tggcgacaac 660  
 aacaacaatt tcgatgagca ggacttcggt gaacaagact ttcgctatgt ctatggcttt 720  
 gagtgtgcaa gatttctct tcaaaatgga agaattgttg gacttaacac ggatagaaga 780  
 tactagtaag agtcaactgt agctcaaagt ggttcgagct acgaacagca tgccatcatc 840  
 acctcct 847

<210> SEQ ID NO 16

-continued

&lt;211&gt; LENGTH: 223

&lt;212&gt; TYPE: PRT

<213> ORGANISM: *Cylicostephanus goldi*

&lt;400&gt; SEQUENCE: 16

Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Ser Ala Val Val Leu  
 1 5 10 15

Ala Gln Gly Val Met Asp Leu Leu Asp Glu Glu Ala Arg Gly Glu His  
 20 25 30

Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val  
 35 40 45

Ser Cys Val Ala Lys Trp Glu Tyr Phe Lys Ile Val Gly Asn Arg Ser  
 50 55 60

Leu Thr Phe Ala Glu Lys Lys Lys Glu Ile Ser Glu Trp Ala Lys Lys  
 65 70 75 80

Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys  
 85 90 95

Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Asp Leu  
 100 105 110

Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln  
 115 120 125

Thr Pro Arg Gln Leu Tyr Val Ala Leu Arg Glu Leu Gly Arg Gln Asn  
 130 135 140

Pro Ala Leu Tyr Arg Val Val Glu Leu Ile Asn Val Ala Val Arg Leu  
 145 150 155 160

Arg Arg Glu Asp Leu Asp Glu Gln Glu Gln Arg Thr Arg Leu Ser Thr  
 165 170 175

Leu Pro Phe Gly Asp Asn Asn Asn Asn Phe Asp Glu Gln Asp Phe Gly  
 180 185 190

Glu Gln Asp Phe Arg Tyr Val Tyr Gly Phe Glu Cys Ala Arg Phe Leu  
 195 200 205

Leu Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr  
 210 215 220

&lt;210&gt; SEQ ID NO 17

&lt;211&gt; LENGTH: 835

&lt;212&gt; TYPE: DNA

<213> ORGANISM: *Cylicostephanus goldi*

&lt;400&gt; SEQUENCE: 17

atgaacaaaa cgtaacatt tctcacagtc gttagtgccg ttgtcctggc gcaaggtgtc 60

atggccctat ttggtaaga gagtcgtgaa gaacaccgcc gtcaccatcg tcattcactc 120

ttaccaccat atctccacaa cgtgagctgt gtggctaaat gggagtactt caaaaattgtg 180

gggaacagga gtttgacgtt tgctgagaaa aagaaagaaa tcagcgagtg ggctaaaaaa 240

tacaatggtg tgtaagctt ttttgacttg atgtaaatgc actcgcagtc cggcctttat 300

aggatgaagt tgcaagctac aatgcttata gagaaaaact caagcaggaa cacaggaaaa 360

acgttagcga acttgtttct gatcttccca acgcagtaaa gaaagtcagc gatcttttgg 420

acaacgaaaa tcagacttct aggcaacttt atgttgcact cagagaactt ggtagacaaa 480

atccggcagt aagttgaaga ggctccaatt ttgggctcaa gcaaaaataa ttatttttagc 540

tataccgtgt cgtcgagtat atcaatgtgg ctgtgagatt aagacgaaaa gaacaggatg 600

aacaagaacg acaaggaacg ctgtcagctc taccttttgg cgagaataac gacaatttgg 660

aagagcagga ctttggtgaa caagacttcc gctatgtcta tggttttgag tgtgcaagat 720

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 ttctccttca aatggaaga atgtttggac tcaacacgga tagaagatac cagtaagagt 780

caactgtagc tcaaagtggg tttgagctac gaacagcatg ccatcatcac ctctt 835

&lt;210&gt; SEQ ID NO 18

&lt;211&gt; LENGTH: 223

&lt;212&gt; TYPE: PRT

<213> ORGANISM: *Cylicostephanus goldi*

&lt;400&gt; SEQUENCE: 18

 Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Ser Ala Val Val Leu  
 1 5 10 15

 Ala Gln Gly Val Met Ala Leu Phe Gly Glu Glu Ser Arg Glu Glu His  
 20 25 30

 Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val  
 35 40 45

 Ser Cys Val Ala Lys Trp Glu Tyr Phe Lys Ile Val Gly Asn Arg Ser  
 50 55 60

 Leu Thr Phe Ala Glu Lys Lys Lys Glu Ile Ser Glu Trp Ala Lys Lys  
 65 70 75 80

 Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys  
 85 90 95

 Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Asp Leu  
 100 105 110

 Pro Asn Ala Val Lys Lys Val Ser Asp Leu Leu Asp Asn Glu Asn Gln  
 115 120 125

 Thr Ser Arg Gln Leu Tyr Val Ala Leu Arg Glu Leu Gly Arg Gln Asn  
 130 135 140

 Pro Ala Val Tyr Arg Val Val Glu Tyr Ile Asn Val Ala Val Arg Leu  
 145 150 155 160

 Arg Arg Lys Glu Gln Asp Glu Gln Glu Arg Gln Gly Thr Leu Ser Ala  
 165 170 175

 Leu Pro Phe Gly Glu Asn Asn Asp Asn Leu Glu Glu Gln Asp Phe Gly  
 180 185 190

 Glu Gln Asp Phe Arg Tyr Val Tyr Gly Phe Glu Cys Ala Arg Phe Leu  
 195 200 205

 Leu Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr  
 210 215 220

&lt;210&gt; SEQ ID NO 19

&lt;211&gt; LENGTH: 830

&lt;212&gt; TYPE: DNA

<213> ORGANISM: *Cylicostephanus longibursatus*

&lt;400&gt; SEQUENCE: 19

atgaacaaaa cgtaacatt tctcaccgtc gtctatgccc tagtoctggc ccaaggtgtc 60

atggaccttt ttggtgaaga gggctgtgaa gaacatcgcc gtcaccatcg tcattcactc 120

ttaccacat atctccacaa tgtgagctgt gtggctaaat ggaataactt caaaattgtg 180

gggaacagga gtttgacggt ttgctgagaaa aaggaagaaa ttagcaagtg ggcaaaaaa 240

tacaatggtt tggtacgctt ttgtaacccc gtataatata ctctgcata ctggccggtt 300

caggatgaag ttgcaagcta cagtgcctgc agggaaaagc ttaagcagga acacaggaaa 360

aacgttagcg aaattgtttc taatcttccc aatgcagtga agaaagtaaa cgatcttttg 420

gacaatgaaa atcagacccc caggcaactt tacgttgctt tcagaaaact tggtaaaaa 480

aatccggcag taagttgaaa gagctgcaat tttgggtttg aggagaaaaa actattttag 540

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ctttatcgtg ttgtcgagta tatcaatggtg cttgtgagac taagacgtga agaatttgat 600
gaagatcagc gaagatcgct gtcagcttta ccttttgccg acaataacga cgatttgaa 660
gagcaggact ttggtaaca ggactttogc tatatctatg gctttgagtg tgcaagattt 720
atccttcaaa atggaagaat gttcggactc aacacggata gaagatatta gtaagagtca 780
actgtagctc gagggtttga gctacgaact gcatgccatc atcacctect 830

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<210> SEQ ID NO 20
<211> LENGTH: 222
<212> TYPE: PRT
<213> ORGANISM: Cylicostephanus longibursatus

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<400> SEQUENCE: 20

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```

Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Tyr Ala Val Val Leu
1          5          10          15
Ala Gln Gly Val Met Asp Leu Phe Gly Glu Glu Gly Arg Glu Glu His
          20          25          30
Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val
          35          40          45
Ser Cys Val Ala Lys Trp Glu Tyr Phe Lys Ile Val Gly Asn Arg Ser
          50          55          60
Leu Thr Phe Ala Glu Lys Lys Glu Glu Ile Ser Lys Trp Ala Lys Lys
65          70          75          80
Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Ser Ala Cys Arg Glu Lys
          85          90          95
Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Ile Val Ser Asn Leu
          100          105          110
Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln
          115          120          125
Thr Pro Arg Gln Leu Tyr Val Ala Phe Arg Lys Leu Gly Lys Gln Asn
          130          135          140
Pro Ala Leu Tyr Arg Val Val Glu Tyr Ile Asn Val Leu Val Arg Leu
          145          150          155          160
Arg Arg Glu Glu Phe Asp Glu Asp Gln Arg Arg Ser Leu Ser Ala Leu
          165          170          175
Pro Phe Gly Asp Asn Asn Asp Asp Leu Glu Glu Gln Asp Phe Gly Glu
          180          185          190
Gln Asp Phe Arg Tyr Ile Tyr Gly Phe Glu Cys Ala Arg Phe Ile Leu
          195          200          205
Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr
          210          215          220

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<210> SEQ ID NO 21
<211> LENGTH: 829
<212> TYPE: DNA
<213> ORGANISM: Cylicocyclus insignis

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<400> SEQUENCE: 21

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```

atgaacaaaa cgtaaacatt tctcaccgtc gtctgtgccg tagtctctggc ccaagtggtc 60
atggaccttt ttgggtgaaga aggtcgtgaa gaacatcgcc gtcaccatcg tcattcactc 120
ttaccaccat atctccacaa tgtgagctgt gtggctaaat gggaaactt caaaattctg 180
gggaacagaa gttttgagctt tgctgagaaa aaggaaaaaa tcagcgagtg ggcaaaaaag 240
tacaatggtg tggtagctt ttgtaactcc gtataatata cctcgcgatg ctggccggtt 300
caggatgaag ttgcaagcta caatgcttgc agggaaaaagc ttaagcagga acacaggaaa 360

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aacgttagcg aaattgttcc taatcttccc aatgcagtaa agaaagtaaa cgatcttttg 420
gacaatgaaa atcagactcc caggcaactt tacgttgccc tcagaaaact cgtaaacaa 480
aatccgccag taagttgaaa gactgcaact ttgggtttaa gggaaaaaaaa ctatttttagc 540
tttaccgcgt tgctgagtat atcaatgtgg ttgtgagact aagacgtgaa gaatctgatg 600
aagaacaacg aagaacgctg tcagctttac cttttggcga caataacgac aacttgggaag 660
agcaagactt tgggtgaagaa gactttcget atatttatgg ctttgagtgt gcaagattta 720
tccttcaaaa tgggagaatg ttcggactca acacggatag aagatatcag taagagtcaa 780
ctgtagctta aaagtttgag ctacgaacag catgccatca tcacctcct 829

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<210> SEQ ID NO 22
<211> LENGTH: 222
<212> TYPE: PRT
<213> ORGANISM: Cylicocyclus insigne

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<400> SEQUENCE: 22

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```

Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Cys Ala Val Val Leu
1          5          10          15
Ala Gln Gly Val Met Asp Leu Phe Gly Glu Glu Gly Arg Glu Glu His
20          25          30
Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val
35          40          45
Ser Cys Val Ala Lys Trp Glu Tyr Phe Lys Ile Leu Gly Asn Arg Ser
50          55          60
Leu Thr Phe Ala Glu Lys Lys Glu Lys Ile Ser Glu Trp Ala Lys Lys
65          70          75          80
Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Cys Arg Glu Lys
85          90          95
Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Ile Val Ser Asn Leu
100         105         110
Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln
115         120         125
Thr Pro Arg Gln Leu Tyr Val Ala Leu Arg Lys Leu Gly Lys Gln Asn
130         135         140
Pro Pro Leu Tyr Arg Val Val Glu Tyr Ile Asn Val Val Val Arg Leu
145         150         155         160
Arg Arg Glu Glu Ser Asp Glu Glu Gln Arg Arg Thr Leu Ser Ala Leu
165         170         175
Pro Phe Gly Asp Asn Asn Asp Asn Leu Glu Glu Gln Asp Phe Gly Glu
180         185         190
Glu Asp Phe Arg Tyr Ile Tyr Gly Phe Glu Cys Ala Arg Phe Ile Leu
195         200         205
Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr
210         215         220

```

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<210> SEQ ID NO 23
<211> LENGTH: 830
<212> TYPE: DNA
<213> ORGANISM: Cylicostephanus longibursatus

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<400> SEQUENCE: 23

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```

atgaacaaaa cgtaaacatt tctcaccgtc gtctatgccg tagtctctggc ccaagggtgc 60
atggaccttt ttggtgaaga gggctctttaa gaacatcgcc gtcaccatcg tcattcactc 120
ttaccacat atctccacaa tgtgagctgt gtggctaaat gggaaactt caaaattctg 180

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gggaacagga gtttgacgtt tgctgagaaa aaggaaaaaa tcagcgagtg ggcaaaaaag 240
tacaatggtg tggtagcgtt ttgtaactca gtataatata tctcgcata ctggccggtt 300
caggatgaag ttgcaagcta caatgcttgc agggaaaagc ttaagcagga acacaggaaa 360
aacgttagcg aaattgttcc taatcttccc aatgcagtga agaaagtaaa cgatcttttg 420
gacaaatgaaa atcagacccc caggcaactt tacgttgccc tcagaaaact tggtaaacaa 480
aatccggcag taagttgaaa gagctgcaat tttgggtttg aggaaaaaaa actattttag 540
ctttatcgtg ttgtcgagta tatcaatgtg cttgtgagac taagacgtga agaatttgat 600
gaagatcagc gaagatcgtc gtcagcttta ccttttggcg acaataacga cgatttgaa 660
gagcaggact ttggatgaaca ggactttogc tatatctatg gctttgagtg tgcaagattt 720
atccttcaaa atggaagaat gttcggactc aacacggata gaagatatta gtaagagtca 780
actgtagctc aagggtttga gctacgaact gcatgccatc atcacctctc 830

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&lt;210&gt; SEQ ID NO 24

&lt;211&gt; LENGTH: 222

&lt;212&gt; TYPE: PRT

<213> ORGANISM: *Cylicostephanus longibursatus*

&lt;400&gt; SEQUENCE: 24

```

Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Tyr Ala Val Val Leu
1           5           10          15
Ala Gln Gly Val Met Asp Leu Phe Gly Glu Glu Gly Leu Glu Glu His
                20           25           30
Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val
            35           40           45
Ser Cys Val Ala Lys Trp Glu Tyr Phe Lys Ile Leu Gly Asn Arg Ser
            50           55           60
Leu Thr Phe Ala Glu Lys Lys Glu Lys Ile Ser Glu Trp Ala Lys Lys
65           70           75           80
Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Cys Arg Glu Lys
            85           90           95
Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Ile Val Ser Asn Leu
            100          105          110
Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln
            115          120          125
Thr Pro Arg Gln Leu Tyr Val Ala Leu Arg Lys Leu Gly Lys Gln Asn
            130          135          140
Pro Ala Leu Tyr Arg Val Val Glu Tyr Ile Asn Val Leu Val Arg Leu
            145          150          155          160
Arg Arg Glu Glu Phe Asp Glu Asp Gln Arg Arg Ser Leu Ser Ala Leu
            165          170          175
Pro Phe Gly Asp Asn Asn Asp Asp Leu Glu Glu Gln Asp Phe Gly Glu
            180          185          190
Gln Asp Phe Arg Tyr Ile Tyr Gly Phe Glu Cys Ala Arg Phe Ile Leu
            195          200          205
Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr
            210          215          220

```

&lt;210&gt; SEQ ID NO 25

&lt;211&gt; LENGTH: 832

&lt;212&gt; TYPE: DNA

<213> ORGANISM: *Cylicocyclus nassatus*

&lt;400&gt; SEQUENCE: 25

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atgaacaaaa cgtaacatt tctcatcgtc gttagtgccg tagtctgac ccaaagtgtt   60
atggactttt tcgatgaaga cggctcgtgaa gaacatcgcc gtcacatcgc tcattccctt   120
ttaccaccgt atctccacaa tatgagctgc gtggcctaat gggataactt cgagattgtg   180
ggggacagga gtctgacgtt tgctgaaaag aaggaaaaaa tcggcgcgagtg ggctaaaaaa   240
tacaatgttg tggaagatt ttgtaactct atgtaaatg acccccgtag gtcgcctctg   300
ttaggatgaa gttgcaagct acaatgctta tagagaaaaa ctaaagcagg agcacaggaa   360
aaacgttagc gagcttctct ctggctctcc caatgctgtg aagaaaataa acgaactttt   420
agacaatgaa aatcagactg ttaggcaact ttatgttgct ttaagagaac ttggtaaaca   480
aaatccagca gtaagttaaa agaagtgcaa ttttgggctt aactaatgag acaattttag   540
ctctaccgtg ttgtcgagta tatcaatgtg gttgtgagac ttagacgtga agatttggat   600
gagcaggaac aacagagaac gctgtcaacc ccacctttcg gcgagaataa cgaagagcaa   660
gactttggtg aacaagactt tcaatatatc tatggttttg agtgtgccag attcactcct   720
caaaatggaa gaatgtttgg acttaacacg gatagaagat attagtaaga gttaactgca   780
gctcaatgtg atagagattg agccacaacc caacatgcca tcatcacctc ct         832

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&lt;210&gt; SEQ ID NO 26

&lt;211&gt; LENGTH: 220

&lt;212&gt; TYPE: PRT

<213> ORGANISM: *Cylicocyclus nassatus*

&lt;400&gt; SEQUENCE: 26

```

Met Asn Lys Thr Leu Thr Phe Leu Ile Val Val Ser Ala Val Val Leu
1           5           10          15
Thr Gln Ser Val Met Asp Phe Phe Asp Glu Asp Gly Arg Glu Glu His
          20          25          30
Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Met
          35          40          45
Ser Cys Val Ala Lys Trp Glu Tyr Phe Glu Ile Val Gly Asp Arg Ser
          50          55          60
Leu Thr Phe Ala Glu Lys Lys Glu Lys Ile Gly Glu Trp Ala Lys Lys
65          70          75          80
Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
          85          90          95
Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Gly Leu
          100         105         110
Pro Asn Ala Val Lys Lys Ile Asn Glu Leu Leu Asp Asn Glu Asn Gln
          115         120         125
Thr Val Arg Gln Leu Tyr Val Ala Leu Arg Glu Leu Gly Lys Gln Asn
          130         135         140
Pro Ala Leu Tyr Arg Val Val Glu Tyr Ile Asn Val Val Val Arg Leu
145         150         155         160
Arg Arg Glu Asp Leu Asp Glu Gln Glu Gln Gln Arg Thr Leu Ser Thr
          165         170         175
Pro Pro Phe Gly Glu Asn Asn Glu Glu Gln Asp Phe Gly Glu Gln Asp
          180         185         190
Phe His Tyr Ile Tyr Gly Phe Glu Cys Ala Arg Phe Ile Leu Gln Asn
          195         200         205
Gly Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr
          210         215         220

```

&lt;210&gt; SEQ ID NO 27

-continued

&lt;211&gt; LENGTH: 762

&lt;212&gt; TYPE: DNA

<213> ORGANISM: *Cylicocyclus nassatus*

&lt;400&gt; SEQUENCE: 27

```

atgaacaaaa cgtaacatt tctcatcgtc gttagtgcc tagtcctggc ccaaagtgtt      60
atggactttt tcgatgaaga aggtcgtgag ggacatcgcc gtcacatcgc tcattcactt    120
ttaccacccat atctccacaa tatgagctgc gtggcctaat gggaaatactt cgagattgtg    180
ggggacagga gctcgtcgtt tgctgaaaag aaggaaaaaa tcggcgagtg ggctaaaaaa    240
tacaatgttg tgtaagatt ttgtaactcc atgttaggat acctccgcac gtcgccctgt    300
ttaggatgaa gttgcaagct acaatgctta tagagaaaaa ctaaagcagg agcacaggaa    360
aaacgttagc gagcttgtct ctggtcttcc caatgctgtg aagaaagtaa acgaactttt    420
agacaatgaa aatcagactg ttaggcaact ttatgttgct ttaagagaac ttggtaaaca    480
aatccagca gtaagttaa agaagtacaa ttttgagctc aactaatgag acaattttag    540
ctctaccgtg ttgtcagata tatcaatgtg gttgtgagac ttagacgtga agattcggat    600
gagcaggaac aacgaagaac tctgtcaacc tcacctttcg gcgagaataa cgaagagcaa    660
gattttggtg aacaagattt tcaatatatc tatggttttg agtgtgcaag attcatcctt    720
caaaatggaa gaatgtttgg actcaatcag gatagaagat at                          762

```

&lt;210&gt; SEQ ID NO 28

&lt;211&gt; LENGTH: 220

&lt;212&gt; TYPE: PRT

<213> ORGANISM: *Cylicocyclus nassatus*

&lt;400&gt; SEQUENCE: 28

```

Met Asn Lys Thr Leu Thr Phe Leu Ile Val Val Ser Ala Ile Val Leu
 1             5             10             15
Ala Gln Ser Val Met Asp Phe Phe Asp Glu Glu Gly Arg Glu Gly His
 20             25             30
Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Met
 35             40             45
Ser Cys Val Ala Lys Trp Glu Tyr Phe Glu Ile Val Gly Asp Arg Ser
 50             55             60
Leu Thr Phe Ala Glu Lys Lys Glu Lys Ile Gly Glu Trp Ala Lys Lys
 65             70             75             80
Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
 85             90             95
Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Gly Leu
100            105            110
Pro Asn Ala Val Lys Lys Val Asn Glu Leu Leu Asp Asn Glu Asn Gln
115            120            125
Thr Val Arg Gln Leu Tyr Val Ala Leu Arg Glu Leu Gly Lys Gln Asn
130            135            140
Pro Ala Leu Tyr Arg Val Val Glu Tyr Ile Asn Val Val Val Arg Leu
145            150            155            160
Arg Arg Glu Asp Ser Asp Glu Gln Glu Gln Arg Arg Thr Leu Ser Thr
165            170            175
Ser Pro Phe Gly Glu Asn Asn Glu Glu Gln Asp Phe Gly Glu Gln Asp
180            185            190
Phe His Tyr Ile Tyr Gly Phe Glu Cys Ala Arg Phe Ile Leu Gln Asn
195            200            205
Gly Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr

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210	215	220	
<210> SEQ ID NO 29			
<211> LENGTH: 829			
<212> TYPE: DNA			
<213> ORGANISM: <i>Cylicocyclus nassatus</i>			
<400> SEQUENCE: 29			
atgaacaaaa	cgtaacatt	tctcatcgcc	gtagtgcca tagtctggc ccaaagtatg 60
gactttttcg	atgaagacgg	tcgtgaagaa	catcgccgtc atcatcgta ttcactttta 120
ccaccatata	cccacaatat	gagctgcgcg	gccaaatggg aatacttga gattgtaggg 180
gacaggagtc	tgacgtttgc	tgaaaagaag	gaaaaaatcg gcgagtgggc taaaaaatac 240
aatgttggtg	taagattttg	taactccatg	taaagatacc cctccatgtc gtcccgttta 300
ggatgaagtt	gcaagctaca	atgcttgtag	agaaaaactg aagcaagagc acaggaaaaa 360
cgtcagcgag	cttgctctctg	gtcttcccaa	tgctgtgaag aaagtaaagc aacttttaga 420
caatgaaaat	cagactgtta	ggcaacttta	tgttgcttta agagaacttg gtaaacaaaa 480
tccagcagta	agttgaaaga	agtgcatttt	gggcttaact aacgagacaa ttttagctct 540
accgtgttgt	cgagtatata	aatgtggctg	tgagacttag acgtgaagat tcggatgagc 600
aggaaaaaac	gaaagcgtg	tcaacctcac	ctttcggcga gaataacgaa gagcaggacc 660
ttggtgaaca	agattttcac	tatatctatg	gctttgagtg tgcaagattc atccttcaaa 720
atggaagaat	gtttggactt	aacacggata	gaagatatta gtaaaatttg actgcagctc 780
aaagtggtag	agattgagct	accaacccaa	catgccatca tcaactcct 829

<210> SEQ ID NO 30															
<211> LENGTH: 219															
<212> TYPE: PRT															
<213> ORGANISM: <i>Cylicocyclus nassatus</i>															
<400> SEQUENCE: 30															
Met	Asn	Lys	Thr	Leu	Thr	Phe	Leu	Ile	Ala	Val	Ser	Ala	Ile	Val	Leu
1			5						10					15	
Ala	Gln	Ser	Met	Asp	Phe	Phe	Asp	Glu	Asp	Gly	Arg	Glu	Glu	His	Arg
			20					25						30	
Arg	His	His	Arg	His	Ser	Leu	Leu	Pro	Pro	Tyr	Leu	His	Asn	Met	Ser
			35				40						45		
Cys	Ala	Ala	Lys	Trp	Glu	Tyr	Phe	Glu	Ile	Val	Gly	Asp	Arg	Ser	Leu
			50				55						60		
Thr	Phe	Ala	Glu	Lys	Lys	Glu	Lys	Ile	Gly	Glu	Trp	Ala	Lys	Lys	Tyr
65					70					75					80
Asn	Val	Val	Asp	Glu	Val	Ala	Ser	Tyr	Asn	Ala	Cys	Arg	Glu	Lys	Leu
				85					90					95	
Lys	Gln	Glu	His	Arg	Lys	Asn	Val	Ser	Glu	Leu	Val	Ser	Gly	Leu	Pro
			100						105					110	
Asn	Ala	Val	Lys	Lys	Val	Asn	Glu	Leu	Leu	Asp	Asn	Glu	Asn	Gln	Thr
			115					120						125	
Val	Arg	Gln	Leu	Tyr	Val	Ala	Leu	Arg	Glu	Leu	Gly	Lys	Gln	Asn	Pro
			130				135							140	
Ala	Leu	Tyr	Arg	Val	Val	Glu	Tyr	Ile	Asn	Val	Ala	Val	Arg	Leu	Arg
145					150						155				160
Arg	Glu	Asp	Ser	Asp	Glu	Gln	Glu	Lys	Arg	Arg	Thr	Leu	Ser	Thr	Ser
				165					170						175
Pro	Phe	Gly	Glu	Asn	Asn	Glu	Glu	Gln	Asp	Leu	Gly	Glu	Gln	Asp	Phe

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180	185	190	
His Tyr Ile Tyr Gly Phe Glu Cys Ala Arg Phe Ile Leu Gln Asn Gly			
195	200	205	
Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr			
210	215		

<210> SEQ ID NO 31  
 <211> LENGTH: 840  
 <212> TYPE: DNA  
 <213> ORGANISM: Cyathostomum pateratum

<400> SEQUENCE: 31

atgaacaaaa cgtaaactt tctcacagtc gttagtgccg tagttctggc ccaaggtgtc	60
atggaccttt ttggtgaaga gggctgtgaa gaacatcgtc gtcaccatcg tcattcactc	120
ttaccacccat atctccacaa tgtgagctgt gaggctaaat gggagtactt caaaattgtg	180
gggaacagga gtttgacgtt tgctgagaaa aaggagaaaa ttagcgagtg ggcaaaaaaa	240
tacaatgttg tgtaagctt ttttgaattg atgtaaattc actcgcatgc tggccttttt	300
aggatgaagt tgcaagctac aatgcttaca gggaaaaact caagcaggag cacagaaaaa	360
acgttagcga acttgtttct gctcttccaa acgcagtaaa gaaagtcaac gatcttctag	420
acaatgaaaa tcgactctt aggcaacttt acgttgccct tagaaaaact ggtagacaaa	480
atccggcagt aagtcgaaag agctgctgcc ttggacttaa gcgaaaaat tatttcagct	540
ttaccgtatt gtgcagtaca ttaatgtggc tgtaagacta agaagtgaag aagtggatga	600
gcaagaacaa cgaagaaggc tgtcagctct accttttggc gaccataacg ataatttggg	660
agagcaggac ttccgtgaaac aagactttcg ctatgtctat ggctttgagt gtgcaagatt	720
tctccttcaa aatggaagaa tgttcggact caacacggat ggaagatatt agtaagaaa	780
aagtgtagct caaagtggta gagtttgagc tacgaactca acatgccatc atcacctcct	840

<210> SEQ ID NO 32  
 <211> LENGTH: 223  
 <212> TYPE: PRT  
 <213> ORGANISM: Cyathostomum pateratum

<400> SEQUENCE: 32

Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Ser Ala Val Val Leu		
1	5	10
Ala Gln Gly Val Met Asp Leu Phe Gly Glu Glu Gly Arg Glu Glu His		
20	25	30
Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val		
35	40	45
Ser Cys Glu Ala Lys Trp Glu Tyr Phe Lys Ile Val Gly Asn Arg Ser		
50	55	60
Leu Thr Phe Ala Glu Lys Lys Glu Lys Ile Ser Glu Trp Ala Lys Lys		
65	70	75
Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys		
85	90	95
Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Ala Leu		
100	105	110
Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln		
115	120	125
Thr Leu Arg Gln Leu Tyr Val Ala Leu Arg Lys Leu Gly Arg Gln Asn		
130	135	140
Pro Ala Leu Tyr Arg Ile Val Glu Tyr Ile Asn Val Ala Val Arg Leu		

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145	150	155	160
Arg Ser Glu Glu Val Asp Glu Gln Glu Gln Arg Arg Arg Leu Ser Ala	165	170	175
Leu Pro Phe Gly Asp His Asn Asp Asn Leu Glu Glu Gln Asp Phe Gly	180	185	190
Glu Gln Asp Phe Arg Tyr Val Tyr Gly Phe Glu Cys Ala Arg Phe Leu	195	200	205
Leu Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Gly Arg Tyr	210	215	220

<210> SEQ ID NO 33  
 <211> LENGTH: 734  
 <212> TYPE: DNA  
 <213> ORGANISM: *Cylicostephanus goldi*

<400> SEQUENCE: 33

```

atgaacaaaa cgtaaacatt tctcacagtc gttagtgcg ttgtcctggc ccaaggtgtc    60
atggccctat ttggtgaaga gagtogtgaa gaacaccgcc gtcaccatcg tcattcactc    120
ttaccaccat atctccacaa cgtgagctgt gtggctaaat gggagtactt caaaattgtg    180
gggaacagga gttttacgctt tgctgagaaa aagaaagaaa tcagcagagtg ggctaataaaa    240
tacaatggtg tggatgaagt tgcaagctac aatgcttata gagaaaaact caagcaggaa    300
cacaggaaaa acgtttagcga acttgtttct gatcttccca acgcagtaaa gaaagtcaac    360
gatcttttgg acaacgaaa tcagacttct aggcaacttt atgttgact cagagaactt    420
ggtagacaaa atccggcact ataccgtgct gtcgagtata tcaatgtggc tgtgagatta    480
agacgaaaag aacaggatga acaagaacga caaggaacgc tgcagctct accttttggc    540
gagaataacg acaatttggg agagcaggac tttggtgaac aagactttcg ctatgtctat    600
ggctttgagt gtgcaagatt tctccttcaa aatggaagaa tgtttggact caacacggat    660
agaagatacc agtaagagtc aactgtagct caaagtgggt ttgagctacg aacagcatgc    720
catcatcacc tcct                                     734
    
```

<210> SEQ ID NO 34  
 <211> LENGTH: 223  
 <212> TYPE: PRT  
 <213> ORGANISM: *Cylicostephanus goldi*

<400> SEQUENCE: 34

Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Ser Ala Val Val Leu	1	5	10	15
Ala Gln Gly Val Met Ala Leu Phe Gly Glu Glu Ser Arg Glu Glu His	20	25	30	
Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val	35	40	45	
Ser Cys Val Ala Lys Trp Glu Tyr Phe Lys Ile Val Gly Asn Arg Ser	50	55	60	
Leu Thr Phe Ala Glu Lys Lys Lys Glu Ile Ser Glu Trp Ala Lys Lys	65	70	75	80
Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys	85	90	95	
Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Asp Leu	100	105	110	
Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln	115	120	125	

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Thr Ser Arg Gln Leu Tyr Val Ala Leu Arg Glu Leu Gly Arg Gln Asn  
 130 135 140

Pro Ala Leu Tyr Arg Val Val Glu Tyr Ile Asn Val Ala Val Arg Leu  
 145 150 155 160

Arg Arg Lys Glu Gln Asp Glu Gln Glu Arg Gln Gly Thr Leu Ser Ala  
 165 170 175

Leu Pro Phe Gly Glu Asn Asn Asp Asn Leu Glu Glu Gln Asp Phe Gly  
 180 185 190

Glu Gln Asp Phe Arg Tyr Val Tyr Gly Phe Glu Cys Ala Arg Phe Leu  
 195 200 205

Leu Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr  
 210 215 220

<210> SEQ ID NO 35  
 <211> LENGTH: 728  
 <212> TYPE: DNA  
 <213> ORGANISM: *Cylicostephanus longibursatus*

<400> SEQUENCE: 35

atgaacaaaa cgtaaactt tctcaccgtc gtctatgccg tagtctggc ccaaggtgtc 60  
 atggaccttt ttggtgaaga gggctgtgaa gaacatcgcc gtcaccatcg tcattcactc 120  
 ttaccaccat atctccacaa tgtgagctgt gtggctaaat gggaaactt caaaattctg 180  
 gggaaacagga gtttgacgtt tgctgagaaa aaggaaaaaa tcagcgagtg ggcaaagaag 240  
 tacaatgttg tggatgaagt tgcaagctat aatgcttgca gggaaaagct taagcaggaa 300  
 cacaggaaaa acgtttagca aattgtttct aatcttccca atgcagttaa gaaagtaaac 360  
 gatcttttgg acaatgaaaa tcagaccccc aggcaacttt acgttgcctt cagaaaactt 420  
 ggtaaacaaa atccggcact ttatcgtggt gtcgagtata tcaatgtgct tgtgagacta 480  
 agacgtgaag aatttgatga agatcaacga agatcgctgt cagctttacc ttttgcgac 540  
 aataacgacg atttggaaga gcaggacttt ggtgaacagg actttcgcta tatctatggc 600  
 tttgagtgtg caagatttat ccttcaaaaat ggaagaatgt tcggaatcaa cacggataga 660  
 agatattagt aagagtcaac tgtagctcaa gggtttgagc tacgaactgc atgccatcat 720  
 cacctect 728

<210> SEQ ID NO 36  
 <211> LENGTH: 222  
 <212> TYPE: PRT  
 <213> ORGANISM: *Cylicostephanus longibursatus*

<400> SEQUENCE: 36

Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Tyr Ala Val Val Leu  
 1 5 10 15

Ala Gln Gly Val Met Asp Leu Phe Gly Glu Glu Gly Arg Glu Glu His  
 20 25 30

Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val  
 35 40 45

Ser Cys Val Ala Lys Trp Glu Tyr Phe Lys Ile Leu Gly Asn Arg Ser  
 50 55 60

Leu Thr Phe Ala Glu Lys Lys Glu Lys Ile Ser Glu Trp Ala Lys Lys  
 65 70 75 80

Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Cys Arg Glu Lys  
 85 90 95

Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Ile Val Ser Asn Leu  
 100 105 110

-continued

Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln  
 115 120 125  
 Thr Pro Arg Gln Leu Tyr Val Ala Leu Arg Lys Leu Gly Lys Gln Asn  
 130 135 140  
 Pro Ala Leu Tyr Arg Val Val Glu Tyr Ile Asn Val Leu Val Arg Leu  
 145 150 155 160  
 Arg Arg Glu Glu Phe Asp Glu Asp Gln Arg Arg Ser Leu Ser Ala Leu  
 165 170 175  
 Pro Phe Gly Asp Asn Asn Asp Asp Leu Glu Glu Gln Asp Phe Gly Glu  
 180 185 190  
 Gln Asp Phe Arg Tyr Ile Tyr Gly Phe Glu Cys Ala Arg Phe Ile Leu  
 195 200 205  
 Gln Asn Gly Arg Met Phe Gly Ile Asn Thr Asp Arg Arg Tyr  
 210 215 220

<210> SEQ ID NO 37  
 <211> LENGTH: 132  
 <212> TYPE: PRT  
 <213> ORGANISM: Cyathostomum sp.

<400> SEQUENCE: 37

Arg Glu Lys Ala Arg Ile Ile Gln Asp Glu Tyr Thr Lys Arg Met Gln  
 1 5 10 15  
 Gln Val Thr Pro Gln Ala Gln Glu Phe Leu Ala Lys Trp Glu Lys Thr  
 20 25 30  
 Trp Phe Thr Asn Val Gln Gln Tyr Ser Gly Asp Lys Lys Ala Phe Phe  
 35 40 45  
 Lys Gln Met Ile Glu Leu Ile Pro Gln Leu Met Glu Glu Val His Gly  
 50 55 60  
 Phe Ser Glu Glu Thr Trp Lys Ser Leu Glu Glu Gln Phe Pro Glu Gln  
 65 70 75 80  
 Thr Ala Ala Trp Lys Asp Asn Glu Asp Arg Leu Lys Gln Phe Tyr Glu  
 85 90 95  
 Phe Ile Lys Ser Leu Pro Lys Gln Asp Leu Ala Glu Asp Pro Glu Ala  
 100 105 110  
 Phe Arg Lys Phe Ala His Leu Gly Leu Gln Lys Leu Leu Pro Ile Glu  
 115 120 125  
 Ala Leu Arg Ala  
 130

<210> SEQ ID NO 38  
 <211> LENGTH: 396  
 <212> TYPE: DNA  
 <213> ORGANISM: Cyathostomum sp.

<400> SEQUENCE: 38

agggagaagg ctgaattat tcaagacgaa tacactaaac gtatgcagca ggtcacacca 60  
 caagctcagg aattctcggc aaaatgggag aagacatggt tcacgaatgt gcagcaatat 120  
 agcggagata agaaagcttt cttcaagcag atgattgagc taatccctca actaatggag 180  
 gaggttcacg ggttctcggg agagacttgg aagagccttg aggagcaatt cccagagcag 240  
 acagccgcat gaaagataa tgaggatcgc ctaaagcaat tttatgagtt tatcaagagc 300  
 ctaccaagc aggacttagc tgaggatcgc gaagattca gaaagtgcgc tcacctcgga 360  
 ctccagaaac ttcttccaat tgaagctctc agagct 396

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<210> SEQ ID NO 39  
 <211> LENGTH: 565  
 <212> TYPE: DNA  
 <213> ORGANISM: *Cyathostomum catinatum*.

<400> SEQUENCE: 39

```
tggtcacacc acaagctcag gagttcctgg ccaaggttaag ctattacctt accaggggtga    60
ggggaaagaa gttggcagcg gtcggaaacc cggtaatcta ctgactttac caattathtt    120
cagtgggaga agacatggtt cacgaatata cagcaatata gtggagacaa gcaagccttc    180
ttaaagcaga tgattgaact aattcctcaa cttatggagg aggttcaggt aagttagccg    240
caaaaathtt taaccaatgg ttgagctoga cattttttca gggattcaca gaggagactt    300
ggaatagcct gagggagcaa ttcccggagc agacagccgc atggaaggat cgtgagtatc    360
tttcataatt actgtacttg gaattatact ttacaatcat aatcctactc ttagacgagg    420
atgcctgaa gcaattctat gagttcatta agagcctacc caaacaacaa ttagctgagg    480
tgattttcat tgatttttcg aaaaatatat ttttgataca ttctttttca ggatccggaa    540
gctttcagaa agttcgctca cctcg                                           565
```

<210> SEQ ID NO 40  
 <211> LENGTH: 569  
 <212> TYPE: DNA  
 <213> ORGANISM: *Cyathostomum catinatum*.

<400> SEQUENCE: 40

```
ttgtcacacc acaagctcag gagttcctgg ctaaggttaag ctattacctt accaggggtga    60
gggggaagaa gttgggagcg gtcggaaacc cggtaatcta ctgactttac caattathtt    120
cagtgggaga ggacatggtt cacgaatata cagcaatata gtggagacaa gcaagccttc    180
ttaaagcaga tgattgaact aattcctcaa cttatggagg aggttcaggt aagttggccc    240
caaaaathtt taaccaatgg ttgagctoga cattttttca gggattcaca gaggagactt    300
ggaatagcct gagggagcaa ttcccggagc agacagccgc atggaaggat cgtaagtatc    360
tttcataatt actgtacttg gaattatact ttacaatcat aatcctactc ttagacgagg    420
atgcctgaa gcaattctat gagttcatta agagcctacc caaacaacaa ttagctgagg    480
tgattttcat tgatttttcg tacgaaaaat atatttttga tacattcttt ttcaggatcc    540
ggaagctttc agaaagtctg ctcacctcg                                           569
```

<210> SEQ ID NO 41  
 <211> LENGTH: 561  
 <212> TYPE: DNA  
 <213> ORGANISM: *Cylicostephanus longibursatus*.

<400> SEQUENCE: 41

```
aggtcacacc acaagctcag gaattcctgg caaaggttaag ctatcacctt accaggggtga    60
ggggtagaag ttaggagcga gggaaaccgg tgatctctta taccattac ttcagtgga    120
gaagatatgg ttcacgaatg tacagcaata tagtgagac aagcaagcct tcttcaagca    180
gatgattgaa ctaattcctc aacttatgga ggaggtagac gtaagtcagc taaagtgatt    240
ttaagaaaaa attaaccttg attttcttt cagggattct cagaggagac ttggaatagc    300
cttaaggagc aattccctga gcagacagcc gcatggaagg atagttagta tttttcataa    360
ttactgtact tgaattata ctttacaatc ataatcctac cctcagacga ggagcgcctg    420
aagcaattct atgagttcat taagagccta cccaacaac aaatagctga ggtgattttc    480
attgattttt cgtacgaaaa gtatatTTTT aatacattct tttgcaggat ccggaagcct    540
```

-continued

tcagaaagtt cgctcacctc g 561

<210> SEQ ID NO 42  
 <211> LENGTH: 589  
 <212> TYPE: DNA  
 <213> ORGANISM: Cylicocyclus nassatus

<400> SEQUENCE: 42

aggtcacacc acaagctcag gaattcctgg caaaggtaag ctacatatt tcgaggggga 60  
 gggcaathtt ggagcgaggg aggagaggaa agggagagaa aactgggttg ggatcactaa 120  
 ctctaccgcg cacttccagt gggagaagac atggttcacg aatgtgcagc aatatagcgg 180  
 agataagaaa gcctttttca aacagatgat tgagctaatc cctcaactaa tggaagaggt 240  
 tcatgtaagt caaccaaagt ggcttttaag cggagattaa actcgaattt ttcttcaggg 300  
 gttctcggag gagacttga agagccttga ggagcaatc ccagagcaga cagcccgatg 360  
 gaaggatagt aagcattctt catagctccc gcctttatca tttatcttca cgatagtaat 420  
 cttathtttta gatgaggatc gcctgaagca attttatgag ttcataaga gcctacccaa 480  
 gcaggactta gctgaggtaa ctttcatggt tttttcctga gctgtaaaaa tgcttgcaac 540  
 taacaacttt tctaggatcc ggaagctttc agaaagtctg ctcactcg 589

<210> SEQ ID NO 43  
 <211> LENGTH: 195  
 <212> TYPE: PRT  
 <213> ORGANISM: Cyathostomum sp.

<400> SEQUENCE: 43

Lys Lys Glu Ser Gln Gly Phe Phe Ser Ile Pro Val Asp Asn Leu Arg  
 1 5 10 15  
 Ala Ser Pro Phe Leu Leu Gln Tyr Ile Lys Glu Tyr Ile Pro Asp Tyr  
 20 25 30  
 Lys Asn Ala Met Glu Lys Phe Glu Asp Ile Pro Lys Gln Tyr Arg Asp  
 35 40 45  
 Leu Ile Pro Glu Glu Val Ala Thr His Leu Lys Ala Ile Thr Ala Glu  
 50 55 60  
 Glu Lys Ala Val Leu Lys Glu Val Met Lys Asp Tyr Ala Lys Tyr Lys  
 65 70 75 80  
 Asp Glu Glu Glu Phe Leu Lys Ala Leu Lys Glu Lys Ser Glu Gly Leu  
 85 90 95  
 His Glu Lys Ala Ser Lys Leu His Asn Phe Ile Lys Gly Lys Val Asp  
 100 105 110  
 Ala Leu Gly Asp Glu Ala Lys Ala Phe Val Lys Lys Val Ile Ala Ala  
 115 120 125  
 Ala Arg Glu Val His Ala Lys Leu Leu Ala Gly Asp Lys Pro Ser Leu  
 130 135 140  
 Glu Asp Ile Lys Lys Lys Ala Lys Glu His Met Gly Glu Phe Glu Lys  
 145 150 155 160  
 Leu Ser Asp Asp Ala Lys Glu Asp Leu Lys Lys Asn Phe Pro Ile Leu  
 165 170 175  
 Thr Ser Val Trp Thr Asn Glu Lys Thr Arg Ala Leu Ile Asp Lys Tyr  
 180 185 190  
 Val Glu Asn  
 195

<210> SEQ ID NO 44

-continued

&lt;211&gt; LENGTH: 573

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Cyathostomum sp.

&lt;400&gt; SEQUENCE: 44

```

atgcttcgaa taactttctt ccttgctctc ttgttgtct acactttttc tgcacctct 60
ggaccgctg aagagaagat agatgtggaa aaaatggaaa aattgaaga tattccaaag 120
caatatcgag accttattcc ggaagaggtg gctacacacc tcaaagccat caccgctgaa 180
gagaaagctg ttctaaaaga ggtaatgaag aattatgcaa agtacaagaa cgaggaggag 240
tttttgaag cgttgaaga aaaatcagag agtttgcag agaaagccag caaacttcac 300
aatttatca aagggaaagt tgacgcactt ggagatgaag caaaggcatt tgtgaagaag 360
gttatcgag ctgctcgaga agtgcagccc aaacttcttg cgggggacaa accatcgctt 420
gaagatatca agaagaaagc caaggagcat atggctgaat tcgagaaact aagcgatgat 480
gccaaggagg atctcaaaaa gaatttccca atccttactt cgtctggac aaatgagaaa 540
acaagagcgt tgattgacaa atatgtggag aac 573

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&lt;210&gt; SEQ ID NO 45

&lt;211&gt; LENGTH: 145

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Cyathostomum sp.

&lt;400&gt; SEQUENCE: 45

```

Gly Lys Met Ser Asp Leu Trp Thr Ala Ile Ser Glu Thr Asn Lys Val
1           5           10           15
Arg Leu Phe Asn Thr Leu Ser Leu Gly Ile Ala Gly Val Leu Cys Ile
                20           25           30
Thr Thr Ala Phe Ile Pro Val Glu Asn Gln Val Val Cys Ala Val Leu
        35           40           45
Ile Thr Leu Leu Gln Gly Val Ile Gly Phe Asn Ser Ala Gly Tyr Asn
        50           55           60
Lys Ala Ala Val Ile Val Ala Arg Gln His Ala His Leu Leu Leu Thr
65           70           75           80
Cys Phe Gly Leu Ile Val Thr Phe Val Pro Leu Val Gln Pro Phe Ile
        85           90           95
Val Gln Leu Val Ala Pro Asp His Ser Trp Asp Gln Trp Phe Tyr Leu
        100          105          110
Phe Val Gly His Gly Leu Val Leu Val Ile Ala Asn Leu Phe Phe Cys
        115          120          125
Leu Thr Ile Glu Ala Lys Pro Ala Ala Phe Thr Gln Lys Thr Asp Ser
        130          135          140
Ser
145

```

&lt;210&gt; SEQ ID NO 46

&lt;211&gt; LENGTH: 435

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Cyathostomum sp.

&lt;400&gt; SEQUENCE: 46

```

ggtaaaatgt cagatttatg gacggcaata agcgaaacaa ataaagtccg cttgttcaac 60
accttgtcgc tgggaattgc tggcgactg tgtataacta ctgctttcat tctgtggaa 120
aatcaggttg tttgcgctgt tttaatcacg ttattgcaag gagttatcgg attcaattca 180
gctggatata acaaagctgc agtcattgtt gctaggcagc atgctcatct tctgttgacc 240

```

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```

tgccttgggc tcattgtcac ttttgtcccc ttggtgcagc cattcatagt tcaacttggtg 300
gcccttgacc atagctggga ccaatggttt tatctgtttg ttgggcatgg tctcgtactt 360
gttatagcga atttattctt ttgtctcact atcgaggcga aaccggcagc gttcacacag 420
aaaactgatt catca 435

```

```

<210> SEQ ID NO 47
<211> LENGTH: 173
<212> TYPE: DNA
<213> ORGANISM: Cyathostomum sp.

```

```

<400> SEQUENCE: 47

```

```

ggtttaatta cccaagtttg aggtactttc taaatctgac cogatcaact gattgtggtc 60
tgattaaatt ttgaaaatct ctccctgaat agggagagta caagagtgca tatccaaaaa 120
aaaaaaaaaa aaaaaaaaaa aaaaacatgt cggccgcctc ggccctctaga ata 173

```

```

<210> SEQ ID NO 48
<211> LENGTH: 287
<212> TYPE: DNA
<213> ORGANISM: Cyathostomum sp.

```

```

<400> SEQUENCE: 48

```

```

ggtttaatta cccaagtttg agtgtcatga agcttgcctg aaaaaagcag agaaccaag 60
aggagatagt ttcacagttc cgccagacag gaaatgcctg ccaagatggt ttgcggaaga 120
ggagaaacgt cgttcaacta gaatgagaag gcattgattc tgtttagtcg ttgagatatt 180
taaaaattct ttgcagaaaa ccttttcaaa tcataaagtc gaagaccaca aaaaaaaaaa 240
aaaaaaaaaa aaaaaaaaaa atgtcggccc cctcggcctc tagaata 287

```

```

<210> SEQ ID NO 49
<211> LENGTH: 620
<212> TYPE: DNA
<213> ORGANISM: Cyathostomum sp.

```

```

<400> SEQUENCE: 49

```

```

ggtttaatta cccaagtttg aggtctcttc aacagtaggt ttagaaatga catcgggat 60
atggcgccgc acccagagcc ctccattatt gctactcctg ttgttgatca gtctaccagt 120
agctgagtggt agtattcgac tatgtggagt gcgactaaca cgaactctta tggctatctg 180
caggaatcaa ttatcgggtt attcgcaaag taaaagatct gctatgtggg aagagcctcg 240
actgaaaacc gtgcactcaa caatgaaacg atcagggatc gccaccgaat gctgcgagaa 300
tcggtgctca ttagctact taaagacata ctgctgcagc acttagcctt ggcactctaa 360
gcccgtttta tctcctctcc atgatctctc ttcggtatct gtataaccga atatagtcac 420
tccggaaatg cggatgctta ggccaatttg ttgacgtttg ccgcatgaat catttgctgt 480
tcgtcattat ctccacagagc tgtaaaagat ctctttttat gaaagtctat tttgtttgag 540
ctgcaccatt aaaccgttca caaaaaaaaa aaaaaaaaaa aaaaaaaaaa acatgtcggc 600
cgccctcggc tetagaataa 620

```

```

<210> SEQ ID NO 50
<211> LENGTH: 172
<212> TYPE: DNA
<213> ORGANISM: Cyathostomum sp.

```

```

<400> SEQUENCE: 50

```

```

ggtttaatta cccaagtttg aggtactttc tagatctgac cogatcaact gattgtggtc 60

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 tgattaaatt ttggaatct cttcctgaac agggagagta caagagtga tatgaaaaaa 120

aaaaaaaaaa aaaaaaaaaa aaaacatgtc ggccgcctcg gcctctagaa ta 172

&lt;210&gt; SEQ ID NO 51

&lt;211&gt; LENGTH: 327

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Cyathostomum sp.

&lt;400&gt; SEQUENCE: 51

ggtttaatta cccaagtttg aggatgctta gtttcaagct cgtttcttc ttcgtacttc 60

tcacagcttg tgtgctaaca gatccaagag tgtaatccg agaaaagcga atggactgga 120

gacgttacta tagcagatgg ggtcgcggaa gctctaattg gggaaaccgc ggaggtacct 180

tcggcggacg aaaatggagt taccgactt ttggacaatg gggacattaa catctgatgt 240

atgaaaagat ctaatgaaat aaagcttoga aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa 300

atgtcggcgc cctcggcctc tagaata 327

&lt;210&gt; SEQ ID NO 52

&lt;211&gt; LENGTH: 324

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Cyathostomum sp.

&lt;400&gt; SEQUENCE: 52

ggtttaatta cccaagtttg agaatgttcg aaaaattcct tctgctactg atcgttgtga 60

tcgccctcat ttctttggcg tctgcagatt tttcatgctt cttcggatgat accatctgca 120

agagcattac atgcaggggc tgcaccgtcg ccacttgctt taatggagac tgtatgtgca 180

cactatgtaa ctgatgatct tcacatgtcg cattaccatt tgtaacaaat acatcttctc 240

ttgttcataa taaatttttc actcaaaaaa aaaaaaaaaa aaaaaaaaaa aaaaaaacatg 300

tcggccgcct cggcctctag aata 324

&lt;210&gt; SEQ ID NO 53

&lt;211&gt; LENGTH: 328

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Cyathostomum sp.

&lt;400&gt; SEQUENCE: 53

ggccgcggga ttttctagag gccgaggcgg gtttttaggt gttcctcaa cttgggtaat 60

taaaccacga ggccgaggcg ggttttaggt tgttctcaa cttgggtaat taaaccacga 120

tggcgaggcg ggttttaggt tgttctcaa cttgggtaat taaaccacga tggcgaggcg 180

ggttttaggt tgttctcaa acttgggtaa ttaaaccaag aggcgaggcg ggttttaggt 240

ttgttctca aacttgggta attaaaccac gatggcgagg cgggttttag gttgttctca 300

aacttgggta attaaaccac tcactagt 328

&lt;210&gt; SEQ ID NO 54

&lt;211&gt; LENGTH: 154

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Cyathostomum sp.

&lt;400&gt; SEQUENCE: 54

ggccgcggga ttattctaga ggccgaggca gtggtatcaa cgcagagtgg ccattacggc 60

cggggagagg gaaaagtctt ttttctctcg gataccaaaa aaaaaaaaaa aaaaaaaaaa 120

aaaaaacatg tcggccgcct cggcctctag aata 154

&lt;210&gt; SEQ ID NO 55

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<211> LENGTH: 387  
 <212> TYPE: DNA  
 <213> ORGANISM: Cyathostomum sp.  
 <400> SEQUENCE: 55  
 ggccgcggga ttttctagag gccgaggcgt cttacttggg tggctcaata actgaaagct 60  
 tagaattcat taaaccttaa cccacagggg ttatttgaca tgcttgactt gaaaatgatg 120  
 ctcttctgct tgtagttggt ttattatgct agctgtaagt atactctggt agaccagaac 180  
 atcaatgtgc tagttgaatg tatcatgtta tcactttgtc aactctata cgaatctagg 240  
 tgtggcaggc cacaccctc tcctgacct gttcaccatc aattagcttt tagctgttat 300  
 ttaataacat cacactgatt gcaaaaaaaaa aaaaaaaaaa aaaaaaaaaa acatgtcggc 360  
 cgctcggcc tctaaaaaat cactagt 387

<210> SEQ ID NO 56  
 <211> LENGTH: 292  
 <212> TYPE: DNA  
 <213> ORGANISM: Cyathostomum sp.  
 <400> SEQUENCE: 56  
 ggccgcggga ttattctaga ggccgaggca gtggtatcaa cgcagagtgg ccattacggc 60  
 cgaagcagtg gtatcaacgc agagtggcca ttacggccgg gtggtgacca cgggtgacgg 120  
 ggaattaggg ttcgattccg gagagggagc ctgagaaacg gctaccacat ccaaggaagg 180  
 cagcagcgcg gcaaatcacc cactcccgc ccggggaggt agtgacgaaa aaaaaaaaaa 240  
 aaaaaaaaaa aaaaacatgt cgccgcctc ggctctaga ataactacta gt 292

<210> SEQ ID NO 57  
 <211> LENGTH: 199  
 <212> TYPE: DNA  
 <213> ORGANISM: Cyathostomum sp.  
 <400> SEQUENCE: 57  
 ggccgcggga ttttctagag gccgaggcgg gttttagctc aaacttgggt aattaaaccg 60  
 gtaggatggc gaggcggggt tctcaactt gggtaattaa accagtagga tggcgaggcg 120  
 ggtttctcaa acttgggtaa ttaaaccggt aggaggccga ggcgggtctc aaacttgggt 180  
 aattaaacca atcactagt 199

<210> SEQ ID NO 58  
 <211> LENGTH: 167  
 <212> TYPE: DNA  
 <213> ORGANISM: Cyathostomum sp.  
 <400> SEQUENCE: 58  
 caagtttgag gtactttcta gatctgacct gatcaactga ttgtggtctg attaaatttt 60  
 ggaaatctct tctgaacag ggagagtaca agagtgtata ttaagaaaaa aaaaaaaaaa 120  
 aaaaaaaaaa catgtcggcc gcctcggcct ctagaataat cactagt 167

<210> SEQ ID NO 59  
 <211> LENGTH: 22  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer  
 <400> SEQUENCE: 59  
 ggttaatta cccaagttag ag 22

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<210> SEQ ID NO 60  
 <211> LENGTH: 17  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer  
  
 <400> SEQUENCE: 60  
  
 attctagagg ccgaggc 17  
  
 <210> SEQ ID NO 61  
 <211> LENGTH: 17  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer  
  
 <400> SEQUENCE: 61  
  
 ttctagaggc cgaggcg 17  
  
 <210> SEQ ID NO 62  
 <211> LENGTH: 20  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer  
  
 <400> SEQUENCE: 62  
  
 aaaaaggagg tgtttggttc 20  
  
 <210> SEQ ID NO 63  
 <211> LENGTH: 23  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer  
  
 <400> SEQUENCE: 63  
  
 cttgaatttg ataaaactac acc 23  
  
 <210> SEQ ID NO 64  
 <211> LENGTH: 18  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer  
  
 <400> SEQUENCE: 64  
  
 aattgtgggg aacaggag 18  
  
 <210> SEQ ID NO 65  
 <211> LENGTH: 21  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer  
  
 <400> SEQUENCE: 65  
  
 aatgaaaatc agactcctag g 21  
  
 <210> SEQ ID NO 66  
 <211> LENGTH: 34  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer

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&lt;400&gt; SEQUENCE: 66

aattcggatc cgcaaggtgt catggacctt tttg

34

&lt;210&gt; SEQ ID NO 67

&lt;211&gt; LENGTH: 38

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Artificial

&lt;220&gt; FEATURE:

&lt;223&gt; OTHER INFORMATION: Primer

&lt;400&gt; SEQUENCE: 67

ccgcaagctt atatctttca tctgtgttga gtccaaac

38

&lt;210&gt; SEQ ID NO 68

&lt;211&gt; LENGTH: 60

&lt;212&gt; TYPE: PRT

<213> ORGANISM: *Cyclicoclyclus ashworthi*

&lt;220&gt; FEATURE:

&lt;221&gt; NAME/KEY: MISC\_FEATURE

&lt;222&gt; LOCATION: (43)..(43)

&lt;223&gt; OTHER INFORMATION: X is E or V

&lt;220&gt; FEATURE:

&lt;221&gt; NAME/KEY: MISC\_FEATURE

&lt;222&gt; LOCATION: (50)..(50)

&lt;223&gt; OTHER INFORMATION: X is G or D

&lt;220&gt; FEATURE:

&lt;221&gt; NAME/KEY: MISC\_FEATURE

&lt;222&gt; LOCATION: (57)..(57)

&lt;223&gt; OTHER INFORMATION: X is E or V

&lt;400&gt; SEQUENCE: 68

Leu Thr Phe Ala Glu Lys Lys Gly Lys Ile Ser Glu Trp Ala Lys Lys  
1 5 10 15Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys  
20 25 30Leu Lys Gln Glu His Arg Lys Asn Val Ser Xaa Leu Val Ser Gly Leu  
35 40 45Pro Xaa Ala Val Lys Lys Val Asn Xaa Leu Leu Asp  
50 55 60

&lt;210&gt; SEQ ID NO 69

&lt;211&gt; LENGTH: 60

&lt;212&gt; TYPE: PRT

<213> ORGANISM: *Cyathostomum catinatum*

&lt;220&gt; FEATURE:

&lt;221&gt; NAME/KEY: misc\_feature

&lt;222&gt; LOCATION: (8)..(8)

&lt;223&gt; OTHER INFORMATION: Xaa can be any naturally occurring amino acid

&lt;400&gt; SEQUENCE: 69

Leu Thr Phe Ala Glu Lys Lys Xaa Glu Ile Ser Glu Trp Ala Lys Lys  
1 5 10 15Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys  
20 25 30Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Ala Leu  
35 40 45Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp  
50 55 60

&lt;210&gt; SEQ ID NO 70

&lt;211&gt; LENGTH: 60

&lt;212&gt; TYPE: PRT

<213> ORGANISM: *Coronocyclus coronatus*

&lt;400&gt; SEQUENCE: 70

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Leu Thr Phe Ala Glu Lys Lys Glu Lys Ile Ser Glu Trp Ala Lys Lys  
 1 5 10 15  
 Tyr Lys Val Glu Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys  
 20 25 30  
 Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Ala Leu  
 35 40 45  
 Pro Gly Ala Val Lys Lys Val Asn Glu Leu Leu Asp  
 50 55 60

<210> SEQ ID NO 71  
 <211> LENGTH: 60  
 <212> TYPE: PRT  
 <213> ORGANISM: *Cylicostephanus goldi*

<400> SEQUENCE: 71

Leu Thr Phe Ala Glu Lys Lys Lys Glu Ile Ser Glu Trp Ala Lys Lys  
 1 5 10 15  
 Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys  
 20 25 30  
 Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Asp Leu  
 35 40 45  
 Pro Ser Ala Val Lys Lys Val Asn Asp Leu Leu Asp  
 50 55 60

<210> SEQ ID NO 72  
 <211> LENGTH: 60  
 <212> TYPE: PRT  
 <213> ORGANISM: *Coronocyclus labiatus*

<400> SEQUENCE: 72

Leu Thr Phe Ala Glu Lys Lys Glu Lys Ile Ser Glu Trp Ala Lys Lys  
 1 5 10 15  
 Tyr Asn Val Val Asp Glu Val Ala Arg Tyr Asn Ala Tyr Arg Glu Lys  
 20 25 30  
 Leu Lys Gln Glu Tyr Arg Lys Asn Val Ser Glu Leu Val Ser Gly Leu  
 35 40 45  
 Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp  
 50 55 60

<210> SEQ ID NO 73  
 <211> LENGTH: 60  
 <212> TYPE: PRT  
 <213> ORGANISM: *Cylicocyclus leptostomum*

<400> SEQUENCE: 73

Leu Thr Phe Ala Glu Lys Lys Gly Lys Ile Ser Glu Trp Ala Lys Lys  
 1 5 10 15  
 Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys  
 20 25 30  
 Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Gly Leu  
 35 40 45  
 Pro Gly Ala Val Lys Lys Val Asn Glu Leu Leu Asp  
 50 55 60

<210> SEQ ID NO 74  
 <211> LENGTH: 60  
 <212> TYPE: PRT  
 <213> ORGANISM: *Cylicostephanus longibursatus*

<400> SEQUENCE: 74

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Leu Thr Phe Ala Glu Lys Lys Glu Glu Ile Ser Lys Trp Ala Lys Lys
1          5          10          15
Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
20          25          30
Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Ile Val Ser Asp Leu
35          40          45
Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp
50          55          60

```

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<210> SEQ ID NO 75
<211> LENGTH: 60
<212> TYPE: PRT
<213> ORGANISM: Cylicostephanus minutus

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<400> SEQUENCE: 75

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Leu Thr Phe Ala Glu Lys Lys Glu Lys Ile Ser Glu Trp Ala Lys Lys
1          5          10          15
Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
20          25          30
Leu Lys Gln Glu His Arg Lys Asn Val Ser Gln Leu Val Ser Ala Leu
35          40          45
Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp
50          55          60

```

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<210> SEQ ID NO 76
<211> LENGTH: 60
<212> TYPE: PRT
<213> ORGANISM: Cylicocyclus nassatus
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (25)..(25)
<223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid

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<400> SEQUENCE: 76

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Leu Thr Phe Ala Glu Lys Lys Glu Lys Ile Gly Glu Trp Ala Lys Lys
1          5          10          15
Tyr Asn Val Val Asp Glu Val Ala Xaa Tyr Asn Ala Tyr Arg Glu Lys
20          25          30
Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Gly Leu
35          40          45
Pro Asn Ala Val Lys Lys Val Asn Glu Leu Leu Asp
50          55          60

```

```

<210> SEQ ID NO 77
<211> LENGTH: 60
<212> TYPE: PRT
<213> ORGANISM: Cyathostomum pateratum
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<222> LOCATION: (8)..(8)
<223> OTHER INFORMATION: X is K or E

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<400> SEQUENCE: 77

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```

Leu Thr Phe Ala Glu Lys Lys Xaa Glu Ile Ser Glu Trp Ala Lys Lys
1          5          10          15
Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
20          25          30
Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Ala Leu
35          40          45
Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp
50          55          60

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<210> SEQ ID NO 78  
 <211> LENGTH: 223  
 <212> TYPE: PRT  
 <213> ORGANISM: *Cyathostomum pateratum*  
 <400> SEQUENCE: 78

Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Ser Ala Val Ala Leu  
 1 5 10 15  
 Ala Gln Gly Val Met Asp Leu Phe Gly Glu Glu Gly Arg Glu Glu His  
 20 25 30  
 Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val  
 35 40 45  
 Ser Cys Glu Ala Lys Trp Glu Tyr Phe Lys Ile Val Gly Asn Arg Ser  
 50 55 60  
 Leu Thr Phe Ala Glu Lys Arg Lys Glu Ile Ser Glu Trp Ala Lys Lys  
 65 70 75 80  
 Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys  
 85 90 95  
 Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Ala Leu  
 100 105 110  
 Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln  
 115 120 125  
 Thr Pro Arg Gln Leu Tyr Val Ala Leu Arg Lys Leu Gly Arg Gln Asn  
 130 135 140  
 Pro Ala Leu Tyr Arg Ile Val Glu Tyr Ile Asn Val Ala Val Arg Leu  
 145 150 155 160  
 Arg Ser Glu Glu Val Asp Glu Gln Glu Gln Arg Arg Arg Leu Ser Ala  
 165 170 175  
 Leu Pro Phe Gly Asp His Asn Asp Asn Leu Glu Glu Gln Asp Phe Gly  
 180 185 190  
 Glu Gln Asp Phe Arg Tyr Val Tyr Gly Phe Glu Cys Ala Arg Phe Leu  
 195 200 205  
 Leu Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Glu Arg Tyr  
 210 215 220

<210> SEQ ID NO 79  
 <211> LENGTH: 203  
 <212> TYPE: PRT  
 <213> ORGANISM: *Nippostrongylus brasiliensis*  
 <400> SEQUENCE: 79

Met Lys Ala Leu Leu Ile Ala Val Leu Ala Leu Thr Ala Ala Ala His  
 1 5 10 15  
 Tyr Arg Gly Lys Glu Ser Glu Thr Gly His Gly Arg His His His His  
 20 25 30  
 Pro Pro Pro Pro Phe Leu Lys Asp Val Asp Lys Ser Ala Arg Lys  
 35 40 45  
 Glu Phe Phe Ala Ile Val Lys Asn Lys Thr Leu Thr Ile Ala Glu Gln  
 50 55 60  
 Lys Ala Ala Val Leu Glu Trp Ala Glu Cys His Gly Ile Lys Asp Glu  
 65 70 75 80  
 Val Glu Gln Phe Gln Gln Lys Met Ala Ser Leu Gly Asp Glu Ile Lys  
 85 90 95  
 Lys Asn Val Ala Glu Leu Ile Ser Lys Leu Pro Ala Ala Phe Gln Ser  
 100 105 110  
 Phe Ser Ala Val Met Glu Ser Glu Asn Gln Thr Arg Arg Glu Gln Lys



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1	5	10	15
Ala Arg His	Asn Glu Gly Cys	Pro Arg Glu Glu Ser	His Arg Ala Gln
	20	25	30
Pro Arg Pro	Lys Phe Leu His	His Val Gly Ile Lys	Ala Arg Arg Glu
	35	40	45
Tyr Phe His	Ile Val Arg Ser	Gly Glu Ile Ile Ala	Lys Gln Asp Glu
	50	55	60
Gln Ile Leu	Asp Trp Ala Lys Lys	Tyr Gly Val Glu Glu	Glu Val Glu
	65	70	75
Glu Phe Asn	Asn Lys Thr Ala Ser	Tyr Val Glu Glu Leu	Val Gln Asn
	85	90	95
Val Thr Asn	Leu Ile Ala Glu Leu	Pro Thr Ala Leu Glu	Ala Phe Leu
	100	105	110
Asn Ile Thr	Gln Asn Lys Asp Gln	Thr Arg Met Glu Met	Lys Lys Ala
	115	120	125
Leu Arg Glu	Met Arg Thr Glu Glu	Phe Glu Val Phe Asp	Ala Leu Lys
	130	135	140
Ala Ala Phe	Lys Val Phe Lys Pro	Asn His Cys Leu Tyr	His Arg Cys
	145	150	155
Thr Asp Ser	Gln Ser Ser Glu Glu	Leu Val Asp Asp Trp	Met Asp Phe
	165	170	175
Gln Glu Lys	Asp Asp Gln Ile Ser	Lys Met Leu Asp Pro	His Asp Glu
	180	185	190
Phe Thr Met	Met Gln Lys Ser Gly	Val	
	195	200	

&lt;210&gt; SEQ ID NO 82

&lt;211&gt; LENGTH: 241

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Caenorhabditis elegans

&lt;400&gt; SEQUENCE: 82

Met Ala Leu	Ser Tyr Ser Phe	Ile Phe Thr Leu	Phe Ala Phe Ser	Ala
1	5	10	15	
Val Val Leu	Ala Gly Pro Gly	Gly Arg His Gly	His Gly Gly Gly	Gly
	20	25	30	
Phe Gly Gly	Ala Pro Gln Leu	Pro Pro Phe Leu	Gln Asn Val Thr	Ala
	35	40	45	
Glu Gly Arg	Gln Ala Phe Phe	Ala Ile Val Ser	Asn Thr Ser Leu	Thr
	50	55	60	
Ile Ser Glu	Thr Glu Ser Gln	Ile Ser Ser Trp	Ala Gln Thr Tyr	Gly
	65	70	75	80
Val Ser Ser	Gln Val Thr Glu	Phe Gln Thr Lys	Val Glu Glu Lys	Leu
	85	90	95	
Asn Glu Ile	Lys Gln Asn Val	Thr Ala Val Ile	Asn Asn Leu Ser	Thr
	100	105	110	
Val Glu Thr	Gln Leu Glu Ala	Ile Phe Ala Asn	Lys Ser Gln Thr	Ile
	115	120	125	
Arg Glu Gln	Phe Gln Ala Leu	Gly Gln Leu Lys	Asp Gln Tyr Pro	Gln
	130	135	140	
Glu Val Gly	Val Leu Leu Phe	Leu Ala Lys Pro	Lys Gly Glu His	Gly
	145	150	155	160
Gly Gln Gly	Pro Phe Gly Gly	Phe Pro Gly Gly	His Gln Gly Gly	Phe
	165	170	175	
Pro Gly Gly	Asn Gln Gly Gly	Phe Gly Gly Asn	Gln Gly Gly Phe	Gly

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180	185	190
Gly Asn Gln Gly Gly Phe Pro Phe Gly Asn Gln Gly Gly Asn Gln Gly		
195	200	205
Gly Phe Pro Phe Gly Asn Pro Gly Asn Gln Gly Gly Phe Gly Gly Asn		
210	215	220
Gln Gly Gly Asn Gln Gly Gly Phe Gly Gly Asn Arg Gly Gly Arg Gly		
225	230	235
240		
Phe		
<210> SEQ ID NO 83		
<211> LENGTH: 235		
<212> TYPE: PRT		
<213> ORGANISM: Caenorhabditis elegans		
<400> SEQUENCE: 83		
Met Ser Tyr Tyr Ser Thr Ser Leu Tyr Ile Phe Ala Ile Thr Met Ala		
1	5	10
15		
Thr Met Val Leu Ala Gly Pro Arg Gly Gly Phe Gly Gly Gly Pro Gly		
20	25	30
Gly Pro Gly Gly Arg Gly Arg His Gly Pro Pro Met Pro Pro Phe Leu		
35	40	45
Gln Asn Val Thr Asp Glu Gly Arg Arg Ala Phe Phe Asp Ile Ala Arg		
50	55	60
Asn Gln Asn Leu Thr Ile Ala Glu Met Glu Ser Gln Thr Ser Thr Trp		
65	70	75
80		
Ala Gln Thr Tyr Gly Val Ser Asp Val Tyr Ser Glu Phe Glu Ala Asn		
85	90	95
Ile Thr Ala His Arg Asn Glu Val Gln Gln Asn Val Thr Gln Val Val		
100	105	110
Ser Gln Leu Ser Ala Ala Gln Thr Ala Leu Glu Ala Val Met Asn Asn		
115	120	125
Lys Asn Gln Thr Arg Gln Gln Met Lys Glu Ala Ile Asp Asn Leu Lys		
130	135	140
Thr Gln Tyr Pro Gln Glu Ile Pro Ala Leu Phe Phe Ile Ser Gly Ser		
145	150	155
160		
Phe Arg Arg Gly Pro Gly Gly Arg His Gly Gly Pro Gly Gly Pro Gly		
165	170	175
Gly Arg Arg Met Gly Pro Gly Gly Arg Gly Gly Asp Ser Arg Glu Gly		
180	185	190
Pro Met Met Gly Gly Met Gly Arg Gly Gly Phe Gly Gly Gln Gly Met		
195	200	205
Gly Gly Met Gly Ala Gly Leu Gly Gln Gly Arg Arg Gly Gly Pro Asp		
210	215	220
Ser Met Asn Glu Ser Ser Asp Val Asn Asp Phe		
225	230	235

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The invention claimed is:

1. A method of diagnosing a *cyathostomin* infection in an animal, said method comprising the steps of:

(a) contacting a sample with a *cyathostomin* larval antigen, wherein the *cyathostomin* larval antigen is at least 60% identical to the sequence of SEQ ID NO: 1; and

(b) identifying a level of anti-*cyathostomin* larval antigen antibodies in the sample, wherein the anti-*cyathostomin* larval antigen antibodies bind to antigen comprising an immunogenic sequence at least 60% identical to the

sequence of SEQ ID NO: 1; wherein a level of anti-*cyathostomin* larval antigen antibodies is indicative of the *cyathostomin* infection.

2. The method of claim 1, wherein the level of anti-*cyathostomin* larval antigen antibodies is evaluated relative to the level of anti-*cyathostomin* larval antigen antibodies present in a reference or control sample obtained from a healthy animal, an animal without a moderate or high mucosal burden of *cyathostomin* parasites, and/or an animal without larval *cyathostomin*osis.

## 109

3. The method of claim 1, wherein the sample is contacted with a *cyathostomin* larval antigen comprising SEQ ID NO: 1.

4. The method of claim 1, wherein the *cyathostomin* larval antigen is bound, conjugated or immobilized on or to a suitable substrate. 5

5. The method of claim 1, wherein the sample is contacted with one or more agent(s) capable of binding:

(a) a *cyathostomin* larval antigen comprising SEQ ID NO: 1; or

(b) a *cyathostomin* larval antigen comprising an amino acid sequence at least 60% identical to the amino acid sequence of SEQ ID NO: 1. 10

6. The method of claim 5, wherein the binding agent(s) are bound, conjugated or immobilized on or to a suitable substrate. 15

7. The method of claim 1, wherein the animal is a member of the Equidae family.

## 110

8. The method of claim 1, wherein the animal is a horse.

9. The method of claim 1, wherein the sample is a biological sample selected from the group consisting of: whole blood, serum, plasma, saliva, sweat, semen, tissue biopsy, tissue scraping, tissue/organ wash/lavage, and fecal preparation.

10. The method of claim 3, wherein the *cyathostomin* larval antigen is bound, conjugated, or immobilized on or to a suitable substrate.

11. The method of claim 1, wherein the level of anti-*cyathostomin* larval antigen antibody is identified using an immunological detection technique.

12. The method of claim 11, wherein the immunological detection technique is selected from the group consisting of enzyme-linked immunosorbent assay (ELISA), Western blot, and dot blot.

\* \* \* \* \*

UNITED STATES PATENT AND TRADEMARK OFFICE  
**CERTIFICATE OF CORRECTION**

PATENT NO. : 8,663,939 B2  
APPLICATION NO. : 13/260935  
DATED : March 4, 2014  
INVENTOR(S) : Matthews et al.

Page 1 of 1

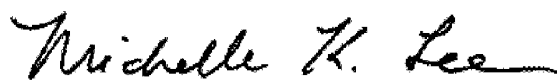
It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

On the Title Page:

The first or sole Notice should read --

Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 8 days.

Signed and Sealed this  
Twenty-ninth Day of September, 2015



Michelle K. Lee  
*Director of the United States Patent and Trademark Office*

专利名称(译)	马寄生虫检测		
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申请号	US13/260935	申请日	2010-03-31
申请(专利权)人(译)	MOREDUN研究所		
当前申请(专利权)人(译)	MOREDUN研究所		
[标]发明人	MATTHEWS JACQUI HODGKINSON JANE PROUDMAN CHRISTOPHER		
发明人	MATTHEWS, JACQUI HODGKINSON, JANE PROUDMAN, CHRISTOPHER		
IPC分类号	G01N33/535 G01N33/569		
CPC分类号	G01N33/569 G01N2333/4353 G01N33/5308		
优先权	2009005511 2009-03-31 GB		
其他公开文献	US20120082992A1		
外部链接	<a href="#">Espacenet</a> <a href="#">USPTO</a>		

摘要(译)

本发明提供了一种诊断cyathostomin感染的方法，所述方法包括鉴定样品中抗cyathostomin幼虫抗原抗体水平的步骤，其中抗cyathostomin幼虫抗原抗体水平指示cyathostomin感染。

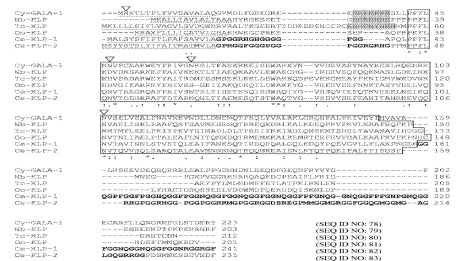


Figure 1

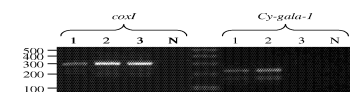


Figure 2