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Matthews et al.

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- (54) **EQUINE PARASITE DETECTION**
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G01N 33/53 (2006.01)
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CPC **G01N 33/569** (2013.01); **G01N 33/5308** (2013.01); **G01N 2333/4353** (2013.01)
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CPC G01N 33/569; G01N 2333/4353;
G01N 33/5308
USPC 435/7.1, 7.92, 252.33, 320.1, 6.12,
435/7.22, 7.95
See application file for complete search history.

References Cited

U.S. PATENT DOCUMENTS

8,663,939 B2 3/2014 Matthews et al.

OTHER PUBLICATIONS

Lederman et al (Molecular Immunology 28:1171-1181, 1991)*
Dowdall, Samantha M.J. et al., "Characterisation of IgG(T) serum antibody responses to two larval antigen complexes in horses naturally- or experimentally-infected with cyathostomins," International Journal for Parasitology, vol. 34, No. 1, Jan. 2004 (pp. 101-108).
Dowdall, S.M.J. et al., "Antigen-specific IgG(T) responses in natural and experimental cyathostominae infection in horses," Veterinary Parasitology, vol. 106, No. 3, Jun. 26, 2002, pp. 225-242.
Dowdall, S.M.J. et al., "Purification and analyses of the specificity of two putative diagnostic antigens for larval cyathostomin infection in horses," Research in Veterinary Science, vol. 75, No. 3, Dec. 2003 (pp. 223-229).

Matthews, Jacqueline B. et al., "Recent developments in research into the Cyathostominae and Anoplocephala perfoliata," Veterinary Research (Les Ulis), vol. 35, No. 4, Jul. 2004 pp. 371-381.
McWilliam, Hamish E.G. et al., "Identification and characterization of an immunodiagnostic marker for cyathostornin developing stage larvae," International Journal for Parasitology, vol. 40, No. 3, Mar. 2010, pp. 265-275.
Abbott, J.B. et al., "Science: Overview—The problem of diagnosing tapeworm infections in horses," Equine Veterinary Journal (2008), vol. 40, No. 1, pp. 5-6.
Abbott, J.B. et al., "Serological changes observed in horses infected with *Anoplocephala perfoliata* after treatment with praziquantel and natural reinfection," The Veterinary Record, Jan. 12, 2008, pp. 51-53.
Barrett, E.J. et al., "Field trial of the efficacy of a combination of ivermectin and praziquantel in horses infected with roundworms and tapeworms," The Veterinary Record, Mar. 13, 2004, pp. 323-325.
Bendtsen, Jannick Dyrlev et al., "Improved Prediction of Signal peptides: SignalP 3.0," J. Mol. Biol., (2004), vol. 340, pp. 783-795.
Bucknell, D.G. et al., "The Prevalence and Epidemiology of Gastrointestinal Parasites of Horses in Victoria, Australia," International Journal for Parasitology, vol. 25, No. 6, 1995, pp. 711-724.
Chapman, M.R. et al., "Equine cyathostome populations: accuracy of species composition estimations," Veterinary Parasitology 116 (2003), pp. 15-21.
Clark, H.J. et al., "Isolation and characterisation of a beta tubulin isotype 2 gene from two species of *cyathostomin*," International Journal for Parasitology, vol. 35 (2005), pp. 349-358.
Eysker, M. et al., "The effect of ivermectin treatment against inhibited early third stage, late third stage and fourth stage larvae and adult stages of the cyathostomes in Shetland ponies and spontaneous expulsion of these helminthes," Vet. Parasitol., vol. 42, (1992) pp. 295-302.
Eysker, M. et al., "Mucosal larval recovery techniques of cyathostomes: can they be standardized?" Veterinary Parasitology, vol. 85 (1999), pp. 137-149.
Geldhof, P. et al., "Protein disulphide isomerase of *Ostertagia ostertagi*: an excretory-secretory product of L4 and adult worms?" International Journal for Parasitology, vol. 33 (2003), pp. 129-136.
Giles, C.J. et al., "Larval cyathostomiasis (immature trichonema-induced enteropathy): A report of 15 clinical cases", Equine Veterinary Journal, vol. 17, No. 3 (1985), pp. 196-201.
Kaplan, Ray M., "Anthelmintic resistance in nematodes of horses," Vet. Res. 33 (2002), pp. 491-507.
Hodgkinson, J.E. et al., "Evaluation of the specificity of five oligoprobes for identification of *cyathostomin* species from horses," International Journal for Parasitology, vol. 31 (2001), pp. 197-204.
Kjaer, L.N. et al., "Interpretation of serum antibody response to *Anoplocephala perfoliata* in relation to parasite burden and faecal egg count," Equine Veterinary Journal, vol. 39, No. 6 (2007), pp. 529-533.

(Continued)

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(57) **ABSTRACT**

The present invention provides a method of diagnosing a cyathostomin infection, said method comprising the step of identifying a level of anti-cyathostomin larval antigen antibodies in a sample, wherein a level of anti-cyathostomin larval antigen antibodies is indicative of a cyathostomin infection.

16 Claims, 11 Drawing Sheets

(56)

References Cited

OTHER PUBLICATIONS

- Klci, T.R. et al., "Evaluation of ivermectin at an elevated dose against encysted equine cyathostome larvae," *Vet. Parasitol.*, vol. 47 (1993) pp. 99-106.
- Larkin, M.A. et al., "Clustal W and Clustal X version 2.0," *Bioinformatics*, vol. 23, No. 21, (2007) pp. 2947-2948.
- Love, S. et al., "The development of naturally acquired cyathostome infection in ponies," *Vet. Parasitol.*, vol. 44, (1992) pp. 127-142.
- Lyons, E.T. et al., "Historical perspective of cyathostomes: prevalence, treatment and control programs," *Veterinary Parasitology*, vol. 85 (1999), pp. 97-112.
- Lyons, E.T. et al., "A study (1977-1992) of population dynamics of endoparasites featuring benzimidazole-resistant small strongyles (Population S) in Shetland ponies," *Veterinary Parasitology*, 1996; 66:75-86.
- Marchier-Bauer, Aron et al., "CDD: a conserved domain database for interactive domain family analysis," *Nucleic Acids Research*, vol. 35, 2007, D237-D240.
- Murphy, D. et al., "The pathogenic effects of experimental cyathostome infections in ponies," *Veterinary Parasitology*, vol. 70 (1997), pp. 99-110.
- Proudman, Chris et al., "Control of intestinal parasites in horses," *In Practice*, Feb. 2000, pp. 90-97.
- Proudman, C.J. et al., "Correlation of antigen specific IgG and IgG(T) responses with *Anoplocephala pefoliata* infection intensity in the horse," *Parasite Immunology*, vol. 18 (1996), pp. 499-506.
- Proudman, C.J. et al., "Use of excretory/secretory antigens for the serodiagnosis of *Anoplocephala pefoliata* cestodosis," *Vet Parasitol.* 1996; 61(3-4):239-47.
- Reid, S.W.J. et al., "Epidemiological risk factors associated with a diagnosis of clinical cyathostomiasis in the horse," *Equine Veterinary Journal*, vol. 27, No. 2 (1995) pp. 127-130.
- Reinemeyer, C.R. et al., "The Prevalence and Intensity of Internal Parasites of Horses in the U.S.A.," *Vet. Parasitol.*, vol. 15, (1984) pp. 75-83.
- Shibui, Akiko et al., "Cloning and characterization of a novel gene encoding keratin-like protein from nematode *Nippostrongylus brasiliensis*," *Biochimica et Biophysica Acta*, 1522 (2001), pp. 59-61.
- Martin, Samuel A.M. et al., "The construction of spliced leader cDNA libraries from the filarial nematode *Brugia pahangi*," *Molecular and Biochemical Parasitology*, vol. 70 (1995) pp. 241-245.
- Altschul, S.F. et al., "Basic local alignment search tool," *J. Mol. Biol.* 1990:215(3): 403-10 (abstract only).
- Klei, T.R. et al., "Irradiated larval vaccination of ponies against strongylus vulgaris," *J. Parasitol.* 1982:68(4): 561-9 (abstract only).
- Lichtenfels, J.R. et al., "Illustrated identification keys to strongylid parasites (Strongylidae: Nematoda) of horses, zebras and asses (Equidae)," *Vet Parasitol.* 2008:156(1-2):4-161, Epub May 21, 2008 (abstract only).
- Ogbourne, C.P., "The prevalence, relative abundance and site distribution of nematodes of the subfamily Cyathostominae in horses killed in Britain," *J. Helminthol.* 1976:50(3):203-14 (abstract only).
- Love S et al., "Pathogenicity of cyathostome infection," *Veterinary Parasitology*, 1999; 85: 113-122.
- International Search Report prepared for PCT/GB2010/000616, issued Sep. 7, 2010.

* cited by examiner

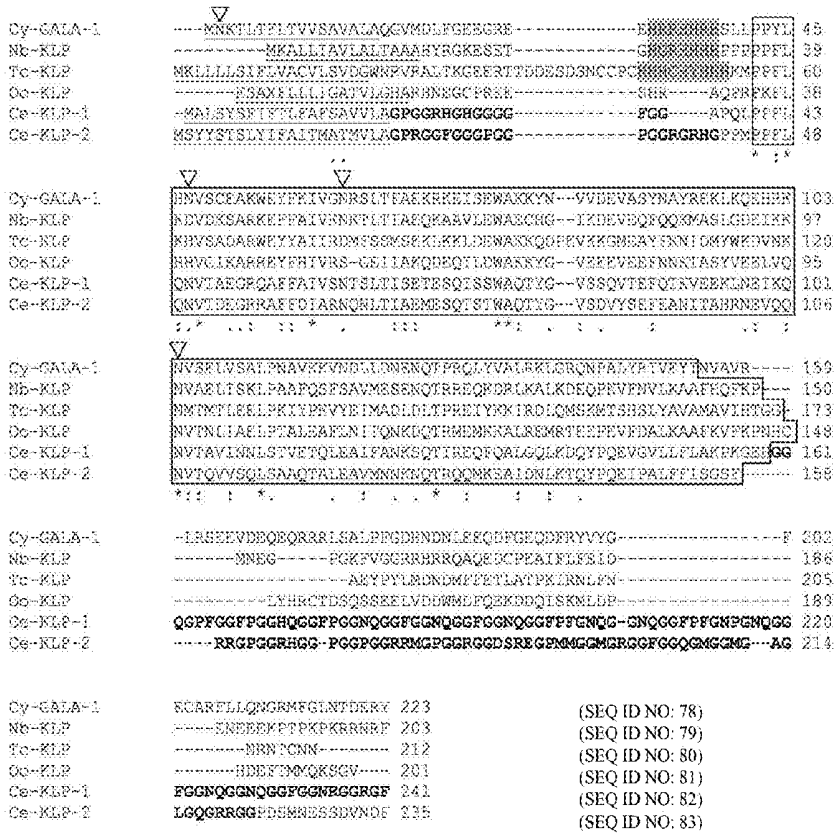


Figure 1

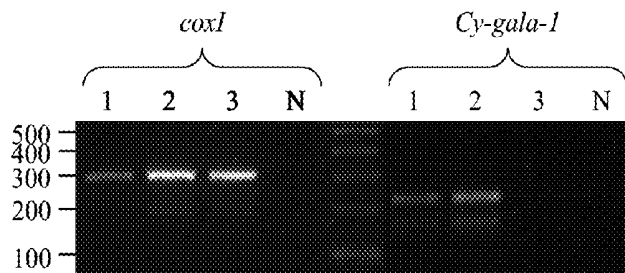


Figure 2

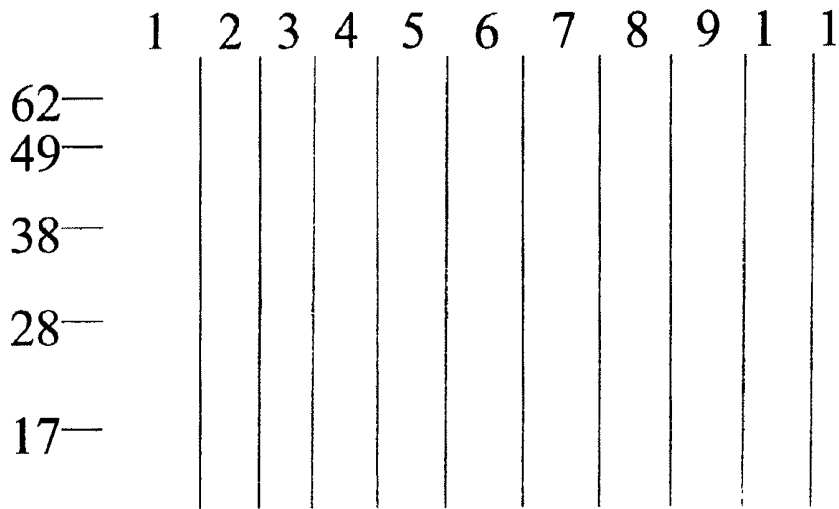


Figure 3A

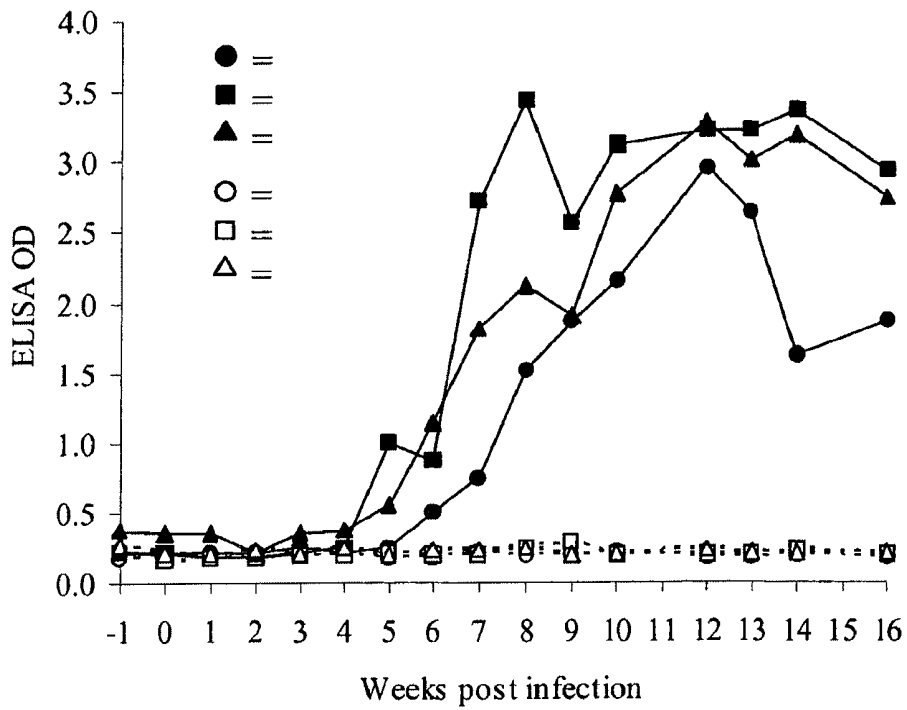


Figure 3B

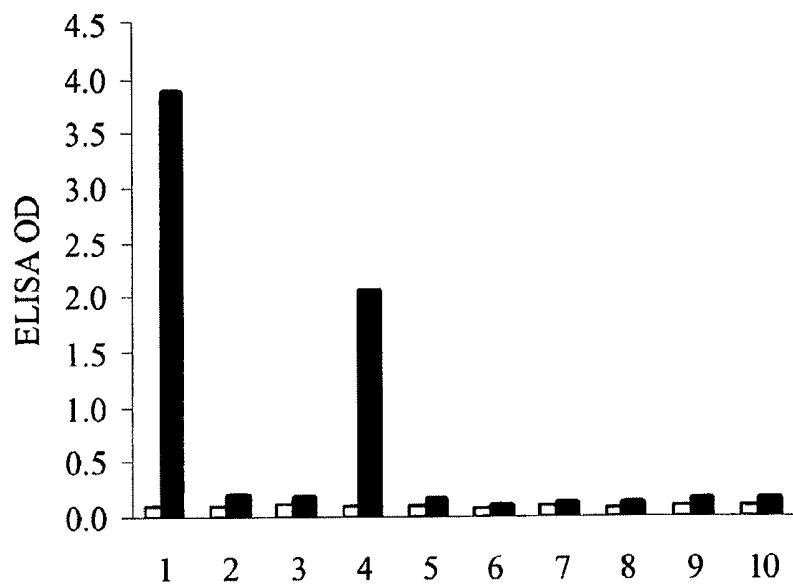


Figure 4A

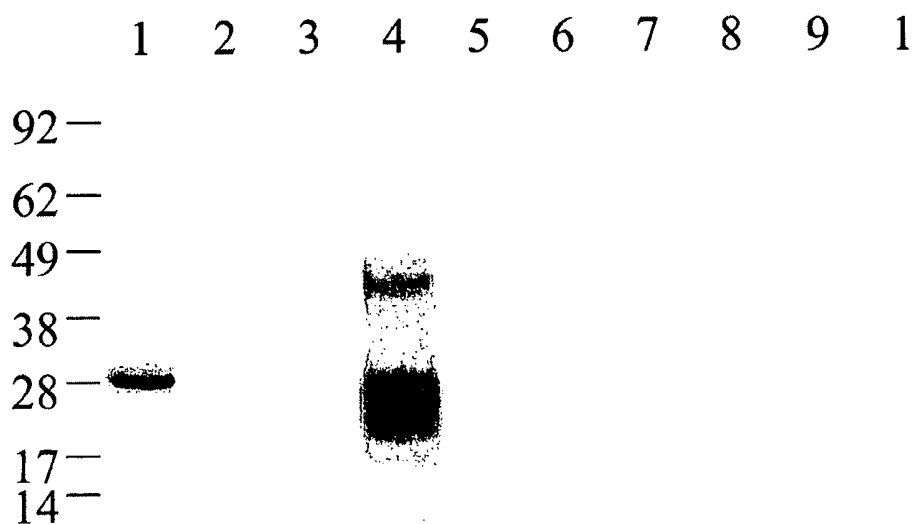


Figure 4B

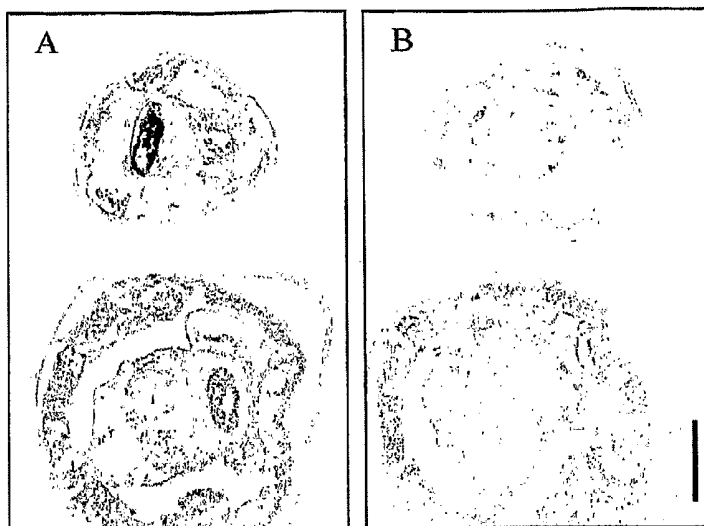


Figure 5

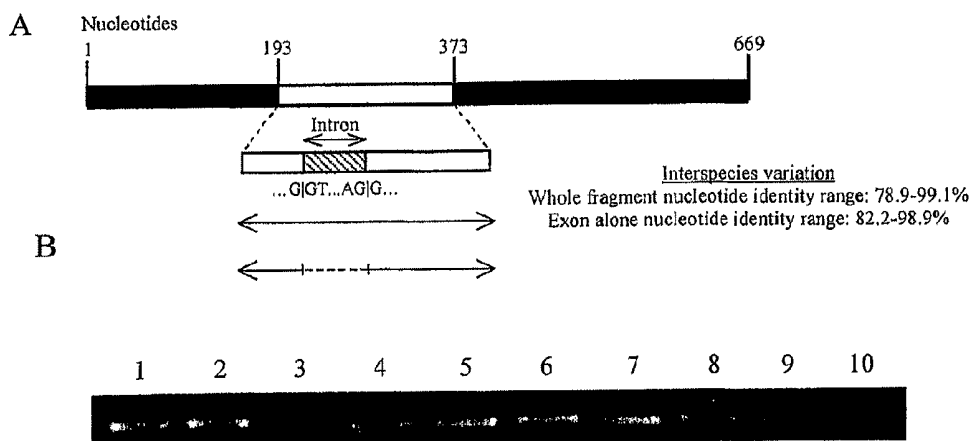


Figure 6

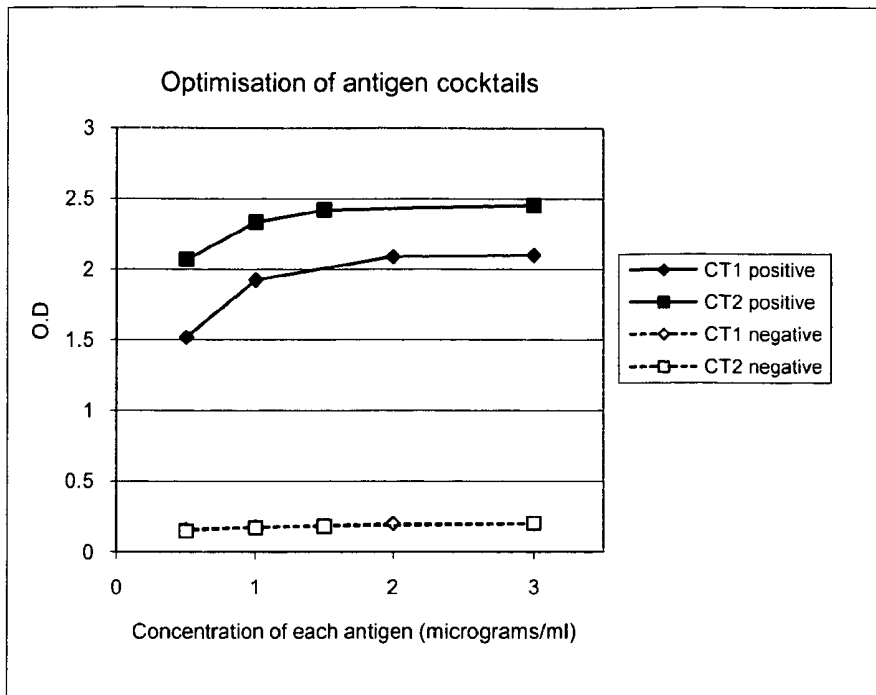


Figure 7A

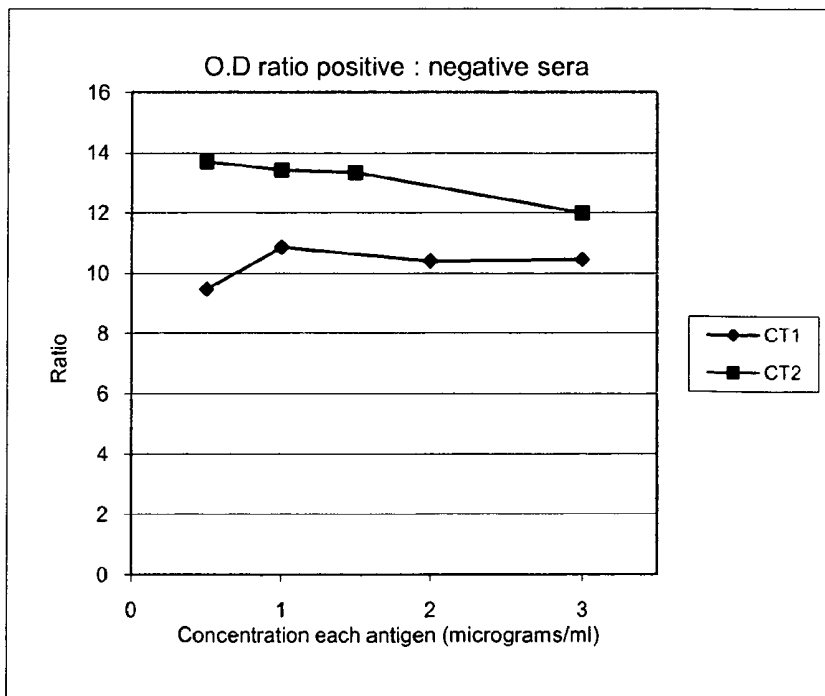


Figure 7B

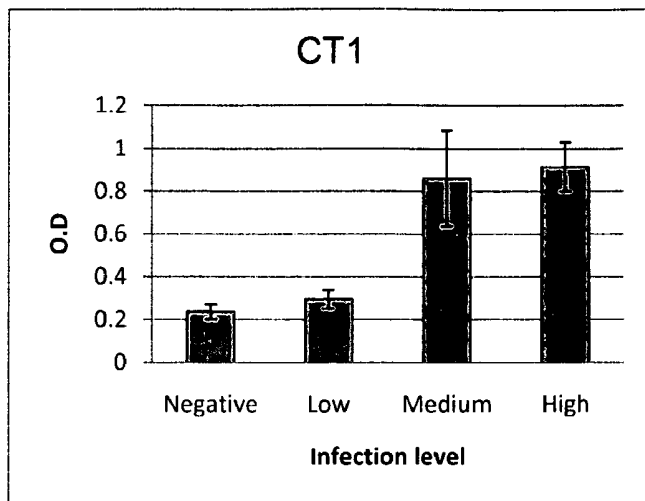


Figure 7C

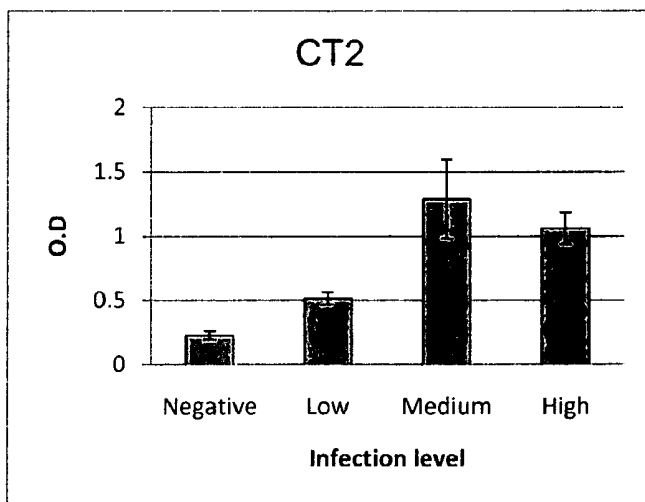


Figure 7D

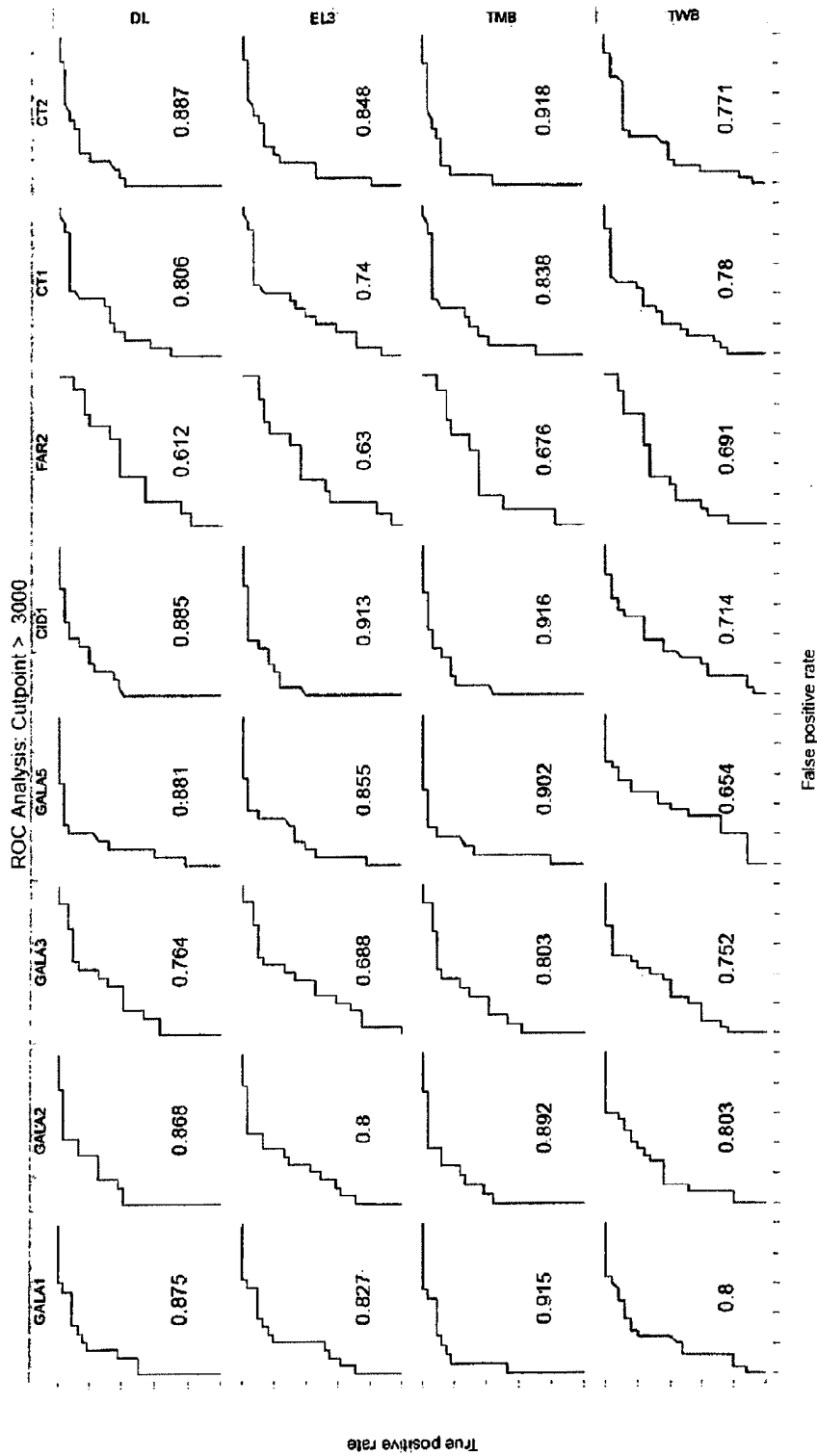


Figure 8A

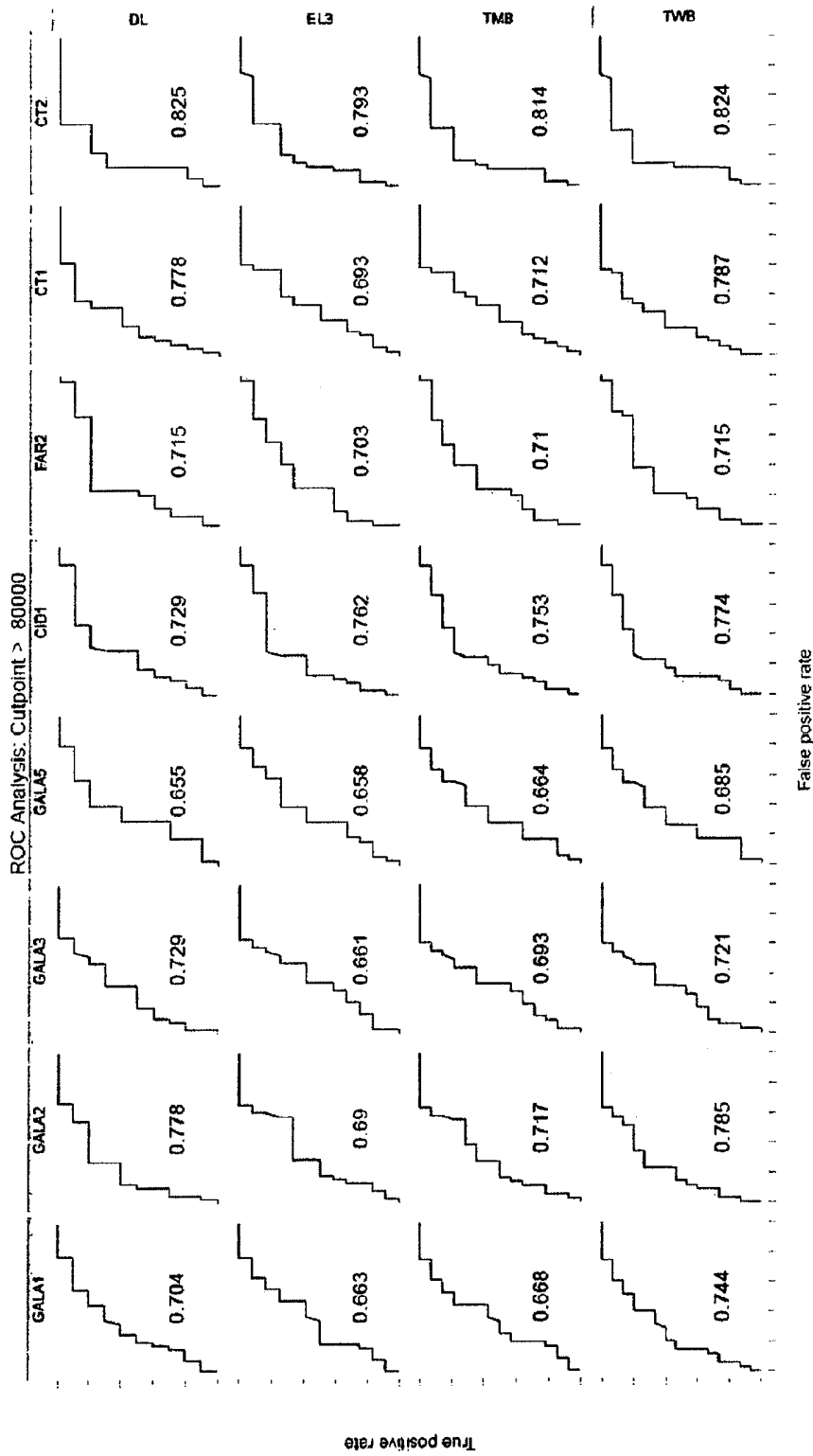


Figure 8B

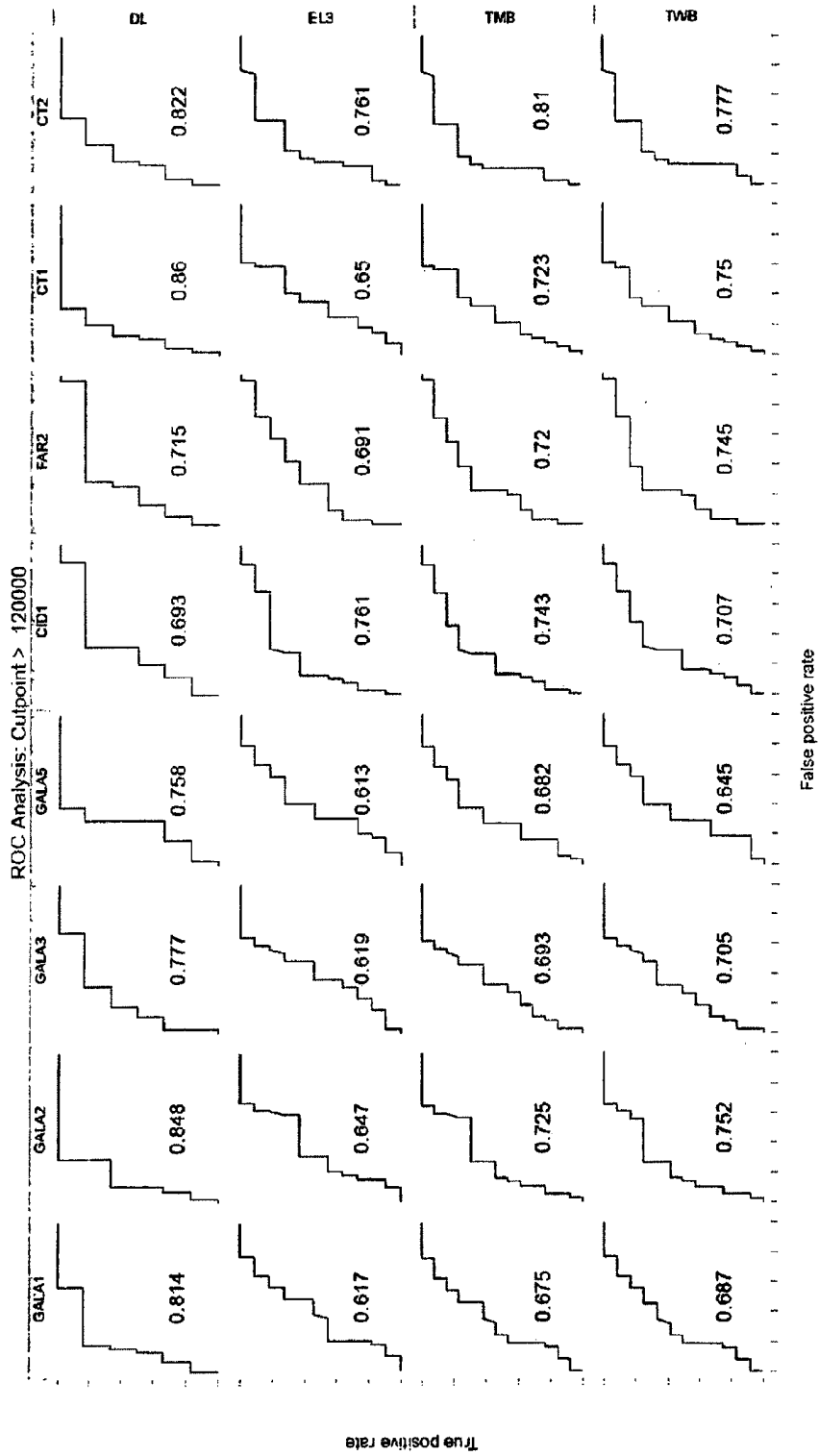


Figure 8C

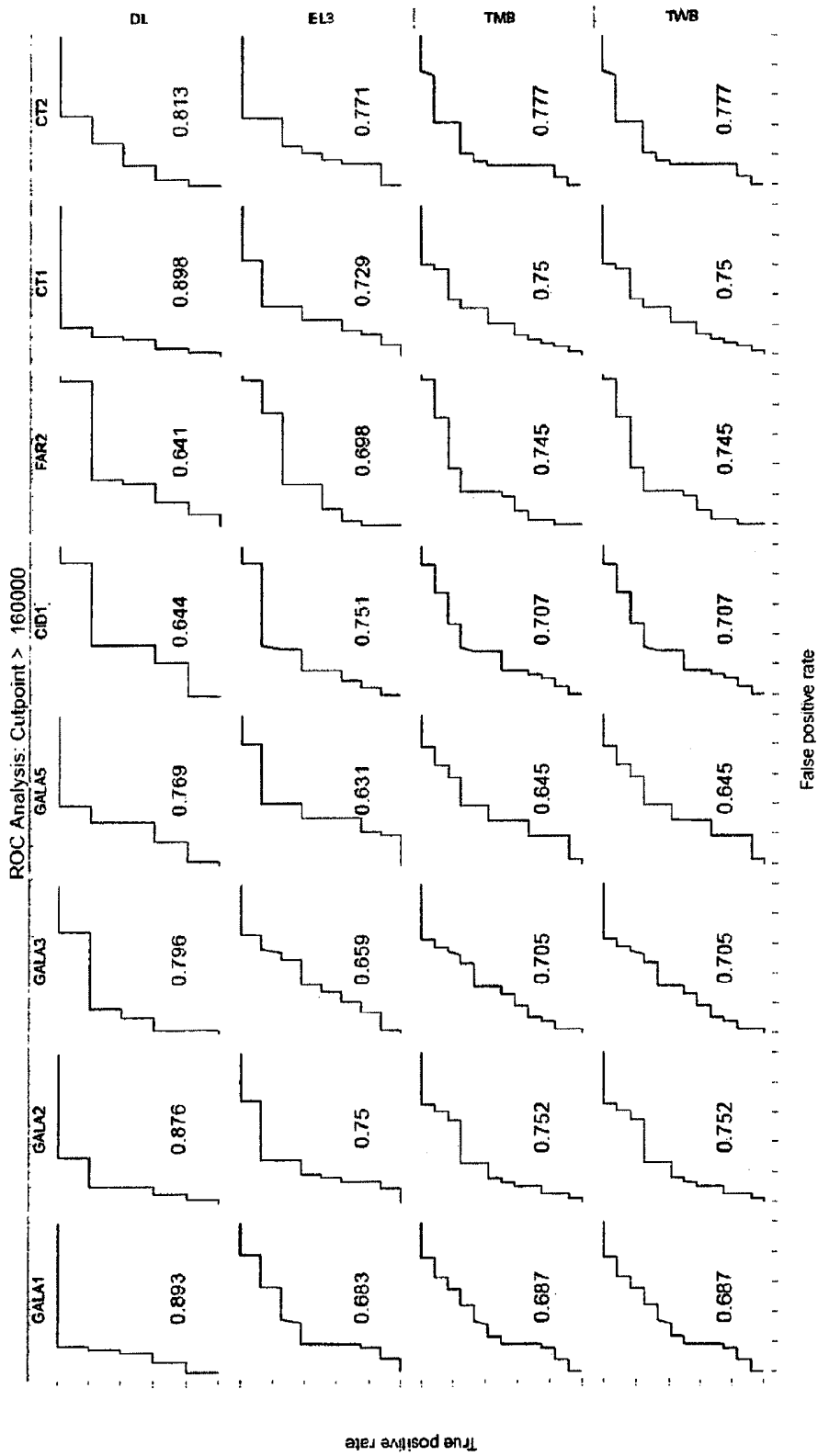


Figure 8D

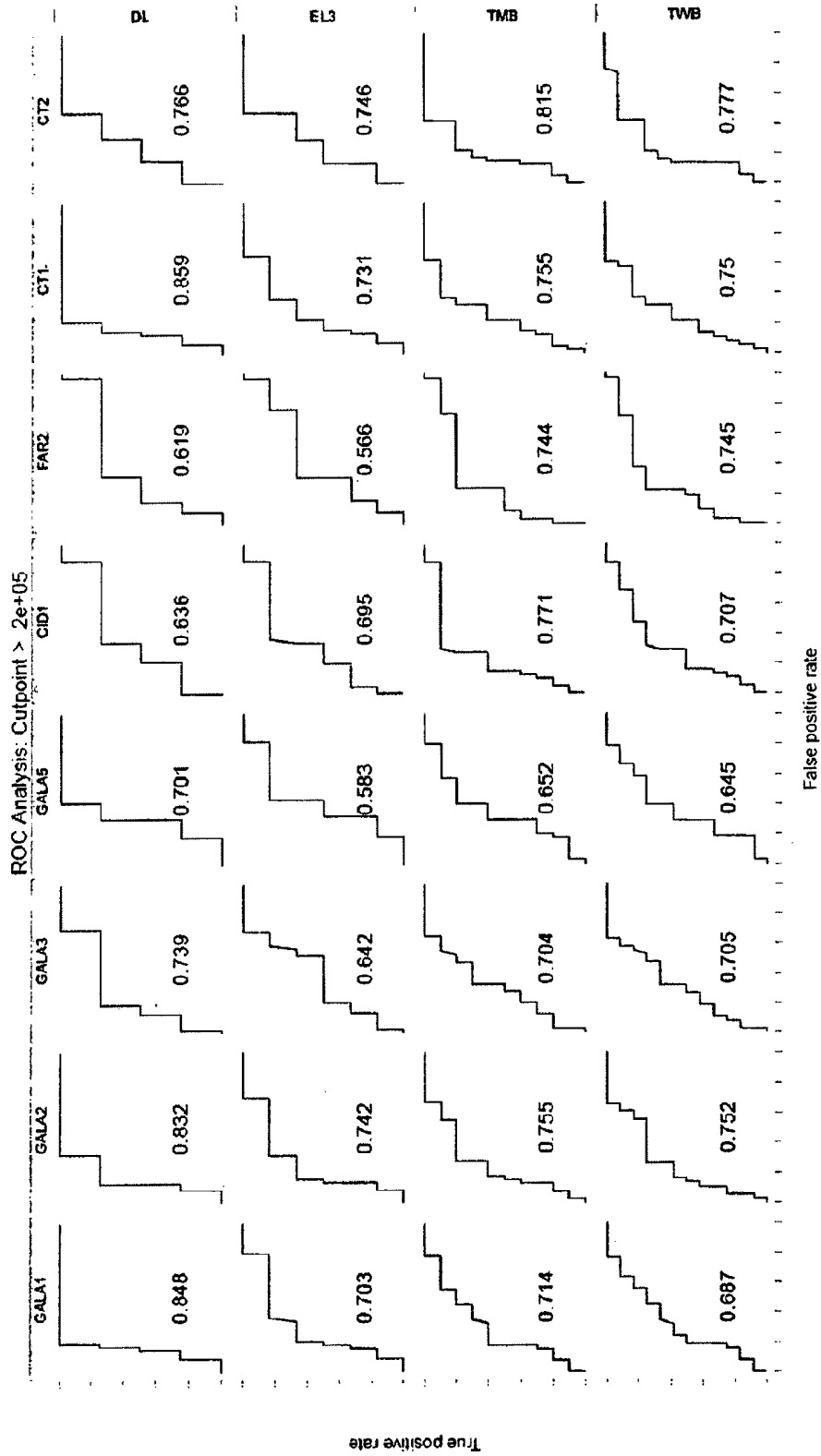


Figure 8E

EQUINE PARASITE DETECTION**CROSS-REFERENCE TO RELATED APPLICATIONS**

This application is a divisional application of U.S. patent application Ser. No. 13/260,935, now U.S. Pat. No. 8,663,939, which is a U.S. national counterpart application of PCT International Application Serial No. PCT/GB2010/000616, filed Mar. 31, 2010, which claims priority to United Kingdom Patent Application Serial Number 0905511.2, filed Mar. 31, 2009, the disclosures of all which are hereby incorporated herein by reference.

FIELD OF THE INVENTION

The present invention provides a means of detecting infections caused by parasitic nematodes belonging to the Cyathostominae group in Equine subjects and in particular in horses.

BACKGROUND OF THE INVENTION

Members of the Cyathostominae group of nematodes infect almost all grazing horses. Most horses have burdens to the order of tens of thousands of cyathostomins and usually do not exhibit clinical disease, however, in some animals, infection leads to a severe inflammatory enteropathy [15]. This disease occurs following accumulation of cyathostomin larvae that encyst and undergo inhibited development as early third larvae (EL3) in the large intestinal wall. Vast numbers of encysted larvae can accumulate and these can reactivate simultaneously to cause an inflammatory enteropathy known as larval cyathostominosis. The principal effect of this syndrome is weight loss, but horses can exhibit other signs including diarrhea, colic, subcutaneous oedema and/or pyrexia [25]. Up to 50% of animals with larval cyathostominosis die as a result of the condition [15]. This disease most commonly occurs in younger horses, however horses have a life-long susceptibility to infection and disease may occur at any age [15, 35]. Encysted larvae can persist for prolonged periods (up to two years in some cases) and it has been proposed that encystment is favoured by a variety of factors including; negative feedback from mature worms in the large intestinal lumen, a large larval challenge or a 'trickle' infection [29]. cyathostomin EL3 have limited susceptibility to several currently available anthelmintics [12, 19] and drug resistance is common, particularly with regard to benzimidazole and pyrantel compounds [17]. Moxidectin is now only drug available that has high efficacy against EL3, but for which resistance is not yet widespread. It is therefore important that the high efficacy of this anthelmintic be maintained for as long as possible.

To reduce the spread of anthelmintic resistance, it is important that only animals with moderate to high cyathostomin burdens are targeted strategically for treatment [32]. Targeted treatments can be undertaken on the basis of faecal egg counts however the latter have no value in estimating burdens of mucosal larvae. Indeed, horses with high mucosal burdens often have low or negative faecal egg counts [31] and there is no specific, non-invasive method to diagnose pre-patent cyathostomin infection. A diagnostic test for mucosal larvae would allow veterinarians to identify horses that require larvicidal anthelmintic treatments. Recently, we identified two larval antigen complexes (observed to migrate at 20 and 25 kDa by 1-dimensional SDS PAGE) that have diagnostic potential [9-11]. Significant increases in serum IgG(T) spe-

cific to these antigen complexes were observed as early as 6 weeks post infection (PI) in experimentally-infected ponies [11]. Antigens present in both complexes appeared to be specific for mucosal larval cyathostomins, indicating their utility as markers of pre-patent infection [11]. When serum IgG(T) levels were compared amongst groups of naturally- and experimentally-infected horses, there was a strong significant correlation of anti-25 kDa serum IgG(T) responses with total mucosal burden, particularly EL3 burden [10]. In naturally infected horses, IgG(T) responses to both larval complexes were significantly greater than those in uninfected individuals [10] and IgG(T) levels to both complexes were significantly higher in larval cyathostominosis clinical cases than in helminth-naïve ponies and parasite-negative horses from an abattoir [10]. These results indicate that an immunoassay based on antigens present in these complexes could ultimately be used to differentially diagnose larval cyathostominosis, or used to target horses with high mucosal burdens for treatment. The native mucosal larval preparations are extremely time-consuming to prepare and rely on a continuous source of infected mucosa. Therefore, it would be advantageous if genes encoding proteins present in these complexes were isolated and cloned and the associated proteins expressed in recombinant form.

SUMMARY OF THE INVENTION

The present invention is based upon the finding that parasitic nematodes belonging to the Cyathostominae group express proteins which can be used to diagnose, detect or identify incidences of cyathostomin infection in animals, particularly horses. Although cyathostomin infections are treatable, the range of effective drugs is rapidly diminishing and at present only moxidectin exhibits a high efficacy against the encysted cyathostomin parasite.

In order to ensure that the development of resistance to moxidectin can be delayed for as long as possible, it is essential that only animals with moderate to high cyathostomin burdens are targeted for treatment. However, the encysted larval stages of this parasite can remain undetected for months or even years eventually emerging from the intestinal wall to cause severe pathology (including symptoms of diarrhea, weight loss, colic, oedema and pyrexia); as such, it is often difficult to know whether or not a particular animal should be treated.

The inventors have identified a number of proteins that are expressed predominantly during the mucosal larval stages (i.e. the early third larval (EL3) and late third (LL3)/developing fourth (DL4) stages). These proteins are highly immunogenic and exhibit low cross-reactivity to proteins present in other helminth species.

Accordingly, a first aspect of this invention provides a method of diagnosing a cyathostomin infection, said method comprising the step of identifying a level of anti-cyathostomin larval antigen antibodies in a sample, wherein a level of anti-cyathostomin larval antigen antibodies is indicative of a cyathostomin infection.

Animals positively diagnosed as having a "cyathostomin infection" by the method provided by the first aspect of this invention may harbour high numbers of encysted cyathostomin in the gut mucosa, particularly the large intestinal wall, as such they may generate a significant immune response to cyathostomin antigens including any antigens produced by the EL3, LL3 and DL4 stages. Animals with infections of this type may otherwise be referred to as having high mucosal burdens. In other instances, positive diagnoses may indicate animals with larval cyathostominosis, an inflammatory enter-

opathy manifesting with symptoms of weight loss, diarrhoea, colic, subcutaneous oedema and/or pyrexia. Conditions of this type are often fatal if untreated.

In one embodiment, "a level" of anti-cyathostomin larval antigen antibodies may be evaluated relative to the "a level" of anti-cyathostomin larval antigen antibodies present in reference or control samples derived from healthy animals or animals not having high mucosal burdens of cyathostomin parasites or larval cyathostominosis. In this way levels and, in particular high levels, of anti-cyathostomin larval antigen antibodies, may easily be detected. Accordingly, the term "a level" may be taken to include levels of anti-cyathostomin larval antigen antibodies which are less or greater than levels of anti-cyathostomin larval antigen antibodies identified in reference or control samples.

It should be understood that in addition to providing methods in which levels of anti-cyathostomin larval antigen antibodies are detected in samples, the present invention might also be adapted to provide methods in which levels of cyathostomin larval antigens are detected in samples. Methods of this type, rather than "indirectly" diagnosing cyathostomin infections via immune responses, may provide a more direct means of diagnosing cyathostomin infections. As above, "a level" of cyathostomin larval antigens may be taken to include levels of anti-cyathostomin larval antigen which are less or greater than levels of anti-cyathostomin larval antigen identified in reference or control samples.

The cyathostomin larval antigens described/mentioned herein may be derived from larval antigen complexes having a molecular weight of about 20 to about 25 kDa. An exemplary larval antigen is obtained from the parasite cyathostomin pateratum and comprises or consists of the following amino acid sequence (designated SEQ ID NO: 1):

SEQ ID NO: 1
 MNKTLTFLTVVSAVALAQQVMDLFGEEGEEHRRHRSLLPPYLHNVSC
 EAKWEYFKIVGNRSLTFAEKRKEISEWAKKYNVVDEVASYNAYREKLEKQ
 HRKNVSELVSLPNAVKVNDLLDNENQTPRQLYVALRKLGRQNPALYRI
 VEYINVAVRLRSEEVDEQEQRRRLSALPFGDHDNLEEQDFGEQDFRYVY
 GFECARFLQNGRMFGLNTERY

The nucleic acid sequence encoding the protein provided by SEQ ID NO: 1 has also been determined and is given as SEQ ID NO: 2 below.

SEQ ID NO: 2
 Atgaacaaaacggttaacatttctcacagtcgttagtcgctagctctggc
 ccaaggtgtcatggaccttttgggtgaagagggctggaagaacatcgtc
 gtcaccatcgtcattcacttttaccaccatctccacaatgtgagctgt
 gaggctaaatgggagtagctcaaaatgtggggaacaggagtttgacctt
 tgctgagaaaaagaaagaaattagcagtgaggcaaaaaatacaatgttg
 tggatgaagtgcaagctacaatgcttacagggaaaaactcaagcaggag
 cacagaaaaacgtagcgaactgtttctgctcttccaaacgcagtgaa
 gaaagtcaatgatctcttagacaatgaaatcagactcctaggcaacttt
 acgttgcccttagaaaacttgtagacaaaatccggcactttaccgtatt
 gtcgagtacatataatgtggctgaagactaagaagtgaagaagtgatga
 gcaagaacaacgaagaaggtgtcagctctaccttttgccgaccataacg

-continued

ataatttggagagcaggacttcggtgaacaagactttcgctatgtctat
 ggctttgagtgcaagatttctccttcaaaatggaagaatgtttggact
 5 taacacagatgaaagatat

One of skill in the art will appreciate that while SEQ ID NO: 1 represents the entire coding sequence of an exemplary cyathostomin larval antigen, after removal of the signal peptide the mature antigen may comprise 206 amino acids yielding a protein having a molecular weight of approximately 25.6 kDa.

In addition, the inventors have isolated homologous antigens from other cyathostomin spp., and the amino acid sequences of these are provided below as SEQ ID NOS: 3, 5 and 7 respectively. In addition, the nucleic acid sequences encoding each of the proteins encoded by SEQ ID NOS: 3, 5 and 7 have been designated SEQ ID NOS: 4, 6 and 8 respectively and each is detailed below.

SEQ ID NO: 3
 HEELRRHHRHSLPPYLHNVSCEAKWEYFKIVGNRSLTFAEKKGKSSEWA
 KKYNVVDEVASYNAYREKLEKQHRKNVSELVSLPGAVKKNVLLDNENQ
 25 TPRQLYVALRKLGRQNPVLYRVVEFVNLVVRFRREDSDEQEQREMLSTLP
 FSENNEEQDLGEQDFQYIYGFECARFIFQNGRMFGLNTERY

The antigen encoded by SEQ ID NO: 3 was isolated from *Cylicocyclus nassatus*.

SEQ ID NO: 4
 Catgaagaacttcgctcgtcaccatcgtcattcacttttaccaccctatct
 35 ccacaatgtgagctgtgaagccaaatgggaataacttcaagattgtgggga
 acaggagcttgacttttctgctgaaaagaagggaaaaagtagcagtgaggca
 aaaaaatacaatgttggtggaagtgcaggtacaatgcctatagaga
 40 aaaaacttaagcaggagcacaggaaaaacgtagcgaactgtttctggtc
 ttcccggctgctgtgaagaaagtaaacgaactcctggataatgagaatcag
 actcctaggcaactttacgttgctctaaagaaagcttggtaaacaaaatcc
 agtactctaccgtgtgtcgagttgtgcaatttggtgtgagatttagac
 45 gtgaagattcggatgagcaagaacaaagagaatgctgtcaactttacct
 ttcagcgaataatgaagagcaggaccttggtgaacaagacttccagta
 catctatggttttgaatgtgcaagattcatcttcaaaatgggagaatgt
 50 ttggactcaacacggatagaagatat

SEQ ID NO: 5
 SCVAKWEYFKIVINRSLTFAQRKEEISKWAKKYKVEDEVASYNAYREKLEK
 55 QHRKNVSELVSNLPGAVERVKNLLDNENQTPKQLYLALRELKQNPALY
 HVVEYVNVVRLKREELDQQDQRRALSGSLFGENNDNLEEQDFGEEDFRY
 VYGFECARFILQNGRMFGLNMDRNY

The antigen encoded by SEQ ID NO: 5 was isolated from *Coronocyclus coronatus*.

SEQ ID NO: 6
 Agctgtgtggctaagtgaggtagcttcaagatcgtgatgaacaggagctc
 65 gacgtttgctcaaaagaaagaaatagcaagtgggcgaaaaataca

5

-continued

aagttgaggatgaagttgcaagctacaatgcttatagagaaaaactcaag
 cagcagcacaggaaaaacgtagcgaactgtttctagtcttcccggtgc
 aatggaagagtgaaacaaacttttgacaatgaaaaccagaccctaaagc
 aactttaccttgccctacgagaacttgcaaaaaaatccggcactttac
 catgttgtcgagtatgtcaatgtggtgtgagacttaaacgagaagaatt
 ggtgaaacaagatcaatgaagagcgctgtcggttcactttttggcgaga
 ataacgacaatctagaagagcaggactttggtgaaagaagactttcgctat
 gtctatgggtttgaatgtgcaagattcctcctcaaaatggaagaatgtt
 tgggtctaacaatggataggaattat

SEQ ID NO: 7

GEEDREHRRHRHSLPPYLHNVCVAKWEYFRIVGNRSLTFAEKKKEI
 SEWAKKYNVLDEVASYNAYREKLQEHKRVSELVSDLPKAVKVNDDLLD
 NENQTPRQLYVALRELGRQNPTLYRIVEYINVAVRRRSEELDEQEGRRL
 SALPFGDNNDNLEEQDFGEQDFRYVYGFECARFLLQNGRMFGLNTERD

The antigen encoded by SEQ ID NO: 7 was isolated from *Cyathostomum catinatum*.

SEQ ID NO: 8

Gaggatcgtgaagaacatcgccgtcaccatcgctcattcactcttgccacc
 atatctccacaactgagctgtgtggccaaatgggaatactttagaattg
 tggggaacaggagtttaacgtttgctgagaaaaagaaagaattagcgag
 tgggcaaaaaatacaatgttctggatgaagtgaagctacaatgctta
 tagggaaaaactcaagcaggagcacagaaaaacgttagcgaactgttt
 ctgatcttcccaaggcagtaaagaaagctcaacgatcttctagacaatgaa
 aatcagactcctaggcaactttatgttgcctctagagagcttggtagaca
 aaatccgacactttaccgtatgtcgagtacatcaatgtggtgtgaagc
 gaagaagtgaagaactggatgagcaagaacaaggaagaaggctgcagct
 ttacctttcggcgacaacaacgataatttggagagcaggacttcggtga
 acaagactttcgctatgtctacggctttgagtggtgcaagatttctcctc
 aaaatggaagaatgttcggactcaacacagatgaaagagat

SEQ ID NOS 9-35 have been translated into amino acid sequences by removing the non-coding regions (introns) identified by comparison with the coding sequence of Gala-1.

The antigen encoded by SEQ ID NO: 9 was isolated from *Cylicocyclus ashworthi*.

SEQ ID NO: 9

ATGAACAAAACGTTAACAATTTCTCACAGTCGTTAGTGCCGTAGTTCTGGC
 CCAAGGTGTCATGGACCTTTTGGTGAAGAGGGTCGTGAAGAACATCGCC
 GTCACCATCGTCATTCACTCTTACCACCATATCTCCACAACGTGAGCTGT
 GTGGCTAAATGGGAGTACTTCAAAATTTAGGGAACAGGAGTTTAAACGTT
 TGCTGAGAAAAAGAAGAAATTAGCCAGTGGGCAAAAAATACAATGTTG
 TGGTAAGCTTTTCTGAATTAATGTAATAACACTCGCATGCTGGCCTTTT
 AGGATGAAGTTGCAAGCTACAATGCTTACAGGAGAAAACCTCAAGCAGGAG

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CACAGAAAAACGTTAGCGAACTGTTTTCTGCTCTTCCAAACGAGTAAA
 GAAAGTCAACAATCTTCTAGACAATGAAAAATCAGACTCTTAGGCAACTTT
 5 ACGTTGCCCTTAGAGAACTTGGTAGACAAAAATCCGGCAGTAAGTAGAAAG
 AGCTGCACTCTGGGCTTAATAAAACAAATATTTAAGCTTTACCCTATT
 GTCGAGTACATCAATGTGGCTGTAAGACGAAGAAGTGAAGGACTGGATGA
 10 GCAAGAACAACGAAGAAAGCTATCAGCTTTACCTTTCCGGCACAACAACG
 ATAATATGGAAGAGCAGGACTTCGGTGAACAAGACTTTCGCTATGTCTAC
 GGCTTTGAGTGTGCAAGATTTCTCCTTCAAAAATGGAAGAATGTTTGGGCT
 15 CAACACAGATGAAAGAGATTAGCAAGAATCAATGTAGTTCAAAGCGGT
 AGAGTTTGAGCTGCAAACTCAGCATGCCATCATCACCTCTCT

SEQ ID NO: 10 (i.e., SEQ ID NO: 9 translated)

MNKLTFLTVVSAVVLAQGVMDLFGEEGREHRRHRSLLPPYLHNVC
 20 VAKWEYFKIVGNRSLTFAEKKKEISQWAKKYNVDEVASYNAYREKLQBE
 HRKNVSELVSLPNAVKVNDDLNENQTPRQLYVALRELGRQNPLALYRI
 VEYINVAVRRRSEGLDEQEQRRLSALPFGDNNDNMEEQDFGEQDFRYVY
 25 GFECARFLLQNGRMFGLNTERD

The antigen encoded by SEQ ID NO: 11 was isolated from *Cyathostomum catinatum*.

SEQ ID NO: 11

ATGAACAAAACGTTAACAATTTCTCACAGTCGTTAGTGCCGTAGTTCTGGC
 TCAAGGTGTCATGGACCTTTTGGTGAAGAAGCCGTGAAGAACATCGCC
 35 GTCACCCTGTCATTCACTCTTGCCACCATATCTCCACAACGTGAGCTGT
 GTGGCTAAATGGGAATACTTCAAGAAATGTTGGGGAACAGGAGTTTGACGTT
 TGCTGAGAAAAAGGAAGAGATTAGCGAGTGGGCAAAAAGTACAATGTTG
 40 TGTAAGCTTTTCTGAATTGATGTAATACTCGCATGCTGGCCTTTTT
 AGGATGAAGTTGCAAGCTACAATGCTTACAGGAAAAACTCAAGCAGGAG
 CACAGAAAAACGTTAGCGAACTGTTTTCTGCTCTTCCAAACGAGTAAA
 GAAAGTCAACGATCTTCTAGACAATGAAAAATCAGACTCTTAGGCAACTTT
 45 ACGTTGCCCTTAGAGAACTTGGTAGACAAAAATCCGGCAGTAAGTCGAAAG
 AGCTGCACTCTGGGCATAAGTAAAAAAGTATTTTAGCTTTACCCTATT
 TGTGGAGTACATCAATGTGGCTGTAAGACTAAGAAGTGAAGAAGTGGATG
 AGCAAGACAACGAAGAAGGCTATCAGCTTTACCTTTTGGTGACCATAAC
 GATAATATGGAAGAGCAGGACTTGGTGATCAAGACTTTCGCTATGTCTA
 50 CGGCTTTGAGTGTGCAAGATTTCTCCTTCAAAAATGGAAGAATGTTTGGAC
 55 TTAACACAGATGAAAGATATTAGTAAAAATTAACGTAGCTCAAAGCGGT
 AGAGTTTGAGCTGCAAACTCAGCATGCCATCATCACCTCTCT

SEQ ID NO: 12 (i.e., SEQ ID NO: 11 translated)

MNKLTFLTVVSAVVLAQGVMDLFGEEGREHRRHRSLLPPYLHNVC
 60 VAKWEYFRIVGNRSLTFAEKKKEISEWAKKYNVDEVASYNAYREKLQBE
 HRKNVSELVSLPNAVKVNDDLNENQTPRQLYVALRELGRQNPLALYRI
 VEYINVAVRLRSEEVDEQEQRRLSALPFGDNDNMEEQDFGDQDFRYVY
 65 GFECARFLLQNGRMFGLNTERD

The antigen encoded by SEQ ID NO: 13 was isolated from *Cylicostephanus goldi*.

SEQ ID NO: 13

ATGAACAAAACGTTAACATTTCTCACAGTCGTTAGTGCCGTAGTCCTGGC
 TCAAGGTGTCGTGGACCTTTTTGGTGAAGAGGGTCGTGAAGAACATCGCC
 GTCACCATCGTCATTCACTCTTACCACCATATCTCCACAACGTCAGCTGT
 GTGGCTAAATGGGAATACTTCAAAAATGTGGGGAAATAGGAGTTGACATT
 TGCTGAGAAAAAGAAAGAAATAGCGAGTGGGCTAAAAATAACAATGTAG
 TGGTAAGCTTTTTTGTACTTGATGTAATGCACCTCGTATGCCGGCCCTTTT
 AGGATGAAGTGCAGGTACAATGCTTATAGAGAAAACTTAAGCAGGAA
 CACAGGAAAACGTCAGCGAACTGTTTCTGATCTTCCCAACGCAGTAAA
 GAAAGTGAATGATCTCTGGACAATGAGAATCAAACCTCCTAGGCAACTTT
 ACATTGCCCTCAGAGAATTTGGTAGACAAAATCCAGAAGTAAGTTGAAAG
 TGCTGCAATTTTAGGCTTAGATAAAACAGTTGTTAAGCTTTACCCTGTT
 GTCGAGTTTATCAATGTGGCTGTAAGAATAAGACGTGAAGATTTGGATGA
 GCAAGAACAACGAACAGGCTGTCAACTTTACCTTTTGGCGACAACAACG
 ACAATTTCCAAGAGCAAGACTTCGGTGAACAAGACTTTTCGCTATGCTAT
 GGCTTTGAGTGTCAAGATTTCTCCTTCAAAAATGGAAGAATGTTGGACT
 TAACACGGATAGAAAGATAC

SEQ ID NO: 14 (i.e., SEQ ID NO: 13 translated)
 MNKTLTFLTVVSAVFLAQGVVDFGEEGREHRRHRHSLPPLYLHNVC
 VAKWEYFKIVGNRSLTFAEKKKEISEWAKKYNVDEVARYNAYREKLKQE
 HRKNVSELVSDLPNAVKKVNDLLDNENQTPRQLYIALRELGRQNPPLYRV
 VEFINAVRIRREDLDEQEQRTLSTLFPDNDNDFEQDFGEQDFRYVY
 GFECARFLLQNGRMFGLNTDRRY

The antigen encoded by SEQ ID NO: 15 was isolated from *Cylicostephanus goldi*

SEQ ID NO: 15

ATGAACAAAACGTTAACATTTCTCACAGTCGTTAGTGCCGTAGTCCTG
 GCCCAAGGTGTCATGGACCTTCTTGATGAAGAGGCTCGTGGAGAGCAT
 CGCCGTACCATCGTCATTCACTCTTACCACCATATCTCCACAACGTC
 AGCTGTGTGGCTAAATGGGAATACTTCAAAAATGTGGGGAACAGGAGT
 TTGACGTTTGCTGAGAAAAAGAAAGAAATAGCGAGTGGGCAAAAAA
 TACAACGTTGTGTAAGCTTTTGTGACTCGATGTAGATACCCAGATA
 TTCTAGATACCCATGCTGGCCTTTTTAGGATGAAGTTGCAAGCTACAA
 TGCTTATAGAGAAAACTCAAGCAGGAACACAGGAAAAACGTTAGCGA
 ACTTGTATCTGATCTTCCCAATGCAGTGAAGAAAGTGAATGATCTCCT
 GGACAATGAGAATCAAACCTCCTAGGCAACTTTACGTTGCCCTCAGAGA
 ACTTGGTAGACAAAATCCAGCAGTAAGTTGAAAGTGTGCAATTTGAG
 GCTTAGATAAAACAGTTGTTAAGCTTTTACCGTGTGTCGAGCTCATC
 AATGTGGCTGTAAGATTAAGACGTGAAGATTTGGATGAGCAAGAACAA
 CGAACAAGGCTGTCAACCTTACCTTTTGGCGACAACAACAATTTTC

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GATGAGCAGGACTTCGGTGAACAAGACTTTCGCTATGTCTATGGCTTT
 GAGTGTGCAAGATTTCTCCTTCAAAAATGGAAGAATGTTTGGACTTAAC
 5 ACGGATAGAAGATACTAGTAAGAGTCAACTGTAGCTCAAAGTGGTTCC
 AGCTACGAACAGCATGCCATCATCACCTCCT

SEQ ID NO: 16 (i.e., SEQ ID NO: 15 translated)
 MNKTLTFLTVVSAVFLAQGVMDLDEEARGEHRRHRHSLPPLYLHNVC
 10 SCVAKWEYFKIVGNRSLTFAEKKKEISEWAKKYNVDEVASYNAYREK
 LKQEHKRVSELVSDLPNAVKKVNDLLDNENQTPRQLYVALRELGRQN
 PALYRVVELINAVRLRREDLDEQEQRTLSTLFPDNDNDFEQDFG
 15 EQDFRYVYGFECARFLLQNGRMFGLNTDRRY

The antigen encoded by SEQ ID NO: 17 was isolated from *Cylicostephanus goldi*

SEQ ID NO: 17

ATGAACAAAACGTTAACATTTCTCACAGTCGTTAGTGCCGTGTCCTG
 GCGCAAGGTGTCATGGCCCTATTTGGTGAAGAGAGTGTGAAGAACAC
 25 CGCCGTACCATCGTCATTCACTCTTACCACCATATCTCCACAACGTC
 AGCTGTGTGGCTAAATGGGAGTACTTCAAAAATGTGGGGAACAGGAGT
 TTGACGTTTGCTGAGAAAAAGAAAGAAATCAGCGAGTGGGCTAAAAA
 30 TACAATGTTGTGTAAGCTTTTTGACTTGATGTAATGCACCTCGCAT
 GCCGGCCTTTATAGGATGAAGTTGCAAGCTACAATGCTTATAGAGAAA
 AACTCAAGCAGGAACACAGGAAAAACGTTAGCGAACTTGTCTGATC
 35 TTCCCAACGCAGTAAAGAAAGTCAAGCATCTTTTGGACAACGAAAAATC
 AGACTTCTAGGCAACTTTATGTTGCACTCAGAGAACTTGGTAGACAAA
 ATCCGGCAGTAAGTTGAAGAGGCTCCAATTTTGGGCTCAAGCAAAAAT
 40 AATTATTTTAGCTATACCGTGTGTCGAGTATATCAATGTGGCTGTGA
 GATTAAGACGAAAAGAACAGGATGAACAAGAACGACAAGGAACGCTGT
 CAGCTCTACCTTTTGGCGAGAATAACGACAATTTGGAAGAGCAGGACT
 TTGGTGAACAAGACTTTCGCTATGTCTATGGCTTTGAGTGTGCAAGAT
 45 TTCTCCTTCAAAAATGGAAGAATGTTTGGACTCAACACGGATAGAAGAT
 ACCAGTAAGAGTCAACTGTAGCTCAAAGTGGGTTTGGACTACGAAACAG
CATGCCATCATCACCTCCT

SEQ ID NO: 18 (i.e., SEQ ID NO: 17 translated)
 MNKTLTFLTVVSAVFLAQGVMLFGEESREHRRHRHSLPPLYLHNVC
 SCVAKWEYFKIVGNRSLTFAEKKKEISEWAKKYNVDEVASYNAYREK
 55 LKQEHKRVSELVSDLPNAVKKVSDLLDNENQTSRQLYVALRELGRQN
 PAVYRVVEYINAVRLRREKQEQERQGLSALPFGENNDNLEEQDFG
 EQDFRYVYGFECARFLLQNGRMFGLNTDRRY

The antigen encoded by SEQ ID NO: 19 was isolated from *Cylicostephanus longibursatus*

SEQ ID NO: 19

ATGAACAAAACGTTAACATTTCTCACCGTCGTCATGCCGTAGTCCTG
 65 GCCCAAGGTGTCATGGACCTTTTTGGTGAAGAGGGTCGTGAAGAACAT

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CGCCGTCACCATCGTCATTCACCTTACCACCATATCTCCACAATGTG
 AGCTGTGTGGCTAAATGGGAATACTTCAAAATTTGTGGGGAACAGGAGT
 TTGACGTTTGTCTGAGAAAAAGGAAGAAATAGCAAGTGGGCAAAAAA
 TACAATGTTGTGTACGCTTTTGTAAACCCGATAATATACTCTCGCA
 TACTGGCCGTTTCAGGATGAAGTTGCAAGCTACAGTGTTCGACGGAA
 AAGCTTAAGCAGGAACACAGGAAAAACGTTAGCGAAATGTTTCTAAT
 CTTCCTCAATGCAGTGAAGAAAGTAAACGATCTTTTGGACAATGAAAA
 CAGACCCCAAGCAACTTTACGTTGCCTTCAGAAAACTTGGTAAACAA
 AATCCGGCAGTAAGTTGAAAGAGCTGCAATTTTGGGTTTGAGGAGAAA
 AAATATTTTAGCTTTATCGTGTGTGCGAGTATATCAATGTGCTTGTG
 AGACTAAGACGTGAAGAATTTGATGAAGATCAGCGAAGATCGCTGTCA
 GCTTTACCTTTTGGCGACAATAACGACGATTTTGAAGAGCAGGACTTT
 GGTGAACAGGACTTTCGCTATATCTATGGCTTTGAGTGTGCAAGATTT
 ATCCTTCAAAATGGAAGAAATGTTTCGACTCAACACGGATAGAAGATAT
 TAGTAAGAGTCAACTGTAGCTCGAGGTTTGTAGCTACGAACTGCATGC
CATCATCACCTCCT

SEQ ID NO: 20 (i.e., SEQ ID NO: 19 translated)
 MNKTLTFLTVVYAVVLAQGVMDLFGEEGREGREHRRHRSLLPPYLHNV
 SCVAKWEYFKIVGNRSLTFAEKKEEISKWAKKYNVDEVASYSACREK
 LKQEHKKNVSEIVSNLPAVKKVNDLLDNENQTPRQLYVAFRKLKQKQ
 PALYRVVEYINVLVRLRREEFDEQRRSLSALPFGDNNDLLEEQDFGE
 QDFRYIYGFECAFILQNGRMFGLNTRRY

The antigen encoded by SEQ ID NO: 21 was isolated from
Cylicocyclus insigne.

SEQ ID NO: 21
ATGAACAAAACGTTAACATTTCTCACCGTCGTCGTGCGTAGTCCTG
 GCCCAAGGTGTCATGGACCTTTTGGTGAAGAAGGTCGTGAAGAACAT
 CGCCGTCACCATCGTCATTCACCTTACCACCATATCTCCACAATGTG
 AGCTGTGTGGCTAAATGGGAATACTTCAAAATTTCTGGGGAACAGAAGT
 TTGACGTTTGTCTGAGAAAAAGGAAAAATCAGCGAGTGGGCAAAAAAG
 TACAATGTTGTGTACGCTTTTGTAACTCCGATAATATACTCCGCA
 TGCTGGCCGTTTCAGGATGAAGTTGCAAGCTACAATGCTTGCAGGGAA
 AAGCTTAAGCAGGAACACAGGAAAAACGTTAGCGAAATGTTTCTAAT
 CTTCCTCAATGCAGTAAAGAAAGTAAACGATCTTTTGGACAATGAAAA
 CAGACTCCAGGCAACTTTACGTTGCCTTCAGAAAACTCGTAAACAA
 AATCCGCCAGTAAGTTGAAAGAGCTGCAACTTTGGGTTTAAAGGAAAA
 AACTATTTTAGCTTTTACCGCTTGTGCGAGTATATCAATGTGGTGTGA
 GACTAAGACGTGAAGAATCTGATGAAGAACAACGAAGACGCTGTGAG
 CTTTACCTTTTGGCGACAATAACGACAACCTTGAAGAGCAAGACTTTG
 GTGAAGAAGACTTTCGCTATATTTATGGCTTTGAGTGTGCAAGATTTA
 TCCTTCAAAATGGGAGAATGTTTCGACTCAACACGGATAGAAGATATC

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AGTAAGAGTCAACTGTAGCTTAAAAGTTTGAGCTACGAACAGCATGCC
ATCATCACCTCCT
 5 SEQ ID NO: 22 (i.e., SEQ ID NO: 21 translated)
 MNKTLTFLTVVCAVVLQGVMDLFGEEGREGREHRRHRSLLPPYLHNV
 SCVAKWEYFKILGNRSLTFAEKKEKISEWAKKYNVDEVASYNACREK
 LKQEHKKNVSEIVSNLPAVKKVNDLLDNENQTPRQLYVALRKLKQKQ
 10 PPLYRVVEYINVVRLRREESDEEQRRTLSALPFGDNNDLLEEQDFGE
 EDFRYIYGFECAFILQNGRMFGLNTRRY

The antigen encoded by SEQ ID NO: 23 was isolated from
 15 *Cylicostephanus longibursatus*.

SEQ ID NO: 23
ATGAACAAAACGTTAACATTTCTCACCGTCGTCATGCCGTAGTCCTG
 20 GCCCAAGGTGTCATGGACCTTTTGGTGAAGAGGGTCTTGAAGAACAT
 CGCCGTCACCATCGTCATTCACCTTACCACCATATCTCCACAATGTG
 AGCTGTGTGGCTAAATGGGAATACTTCAAAATTTCTGGGGAACAGGAGT
 25 TTGACGTTTGTCTGAGAAAAAGGAAAAATCAGCGAGTGGGCAAAAAAG
 TACAATGTTGTGGTACGCTTTTGTAACTCAGTATAATATATCTCCGCA
 TACTGGCCGTTTCAGGATGAAGTTGCAAGCTACAATGCTTGCAGGGAA
 30 AAGCTTAAGCAGGAACACAGGAAAAACGTTAGCGAAATGTTTCTAAT
 CTTCCTCAATGCAGTGAAGAAAGTAAACGATCTTTTGGACAATGAAAA
 CAGACCCCAAGCAACTTTACGTTGCCTTCAGAAAACTTGGTAAACAA
 35 AATCCGGCAGTAAGTTGAAAGAGCTGCAATTTTGGGTTTGAAGAAAA
 AAATATTTTAGCTTTTATCGTGTGTGCGAGTATATCAATGTGCTTGTG
 AGACTAAGACGTGAAGAATTTGATGAAGATCAGCGAAGATCGCTGTCA
 40 GCTTTACCTTTTGGCGACAATAACGACGATTTTGAAGAGCAGGACTTT
 GGTGAACAGGACTTTCGCTATATCTATGGCTTTGAGTGTGCAAGATTT
 ATCCTTCAAAATGGAAGAAATGTTTCGACTCAACACGGATAGAAGATAT
 TAGTAAGAGTCAACTGTAGCTCAAGGTTTGTAGCTACGAACTGCATGC
 45 CATCATCACCTCCT

SEQ ID NO: 24 (i.e., SEQ ID NO: 23 translated)
 MNKTLTFLTVVYAVVLAQGVMDLFGEEGLEHRRHRSLLPPYLHNV
 50 SCVAKWEYFKILGNRSLTFAEKKEKISEWAKKYNVDEVASYNACREK
 LKQEHKKNVSEIVSNLPAVKKVNDLLDNENQTPRQLYVALRKLKQKQ
 PALYRVVEYINVLVRLRREEFDEQRRSLSALPFGDNNDLLEEQDFGE
 55 QDFRYIYGFECAFILQNGRMFGLNTRRY

The antigen encoded by SEQ ID NO: 25 was isolated from
Cylicocyclus nassatus.

SEQ ID NO: 25
ATGAACAAAACGTTAACATTTCTCATCGTCGTTAGTCCGTAGTCCTG
 60 ACCCAAGTGTATGGACTTTTTCGATGAAGACGGTCTGGAAGAACAT
 CGCCGTCATCATCGTCATTCCTTTTACCACCGTATCTCCACAATATG
 65 AGCTCGTGGCCAAATGGGAATACTTCGAGATTGTGGGGACAGGAGT

11

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CTGACGTTTGTCTGAAAAGAAGGAAAAATCGGCGAGTGGGCTAAAAAA
 TACAATGTTGTGGTAAGATTTTGTAACTCTATGTAAGATAACCCCGT
 ACGTCGCCCTGTTTAGGATGAAGTTGCAAGCTACAATGCTTATAGAGA
 AAAACTAAAGCAGGAGCACAGGAAAAACGTTAGCGAGCTTGTCTCTGG
 TCTTCCCAATGCTGTGAAGAAAAATAACGAACCTTTAGACAATGAAAA
 TCAGACTGTTAGGCAACTTTATGTTGCTTTAAGAGAACCTGGTAAACA
 AAATCCAGCAGTAAGTTAAAAGAAGTGCAATTTTGGGCTTAACTAATG
 AGACAATTTTAGCTCTACCGTGTGTGCGAGTATATCAATGTGGTTGTG
 AGACTTAGACGTGAAGATTTGGATGAGCAGGAACAACAGAGAACGCTG
 TCAACCCACCTTTCGGCGAGAATAACGAAGAGCAAGACTTTGGTGAA
 CAAGACTTTCACTATACTATGGTTTTGAGTGTGCCAGATTTCATCCTT
 CAAAATGGAAGAATGTTTGGACTTAACACGGATAGAAGATATTAGTAA
 GAGTTAACTGCAGCTCAATGTGATAGAGATTGAGCCACAACCCAACT
GCCATCATCACCTCCT

SEQ ID NO: 26 (i.e., SEQ ID NO: 25 translated)
 MNKTLTFLIVVSAVVLTSVMDFFDEEDGREHRRHHRHSLPPLYLHNM
 SCVAKWEYFEIVGDRSLTFAEKKEKIGEWAKKYNVVEVASYNAYREK
 LKQEHRKNVSELVSGLPNAVKKINELLDNENQTVRQLYVALRELKQON
 PALYRVVEYINVVRLRREDLDEQEQRRLSTSPFGENNEEQDFGEQD
 FHYIYGFECARFILQNGRMFGLNTDRRY

The antigen encoded by SEQ ID NO: 27 was isolated from
Cylicocyclus nassatus.

SEQ ID NO: 27
ATGAACAAAACGTTAACATTTCTCATCGTCGTTAGTGCCATAGTCTCG
 GCCCAAAGTGTATGGACTTTTTTCGATGAAGAAGGTCGTGAGGGACAT
 CGCCGTCATCATCGTCATTTTACCACCATATCTCCACAATATG
 AGCTCGCTGGCCAAATGGGAATACTTCGAGATTGTGGGGACAGGAGT
 CTGACGTTTGTCTGAAAAGAAGGAAAAATCGGCGAGTGGGCTAAAAAA
 TACAATGTTGTGGTAAGATTTTGTAACTCCATGTTAGGATACCTCCGC
 ACGTCGCCCTGTTTAGGATGAAGTTGCAAGCTACAATGCTTATAGAGA
 AAAACTAAAGCAGGAGCACAGGAAAAACGTTAGCGAGCTTGTCTCTGG
 TCTTCCCAATGCTGTGAAGAAAGTAAACGAACCTTTAGACAATGAAAA
 TCAGACTGTTAGGCAACTTTATGTTGCTTTAAGAGAACCTGGTAAACA
 AAATCCAGCAGTAAGTTAAAAGAAGTACAATTTTGAGCTCAACTAATG
 AGACAATTTTAGCTCTACCGTGTGTGCGAGTATATCAATGTGGTTGTG
 AGACTTAGACGTGAAGATTCGGATGAGCAGGAACAACGAAGACTCTG
 TCAACCTCACCTTTCGGCGAGAATAACGAAGCAAGATTTTGGTGAA
 CAAGATTTTCACTATACTATGTTGTTTTGAGTGTGCAAGATTTCATCCTT
 CAAAATGGAAGAATGTTTGGACTCAATACGGATAGAAGATAT

SEQ ID NO: 28 (i.e., SEQ ID NO: 27 translated)
 MNKTLTFLIVVSAIVLAQSVMDFFDEEGREGHRRHHRHSLPPLYLHNM
 SCVAKWEYFEIVGDRSLTFAEKKEKIGEWAKKYNVVEVASYNAYREK

12

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LKQEHRKNVSELVSGLPNAVKKVNEELLDNENQTVRQLYVALRELKQON
 PALYRVVEYINVVRLRREDSDEQEQRRLSTSPFGENNEEQDFGEQD
 5 FHYIYGFECARFILQNGRMFGLNTDRRY

The antigen encoded by SEQ ID NO: 29 was isolated from
Cylicocyclus nassatus.

10
 SEQ ID NO: 29
ATGAACAAAACGTTAACATTTCTCATCGCCGTTAGTGCCATAGTCTCG
 GCCCAAAGTATGGACTTTTTTCGATGAAGACGGTCGTGAAGAACATCGC
 15 CGTCATCATCGTCATTTTACCACCATATCTCCACAATATGAGC
 TCGCGCGCCAAATGGGAATACTTCGAGATTGTAGGGGACAGGAGTCTG
 ACGTTTGTCTGAAAAGAAGGAAAAATCGGCGAGTGGGCTAAAAATAC
 20 AATGTTGTGGTAAGATTTTGTAACTCCATGTAAGATACCCCTCCATG
 TCGTCCCCTTTAGGATGAAGTTGCAAGCTACAATGCTTCAGAGAAAA
 ACTGAAGCAAGAGCACAGGAAAAACGTCAGCGAGCTTGTCTCTGGTCT
 25 TCCCAATGCTGTGAAGAAAGTAAACGAACCTTTAGACAATGAAAAATCA
 GACTGTTAGGCAACTTTATGTTGCTTTAAGAGAACTTGGTAAACAAAA
 TCCAGCAGTAAGTTGAAGAAGTGCATTTTGGGCTTAACTAACGAGAC
 30 AATTTTAGCTCTACCGTGTGTGCGAGTATATCAATGTGGCTGTGAGAC
 TTAGACGTGAAGATTTCGGATGAGCAGGAAAAACGAAGAACGCTGTCAA
 CCTCACCTTTCGGCGAGAATAACGAAGAGCAGGACCTTGGTGAACAAG
 35 ATTTTCACTATACTATGGCTTTGAGTGTGCAAGATTTCATCCTTCAA
 ATGGAAGAATGTTTGGACTTAACACGGATAGAAGATATTAGTAAAATT
 TGACTGCAGCTCAAAGTGGTAGAGATTGAGCTACCAACCAACATGCC
 40 ATCATCACCTCCT

SEQ ID NO: 30 (i.e., SEQ ID NO: 29 translated)
 MNKTLTFLIVSAIVLAQSMDFDEEDGREHRRHHRHSLPPLYLHNM
 CAKWEYFEIVGDRSLTFAEKKEKIGEWAKKYNVVEVASYNACREKL
 45 KQEHRKNVSELVSGLPNAVKKVNEELLDNENQTVRQLYVALRELKQONP
 ALYRVVEYINAVRLRREDSDEQEKRRLSTSPFGENNEEQDLGEQDF
 HYIYGFECARFILQNGRMFGLNTDRRY

50 The antigen encoded by SEQ ID NO: 31 was isolated from
Cyathostomum pateratum.

55
 SEQ ID NO: 31
ATGAACAAAACGTTAACATTTCTCACAGTCGTTAGTGCCGATGTTCTG
 GCCCAAAGTGTATGGACTTTTTTGGTGAAGAGGGTCGTGAAGAACAT
 CGTCGTACCATCGTCATTTTACCACCATATCTCCACAATGTG
 60 AGCTGTGAGGCTAAATGGGAGTACTTCAAATTTGGGGAACAGGAGT
 TTGACGTTTGTCTGAGAAAAAGGAGAAAATTAGCGAGTGGGCAAAAAAA
 TACAATGTTGTGGTAAGCTTTTTTGAATGATGTAATTTCACTCGCAT
 GCTGGCCTTTTGGATGAAGTTGCAAGCTACAATGCTTACAGGGAAA
 65 AACTCAAGCAGGAGCACAGAAAAACGTTAGCGAAGCTTGTCTCTGCTC

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TTCCAAACGCAGTAAAGAAAAGTCAACGATCCTTAGACAAATGAAAATC
 AGACTCTTAGGCAACTTTACGTTGCCTTAGAAAACCTTGGTAGACAAA
 ATCCGGCAGTAAGTCGAAAGAGCTGCGTCTTGGACTTAAGCGGAAA
 ATTATTTACAGCTTTACCGTATGTGCGAGTACATTAATGTGGCTGTAAG
 ACTAAGAAGTGAAGAAGTGGATGAGCAAGAACAACGAAGAAGGCTGTC
 AGCTCTACCTTTTGGCGACCATAACGATAATTTGGAAGAGCAGGACTT
 CGGTGAACAAGACTTTTCGCTATGCTATGGCTTTGAGTGTGCAAGATT
 TCTCCTTCAAAATGGAAGAATGTTCCGACTCAACACGGATGGAAGATA
 TTAGTAAGAAAACAGTGTAGCTCAAAGTGGTAGAGTTTGGACTACGAA
 CTCAACATGCCATCATCACCTCCT

SEQ ID NO: 32 (i.e., SEQ ID NO: 31 translated)
 MNKTLTFLTVVSAVLAQGVMDLFGEEGREHRRHRHSLPPYLHN
 SCEAKWEYFKIVGNRSLTFAEKKEKISEWAKKYNVDEVASYNAYREK
 LKQEHKKNVSELVSNLPAVKKVNDLLDNENQTLRQLYVALRKLGRQN
 PALYRIVEYINAVRLRSEEVDEQEQRRLSALPFGDHDNDLLEEQDFG
 EQDFRYVYGFECARFLLQNGRMFGLNLDGRY

The following sequences (SEQ ID NOS: 33 and 35) represent Cyathostomin GALA sequences obtained from cDNA clones.

The antigen encoded by SEQ ID NO: 33 was isolated from *Cylicostephanus goldi*.

SEQ ID NO: 33
ATGAACAAAACGTTAACATTTCTCACAGTCGTTAGTGCCTGTGCTCTG
 GCCCAAGGTGTCATGGCCCTATTTGGTGAAGAGAGTTCGTGAAGAACAC
 CGCCGTCACCATCGTCATCTCACTCTTACCACCATATCTCCACAACGTG
 AGCTGTGTGGCTAAATGGGAGTACTTCAAAATGTGGGAAACAGGAGT
 TTGACGTTTGTGAGAAAAAGAAAGAAATCAGCGAGTGGGCTAAAAAA
 TACAATGTTGTGGATGAAGTTGCAAGCTACAATGCTTATAGAGAAAAA
 CTCAAGCAGGAACACAGGAAAAACGTTAGCGAARTGTTTCTGATCTT
 CCCAACGCAGTAAAGAAAAGTCAACGATCTTTTGGACAACGAAAATCAG
 ACTTCTAGGCAACTTTATGTTGCACTCAGAGAACTTGGTAGACAAAAT
 CCGGCACTATACCGTGTGTCGAGTATATCAATGTGGCTGTGAGATTA
 AGACGAAAAGAACAGGATGAACAAGAACGACAAGGAACGCTGTGAGCT
 CTACCTTTTGGCGAGAATAACGACAATTTGGAAGAGCAGGACTTTGGT
 GAACAAGACTTTTCGCTATGCTATGGCTTTGAGTGTGCAAGATTTCTC
 CTTCAAAATGGAAGAATGTTTGGACTCAACACGGATAGAAGATACCAG
 TAAGAGTCAACTGTAGCTCAAAGTGGGTTTGGACTACGAACAGCATGC
 CATCATCACCTCCT

SEQ ID NO: 34 (i.e., SEQ ID NO: 33 translated)
 MNKTLTFLTVVSAVLAQGVMDLFGEEGREHRRHRHSLPPYLHN
 SCVAKWEYFKIVGNRSLTFAEKKEKISEWAKKYNVDEVASYNAYREK
 LKQEHKKNVSELVSNLPAVKKVNDLLDNENQTSRQLYVALRELGRQN

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PALYRVVEYINAVRLRRRKEQDEQERQGTLSALPFGENNDNLEEQDFG
 EQDFRYVYGFECARFLLQNGRMFGLNLDGRY

5 The antigen encoded by SEQ ID NO: 35 was isolated from *Cylicostephanus longibursatus*.

SEQ ID NO: 35
 10 C. lon91- GALA
ATGAACAAAACGTTAACATTTCTCACAGTCGTTAGTGCCTGTGCTCTG
 GCCCAAGGTGTCATGGCCCTTTTGGTGAAGAGGGTTCGTGAAGAACAT
 CGCCGTCACCATCGTCATCTCACTCTTACCACCATATCTCCACAATGTG
 15 AGCTGTGTGGCTAAATGGGAATACTTCAAAATCTGGGGAACAGGAGT
 TTGACGTTTGTGAGAAAAAGGAAAAATCAGCGAGTGGGCAAGAAG
 TACAATGTTGTGGATGAAGTTGCAAGCTATAATGCTTGCAGGGAAG
 20 CTTAAGCAGGAACACAGGAAAAACGTTAGCGAAATGTTTCTAATCTT
 CCCAATGCAGTGAAGAAAGTAAACGATCTTTTGGACAATGAAAATCAG
 ACCCCAGGCAACTTTACGTTGCCTCAGAAAACCTGGTAAACAAAAT
 25 CCGGCACTTTATCGTGTGTCGAGTATATCAATGTGCTTGTGAGACTA
 AGACGTGAAGAATTTGATGAAGATCAACGAAGATCGTGTGAGCTTTA
 CCTTTTGGCGACAATAACGACGATTTGGAAGAGCAGGACTTTGGTGAA
 30 CAGGACTTTTCGCTATATCTATGGCTTTGAGTGTGCAAGATTTATCCTT
 CAAAATGGAAGAATGTTTCGGAATCAACACGGATAGAAGATATTAGTAA
 GAGTCAACTGTAGCTCAAGGGTTTGGACTACGAACCTGCATGCCATCAT
 35 CACCTCCT

SEQ ID NO: 36 (i.e., SEQ ID NO: 35 translated)
 MNKTLTFLTVVSAVLAQGVMDLFGEEGREHRRHRHSLPPYLHN
 SCVAKWEYFKILGNRSLTFAEKKEKISEWAKKYNVDEVASYNACREK
 40 LKQEHKKNVSEIVSNLPAVKKVNDLLDNENQTPRQLYVALRKLGRQN
 PALYRVVEYINVLVRLRREEFDEQRRLSALPFGDNDLLEEQDFGE
 QDFRYIYGFECARFILQNGRMFGINTDRRY

45 Each of the proteins provided by SEQ ID NO: 1, 3, 5, 7, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34 and 36 (or encoded by the nucleic acid sequences of SEQ ID NOS: 2, 4, 6, 8, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33 and 35) may be classified as a member of the "keratin-like" proteins although, because they lack the glycine-rich domains characteristic of other KLP proteins and are localised to the gut of larval cyathostomin, the inventors have chosen to designate these proteins cyathostomin gut-associated larval antigens (Cy-GALA).

55 Using any of the Cy-GALA sequences described herein, one of skill in the art could readily identify related or homologous sequences in other species, such as, for example, other cyathostomin spp. etc. For example, the nucleic acid sequence encoding these proteins could be used to probe for homologous sequences in other cyathostomin species.

60 Other potentially useful cyathostomin larval antigens include those encoded by the following sequences, designated SEQ ID NOS: 37-58. (SEQ ID NOS: 37, 43, and 45 are amino acid sequences and SEQ ID NOS: 38-42, 44, and 46-58 are nucleic acid sequences, SEQ ID NOS: 38, 44, and 46 encoding SEQ ID NOS: 37, 43, and 45 respectively). It should be understood that the invention further encompasses pro-

teins, peptides and amino acids having sequences encoded by
SEQ ID NOS: 39-42 and 46-58.

(CID-1):

SEQ ID NO: 37
REKARI IQDEYTKRMQQVTPQAQEF LAKWEKTWFTNVQYSGDKKAF
KQMIELIPQLMEEVHGFSEETWKSLEEQFPPEQTAANKDNEDRLKQFYE
FIKSLPKQDLAEDPEAFRKF AHLGLQLLPIEALRA

(nucleic acid sequence encoding CID-1)

SEQ ID NO: 38
AGGGAGAAGGCTAGAATTATTCAAGACGAATACACTAAACGTATGCAG
CAGGTCACACCACAAGCTCAGGAATTCCTGGCAAAATGGGAGAAGACA

-continued

TGGTTCACGAATGTGCAGCAATATAGCGGAGATAAGAAAGCTTTCCTC
AAGCAGATGATTGAGCTAATCCCTCAACTAATGGAGGAGTTCATGGG
5 TTCTCGGAAGAGACTTGGAAAGCCTTGAGGAGCAATCCAGAGCAG
ACAGCCGCATGGAAGATAATGAGGATCGCCTAAAGCAATTTATGAG
TTTATCAAGAGCCTACCCAAGCAGGACTTAGCTGAGGATCCGGAAGCA
10 TTCAGAAAGTTCGCTCACCTCGGACTCCAGAACTTCTTCCAATTGAA
GCTCTCAGAGCT

CID antigens from other Cyathostomin organisms may
include those encoded by the genomic DNA sequences pro-
vided as SEQ ID NOS: 39-42 provided below.

SEQ ID NO: 39

C. cat01 - CID
TGGTTCACACCACAAGCTCAGGAGTTCCTGGCCAAGGTAAGCTATTACCTTACCAGGGT
AGTTGGCAGCGGTGCGAAACCCGGTAATCTACTGACTTTACCAATATTTTCAGTGGGAG
TTCACGAATATACAGCAATACAGTGGAGACAAGCAAGCCTTCTTTAAGCAGATGATTGAACTAATTCCT
CAACTTATGGAGGAGGTTTCAGGTAAGTTAGCCGAAAAATTTTAAACCAATGGTTGAGCTCGACATTTT
TTCAGGATTCACAGAGGAGACTTGGAAATAGCCTGAGGAGCAATTCAGGAGCAGACAGCCGATGGA
AGGATCGTGAATATCTTTCATAATTACTGTACTTGGAAATATACTTTACAATCATAATCCTACTCTTAG
ACGAGGATCGCCTGAAGCAATTCATGAGTTCATTAAGAGCCTACCCAAACAACAATTAGCTGAGGTGA
TTTTATTGATTTTTTCGAAAAATATATTTTTGATACATTCTTTTCAGGATCCGGAAGCTTTCAGAAAG
TTCGCTCACCTCG

SEQ ID NO: 40

C. cat02 - CID
TTGTCACACCACAAGCTCAGGAGTTCCTGGCTAAGGTAAGCTATTACCTTACCAGGGT
AGTTGGGAGCGGTGCGAAACCCGGTAATCTACTGACTTTACCAATATTTTCAGTGGGAGAGGACATGG
TTCACGAATATACAGCAATACAGTGGAGACAAGCAAGCCTTCTTTAAGCAGATGATTGAACTAATTCCT
CAACTTATGGAGGAGGTTTCAGGTAAGTTGGCCGAAAAATTTTAAACCAATGGTTGAGCTCGACATTTT
TTCAGGATTCACAGAGGAGACTTGGAAATAGCCTGAGGAGCAATTCAGGAGCAGACAGCCGATGGA
AGGATCGTAAGTATCTTTCATAATTACTGTACTTGGAAATATACTTTACAATCATAATCCTACTCTTAG
ACGAGGATCGCCTGAAGCAATTCATGAGTTCATTAAGAGCCTACCCAAACAACAATTAGCTGAGGTGA
TTTTATTGATTTTTTCGTAACGAAAAATATATTTTTGATACATTCTTTTCAGGATCCGGAAGCTTTCAG
AAAGTTCGCTCACCTCG

SEQ ID NO: 41

C. lon91 - CID
AGGTCACACCACAAGCTCAGGAATTCCTGGCAAAGGTAAGCTATCACCTTACCAGGGT
GTTAGGAGCGAGGGAACCCGGTATCTCTTATACCCATTACTTCAGTGGGAGAAAGATATGGTTACAGAA
TGTACAGCAATATAGTGGAGACAAGCAAGCCTTCTTCAAGCAGATGATTGAACTAATTCCTCAACTTAT
GGAGGAGGTACAGTAAGTACAGTAAAGTGATTTTAAAGAAAAATTAAGCCTGATTTTCTTTTCAGGGA
TTCTCAGAGGAGACTTGGAAATAGCCTTAAGGAGCAATTCCTGAGCAGACAGCCGATGGAAGGATAGT
GAGTATTTTTTCATAATTACTGTACTTGGAAATATACTTTACAATCATAATCCTACCTCAGACGAGGAG
CGCCTGAAGCAATTCATGAGTTCATTAAGAGCCTACCCAAACAACAATAGCTGAGGTGATTTTCATT
GATTTTTTCGTACGAAAAGTATATTTTTAATACATTCTTTTCAGGATCCGGAAGCCTTTCAGAAAGTTCG
CTCACCTCG

-continued

SEQ ID NO: 42

C.nas07-CID

AGGTCACACCACAAGCTCAGGAATTCTTGCCAAAGGTAAGCTACCATATTTTCGAGGGGGAGGGCAATTT
 TGGAGCGAGGGAGGAGAGGAAAGGGGAGAGAAACACTGGTTGGGATCACTAACTCTACCCGCCACTTCCA
 GTGGGAGAAGACATGGTTCACGAATGTGCAGCAATATAGCGGAGATAAGAAAGCCTTTTCAAACAGAT
 GATTGAGCTAATCCCTCAACTAATGGAAGAGGTTTCATGTAAGTCAACCAAGTGGCTTTTAAGCGGAGA
 TTAAACTCGAATTTTCTTTCAGGGTCTCGGAGGAGACTTGAAGAGCCTTGAGGAGCAATTCACAGA
 GCAGACAGCCGCATGGAAGGATAGTAAGCATTCTTCATAGCTCCCGCCTTTATCATTTATCTTCACGAT
 AGTAATCTTATTTTATGATGAGGATCGCCTGAAGCAATTTTATGAGTTCATCAAGAGCCTACCCAAGCA
 GGACTTAGCTGAGGTAACCTTCATGGTTTTTCTGAGCTGTAATAATGCTTGCAACTAACAACTTTTC
 TAGGATCCGGAAGCTTTTCAGAAAGTTCGCTCACCTCG

(FAR-2):

SEQ ID NO: 43

KKESQGFFSIPVDNLRASPFLQYIKEYIPDYKNAMEKFEDIPKQYRDLIPEEVATHLKAITAEKAVL
 KEVMKYAKYKDEEFLKALKEKSEGLHEKASKLHNFIKGVKDALGDEAKAFVKKVIAAAREVHAKLLA
 GDKPSLEDIKKKAKEHMGEFEKLSDDAKEDLKNFPILTSVWNEKTRALIDKYVEN

(nucleic acid sequence encoding (FAR-2))

SEQ ID NO: 44

ATGCTTCGAATAACTTTCTTCTTGCTCTCTTTGTTGTCTACACTTTTCTGCACCTCTGGACCCGCT
 GAAGAGAAGATAGATGTGGAATAATGGAATAATTTGAAGATATCCAAAGCAATATCGAGACCTTATT
 CCGGAAGAGGTAGCTACACACCTCAAAGCCATCACCGCTGAAGAGAAAGCTGTTCTAAAAGAGGTAATG
 AAGAATTATGCAAGTACAAGAACGAGGAGGAGTTTTTGAAGCGTTGAAAGAAAAATCAGAGAGTTTG
 CATGAGAAAGCCAGCAAACCTCACAAATTTATCAAAGGAAGGTTGACGCACTTGGAGATGAAGCAAAG
 GCATTTGTGAAGAAGTTATCGCAGCTGCTCGAGAAGTGCATGCCAACTCTTGC CGGGGACAAACCA
 TCGCTTGAAGATATCAAGAAGAAAGCAAGGAGCATATGGCTGAATTCGAGAACTAAGCGATGATGCC
 AAGGAGGATCTCAAAAAGAAATTTCCAATCCTTACTTCCGTCTGGACAAATGAGAAAAACAAGAGCGTTG
 ATTGACAAATATGTGGAGAAC

(UNK-50a):

SEQ ID NO: 45

GKMSDLWTAISETNKVRLFNLTLSLGIAGVLCITTAFFIPVENQVCAVLI TLLQGVIGFNSAGYNKAAVI
 VARQHAHLLLTCFGLIVTFVPLVQPFIVQLVAPDHSWDQWFYLFVGHGLVLVIANLFFCLTIEAKPAF
 TQKTDSS

(nucleic acid sequence encoding UNK-50a)

SEQ ID NO: 46

GGTAAAATGTCAGATTTATGGACGGCAATAAGCGAAACAATAAAGTCCGCTTGTTC AACACCTTGTTCG
 CTGGGAATTGCTGGCGTACTGTGTATAACTACTGCTTTCATTCCTGTGGAAATCAGGTTGTTGCGCT
 GTTTTAATCAGTTATGCAAGGAGTTATCGGATTC AATTCAGCTGGATATAACAAGCTGCAGTCATT
 GTTGCTAGGCAGCATGCTCATCTTCTGTTGACCTGCTTTGGGCTCATTGTC ACTTTTGTCCCCTGGTG
 CAGCCATT CATAGTTCAACTTGTGGCCCTGACCATAGCTGGGACCAATGGTTTTATCTGTTTGTGGG
 CATGGTCTCGTACTTGTATAGCGAATTTATTCTTTTGTCTCACTATCGAGGCGAAACCGGCAGCGTTC
 ACACAGAAAATGATTCATCA

The following sequences represent nucleic acid sequences encoding potentially useful EL3 antigens (or fragments or portions thereof). As above, it should be understood that the in addition to these nucleic acid sequences, the present invention

relates to amino acid sequences comprising sequences encoded by SEQ ID NOS: 47-58 or derivatives, variants or homologues thereof.

SEQ ID NO: 47

EL3sequence1
 GGTTTAATTACCCAAGTTTGAGGTAAGTTTCTAAATCTGACCCGATCAACTGATTGTGGTCTGATTAAT
 TTTGAAAAATCTCTCCTGAATAGGGAGAGTACAAGAGTGCATATCCAAAAAAAAAAAAAAAAAAAAAAAA
 AAAAAACATGTCGGCCGCCTCGGCCTCTAGAATA

SEQ ID NO: 48

EL3sequence2
 GGTTTAATTACCCAAGTTTGAGTGTTCATGAAGCTTGCCCTGAAAAAGCAGAGAAACCAAGAGGAGATAG
 TTTACAGTTCCCGCAGACAGGAAATGCGTGCAGATGTTTTCGCGAAGAGGAGAAACGTCGTTCACT
 TAGAATGAGAAGGCATTGATTCTGTTTGTAGTGTGAGATATTTAAAAATCTTTGCAGAAAAACCTTTTC
 AAATCATAAAGTCGAAGACCACAAAAAAAAAAAAAAAAAAAAAAAAAAAAACATGTCGGCCGCCTCGG
 CCTCTAGAATA

SEQ ID NO: 49

EL3sequence3 (Cy-Ins-1)
 GGTTTAATTACCCAAGTTTGAGGCTGCTTCAACAGTAGGTTTAGAAATGACATCGCGGATATGGCGCCG
 CACCCAGAGCCCTCCATTATGTACTCTGTTGTTGATCAGTCTACCAGTAGCTGAGTGTAGTATTCG
 ACTATGTGGAGTGGCACTAACACGAACTCTTATGGCTATCTGCAGGAATCAATTATGCGGTTATTCGCA
 AAGTAAAAGATCTGCTATGTGGGAGAGCCTCGACTGGAAACCGTCACTCAACAATGAAACGATCAGG
 GATCGCCACCGAATGCTGCGAGAATCGGTGCTCATTAGTACTTAAAGACATACTGCTGCAGCACTTA
 GCCTTGGCATCTTAAGCCGCTTTTATCTCCTCCTCCATGATCTCTCTCGTTATCTGTATAACCGAATAT
 AGTCATTCCGGAATGCGGATGCTTAGGCCAATTTGTTGACGTTTCCCGCATGAATCATTGCTGTTTCG
 TCATTATCTCACAGAGTGTAAAAGATCTCTTTTATGAAAGTCTATTTGTTTGTAGCTGCACCATTAA
 ACCGTTTACAAAAAAAAAAAAAAAAAAAAAAAAAAAAACATGTCGGCCGCCTCGGCCTCTAGAATAA

SEQ ID NO: 50

EL3sequence4
 GGTTTAATTACCCAAGTTTGAGGTAAGTTTCTAGATCTGACCCGATCAACTGATTGTGGTCTGATTAAT
 TTTGAAAAATCTCTCCTGAACAGGGAGAGTACAAGAGTGTATGAAAAAAAAAAAAAAAAAAAAAAAA
 AAAAAACATGTCGGCCGCCTCGGCCTCTAGAATA

SEQ ID NO: 51

EL3sequence5
 GGTTTAATTACCCAAGTTTGAGGATGCTTAGTTTCAAGCTCGTCTTCTCTCTGACTTCTCACAGCTT
 GTGTGCTAACAGATCCAAGAGTGTAAATCCGAGAAAAGCGAATGGACTGGAGACGTTACTATAGCAGAT
 GGGGTCGCGGAAGCTCTAATTGGGAAACCGCGGAGGTACCTTCGCGGACGAAAATGGAGTTACCCGA
 CTTTTGAGCAATGGGGACATTAACATCTGATGTATGAAAAGATCTAATGAAATAAAGCTTCGAAAAAAA
 AAAAAAAAAAAAAAAAAAAAAACATGTCGGCCGCCTCGGCCTCTAGAATA

SEQ ID NO: 52

EL3sequence6 (Cy-Cbg-1)
 GGTTTAATTACCCAAGTTTGAGAAATGTCGAAAAATTCCTTCTGCTACTGATCGTTGTGATCGCCCTCA
 TTTCTTTGGCGTCTGCAGATTTTTTCATGCTTCTCGGTGATACCATCTGCAAGAGCATTACATGCAGGG
 GCTGCACCCGTCGCCACTTGCCCTTAATGGAGACTGTATGTGCACACTATGTAACCTGATGATCTTCACATG
 TCGCATTACCATTTGTAACAAATACATTTTCTCTTGTTCATAATAAATTTTCACTCAAAAAAAAAAAAA
 AAAAAAAAAAAAAAAAAAAAAACATGTCGGCCGCCTCGGCCTCTAGAATA

SEQ ID NO: 53

EL3sequence7
 GGCCGCGGATTTTCTAGAGCCGAGGCGGTTTGTAGTTGTTCTCAAACCTGGGTAATTAACCACG
 AGCCGAGGCGGGTTTTAGTTGTTCTCAAACCTGGGTAATTAACCACGATGGCGAGGCGGGTTTTAG
 GTTGTCTCAAACCTGGGTAATTAACCACGATGGCGAGGCGGGTTTTAGTTGTTCTCAAACCTGGG

- continued

TAATTAACAACGAGGCGGAGGCGGGTTTATAGGTGTTCTCAAACCTGGGTAATTAACCAACGATGGC
GAGGCGGGTTTATAGGTGTTCTCAAACCTGGGTAATTAACCAATCACTAGT

SEQ ID NO: 54

EL3sequence8
GGCCGCGGGATTATTCTAGAGGCGGAGGCAGTGGTATCAACGCAGAGTGGCCATTACGGCCGGGGAGAG
GGAAAAGTTTCTTTCTCTCGGATACCAAAAAAAAAAAAAAAAAAAAAAAAAAACAATGTCGGCCGC
CTCGGCCTCTAGAATA

SEQ ID NO: 55

EL3sequence9
GGCCGCGGGATTCTAGAGGCCGAGGCGTCTTACTTGGGTGGCTCAATAACTGAAAGCTTAGAATTCA
TTAAACCTTAACCCACAGGGTTATTTGACATGCTTGACTTGAAAATGATGCTCTTCTGCTTGTAGTTG
TTTTATTATGCTAGCTGTAAGTATACTCTGGTAGACCAGAACAATCAATGCTAGTTGAATGTATCATG
TTATCACTTTGTCACTCTATACGAATCTAGGTGTGGCAGGCCACCCCTCTCTGACCCCTGTTTAC
CATCAATTAGCTTTAGCTGTTATTTAATAACATCACACTGATGCAAAAAAAAAAAAAAAAAAAAAA
AAAAAACATGTCGGCCCTCGGCCTCTAAAAATCACTAGT

SEQ ID NO: 56

EL3sequence10
GGCCGCGGGATTATTCTAGAGGCGGAGGCAGTGGTATCAACGCAGAGTGGCCATTACGGCCGAGCAGT
GGTATCAACGCAGAGTGGCCATTACGGCCGGGTGGTGACCAGGGTGACGGGAATTAGGGTTCGATTC
CGGAGAGGGAGCCTGAGAAACGGCTACCACATCCAAGGAAGGCAGCAGGCGCGCAAAATACCCACTCCC
GACCCGGGAGGTAGTGACGAAAAAAAAAAAAAAAAAAAAAAAAAACAATGTCGGCCCTCGGCCTC
TAGAATAATCACTAGT

SEQ ID NO: 57

EL3sequence11
GGCCGCGGGATTCTAGAGGCCGAGGCGGGTTTATAGCTCAAACCTGGGTAATTAACCGGTAGGATGG
CGAGGCGGGTTTCTCAAACCTGGGTAATTAACCCAGTAGGATGGCGAGGCGGGTTTCTCAAACCTGGGT
AATTAACCGGTAGGAGGCCGAGGCGGGTCTCAAACCTGGGTAATTAACCAATCACTAGT

SEQ ID NO: 58

EL3sequence12
CAAGTTTGAGGTACTTTCTAGATCTGACCCGATCAACTGATTGTGGTCTGATTAATTTGGAAATCTC
TTCTGAAACAGGGAGAGTACAAGAGTGTATATTAAGAAAAAAAAAAAAAAAAAAAAAAAAAACAATGTCGG
CCGCCTCGGCCTCTAGAATAATCACTAGT

45

As such, the present invention relates to the proteins encoded by the sequences designated as SEQ ID NOS: 1, 3, 5, 7, 37, 43, and 45, the corresponding gene sequences (such as, for example, those given as SEQ ID NOS: 2, 4, 6, 8, 38, 44, and 46) and proteins, peptides and/or amino acids comprising sequences encoded by SEQ ID NOS 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35-42, and 47-58, as well as any fragments, portions, mutants, variants, derivatives, analogues and/or homologues/orthologues thereof. Furthermore, the methods described herein may provide means for detecting levels of antibodies which bind to proteins comprising (or encoded by) any of SEQ ID NOS: 1-58 (or fragments, portions, mutants, derivatives, analogues or variants thereof).

Typically the fragments, portions, mutants, variants, derivatives, analogues and/or homologues/orthologues mentioned in this invention are immunogenic or encode immunogenic cyathostomin larval antigens—that is, they are capable of generating immune, preferably humoral, responses.

The term “mutants” may encompass naturally occurring mutants or those artificially created by the introduction of one or more amino acid/nucleic acid additions, deletions, substitutions or inversions.

One of skill in this field will readily understand that proteins or nucleic acids homologous to the proteins encoded by SEQ ID NOS: 1, 3, 5, 7, 37, 43, and 45 or nucleic acid sequences of SEQ ID NOS: 2, 4, 6, 8, 38, 39-42, 44, 46, and 47-58, may exhibit as little as 20 or 30% sequence homology or identity thereto (or to a portion thereof). In other instances however, homologous proteins or nucleic acid sequences may exhibit at least 40, 50, 60, 65, 70, 75, 80, 85, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99% homology or identity the whole or part of SEQ ID NOS: 1-20 detailed above. As such, proteins or nucleic acids homologous to (or partially identical with) the proteins and/or nucleic acid sequences provided by SEQ ID NO: 1-20 are also included within the scope of this invention.

It should also be understood that natural variations due to, for example, polymorphisms, may exist between related (or homologous) proteins/genes from any given cyathostomin species. These variants may manifest as proteins/genes which exhibit one or more amino/nucleic acid substitutions, additions, deletions and/or inversions relative to a reference sequence (for example any of the sequences provided by SEQ ID NOS: 1-58 described above). All such variants, especially those which are functional and/or are immunogenic (or

65

encode functional/immunogenic proteins or peptides) are to be included within the scope of this invention.

Additionally, or alternatively, analogues of the various peptides described herein may be made by introducing one or more amino acid substitutions into the primary sequence. In certain embodiments, one or more of these substitutions may represent a "conservative substitution". One of skill in this field will understand that the term "conservative substitution" is intended to embrace the act of replacing one or more amino acids of a protein or peptide sequence with an alternate amino acid with similar properties and which does not substantially alter the physio-chemical properties and/or structure or function of the native (or wild type) protein.

As is well known in the art, the degeneracy of the genetic code permits substitution of one or more bases in a codon without changing the primary amino acid sequence. Consequently, although the nucleic acid sequences described in this application are known to encode potentially useful cyathostomin larval antigens, the degeneracy of the code may be exploited to yield variant nucleic acid sequences which encode the same primary amino acid sequences.

Also encompassed by this invention are splice variants of the primary gene transcripts encoded by any of the gene sequences described herein, as well as and the translated Cyathostomin larval antigen splice variant proteins which are encoded thereby. By way of example, splice variants of the Cy-GALA proteins described herein, including, for example, variants encoded by transcripts having 115 bp segment deletions, are within the scope of this invention. Furthermore, one of skill in this field will readily appreciate that polyadenylation variants and start codon variants, including cDNA sequences encoding the same, may also be included within the scope of this invention.

As stated, this invention finds particular application in the identification or diagnosis of cyathostomin infections in horses but may be more generally be used to diagnose or identify cyathostomin infections present in other species of the Equidae family including, for example, donkeys and zebra

The term "sample" should be understood as including any samples comprising antibodies and/or cyathostomin larval antigens. For example, suitable samples may include fluids such as whole blood, plasma, serum, saliva, sweat and/or semen. In other instances "samples" such as tissue biopsies and/or scrapings may be used. In particular biopsies or scrapings from the gut may be used. In addition, a sample may comprise a tissue or gland secretion and washing protocols may be used to obtain samples of fluid secreted into, for example, the gut. In other embodiments, faecal samples may be used. One of skill will understand that in order to prepare a faecal sample for use, it may be necessary to add buffers and various protease inhibitors and subject the sample to procedures such as centrifugation, to remove particulate material. As such, "faecal samples" may represent suitable samples for use in the methods provided by this invention. As stated, a "reference" or "control" sample may be derived from healthy animals or from animals not having high mucosal burdens of cyathostomin parasites or larval cyathostominosis.

In order to identify a level of anti-cyathostomin larval antigen antibodies present in a sample, the sample may be contacted with one or more cyathostomin larval antigen(s) (such as those provided, comprising or encoded by SEQ ID NOS: 1-58) under conditions which permit binding between any anti-cyathostomin larval antigen antibodies present in the sample and the cyathostomin larval antigen(s). Anti-cyathostomin larval antigen antibodies bound to cyathostomin larval antigen may easily be detected with the use of agents capable

of binding anti-cyathostomin larval antigen antibodies. In one embodiment, the agents capable of binding anti-cyathostomin larval antigen antibodies may be conjugated or linked to a detectable moiety.

One of skill will appreciate that while the methods provided by this invention may provide a means of detecting antibodies having affinity for, or specificity/selectivity to a single cyathostomin antigen (such as any described herein), in certain embodiments, the methods may exploit the use of one or more of the cyathostomin antigens. Since, for example, horses tend to be infected with one or more different cCyathostomin species, assays/methods which utilise cocktails of cyathostomin antigens provide a means of increasing the likelihood of a positive diagnosis. Accordingly, it should be understood that the methods described herein may use one or more of the cyathostomin antigens described herein.

In one embodiment, the methods provided by this invention may utilise substrates to which one or more cyathostomin larval antigens have been bound, conjugated or immobilised. One of skill in that art will appreciate that in addition to techniques which allow antigens to be bound, conjugated or immobilised "directly" on to the surface of substrates, other techniques may involve the use of substrates which have been coated with agents capable of binding cyathostomin larval antigens.

It is to be understood that the term "agents capable of binding cyathostomin larval antigens" may include, for example, antibodies such as monoclonal or polyclonal antibodies and/or other types of peptide or small molecule capable of binding to cyathostomin larval antigens. It should be noted that this definition applies to all types of binding agent mentioned herein. Furthermore, references to "antibodies" herein are intended to encompass "anti-cyathostomin larval antigen antibodies".

The techniques used to generate antibodies (either monoclonal or polyclonal) are well known to one of skill and may involve the use of cyathostomin antigens (or fragments or portions thereof) either isolated or purified from cyathostomin parasites or recombinantly generated as described herein.

Suitable substrates may include, for example, glass, nitrocellulose, paper, agarose and/or plastics. A substrate such as, for example, a plastic material, may take the form of a microtitre plate.

In order to detect a level of antibody present in a sample, immunological detection techniques such as, for example, enzyme-linked immunosorbent assays (ELISA) may be used. One of skill in this field will appreciate that ELISAs may use substrates to which cyathostomin larval antigens have been "captured" or bound by binding agents (capable of binding cyathostomin larval antigens) bound or immobilised to the substrate. Alternatively, substrates may comprise cyathostomin larval antigens, which have been directly bound or immobilised to the substrate.

An ELISA may involve contacting the sample to be tested with a substrate under conditions which permit binding between any antibodies present in the sample and the cyathostomin larval antigens bound or immobilised to the substrate as described above. One familiar with these techniques will appreciate that prior to contacting the sample to be tested with the substrate, a blocking step may be introduced to reduce incidences of non-specific binding.

An ELISA may comprise the further step of contacting the substrate with a further binding agent capable of binding one or more of the antibodies present in the sample. Such agents may otherwise be known as "secondary antibodies" and may

take the form of rodent or ruminant antibodies specific to particular forms of equine antibody.

Secondary antibodies useful in the present invention may be conjugated to moieties which permit them to be detected (referred to hereinafter as “detectable moieties”). For example, the secondary antibodies may be conjugated to an enzyme capable of reporting a level via a colourmetric chemiluminescent reaction. Such conjugated enzymes may include but are not limited to Horse Radish Peroxidase (HRP) and Alkaline Phosphatase (AlkP). Additionally, or alternatively, the secondary antibodies may be conjugated to a fluorescent molecule such as, for example a fluorophore, such as FITC, rhodamine or Texas Red. Other types of molecule which may be conjugated to binding agents include radiolabelled moieties.

The amount of secondary antibody (identifiable by means of the detectable moiety) bound to the anti-cyathostomin larval antibodies, may be representative of the anti-cyathostomin larval antibodies present in the sample tested.

Alternatively, in order to identify a level of cyathostomin larval antigen present in a sample, a substrate or substrate comprising one or more agents capable of binding one or more cyathostomin larval antigens, may first be contacted with a sample to be tested. Any cyathostomin larval antigen bound to the substrate or to the agents capable of binding the cyathostomin larval antigen, may be detected with the use of a further agent capable of binding the cyathostomin larval antigen (referred to hereinafter as the “primary binding agent”). Additionally, or alternatively, the primary binding agents may have affinity for, or bind to cyathostomin larval antigen:substrate complexes or complexes comprising cyathostomin larval antigen and the abovementioned agents capable of binding the cyathostomin larval antigen. In one embodiment, the primary binding agent may be an antibody conjugated to a detectable moiety as described above.

Alternatively, any cyathostomin larval antigen bound to the substrate or agents capable of binding the cyathostomin larval antigen, may be detected by means of a yet further binding agent having affinity for the primary binding agents. In certain embodiments, the further binding agents may be conjugated to detectable moieties.

In one embodiment, the methods for identifying a level of cyathostomin larval antigen or a level of anti-cyathostomin larval antigen antibodies, may take the form of “dip-stick” test, wherein a substrate (or portion thereof) is contacted with a sample to be tested under conditions which permit the binding of any cyathostomin larval antigen or anti cyathostomin larval antigen antibodies present in the sample, to the substrate or a binding agent bound or immobilised thereto.

Other techniques which exploit the use of agents capable of binding the cyathostomin larval antigen or antibodies which bind thereto include, for example, techniques such as western blot or dot blot. A western blot may involve subjecting a sample to electrophoresis so as to separate or resolve the components, for example the proteinaceous components, of the sample. In other embodiments, electrophoresis techniques may be used to separate proteins purified from cyathostomin parasites and/or proteins generated in a recombinant form. The resolved components/proteins may then be transferred to a substrate, such as nitrocellulose.

In order to identify any cyathostomin larval antigen present in a sample, the substrate (for example nitrocellulose substrate) to which the resolved components and/or proteins have been transferred, may be contacted with a binding agent capable of binding cyathostomin larval antigens under conditions which permit binding between any cyathostomin lar-

val antigen in the sample (or transferred to the substrate) and the agents capable of binding the cyathostomin larval antigen.

Advantageously, the agents capable of binding the cyathostomin larval antigen may be conjugated to a detectable moiety.

Additionally, the substrate may be contacted with a further binding agent having affinity for the binding agent(s) capable of binding the cyathostomin larval antigen. Advantageously, the further binding agent may be conjugated to a detectable moiety.

Similar techniques may also be used to detect levels of anti-cyathostomin larval antigen antibodies present in samples. Techniques of this type may be known as “immunoblots” or “dotblots” or ‘dipsticks’ where cyathostomin antigen(s) is/are immobilised onto suitable substrates (for example a nitrocellulose substrate) and contacted with agents capable of binding cyathostomin antigen(s). In certain embodiments any of the samples described above may be used a source of cyathostomin antigen. Additionally or alternatively, the cyathostomin larval antigen may be isolated or purified from the parasite, or produced in recombinant form.

Other immunological techniques which may be used to identify a level of cyathostomin larval antigen in a sample include, for example, immunohistochemistry wherein binding agents, such as antibodies capable of binding cyathostomin larval antigens, are contacted with a sample such as those described above, under conditions which permit binding between any cyathostomin larval antigen present in the sample and the cyathostomin larval antigen binding agent. Typically, prior to contacting the sample with the binding agent, the sample is treated with, for example a detergent such as Triton X100. Such a technique may be referred to as “direct” immunohistochemical staining.

Alternatively, the sample to be tested may be subjected to an indirect immunohistochemical staining protocol wherein, after the sample has been contacted with a cyathostomin larval antigen binding agent, a further binding agent (a secondary binding agent) which is specific for, has affinity for, or is capable of binding the cyathostomin larval antigen binding agent, is used to detect cyathostomin larval antigen/binding agent complexes.

The skilled person will understand that in both direct and indirect immunohistochemical techniques, the binding agent or secondary binding agent may be conjugated to a detectable moiety. Preferably, the binding agent or secondary binding agent is conjugated to a moiety capable of reporting a level of bound binding agent or secondary binding agent, via a colourmetric chemiluminescent reaction.

In order to identify the levels of cyathostomin larval antigen present in the sample, one may compare the results of an immunohistochemical stain with the results of an immunohistochemical stain conducted on a reference sample. By way of example, a sample revealing more bound cyathostomin larval antigen binding agent (or secondary binding agent) than in a reference sample, may have been provided by a subject with a cyathostomin infection.

In addition to the methods and techniques described above, the present invention also contemplates the use of a range of PCR based techniques which may be used to detect levels of cyathostomin antigen gene expression or gene quantity in a given sample. Useful techniques may include, for example, polymerase chain reaction (PCR) using genomic DNA as template or reverse transcriptase (RT)-PCR (see below) based techniques in combination with real-time PCR (otherwise known as quantitative PCR). In the present case, real time-PCR may be used to determine the level of expression of the genes encoding any of the cyathostomin larval antigens

described herein. Typically, and in order to quantify the level of expression of a particular nucleic acid sequence, RT-PCR may be used to reverse transcribe the relevant mRNA to complementary DNA (cDNA). Preferably, the reverse transcriptase protocol may use primers designed to specifically amplify an mRNA sequence of interest (in this case a cyathostomin mRNA encoding a cyathostomin larval antigen). Thereafter, PCR may be used to amplify the cDNA generated by reverse transcription. Typically, the cDNA is amplified using primers designed to specifically hybridise with a certain sequence and the nucleotides used for PCR may be labelled with fluorescent or radiolabelled compounds.

One of skill in the art will be familiar with the technique of using labelled nucleotides to allow quantification of the amount of DNA produced during a PCR. Briefly, and by way of example, the amount of labelled amplified nucleic acid may be determined by monitoring the amount of incorporated labelled nucleotide during the cycling of the PCR.

Further information regarding the PCR based techniques described herein may be found in, for example, PCR Primer: A Laboratory Manual, Second Edition Edited by Carl W. Dieffenbach & Gabriela S. Dveksler: Cold Spring Harbour Laboratory Press and Molecular Cloning: A Laboratory Manual by Joseph Sambrook & David Russell: Cold Spring Harbour Laboratory Press.

Other techniques that may be used to determine the level of cyathostomin larval antigen gene expression in a sample, include, for example, northern and/or Southern blot techniques. A northern blot may be used to determine the amount of a particular mRNA present in a sample and as such, could be used to determine the amount of cyathostomin larval antigen gene expression. Briefly, total or messenger (m)RNA may be extracted from any of the samples described above using techniques known to the skilled artisan. The extracted RNA may then be subjected to electrophoresis. A nucleic acid probe, designed to hybridise (i.e. complementary to) an RNA sequence of interest—in this case the mRNA encoding a cyathostomin larval antigen, may then be used to detect and quantify the amount of a particular mRNA present in a sample.

Additionally, or alternatively, a level of cyathostomin larval antigen gene expression may be identified by way of microarray analysis. Such a method would involve the use of a DNA micro-array which comprises nucleic acid derived from cyathostomin larval antigen genes. To identify a level of cyathostomin larval antigen gene expression, one of skill in the art may extract the nucleic acid, preferably the mRNA, from a sample and subject it to an amplification protocol such as, RT-PCR to generate cDNA. Preferably, primers specific for a certain mRNA sequence—in this case sequences encoding cyathostomin larval antigen genes may be used.

The amplified cyathostomin larval antigen cDNA may be subjected to a further amplification step, optionally in the presence of labelled nucleotides (as described above). Thereafter, the optionally labelled amplified cDNA may be contacted with the microarray under conditions which permit binding with the DNA of the microarray. In this way, it may be possible to identify a level of cyathostomin larval antigen gene expression.

In addition, other techniques such as deep sequencing and/or pyrosequencing may be used to detect cyathostomin larval antigen sequences in any of the samples described above, particularly faecal matter extracts. Further information on these techniques may be found in “Applications of next-generation sequencing technologies in functional genomics”, Olena Morozovaa and Marco A. Marra, Genomics Volume 92, Issue 5, November 2008, Pages 255-264 and “Pyrosequencing sheds light on DNA sequencing”, Ronaghi, Genome Research, Vol. 11, 2001, pages 3-11.

quencing sheds light on DNA sequencing”, Ronaghi, Genome Research, Vol. 11, 2001, pages 3-11.

The present invention also extends to kits comprising reagents and compositions suitable for diagnosing cyathostomin infections. For example, depending on whether or not the kits are intended to be used to identify levels of cyathostomin larval antigen or antibodies thereto in samples, the kits may comprise substrates having cyathostomin larval antigens or agents capable of binding cyathostomin larval antigens, bound thereto. In addition, the kits may comprise agents capable of binding cyathostomin larval antigens—particularly where the kit is to be used to identify levels of cyathostomin larval antigens in samples. In other embodiments, the kit may comprise agents capable of binding the cyathostomin larval antigens, for example specifically raised polyclonal antibodies or monoclonal antibodies. Where the kits are intended to diagnose equine cyathostomin larval infections, these binding agents may take the form of antibodies capable of binding equine antibodies. The antibodies may be conjugated to detectable moieties. Kits for use in detecting the expression of genes encoding cyathostomin larval antigen gene may comprise one or more oligonucleotides/primers for detecting/amplifying/probing cyathostomin larval antigen encoding sequences. The kits may also comprise other reagents to facilitate, for example, sequencing, PCR and/or RFLP analysis. All kits described herein may further comprise instructions for use.

It will be appreciated that the uses, medicaments and methods of treatment described herein may require the generation of recombinant cyathostomin larval antigens (or genes encoding the same) and as such, the present invention further contemplates methods of generating and/or expressing recombinant cyathostomin larval antigen genes and/or proteins (such as for example those described above as SEQ ID NOS: 1-58). One of skill in this field will appreciate that PCR techniques may be exploited to selectively obtain cyathostomin larval antigen gene sequences from a variety of sources including, for example, equine gut tissue, faecal matter or extracts prepared from cyathostomin nematodes. In one embodiment, molecular cloned cyathostomin larval antigen gene sequences may be introduced into a vector (such as a plasmid or expression cassette). In one embodiment, the vector may further comprise a nucleotide sequence of a tag or label to assist in protein purification procedures.

A host cell may be transformed with the vector and maintained under conditions suitable to induce expression of the cyathostomin larval antigen gene sequence and production of recombinant cyathostomin larval antigen. Techniques used to purify recombinant proteins generated in this way are known and, where the recombinant protein is tagged or labelled, these may include the use of, for example, affinity chromatography techniques.

In view of the above, further aspects of this invention provide an expression vector comprising a cyathostomin larval antigen gene sequence and a host cell transformed therewith, respectively.

In a further aspect, the present invention provides a method for determining whether or not an equine subject should be treated with anthelmintic drug, said method comprising the step of detecting a level of anti-cyathostomin larval antigen antibodies in a sample as per the first aspect of this invention and/or a level of cyathostomin larval antigen in a sample, wherein a level of anti-cyathostomin larval antigen antibodies and/or antigen, is indicative of an equine subject that should

be administered a anthelmintic drug. In one embodiment, the anthelmintic drug may be Moxidectin.

DETAILED DESCRIPTION

The present invention will now be described in detail and with reference to the following Figures which show:

FIG. 1. ClustalW alignment of Cy-GALA-1 with its orthologues in other nematode species. Cyathostomin (Cy) GALA-1 is compared to *N. brasiliensis* keratin-like protein (Nb-KLP) (accession number: BAB68205); *T. circumcincta* (Tc) (AAM45145); *O. ostertagi* (Oo) (CAD22110); *C. elegans* (Ce) KLP-1 (NP 502026) and Ce-KLP-2 (NP 501448). The signal peptide for each sequence is underlined and the domain of unknown function (DUF148) is boxed. The histidine-rich region is highlighted in grey and the glycine-rich regions of the *C. elegans* sequences are shown in bold.

FIG. 2: Development transcription pattern of Cy-gala-1. RT-PCR was performed using gene-specific primers for Cy-gala-1 and the housekeeping gene cytochrome oxidase c subunit I (cox1), from mixed-species pools of EL3 (lane 1), DL (lane 2) and LP (lane 3) cDNA. For each reaction no-template controls were performed (N). Sizes in base pairs (bp) are labelled on the left-hand side.

FIGS. 3A and 3B. Immunoreactivity of rCy-GALA-1. IgG (T) reactivity to rCy-GALA-1 in horses infected with cyathostomins or other helminths as assessed by (A) immunoblot and (B) ELISA. FIG. 3A. Lane 1: Coomassie blue. Lanes 2-11: IgG(T) reactivity of specific equine sera: HF (2); CI (3); a pool of sera from cyathostomin-free horses (n=5) from an abattoir (4); a pool of sera from cyathostomin-infected horses which harboured total mucosal larval burdens of >100,000 (n=6) from an abattoir (5); horses mono-specifically infected with *P. equorum* (6), *S. edentatus* (7), *S. westeri* (8) or *S. vulgaris* (9). Also shown is IgG reactivity in sera from a rabbit before (lane 10) and after two immunisations (lane 11) with a 20 kDa complex purified from EL3/DL somatic extracts [11]. FIG. 3B. ELISA indicating IgG(T) reactivity to rCy-GALA-1 antigen in equine sera over an experimental infection [29]. Responses in the CI group are depicted by the solid lines and black shapes and in the HF group by dashed lines and white shapes.

FIG. 4: Reactivity of anti-rCy-GALA-1 antiserum to cyathostomins and other equine helminths. IgG(T) responses were assessed by (A) ELISA and (B) immunoblot. ELISA results depict binding of anti-rCy-KLP-1 anti-sera (black) and pre-immunisation serum (white). For both assays, the antigens were as follows: 1=rCy-GALA-1; 2=cyathostomin IL3; 3=cyathostomin EL3; 4=cyathostomin DL; 5=cyathostomin LP; 6=adult *A. perfoliata*; 7=adult *P. equorum*; 8=adult *S. edentatus*; 9=adult *S. vulgaris*; 10=adult *S. equinus*.

FIG. 5: Immunolocalisation of Cy-GALA. Transverse sections of DL cyathostomins were probed with anti-rCy-GALA-1 antiserum (A) and pre-immunization serum (B). Specific binding of antiserum in the parasite gut is indicated by the black arrows. The vertical bar represents 40 μ m.

FIG. 6: Schematic representation of Cy-gala-1 and the 220 bp fragment of the gene amplified from 10 cyathostomin species. Cy-gala-1 cDNA sequence is represented by black boxes (A). The 220 bp region PCR amplified from genomic DNA samples from 10 cyathostomin species is represented by the white box. The latter is expanded to indicate the position of the intron (hatched box). The range in interspecies variation for the whole gene fragment (and also without the intron sequence) are depicted. A representative PCR product of Cy-gala-1 is shown for each species (B): *C. catinatum* (1); *C. nassatus* (2); *C. goldi* (3); *C. longibursatus* (4); *C. coro-*

natum (5); *C. pateratum* (6); *C. ashworthi* (7); *C. leptosomum* (8); *C. minutus* (9) and *C. labiatus* (10).

FIG. 7A: Optimisation of antigen cocktails. The antibody response of encysted cyathostomin infected (positive) and non-infected (negative) animals is shown for varying concentrations of antigen and two different cocktails of antigen (CT1 and CT2). CT1 contains Gala 1, Gala 2, Gala 3. CT2 contains Gala 1, Gala 2, Gala 3 and CID 1. Individual antigen concentration is shown on the x axis and optical density (O.D) on the y axis. 7B: Ratio of signal for encysted cyathostomin infected (positive) to uninfected (negative) animals in an ELISA. Individual antigen concentration is on the x axis and ratio of positive to negative optical density on the y axis. C: shows mean serum antibody response to cocktail 1 (CT1) in groups of horses with varying infection levels. CT1 contains Gala 1, Gala 2, Gala 3. Horses were grouped as follows according to total mucosal parasite burden (TMB). Neg; uninfected horses TMB=0 (n=5), Low; TMB=0-20000, (n=8), Medium; TMB=20000-100000, (n=7), High; TMB=>100000 (n=26). Error bars show +/- standard error of the mean. O.D=optical density. D: shows mean serum antibody response to cocktail 2 (CT2) in groups of horses with varying infection levels. CT2 contains Gala 1, Gala 2, Gala 3 and CID 1. Horses were grouped as follows according to total mucosal parasite burden (TMB). Negative; uninfected horses TMB=0 (n=5), Low; TMB=0-20000, (n=8), Medium; TMB=20000-100000, (n=7), High; TMB=>100000 (n=26). Error bars show +/- standard error of the mean. O.D=optical density.

FIG. 8A-E: ROC analysis of ELISA data derived from cocktail (CT) 1 (which includes GALA-1, 2 and -3) and CT2 (which includes GALA-1, -2, -3 and CID-1). The Areas Under the Curve (AUC) are shown on each graph for each CT at the specified cyathostomin burden cut-off value indicated on each set of charts. The results indicate that CT1 and CT2 allow clear discrimination at different levels of cyathostomin mucosal burden, especially developing larval (DL) burdens above 120,000; however, it is likely that the AUC values could be improved by developing the assay to take into account cyathostomin species complexity and by including proteins that specifically relate to EL3. These additional proteins have been identified and will be added systematically to the cocktails to test their effect on AUC in the ROC analysis.'

MATERIALS AND METHODS

Parasite Material

Cyathostomins were collected from equine large intestinal tissue as described previously [9]. Briefly, caecum and ventral colon samples were removed at an abattoir and luminal parasites (LP), consisting of fifth stage larvae and adults, were collected from intestinal washings using sieves. Mucosal larval stages were recovered by pepsin-HCl digestion [9]. The mucosal parasites were separated into two populations based on size following previous recommendations [13]: (i) EL3 and (ii) late third stage (LL3)/developing fourth stage (DL4), collectively termed developing larvae (DL). Nematode samples for RNA extraction were placed into RNAlater (Ambion) at 4°C., while those for protein extraction and genomic DNA isolation were snap frozen in liquid nitrogen and stored at -80°C. For immunolocalisation experiments, DL were fixed in 10% formal saline. Infective third-stage larvae (IL3) were collected from horse feces as described previously [8]. Individual adult cyathostomins were identified to species according to published recommendations [16]. Adult stage large strongyles, *Anoplocephala perfoliata* and *Parascaris equorum*, were also obtained and stored at -80°C.

Construction of a Complementary (c)DNA Library and Immunoscreening

Cyathostomin RNA was extracted from DL populations by homogenisation in a mortar and pestle under liquid nitrogen, then using TRIzol (Invitrogen) according to the manufacturer's instructions. Integrity of RNA samples was assessed using a 2100 Bioanalyser (Agilent Technologies) and RNA stored in RNase-free water at -80°C . A mixed-species DL cDNA library was constructed using a SMART cDNA Library Construction Kit (Clontech Laboratories, Inc) using long distance PCR according to manufacturer's instructions. Briefly, the cDNA was synthesised by reverse transcriptase (RT)-PCR using $1\ \mu\text{g}$ total RNA pooled from 11 separate DL RNA samples collected over a 6-month period from a range of intestinal sites. This was done to maximise cyathostomin species representation within the cDNA library. After ligation into the λ Triplex2 vector, the cDNA was packaged into Giga-pack Gold III packaging extract (Stratagene) and amplified in *Escherichia coli* XL1-Blue strain, (Stratagene). Library quality was assessed by analysing insert size in 40 plaques chosen at random. Length and identity of the inserts were determined by PCR and sequencing; the majority of plaques contained an insert with an average size of 500 base pairs (bp).

An EL3 cDNA library was constructed using the same method as for the construction of the DL cDNA library with the exception being the use of a SL1 primer to amplify nematode specific DNA prior to ligation into the Triplex2 vector. (Martin, et al, 1995). Briefly, the cDNA was synthesised by reverse transcriptase (RT)-PCR using $1\ \mu\text{g}$ total RNA pooled from EL RNA samples from EL3 larvae collected from a range of intestinal sites from 6 individual horses. This cDNA was then used in a PCR with SL1 forward primer sequence: GGTTTAATTACCCAAGTTTGAG and reverse primer sequences: ATTCTAGAGGCCGAGGC and TTCTAGAGGCCGAGGCG. Products of this PCR were then used for packaging into the Triplex2 vector as described for generation of the DL cDNA library.

Immunoscreening was performed according to the manufacturer's protocol. For immunoscreening, two types of sera were used: cyathostomin-infected (CI) and helminth-free (HF) sera [29]. Ponies in the CI group ($n=3$) had been trickle infected with a total of 3.9 million cyathostomin IL3 over a period of 9 weeks, while the HF control group ($n=3$) were maintained helminth-free. Serum was obtained weekly from both groups. For immunoscreening, a pool of CI sera was prepared by combining samples obtained from the three ponies at 12, 13, 14 and 16 weeks PI. The pool of HF sera was made by combining samples obtained from the three ponies at 2, 3, 4 and 6 weeks before the start of the infection period. To reduce background reactivity, both pools of sera were pre-absorbed with *E. coli* lysate by incubating equal volumes of each and rocking for 4 h at room temperature [37]. After centrifugation at $18,000\times g$ for 10 min, the supernatant was retained for probing library filter lifts. The primary immunoscreen consisted of approximately 108,000 cDNA clones in *E. coli* XL1-Blue strain. Plaque lifts were made onto nitrocellulose filters (Hybond-C Extra, GE Healthcare). The membranes were washed [five \times 10 min in Tris-buffered saline (10 mM Tris, 150 mM NaCl, pH 7.4) containing 0.05% Tween-20 (TBST)], then blocked for 1 h with 1% gelatin/TBST. In the first screen, the serum pool from the CI ponies was used at 1:200 in TBST and incubated with the membranes overnight at 4°C . The secondary antibody (goat anti-equine IgG(T), Serotec) and tertiary antibody (rabbit anti-goat[IgG]:HRP, Sigma), were incubated at 1:200 and 1:500 respectively, for 1 h each, with washing (as above) between steps. Filters were developed using SIGMAFAST DAB with Metal Enhancer

(Sigma). Positive clones were isolated by taking agar plugs from the corresponding plate. Plaques that reacted non-specifically with equine sera (false positives) were identified by performing a second screen. Here, clones selected in the first round were screened as described above, except that filters were cut in half and one half probed with the CI serum pool and the other with the HF serum pool. Only plaques that reacted with the CI serum pool and not the HF serum pool were selected for sequence analysis. Vector-specific primers were used to amplify selected phage inserts and the PCR products purified using a QIAquick PCR Purification Kit (Qiagen). Each purified PCR product was sequenced using a commercial service (MWG Biotech). The resultant sequences were translated and searched against the GenBank 'non-redundant protein' database using BLASTp, and then against the 'non-human, non-mouse' EST database using tBLASTn, from the National Centre for Biotechnology Information [3]. Sequence alignments were performed using ClustalW2 [21] from the European Bioinformatics Institute (<http://www.ebi.ac.uk/Tools/clustalw2/>) and analysis for signal peptides performed using SignalP 3.0 [5]. Sequence identities were calculated using MegAlign 8.0.2 (DNASTAR) based on ClustalW alignments. Molecular mass estimations were made using an online tool from the Sequence Manipulation Suite (http://www.bioinformatics.org/sms2/protein_mw.html) and glycosylation sites identified using ExpASY Pro site (<http://ca.expasy.org/prosite/>).

RT-PCR to Determine Temporal Transcription Pattern of the mRNA Encoding the Cyathostomin Gut-Associated Larval Antigen-1 (Cy-GALA-1)

Stage-specific cDNA was synthesised from $1\ \mu\text{g}$ each of EL3, DL and LP total RNA using a SMART cDNA Library Construction Kit (Clontech Laboratories, Inc.). Briefly, first-strand cDNA was synthesised and amplified using the long-distance PCR method (22 cycles). Double-stranded cDNA was purified using a QIAquick PCR Purification Kit (Qiagen), eluted in $50\ \mu\text{l}$ dH₂O and stored at -20°C . until required. Integrity and loading of each cDNA population was assessed by amplifying a portion of the cytochrome oxidase c subunit I (coxI) gene using primers designed to conserved sequences among cyathostomins (sense: 5'-AAAAAGGAGGTGTTTTGGTTC-3' (SEQ ID NO: 62); antisense: 5'-CTTGAATTTGATAAACTACACC-3' (SEQ ID NO: 63)). PCR conditions were as follows: $0.3\ \mu\text{M}$ primers, $0.25\ \mu\text{M}$ dNTPs and $1.5\ \text{mM}$ MgCl₂ with the following cycling: 94°C . for 5 min, 40 cycles at 94°C . for 30 sec, 60°C . for 30 sec and 72°C . for 30 sec, with a final extension at 72°C . for 7 min. PCR was performed using Platinum Taq (Invitrogen) with $1\ \mu\text{l}$ cDNA from each developmental stage. Primers were designed for the most abundant immunoreactive clone identified in Section 2.2 and designated cyathostomin gut-associated larval antigen-1 (Cy-GALA-1). The primer sequences were as follows: sense, 5'-AATTGTGGGGAACAGGAG-3' (SEQ ID NO: 64); antisense, 5'-AATGAAAATCAGACTCCTAGG-3' (SEQ ID NO: 65). PCR conditions were as above, but using 35 cycles. This experiment was repeated twice and the PCR products were analysed on 2% w/v agarose gels using TrackIt 100 bp DNA Ladder (Invitrogen) for size determination. The gels were stained with 1 \times GelRed (Biotium).

Expression of Recombinant Cy-GALA-1

The Cy-GALA-1 clone from the library immunoscreen that contained the largest insert was chosen for expression of recombinant protein. This clone incorporated the full-length coding sequence of Cy-gala-1 including the putative initiating methionine, signal peptide and poly-A tail. Primers were designed to amplify the coding sequence of Cy-gala-1 (minus the sequence that encoded the signal peptide) for sub-cloning

into pET-22b(+) vector (Novagen). Appropriate sequences encoding flanking restriction enzyme sites were incorporated for uni-directional cloning. The primer sequences were as follows (NB: BamHI and HindIII sites underlined): sense 5'-AATTCGGATCCGCAAGGTGTCATGGACCTTTTGG-3' (SEQ ID NO: 66); antisense, 5'-CCGCAAGCT-TATATCTTTCATCTGTGTTGAGTCCAAAC-3' (SEQ ID NO: 67). The PCR step was performed as described above except that the annealing temperature was 58° C. and 30 cycles were used. The PCR product was purified as described above. The pET-22b(+) vector and PCR product were digested with BamHI and HindIII and ligation, using a 1:1 ratio of vector to PCR product, performed according to Novagen's protocol. Plasmids were transformed into *E. coli* JM109 Competent Cells (Promega) following manufacturer's instructions and selected on ampicillin-agar. A selection of colonies was subjected to colony PCR to ensure the presence of the cDNA encoding Cy-GALA-1. A colony which contained an insert of the correct estimated size was subjected to plasmid purification using a Wizard Plus SV Miniprep kit (Promega) and the purified plasmid was both sequenced and transformed into *E. coli* BL21-CodonPlus (DE3)-RIL competent cells (Stratagene) for expression of recombinant protein (rCy-GALA-1). Following induction with 1 mM isopropyl-beta-D-thiogalactopyranoside (Bioline), soluble rCy-GALA-1, present in the bacterial lysate supernatant, was purified on a His-trap HP column (GE Healthcare), following manufacturer's instructions. The purified protein was dialysed into phosphate buffered saline, pH 7.4 (PBS), using cellulose dialysis tubing (Sigma) and stored at -20° C. until required. Purified rCy-GALA-1 (0.5 µg) was separated by SDS-PAGE, and a band at the expected size excised and subjected to matrix-assisted laser desorption/ionization time-of-flight (MALDI-ToF-ToF) mass spectrometry using an Ultraflex II MALDI-ToF-ToF mass spectrometer (Bruker Daltonics). The identity of the protein was confirmed by comparing the peptide mass fingerprint (PMF) generated to the theoretical peptide mass fingerprint (PMF) of Cy-GALA-1.

Preparation of rCy-GALA-1 Antiserum in Rabbits

Anti-rCy-GALA-1 antiserum was generated by injecting a rabbit with 50 µg of rCy-GALA-1, in 0.5 mg/ml QuilA/PBS (1 ml total injection). A secondary injection was administered three weeks later, after which a test bleed indicated a specific antibody response to the recombinant antigen. This experiment was performed under the legislation of a UK Home Office License.

Immunoblotting

Soluble somatic antigen extracts were prepared from cyathostomin stages (IL3, EL3, DL and LP) and adult worms of other helminth species (*A. perfoliata*, *Strongylus equinus*, *Strongylus edentatus*, *Strongylus vulgaris* and *P. equorum*) as described previously [9]. However, IL3 were disrupted using a Ribolyser Fast Prep FP120 (Thermo Scientific) instead of a glass homogeniser. Proteins were separated on 4-12% polyacrylamide Bis-Tris gels (NuPAGE MES system, Invitrogen) according to the manufacturer's protocol. For immunoblotting, proteins were transferred to nitrocellulose membranes. To assess cross-reactivity, 0.1 µg rCy-GALA-1 was loaded onto lanes of a 15-well 12% NuPAGE gel using SeeBlue Plus2 protein standards (Invitrogen) for molecular weight estimations. In one lane, 0.4 µg was loaded and after electrophoresis was cut from the gel and stained with Coomassie blue. After transfer, the blot was sliced into separate lanes and blocked in TNTT (10 mM Tris, 0.5M NaCl, 0.05% Tween-20, 0.01% thimerosal, pH 7.4). Each of the following sera was used, diluted 1:200 in TNTT: CI and HF sera pools (described

above); a pool of 5 horses found to be cyathostomin-free (CF) from a local abattoir and a pool of 12 horses (from the same abattoir) with mucosal cyathostomin burdens of >100,000 (endemic infected—EI); horses mono-specifically infected with *S. edentatus* or *S. vulgaris* [20], *P. equorum* or *Strongylus westeri* [11]. Also tested was rabbit antiserum (and pre-immunisation samples) generated to the native 20 kDa cyathostomin complex [11]. Sera were incubated at room temperature for 1.5 h. Washing consisted of three, 5 min incubations in TNTT. The secondary and tertiary steps were as described for the immunoscreening (above), with the exception that the anti-20 kDa antiserum blots were incubated with goat anti-rabbit Ig:HRP (Dako) at 1:500. The blots were developed as for the library screen.

For detection of Cy-GALA-1 protein in somatic extracts of cyathostomins and other helminth species, somatic extracts (9 µg each antigen) were loaded onto 10-well, 4-12% NuPAGE gels, using 10 ng rCy-GALA-1 for comparison. After transfer to nitrocellulose, periodate treatment of the blots was performed as described previously [9]. The blots were probed with pre-immunisation rabbit serum and anti-rCy-GALA-1 serum at 1:300 in TNTT, followed by goat anti-rabbit(Ig):HRP (Dako) at 1:500, and developed as described above. Three 5 min washes were applied between steps.

Enzyme-Linked Immunosorbant Assay (ELISA)

To test reactivity of experimentally infected pony sera (CI) to rCy-GALA-1 over the course of infection, the following conditions were used. Each well of a Microlon High Binding plate (Greiner Bio-One) was coated with 100 µl of rCy-GALA-1 (1 µgml⁻¹ in bicarbonate coating buffer, 0.1 M, pH 9.6) overnight at 4° C. Plates were washed with 0.05% Tween-20 in PBS (PBST), six times. Block solution (2% soya infant powder (w/v) in PBST) was added, 200 µl per well, and incubated for 1 h at 37° C. Plates were washed six times and CI and HF sera (1:200 in block solution), from weekly time points 2 weeks before infection to 16 weeks PI, added and incubated for 2 h at 37° C. After washing, 100 µl goat anti-equine IgG(T) were added, diluted 1:200 in blocking solution. After 1 h at 37° C. and washing, 100 µl rabbit anti-goat (Ig):HRP were added, diluted in block at 1:500, and incubated for 1 h at 37° C. To develop the reaction, Sigma-FAST OPD tablets (Sigma) were dissolved in H₂O according to the manufacturer's instructions and 100 µl added to each well and incubated for 15 min. Fifty µl of 2.5 M H₂SO₄ were added to stop the reaction and the absorbance read at 490 nm. The same conditions were used to measure the anti-rCy-GALA-1 antiserum response to somatic extracts of cyathostomin stages and other adult helminth species extracts, except that these were coated at 2 µgml⁻¹. The antiserum and goat anti-rabbit:HRP were used at 1:500.

Immunolocalisation

Cyathostomin DL were fixed in 10% formal saline and immobilised in a solidified gelatin plug by mixing with molten 5% gelatin/PBS (<30° C.) and allowing to set. The plugs were then dehydrated with alcohol and xylene and embedded in paraffin wax. Sections were cut at 3 µm using a microtome and the slides stored at 4° C. Immunolocalisation was performed using an EnVision+ System-HRP for rabbit primary antibodies (DakoCytomation) in a Sequenza Slide Rack (Thermo Scientific) at room temperature. After de-waxing, the slides were incubated in 0.5% Tween-80/PBS (PBST80) with 0.3% H₂O₂ for 20 min, to inactivate endogenous peroxidases. Blocking was performed using 100 µl 25% normal goat serum (NGS) in PBST80 for 1 h. Rabbit antisera obtained prior to and after two immunisations with rCy-GALA-1 were diluted 1:100 in 10% NGS/PBST80, and 100 µl incubated on

the slides for 1 h. After two washes in PBS at room temperature, 100 μ l of HRP-labelled polymer conjugated to goat anti-rabbit Ig, was incubated (neat) for 30 min. The reactions were developed in neat 3-amino-9-ethylcarbazole substrate chromogen for 7.5 min. Slides were washed in H₂O and counterstained using haematoxylin.

Single Worm PCR to Identify the Gene Encoding GALA in Different Cyathostomin Species

Genomic DNA was isolated from 54 individually identified adult cyathostomins using the DNeasy Blood and Tissue kit (Qiagen) according to their protocol, but with the addition of a homogenisation step before the proteinase K digestion step; each individual was disrupted briefly using a 1.5 ml microfuge tube homogeniser in 50 μ l ATL buffer supplied with the kit. The following 10 species were examined (NB: numbers of worms used for each species is shown in parenthesis): *Cyathostomum catinatum* (10), *Cylicostephanus goldi* (8), *Coronocyclus coronatus* (6), *Cyathostomum pat-
eratum* (6), *Cylicocyclus nassatus* (6), *Cylicostephanus longibursatus* (5), *Cylicocyclus ashworthi* (4), *Cylicocyclus leptostomum* (3), *Coronocyclus labiatus* (1) and *Cylicostephanus minutus* (1). The same primers used in Section 2.3 for RT-PCR were used to amplify a conserved fragment of Cy-gala in each species. The cycling conditions were: 2 min at 94° C., followed by 40 cycles of 15 sec at 94° C., 30 sec at 58° C. and 60 sec at 72° C., and a final extension at 72° C. for 7 min. PCR products were analysed on agarose gels as described above and PCR products from each of the 54 individuals cloned into pGEMT-Easy (Promega) according to manufacturer's instructions. Each clone was sequenced in forward and reverse directions with vector-specific primers using the commercial sequence facility described above.

Results

Immunoscreening of the Cyathostomin DL cDNA Library and Sequence Analysis of Cy-Gala-1

The primary immunoscreening yielded 33 positive clones; five of which were excluded as false positives on the basis of the secondary screen using HF sera. The remaining 28 clones contained inserts ranging in size from approximately 500 to 1500 bp. Sequence analysis indicated that 15 of these showed high identity to one another (73-100% at the amino acid [aa] level). One of these (Cy-gala-1) represented a full-length coding sequence: i.e. it contained a putative initiation codon, signal peptide and termination codon upstream of a poly-A tail. The entire coding sequence was 223 aa which, after cleavage of the signal peptide, would result in a 206 aa mature protein estimated at 25.6 kDa. Cy-GALA-1 contains a highly conserved domain as revealed by a domain search via BLASTp analysis [28]. The function of this domain is unknown and in *Caenorhabditis elegans* is designated Domain of Unknown Function 148 (DUF148). The Cy-GALA-1 sequence displayed highest aa identity to a sequence from *Nippostrongylus brasiliensis* (accession number: BAB68205; 35% identity over 128 residues), also identified via immunoscreening [38]. Two predicted proteins from *C. elegans* showed 34% identity over 105 residues to Cy-GALA-1. These proteins were 44.5% identical to each other. Also identified, were two trichostrongyloid ESTs: one from *Teladorsagia circumcincta* L3 (accession number: AAM45145), which displayed 32% identity to Cy-GALA-1 over 102 aa, and one from *Ostertagia osteragi* adult worms (accession number: CAD22110) with showed 32% identity over 140 aa. The two *C. elegans* orthologues (referred to here as Ce-KLP-1 (NP_502026) and Ce-KLP-2 (NP_501448)) contain glycine-rich domains which gives them homology to keratin sequences and hence their designation as 'keratin-like' proteins (KLP). All the parasitic nematode sequences

described here lack this glycine rich sequence, despite some being previously designated as 'KLP-like' proteins [38]. Rather than classifying Cy-GALA-1 as a KLP, it was instead named to reflect its localisation to the gut (see below). An alignment of Cy-GALA-1 with its orthologous sequences in *N. brasiliensis* and *C. elegans* is depicted in FIG. 1. In all the parasitic nematode sequences, except that of *T. circumcincta*, a histidine-rich motif precedes DUF148 (FIG. 1); its function is unknown. In addition, four potential N-linked glycosylation sites were identified. Searching Cy-GALA-1 at Nembase gave additional significant hits. All of these EST sequences contained regions with high identity to DUF148 and some had glycine-rich regions. The closest matches were to sequences identified in adult *Ancylostoma ceylanicum* (accession numbers: CB176510, CB190303 and CB339159), with 45-46% aa identity to Cy-GALA-1 over 110 residues.

Temporal Transcription Pattern of Cy-Gala-1

Cy-gala-1 transcript was detected in DL and EL3 cDNA and not in cDNA from LP parasites (FIG. 2). After 40 cycles, similar levels of coxI PCR product were observed in DL and LP cDNA. However, a coxI PCR product from EL3 was less intense, indicating low quality of EL3 cDNA. This was due to degradation of EL3 RNA caused by the extensive digestion method required to harvest these larvae. These results indicate the apparent specificity of this transcript for mucosal stages; hence the gene was selected for expression of recombinant protein for assessment as a diagnostic marker.

Expression of rCy-GALA-1 and its Immunoreactivity

rCy-GALA-1 was obtained from the soluble fraction of the *E. coli* lysate; the purified protein was approximately 28 kDa (FIG. 3A). The identity of this protein as rCy-GALA-1 was confirmed by MALDI-ToF-ToF (data not shown). Its molecular weight was slightly higher than the expected size of native Cy-GALA, calculated to be 25.6 kDa, and was due to addition of the His-tag and *E. coli* signal peptide. Anti-rCy-GALA-1 antiserum predominantly recognised the expected size band in somatic DL extracts (Section 3.4 and FIG. 4). The immunoreactivity of the recombinant antigen is shown in (FIG. 3B). Only IgG(T) in CI and EI sera equine sera bound rCy-GALA-1, indicating that both experimentally and naturally infected horses recognise this antigen. Sera from horses harbouring other parasitic helminths did not contain IgG(T) that bound Cy-GALA-1. The rabbit antiserum to the cyathostomin larval anti-20 kDa complex generated previously [11], showed strong reactivity to rCy-GALA-1.

Levels of rCy-GALA-1-specific IgG(T) in sera from infected vs. non-infected ponies [29] were measured by ELISA (FIG. 3B). Increases in rCy-GALA-1-specific IgG(T) levels were observed in all infected ponies by 6 weeks PI. A more rapid increase was observed in pony 104. Antigen-specific IgG(T) levels plateaued at 8 weeks PI for 104 and 12 weeks PI for 101 and 105; these levels remained elevated until the end of the measurement period at 16 weeks PI. No significant increases in rCy-GALA-1-specific IgG(T) levels were observed in any of the HF ponies throughout the experiment. Murphy and Love (1997) [29] described clinical signs in the infected animals from 4-6 weeks PI. While all showed a slower increase in percentage weight gain than the control group, pony 104 showed a drop in weight gain over weeks 4-8 PI. These signs may indicate a higher level of infection in 104. Anti-rCy-GALA-1 antiserum reactivity was tested against somatic extracts from *A. perfoliata*, *P. equorum*, *S. edentatus*, *S. vulgaris* and *S. equorum* [FIG. 4]. No reactivity was observed except to a band at 38 kDa in the *P. equorum* extract. Binding to this band was less than that seen in the cyathostomin DL lane (FIG. 4).

Detection of Cy-GALA-1 in Different Cyathostomin Stages

Antiserum raised to rCy-GALA-1 was used to investigate the presence of the native protein in different cyathostomin stages (FIG. 4). This antiserum bound the 28 kDa recombinant antigen (FIG. 4, lane 1); an additional band at 53 kDa was bound and may represent a dimeric form of Cy-GALA-1. The anti-rCy-GALA-1 antisera showed reactivity to EL3 and DL somatic extracts but not to adult extract (FIG. 4). Immunoreactivity to antigens in EL3 and DL stages was primarily directed at molecules of approximately 26 kDa, corresponding to the calculated molecular mass of Cy-GALA-1. Two other EL3 and DL antigens were bound by IgG in anti-Cy-GALA-1 antisera, one at approximately 45 kDa and the other at 55 kDa. The ELISA results indicated high reactivity to DL, however no binding was observed in EL3 or adult extract.

Immunolocalisation of Cy-GALA-1

DL were subjected to immunolocalisation studies (FIG. 5). Reactivity was detected in the gut of individual worms, where considerable staining was observed on the gut epithelium and in the gut lumen. No reactivity was detected to any other structures in the nematodes.

Single Worm PCR to Identify the Gene Encoding Cy-GALA in Different Cyathostomin Species

Single worm PCR experiments were performed using primers to amplify a 220 bp fragment of Cy-gala-1 from 50 morphologically-identified adult worms encompassing 10 species. A PCR product was obtained from all nematodes tested and sequencing confirmed that PCR products representative of each species encoded Cy-gala sequence. FIG. 6 shows a schematic representation of this fragment and PCR products from each species. There was variation in size of the PCR product obtained from different species; from 267 bp (for all *C. coronatus* individuals) to 284 bp (for one *C. goldi* individual). This variation was due to a difference in intron size at this site amongst the species. The precise location of the intron was conserved as indicated by splice site analysis (FIG. 6). Nucleotide identities between individuals from different species ranged from 78.9-99.1% for the whole fragment. Higher nucleotide identities were observed in the coding region; interspecies variation ranged from 82.2-98.9% over 180 nt, while the amino acid identities were 80-100% over 60 residues. At the aa level, intra-species variation was as follows: *C. catinatum* 93.3-100%; *Cs. goldi* 90.0-100.0%; *Co. coronatus* 96.7-100.0%; *C. pateratum* 93.3-100%; *Cc. nassatus* 88.3-98.3%; *Cs. longibursatus* 91.7-100%; *Cc. ashworthi* 90.0-100.0%; *Cc. leptostomum* 88.3-100.0%. In an attempt to assign a species for the library clone, Cy-gala-1, the coding sequence from each individual was compared against Cy-gala-1 in this 220 bp fragment. The highest identity was found to a *C. pateratum* individual (97.8% nt identity and 98.3% aa identity). Therefore, with the available sequence data for each species, we have provisionally identified Cy-gala-1 as belonging to *C. pateratum*.

Optimisation of Antigen Cocktails.

The optimum concentration of antigen to use in an ELISA using a cocktail of antigens was evaluated using sera from cyathostomin infected (positive) and non-infected (negative) animals. FIG. 7A shows the serum antibody response to varying concentrations of antigen in two different cocktails of antigen (CT1 and CT2). CT1 contains GALA-1, -Gala 2 and -3. CT2 contains these three antigens plus CID1. Individual antigen concentration is shown on the x-axis and optical density (O.D) on the y-axis. FIG. 7B shows the ratio of the OD signal obtained on cyathostomin infected (positive) vs. uninfected (negative) animals in an ELISA. Individual antigen concentration is on the x-axis and ratio of positive to negative optical density on the y-axis.

Evaluation of Antigen Cocktail for Discriminating Different Levels of Infection.

Two different cocktails of antigen were tested in an ELISA to assess their potential for discriminating different levels of mucosal infection. FIGS. 7C and D shows mean serum antibody response to cocktail 1 (CT1) and cocktail 2 (CT2) respectively in groups of horses with varying infection levels. CT1 and CT2 were as described above. Horses were grouped as follows according to total mucosal parasite burden (TMB). Neg; uninfected horses TMB=0 (n=5), Low; TMB=0-20000, (n=8), Medium; TMB=20000-100000, (n=7), High; TMB=>100000 (n=26). Error bars show +/- standard error of the mean. O.D=optical density.

DISCUSSION

Identification of the cyathostomin GALA sequence is an advance in the development of an ELISA for the diagnosis of larval cyathostominosis. Three important criteria were met by this protein: 1) it appeared to be specific to larval stages; 2) there was no cross reactivity with the other equine helminth species assessed here and 3) the gene encoding the protein was isolated from all cyathostomin species examined with a relatively low level of sequence variation amongst the species. Furthermore, serum IgG(T) responses to rCy-GALA-1 increased within 5 weeks of the administration of an experimental infection and the protein was also the target of IgG(T) responses in naturally infected horses.

The RT-PCR, immunoblot and ELISA results indicated that Cy-GALA-1 is restricted to parasitic larval stages, particularly DL stages. This is a vital feature for a diagnostic marker that specifically indicates mucosal larval burden. Despite numerous attempts, RNA extracted from EL3 was of relatively poor quality so it was difficult to judge precise levels of transcription in these stages. EL3 require extensive digestion in pepsin/HCl at 37° C. to remove them in sufficient quantity from the intestinal mucosa and submucosa and so it is technically difficult to obtain sufficient high quality RNA. The EL3 somatic protein extracts also contained a small amount of contaminating host protein (it is impossible to totally separate every single worm from its host capsule), and this may have resulted in the lower levels of reactivity of EL3 extracts to Cy-GALA-1 antiserum as indicated by the ELISA results. Immunolocalisation was also attempted in EL3, but degradation resulted in a lack of distinct morphology and no specific binding was observed (data not shown). Therefore it remains to be fully elucidated if Cy-GALA is a significant immunogen of EL3, or is predominantly an antigen of the later larval stages. Immunolocalisation studies of diseased equine mucosa are planned, to provide EL3 embedded in their mucosal cysts.

Serum IgG(T) responses to rCy-GALA-1 over the time course of an experimental infection showed that the antigen is a reasonably early indicator of infection and these responses were identified whilst the infections were not patent [29]. Indeed, in these ponies, the infections never progressed to patency even though the experiment was continued until 60 and 62 weeks PI in two of the animals. Substantial increases in reactivity were observed at 5 weeks PI in one animal (pony 104) and by 6 weeks in all ponies. cyathostomin larval-specific serum IgG(T) responses were analysed previously in these animals and similar dynamics of responses were observed to the 20 and 25 kDa complexes purified from EL3/DL mixtures [11]. Furthermore, serum IgG(T) reactivity to crude larval antigen was also observed to increase only after 6 weeks PI in these ponies [9], suggesting that only by this time point do larvae stimulate a detectable serum IgG(T)

response. Pony 104 had the most pronounced increase in IgG(T) to rCy-GALA-1 and this is similar to its response to crude larval antigen and the purified 20- and 25-kDa antigen complexes [9, 11]. The clinical signs observed in this pony (reduced weight gain, lowest plasma fructosamine) indicate that it may have had a greater burden of mucosal larvae [29]. Indeed, when this animal was euthanized at 20 weeks PI it was found to have a high cyathostomin burden. Unfortunately the other two ponies in the group were necropsied at 60 and 62 weeks PI so their burdens cannot be directly compared with pony 104. Nevertheless, the data provides preliminary evidence that this recombinant antigen may be able to distinguish varying degrees of disease.

As mentioned above, there is similarity of the IgG(T) response to rCy-GALA-1 and to the two larval antigen complexes purified and shown to have diagnostic potential previously [10, 11]. The molecular mass of Cy-GALA, estimated at 25.6 kDa, means that it could feasibly be a component of the 25 kDa antigen complex, an observation supported by the results using anti-rCy-GALA-1 against EL and DL somatic extracts in western blots. Antiserum generated to the 20 kDa complex in rabbits also bound rCy-GALA-1 indicating its presence in this complex also. This is not altogether surprising as these complexes were excised rather crudely from SDS-polyacrylamide gels [10, 11].

Specificity of Cy-GALA-1 in the cyathostominae was confirmed by probing the recombinant protein with sera from horses infected mono-specifically with heterologous helminth species. While experimentally infected (CI) and naturally infected (EI) horses recognised rCy-GALA-1, IgG(T) in serum from horses with large strongyle infections (*S. edentatus*, *S. westeri* or *S. vulgaris*) and *P. equorum* infection, did not bind the antigen. Cross-reactivity was further explored by probing somatic extracts of other equine parasites with anti-rCy-GALA-1 serum: extracts from *A. perfoliata*, *P. equorum*, *S. edentatus*, *S. vulgaris* and *S. equorum* were analysed. In the ELISA no binding above background levels was observed in any of the five other parasite extracts. In the immunoblot, there was a degree of binding to a band of approximately 38 kDa in the *P. equorum* extract, but this was of far less intensity than binding observed in the cyathostomin DL samples. Furthermore, there was no cross reactivity to *P. equorum* antigens when the samples were assessed using the ELISA.

The presence of sequences encoding GALA-like proteins was confirmed in 10 cyathostomin species, indicating ubiquity of this gene in the group. There are currently 50 recognised cyathostomin species [23], and while a large number of species are often found in infected individuals [6, 7], the bulk of the burden is consistently found to comprise 5-10 species [26, 27, 36]. Nine of the species explored in this study belong to the 10 most common cyathostomins as identified by Reinemeyer et al. (1984) [36], Ogbourne (1976) [30] and Lichtenfels et al 2001 [22]. The presence of Cy-GALA in these species indicates it is likely to be present in most, if not all, cyathostomins. An analysis of the sequence of Cy-gala-1 amongst the cyathostomins indicated a low level of sequence diversity across the selected 220 bp region. It is possible that greater diversity exists outside this region and the full-length cDNA sequences of Cy-gala are currently being isolated from a number of species to investigate this further. Promisingly, for development of a specific immunoassay, the levels of sequence diversity identified thus far are substantially lower among cyathostomins than they are when the Cy-gala sequences are compared to orthologous sequences in other nematode species, i.e. 80-100% vs. 25-35% identity. The nematodes that were present in the CI pony group unfortunately

had not been identified, so it is difficult to compare levels of rCy-GALA-1 IgG(T) with the species present.

A factor that must be considered in the development of any helminth immunodiagnostic assay is the length of time that circulating specific immunoglobulin levels take to return to normal values after anthelmintic treatment. Since the ponies used in the experimental infection were not treated with anthelmintic before necropsy, this could not be assessed here. Studies on a commercially-available serological ELISA for *A. perfoliata* [33, 34], which is based on the specific binding of IgG(T) to a purified 12/13 kDa antigen complex, indicated that post-treatment IgG(T) levels can take months to reduce to 'non-infection' levels [2, 4]. Also, Kjaer et al. (2007) [18] found that two thirds of horses which had no visible signs of tapeworm infection at necropsy had ELISA ODs higher than the current accepted cut-off for infection (0.2). Despite this, the *A. perfoliata* 12/13 kDa antigen ELISA is still regarded as the most useful diagnostic tool for infection [1, 18]. These observations suggest that circulating IgG(T) levels may remain high for a time after treatment and this will be considered when designing how a cyathostomin diagnostic assay, based on IgG(T), could be used in future.

No function has been ascribed to orthologues of Cy-GALA in other nematode species and only Nb-KLP has been characterised in any detail [38]. It was speculated that Nb-KLP may be a cuticular protein, based on its identity to Ce-KLPs, which are described as 'keratin-like'. However the authors did not explore this further. Ce-KLP-1 and -2 encode hypothetical proteins, and some information regarding these is available in WormBase (www.wormbase.org). Both are predicted to be alpha-helical proteins, and Ce-KLP-1 has been confirmed by transcript evidence, while Ce-KLP-2 has been partially confirmed. Ce-KLP-1 shows no RNAi phenotype, while Ce-KLP-2 displays 'embryonic lethal', indicating that it may play a role in development. An anatomic expression plan is available for Ce-KLP-2, showing expression in pharyngeal muscles and tail neurons which is different to what was observed here with localisation of Cy-GALA to the worm intestinal lumen. The function of this molecule remains to be elucidated.

REFERENCES

- [1] Abbott J. B., Barrett E. J., The problem of diagnosing tapeworm infections in horses, *Equine Vet. J.* (2008) 40:5-6.
- [2] Abbott J. B., Mellor D. J., Barrett E. J., Proudman C. J., Love S., Serological changes observed in horses infected with *Anoplocephala perfoliata* after treatment with praziquantel and natural reinfection, *Vet. Rec.* (2008) 162:50-3.
- [3] Altschul S. F., Gish W., Miller W., Myers E. W., Lipman D. J., Basic Local Alignment Search Tool, *J. Mol. Biol.* (1990) 215:403-410.
- [4] Barrett E. J., Farlam J., Proudman C. J., Field trial of the efficacy of a combination of ivermectin and praziquantel in horses infected with roundworms and tapeworms, *Vet. Rec.* (2004) 154:323-325.
- [5] Bendtsen J. D., Nielsen H., von Heijne G., Brunak S., Improved prediction of signal peptides: SignalP 3.0, *J. Mol. Biol.* (2004) 340:783-95.
- [6] Bucknell D. G., Gasser R. B., Beveridge I., The prevalence and epidemiology of gastrointestinal parasites of horses in Victoria, Australia, *Int. J. Parasitol.* (1995) 25:711-24.

- [7] Chapman M. R., Kearney M. T., Klei T. R., Equine cyathostome populations: accuracy of species composition estimations, *Vet. Parasitol.* (2003) 116:15-21.
- [8] Clark H. J., Kaplan R. M., Matthews J. B., Hodgkinson J. E., Isolation and characterisation of a beta tubulin isotype 2 gene from two species of cyathostomin, *Int. J. Parasitol.* (2005) 35:349-58.
- [9] Dowdall S. M., Matthews J. B., Mair T., Murphy D., Love S., Proudman C. J., Antigen-specific IgG(T) responses in natural and experimental cyathostominae infection in horses, *Vet. Parasitol.* (2002) 106:225-42.
- [10] Dowdall S. M., Proudman C. J., Klei T. R., Mair T., Matthews J. B., Characterisation of IgG(T) serum antibody responses to two larval antigen complexes in horses naturally- or experimentally-infected with cyathostomins, *Int. J. Parasitol.* (2004) 34:101-8.
- [11] Dowdall S. M., Proudman C. J., Love S., Klei T. R., Matthews J. B., Purification and analyses of the specificity of two putative diagnostic antigens for larval cyathostomin infection in horses, *Res. Vet. Sci.* (2003) 75:223-9.
- [12] Eysker M., Boersema J. H., Kooyman F. N. J., The effect of ivermectin treatment against inhibited early 3rd stage, late 3rd stage and 4th stage sarvae and adult stages of the cyathostomes in Shetland ponies and spontaneous expulsion of these helminths, *Vet. Parasitol.* (1992) 42:295-302.
- [13] Eysker M., Klei T. R., Mucosal larval recovery techniques of cyathostomes: can they be standardized?, *Vet. Parasitol.* (1999) 85:137-44.
- [14] Geldhof P., Vercauteren I., Knox D., De Maere V., Van Zeveren A., Berx G., Vercruyse J., Protein disulphide isomerase of *Ostertagia ostertagi*: an excretory-secretory product of L4 and adult worms? *Int. J. Parasitol.* (2003) 33:129-36.
- [15] Giles C. J., Urquhart K. A., Longstaffe J. A., Larval cyathostomiasis (immature trichonema-induced enteropathy): a report of 15 clinical cases, *Equine Vet. J.* (1985) 17:196-201.
- [16] Hodgkinson J. E., Love S., Lichtenfels J. R., Palfreman S., Ramsey Y. H., Matthews J. B., Evaluation of the specificity of five oligoprobes for identification of cyathostomin species from horses, *Int. J. Parasitol.* (2001) 31:197-204.
- [17] Kaplan R. M., Anthelmintic resistance in nematodes of horses, *Vet. Res.* (2002) 33:491-507.
- [18] Kjaer L. N., Lungholt M. M., Nielsen M. K., Olsen S. N., Maddox-Hyttel C., Interpretation of serum antibody response to *Anoplocephala perfoliata* in relation to parasite burden and faecal egg count, *Equine Vet. J.* (2007) 39:529-33.
- [19] Klei T. R., Chapman M. R., French D. D., Taylor H. W., Evaluation of ivermectin at an elevated dose against encysted equine cyathostome larvae, *Vet. Parasitol.* (1993) 47:99-106.
- [20] Klei T. R., Torbert B. J., Chapman M. R., Ochoa R., Irradiated larval vaccination of ponies against *Strongylus vulgaris*, *J. Parasitol.* (1982) 68:561-9.
- [21] Larkin M. A., Blackshields G., Brown N. P., Chenna R., McGettigan P. A., McWilliam H., Valentin F., Wallace I. M., Wilm A., Lopez R., Thompson J. D., Gibson T. J., Higgins D. G., Clustal W and clustal X version 2.0, *Bioinformatics* (2007) 23:2947-8.

- [22] Lichtenfels J. R., McDonnell A., Love S., Matthews J. B., Nematodes of the tribe Cyathostominae (Strongylidae) collected from horses in Scotland, *Comp. Parasitol.*, (2001) 68:265-269.
- [23] Lichtenfels J. R., Kharchenko V. A., Dvojnos G. M., Illustrated identification keys to strongylid parasites (Strongylidae: Nematoda) of horses, zebras and asses (Equidae), *Vet. Parasitol.* (2008) 156:4-161.
- [24] Love S., Duncan J. L., The development of naturally acquired cyathostome infection in ponies, *Vet. Parasitol.* (1992) 44:127-42.
- [25] Love S., Murphy D., Mellor D., Pathogenicity of cyathostome infection, *Vet. Parasitol.* (1999) 85:113-21.
- [26] Lyons E. T., Tolliver S. C., Drudge J. H., Historical perspective of cyathostomes: prevalence, treatment and control programs, *Vet. Parasitol.* (1999) 85:97-111.
- [27] Lyons E. T., Tolliver S. C., Drudge J. H., Stamper S., Swerczek T. W., Granstrom, D. E., A study (1977-1992) of population dynamics of endoparasites featuring benzimidazole-resistant small strongyles (Population S) in Shetland ponies, *Vet. Parasitol.* (1996) 66:75-86.
- [28] Marchler-Bauer A., Anderson J. B., Derbyshire M. K., Weese-Scott C., Gonzales N. R., Gwadz M., Hao, L. N., He S. Q., Hurwitz D. I., Jackson J. D., Ke Z. X., Krylov D., Lanczycki C. J., Liebert C. A., Liu C. L., Lu F., Lu S. N., Marchler G. H., Mullokandov M., Song J. S., Thanki, N., Yamashita, R. A., Yin, J. J., Zhang, D. C., Bryant S. H., CDD: a conserved domain database for interactive domain family analysis, *Nucleic Acids Res.* (2007) 35:D237-D240.
- [29] Murphy D., Love S., The pathogenic effects of experimental cyathostome infections in ponies, *Vet. Parasitol.* (1997) 70:99-110.
- [30] Ogbourne C. P., The prevalence, relative abundance and site distribution of nematodes of the subfamily Cyathostominae in horses killed in Britain, *J. Helminthol.* (1976) 50:203-14.
- [31] Paul J. W., Equine larval cyathostomosis, *Comp. Cont. Educ. Pract. Vet.* (1998) 20:509-515.
- [32] Proudman C. J., Matthews J., Control of intestinal parasites in horses, *In Pract.* (2000) 22:90-97.
- [33] Proudman C. J., Trees A. J., Correlation of antigen specific IgG and IgG(T) responses with *Anoplocephala perfoliata* infection intensity in the horse, *Parasite Immunol.* (1996) 18:499-506.
- [34] Proudman C. J., Trees A. J., Use of excretory/secretory antigens for the serodiagnosis of *Anoplocephala perfoliata* cestodosis, *Vet. Parasitol.* (1996) 61:239-47.
- [35] Reid S. W. J., Mair T. S., Hillyer M. H., Love, S., Epidemiologic risk-factors associated with a diagnosis of clinical cyathostomiasis in the horse, *Equine Vet. J.* (1995) 27:127-30.
- [36] Reinemeyer C. R., Smith S. A., Gabel A. A., Herd R. P., The prevalence and intensity of internal parasites of horses in the USA, *Vet. Parasitol.* (1984) 15:75-83.
- [37] Sambrook J., Fritsch E., Maniatis T., *Molecular cloning: a laboratory manual*, Cold Harbor Spring Laboratory Press, second edition, 1989.
- [38] Shibui A., Takamoto M., Shi Y., Komiyama A., Sugane K., Cloning and characterization of a novel gene encoding keratin-like protein from nematode *Nippostrongylus brasiliensis*, *BBA-Gene Structure and Expression* (2001)
- [40] Martin S. A. M., Thompson F. J., Devaney E., The construction of spliced leader cDNA libraries from the filarial nematode *Brugia pahangi*, *Molecular and Biochemical Parasitology*, (1995)

Additional immunoreactive clones: The following table lists additional sequences encoding immunoreactive cyathostomin antigens which were identified from the larval cDNA library, from two screenings which revealed distinct clones. The first immunoscreen (A) used serum from experimentally cyathostomin-infected ponies (from a previous study¹), and the second immunoscreen (B) used a pool of sera from naturally infected horses; both groups had high parasite burdens. The antigens which have been checked for immunogenicity and cross specificity by recombinant bacterial expression are also indicated.

Antigen	Number of clones per screen		Closest homologues (Accession numbers in brackets)	Amino acid identity	Transcription pattern			Immuno-genicity	Cross specificity
	A	B			EL3	DL	LP		
Gut-associated larval antigen (GALA)	15	1	Keratin-like protein, <i>Nippostrongylus brasiliensis</i> (BAB68205). Keratin-like proteins, <i>Caenorhabditis elegans</i> (NP 502026 and NP 501448)	35% over 128 a.a. 34% over 104 a.a.	+	++	-	+	+
Glutathione-S-transferase (GST)	1	0	Cytosolic GST from <i>Oesophagostomum dentatum</i> (ACA30415)	85% over 209 a.a.	-	+	+		
Galectin-1 (GAL-1)	1	0	Galectin family member, <i>C. elegans</i> (NP 495163)	83% over 279 a.a.	-	++	+		
Galectin-2 (GAL-2)	0	1	Galectin family member, <i>Haemonchus contortus</i> (AAF63406)	91% over 259 a.a.	+	++	++		
Nematode polyprotein allergen/antigen (NPA)	4	0	NPA from <i>Dictyocaulus viviparus</i> (Q24702)	42% over 314 a.a.	-	++	++		
Cyathostomin immunodominant antigen-1 (CID-1)	3	0	EST from larval-stage <i>Necator americanus</i> (BG467549). Function of this is unknown.	59% over 61 a.a.	-	++	++	+	
Surface associated antigen (SAA)	4	8	SAA-2, <i>N. americanus</i> (ACE79378)	71% over 146 a.a.	+	++	++		
Fatty acid/retinol binding protein-1 (FAR-1)	0	1	Putative ES protein with FAR binding domain, <i>Ostertagia ostertagi</i> (CAD20464)	45% over 100 a.a.	+	++	++		
Fatty acid/retinol binding protein-2 (FAR-2)	0	1	FAR binding protein, <i>Ancylostoma ceylanicum</i> (ACC76809)	72% over 160 a.a.	-	++	+		
Globin (GLO)	0	15	Cuticle globin, from <i>Syngamus trachea</i> (AAL56426)	54% over 161 a.a.	+	++	++		
Clone of unknown function-20a (Unk-20a)	0	1	No homology found	NA	-	++	++		
Unk-46a	0	1	Third-stage larval EST, <i>N. brasiliensis</i> (EH359049)	33% over 124 a.a.	-	++	++		
Unk-50a	0	1	Hypothetical protein, <i>C. elegans</i> (NP 490737)	33% over 140 a.a.	-	++	+		

¹Murphy D., Love S., The pathogenic effects of experimental cyathostome infections in ponies, Vet. Parasitol. (1997) 70: 99-110.

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Homologues of cyathostomin gut-associated larval antigen (Cy-GALA)			Homologues of cyathostomin gut-associated larval antigen (Cy-GALA)		
Homologue	Amino acid identity to Cy-GALA-1	Putative species	Homologue	Amino acid identity to Cy-GALA-1	Putative species
Cy-GALA-1	—	<i>C. pateratum</i>	Cy-GALA-3	77.7%	<i>C. coronatus</i>
Cy-GALA-2	83.3%	<i>C. nassatus</i>	Cy-GALA-4	93.0%	<i>C. catinatum</i>

Sequences of a conserved region of GALA from individual cyathostomin species

Species	Sequence	Number of individuals	Intraspecies aa identity range
<i>C. ashworthi</i>	LTFAEKKGKISEWAKKYNVDEVASYNAYREKLKQEHRKNVS (E/V) LVSGLP (G/D) AVKKVN (E/V) LLD (SEQ ID NO: 68)	4	90.0-100%
<i>C. catinatum</i>	LTFAEKK (E/K) EISEWAKKYNVDEVASYNAYREKLKQEHRKNVSELVSALP NAVKKVNDLLD (SEQ ID NO: 69)	10	91.7-100%
<i>C. coronatus</i>	LTFAEKKKISEWAKKYKVEDEVASYNAYREKLKQEHRKNVSELVSALPGA VK KVNELLD (SEQ ID NO: 70)	6	96.7-100%
<i>C. goldi</i>	LTFAEKKKEISEWAKKYNVDEVASYNAYREKLKQEHRKNVSELVSDLP S AVK KVNDDLLD (SEQ ID NO: 71)	8	90.0-100%
<i>C. labiatus</i>	LTFAEKKKISEWAKKYNVDEVARYNAYREKLKQEYRKNVSELVSGLPNAV K KVNDDLLD (SEQ ID NO: 72)	1	—
<i>C. leptostomum</i>	LTFAEKKGKISEWAKKYNVDEVASYNAYREKLKQEHRKNVSELVSGLPGA VK KVNELLD (SEQ ID NO: 73)	3	88.3-100%

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Sequences of a conserved region of GALA from individual cyathostomin species			
Species	Sequence	Number of individuals	Intraspecies aa identity range
<i>C. longibursatus</i>	LTFAEKKEEISKWAKKYNVVEVASYNAYREKLKQEHRKNVSEIVSDLPNAVK KVNDLLD (SEQ ID NO: 74)	5	91.7-100%
<i>C. minutus</i>	LTFAEKKEKISEWAKKYNVVEVASYNAYREKLKQEHRKNVSQLVSALPNAVK KVNDLLD (SEQ ID NO: 75)	1	—
<i>C. nassatus</i>	LTFAEKKEKIGEWAKKYNVVEVAXYNAYREKLKQEHRKNVSELVSGLPNAVK KVNELLD (SEQ ID NO: 76)	6	88.3-100%
<i>C. pateratum</i>	LTFAEKK(K/E)EISEWAKKYNVVEVASYNAYREKLKQEHRKNVSELVSALP NAVKKVNDLLD (SEQ ID NO: 77)	6	93.3-100%

SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 83

<210> SEQ ID NO 1

<211> LENGTH: 223

<212> TYPE: PRT

<213> ORGANISM: cyathostomum pateratum

<400> SEQUENCE: 1

Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Ser Ala Val Ala Leu
1 5 10 15

Ala Gln Gly Val Met Asp Leu Phe Gly Glu Glu Gly Arg Glu Glu His
20 25 30

Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val
35 40 45

Ser Cys Glu Ala Lys Trp Glu Tyr Phe Lys Ile Val Gly Asn Arg Ser
50 55 60

Leu Thr Phe Ala Glu Lys Arg Lys Glu Ile Ser Glu Trp Ala Lys Lys
65 70 75 80

Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
85 90 95

Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Ala Leu
100 105 110

Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln
115 120 125

Thr Pro Arg Gln Leu Tyr Val Ala Leu Arg Lys Leu Gly Arg Gln Asn
130 135 140

Pro Ala Leu Tyr Arg Ile Val Glu Tyr Ile Asn Val Ala Val Arg Leu
145 150 155 160

Arg Ser Glu Glu Val Asp Glu Gln Glu Gln Arg Arg Arg Leu Ser Ala
165 170 175

Leu Pro Phe Gly Asp His Asn Asp Asn Leu Glu Glu Gln Asp Phe Gly
180 185 190

Glu Gln Asp Phe Arg Tyr Val Tyr Gly Phe Glu Cys Ala Arg Phe Leu
195 200 205

Leu Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Glu Arg Tyr
210 215 220

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<210> SEQ ID NO 2
<211> LENGTH: 669
<212> TYPE: DNA
<213> ORGANISM: cyathostomum pateratum

<400> SEQUENCE: 2
atgaacaaaa cgtaaactt tctcacagtc gttagtgcg tagctctggc ccaaggtgtc   60
atggaccttt ttggtgaaga gggctcgtgaa gaacatcgtc gtcaccatcg tcattcactt   120
ttaccacat atctccacaa tgtgagctgt gaggctaaat gggagtactt caaaattgtg   180
gggaacagga gtttgacctt tgctgagaaa agaaaggaaa ttagcgagtg ggcaaaaaa   240
tacaatggtt tggatgaagt tgcaagctac aatgcttaca gggaaaaact caagcaggag   300
cacagaaaaa acgttagcga acttgtttct gctcttccaa acgcagtcaa gaaagtcaat   360
gatcttctag acaatgaaaa tcagactcct aggcaacttt acgttgccct tagaaaactt   420
ggtagacaaa atccggcact ttaccgtatt gtcgagtaca ttaatgtggc tgtaagacta   480
agaagtgaag aagtggatga gcaagaacaa cgaagaaggc tgtcagctct accttttggc   540
gaccataacg ataatttggg agagcaggac ttcggtgaac aagactttcg ctatgtctat   600
ggctttgagt gtgcaagatt tctccttcaa aatggaagaa tgtttggact taacacagat   660
gaaagatat                                     669

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<210> SEQ ID NO 3
<211> LENGTH: 192
<212> TYPE: PRT
<213> ORGANISM: Cyclicocyclus nassatus

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<400> SEQUENCE: 3
His Glu Glu Leu Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr
1          5          10          15
Leu His Asn Val Ser Cys Glu Ala Lys Trp Glu Tyr Phe Lys Ile Val
          20          25          30
Gly Asn Arg Ser Leu Thr Phe Ala Glu Lys Lys Gly Lys Ser Ser Glu
          35          40          45
Trp Ala Lys Lys Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala
          50          55          60
Tyr Arg Glu Lys Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu
          65          70          75          80
Val Ser Gly Leu Pro Gly Ala Val Lys Lys Val Asn Glu Leu Leu Asp
          85          90          95
Asn Glu Asn Gln Thr Pro Arg Gln Leu Tyr Val Ala Leu Arg Lys Leu
          100          105          110
Gly Lys Gln Asn Pro Val Leu Tyr Arg Val Val Glu Phe Val Asn Leu
          115          120          125
Val Val Arg Phe Arg Arg Glu Asp Ser Asp Glu Gln Glu Gln Arg Glu
          130          135          140
Met Leu Ser Thr Leu Pro Phe Ser Glu Asn Asn Glu Glu Gln Asp Leu
          145          150          155          160
Gly Glu Gln Asp Phe Gln Tyr Ile Tyr Gly Phe Glu Cys Ala Arg Phe
          165          170          175
Ile Phe Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr
          180          185          190

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<210> SEQ ID NO 4
<211> LENGTH: 576
<212> TYPE: DNA

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<213> ORGANISM: *Cylicocyclus nassatus*

<400> SEQUENCE: 4

```

catgaagaac ttgctgctca ccactgctcat tcactttttac caccctatct ccacaatgtg      60
agctgtgaag ccaaatggga atacttcaag attgtgggga acaggagctt gacttttgct      120
gaaaagaagg gaaaaagtag cgagtgggca aaaaaataca atgttggtgga tgaagttgca      180
agttacaatg cctatagaga aaaacttaag caggagcaca ggaaaaacgt tagcgaactt      240
gtttctggtc ttcccgggtc tgtgaagaaa gtaaacgaac tcttgataa tgagaatcag      300
actcctaggc aactttacgt tgctctaaga aagcttggtg aacaaaatcc agtactctac      360
cgtgttgctg agtttgctca tttggttggt agatttagac gtgaagattc ggatgagcaa      420
gaacaacgag aaatgctgct aactttacct ttcagcgaaa ataatgaaga gcaggacctt      480
ggggaacaag acttcagta catctatggt tttgaatgtg caagattcat ctttcaaaat      540
gggagaatgt ttggactcaa cacggataga agatat                                  576

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<210> SEQ ID NO 5

<211> LENGTH: 175

<212> TYPE: PRT

<213> ORGANISM: *Coronocyclus coronatus*

<400> SEQUENCE: 5

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Ser Cys Val Ala Lys Trp Glu Tyr Phe Lys Ile Val Ile Asn Arg Ser
1          5          10          15
Leu Thr Phe Ala Gln Arg Lys Glu Glu Ile Ser Lys Trp Ala Lys Lys
          20          25          30
Tyr Lys Val Glu Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
          35          40          45
Leu Lys Gln Gln His Arg Lys Asn Val Ser Glu Leu Val Ser Asn Leu
          50          55          60
Pro Gly Ala Val Glu Arg Val Asn Lys Leu Leu Asp Asn Glu Asn Gln
65          70          75          80
Thr Pro Lys Gln Leu Tyr Leu Ala Leu Arg Glu Leu Gly Lys Gln Asn
          85          90          95
Pro Ala Leu Tyr His Val Val Glu Tyr Val Asn Val Val Val Arg Leu
          100          105          110
Lys Arg Glu Glu Leu Asp Gln Gln Asp Gln Arg Arg Ala Leu Ser Gly
          115          120          125
Ser Leu Phe Gly Glu Asn Asn Asp Asn Leu Glu Glu Gln Asp Phe Gly
          130          135          140
Glu Glu Asp Phe Arg Tyr Val Tyr Gly Phe Glu Cys Ala Arg Phe Ile
          145          150          155          160
Leu Gln Asn Gly Arg Met Phe Gly Leu Asn Met Asp Arg Asn Tyr
          165          170          175

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<210> SEQ ID NO 6

<211> LENGTH: 525

<212> TYPE: DNA

<213> ORGANISM: *Coronocyclus coronatus*

<400> SEQUENCE: 6

```

agctgtgtgg ctaagtggga gtacttcaag atcgtgatga acaggagtct gacgtttgct      60
caaagaaagg aagaaattag caagtgggcg aaaaaataca aagttgagga tgaagttgca      120
agctacaatg cttatagaga aaaactcaag cagcagcaca ggaaaaacgt tagcgaactt      180

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gtttctagtc ttcccggtgc aatggaaaga gtgaacaaac ttttgacaa tgaaccag 240
accocaaagc aactttacct tgcocaaaga gaacttgga aacaaatcc ggcactttac 300
catgttgctg agtatgtcaa tgtggttg agacttaaac gagaagaatt ggatgaacaa 360
gatcaatgaa gagcgctgct gggttcactt tttggcgaga ataacgacaa tctagaagag 420
caggactttg gtgaagaaga ctttcgctat gtctatgggt ttgaatgtgc aagattcatc 480
cttcaaaatg gaagaatggt tggctaaac atggatagga attat 525

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<210> SEQ ID NO 7
<211> LENGTH: 199
<212> TYPE: PRT
<213> ORGANISM: Cyathostomum catinatum

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<400> SEQUENCE: 7

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Gly Glu Glu Asp Arg Glu Glu His Arg Arg His His Arg His Ser Leu
1           5           10           15
Leu Pro Pro Tyr Leu His Asn Val Ser Cys Val Ala Lys Trp Glu Tyr
20           25           30
Phe Arg Ile Val Gly Asn Arg Ser Leu Thr Phe Ala Glu Lys Lys Lys
35           40           45
Glu Ile Ser Glu Trp Ala Lys Lys Tyr Asn Val Leu Asp Glu Val Ala
50           55           60
Ser Tyr Asn Ala Tyr Arg Glu Lys Leu Lys Gln Glu His Arg Lys Asn
65           70           75           80
Val Ser Glu Leu Val Ser Asp Leu Pro Lys Ala Val Lys Lys Val Asn
85           90           95
Asp Leu Leu Asp Asn Glu Asn Gln Thr Pro Arg Gln Leu Tyr Val Ala
100          105          110
Leu Arg Glu Leu Gly Arg Gln Asn Pro Thr Leu Tyr Arg Ile Val Glu
115          120          125
Tyr Ile Asn Val Ala Val Arg Arg Arg Ser Glu Glu Leu Asp Glu Gln
130          135          140
Glu Gln Gly Arg Arg Leu Ser Ala Leu Pro Phe Gly Asp Asn Asn Asp
145          150          155          160
Asn Leu Glu Glu Gln Asp Phe Gly Glu Gln Asp Phe Arg Tyr Val Tyr
165          170          175
Gly Phe Glu Cys Ala Arg Phe Leu Leu Gln Asn Gly Arg Met Phe Gly
180          185          190
Leu Asn Thr Asp Glu Arg Asp
195

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<210> SEQ ID NO 8
<211> LENGTH: 591
<212> TYPE: DNA
<213> ORGANISM: Cyathostomum catinatum

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<400> SEQUENCE: 8

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```

gaggatcgtg aagaacatcg ccgtcacatc cgtcattcac tttgcccacc atatctccac 60
aacgtgagct gtgtggccaa atgggaatac tttagaattg tggggaacag gagtttaacg 120
tttgctgaga aaaagaaaga aattagcgag tgggcaaaaa aatacaatgt tctggatgaa 180
gtagcaagct acaatgctta tagggaaaaa ctcaagcagg agcacagaaa aaacgttagc 240
gaacttgttt ctgatcttcc caaggcagta aagaaagtca acgatcttct agacaatgaa 300
aatcagactc ctaggcaact ttatgttgcc cttagagagc ttggtagaca aaatccgaca 360

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ctttaccgta ttgtcgagta catcaatgtg gctgtaagc gaagaagtga agaactggat 420
gagcaagaac aaggaagaag gctgtcagct ttacctttcg gcgacaacaa cgataatttg 480
gaagagcagg acttcggtga acaagacttt cgctatgtct acggctttga gtgtgcaaga 540
tttctccttc aaaatggaag aatgttcgga ctcaacacag atgaaagaga t 591

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<210> SEQ ID NO 9
<211> LENGTH: 841
<212> TYPE: DNA
<213> ORGANISM: Cylicocyclus ashworthi

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<400> SEQUENCE: 9

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```

atgaacaaaa cgtaacatt tctcacagtc gttagtgcg tagttctggc ccaaggtgtc 60
atggaccttt ttggtaaga gggctgtgaa gaacatcgcc gtcaccatcg tcattcactc 120
ttaccacat atctccacaa cgtgagctgt gtggctaaat gggagtactt caaaattgta 180
gggaacagga gtttaacgtt tgctgagaaa aaagaagaaa ttagccagtg ggcaaaaaaa 240
tacaatggtg tgtaagctt ttctgaatta atgtaatac actcgcatgc tggccttttt 300
aggatgaagt tgcaagctac aatgcttaca gggagaaact caagcaggag cacagaaaaa 360
acgttagcga acttgtttct gctcttccaa acgcagtaaa gaaagtcaac aatcttctag 420
acaatgaaaa tcagactctt aggcaacttt acgttgccct tagagaactt ggtagacaaa 480
atccggcagt aagtagaaa agctgcactc ctgggcttaa taaaacaaat tatttaagct 540
ttaccgtatt gtgcagtaca tcaatgtggc tgtaagacga agaagtgaag gactggatga 600
gcaagaacaa cgaagaaagc tatcagcttt acctttcggc gacaacaacg ataatatgga 660
agagcaggac ttoggtgaac aagactttcg ctatgtctac ggctttgagt gtgcaagatt 720
tctccttcaa aatggaagaa tgtttgggct caacacagat gaaagagatt agcaaagaat 780
caattgtagt tcaaagcggg agagtttgag ctgcaaaactc agcatgccat catcacctcc 840
t 841

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<210> SEQ ID NO 10
<211> LENGTH: 223
<212> TYPE: PRT
<213> ORGANISM: Cylicocyclus ashworthi

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<400> SEQUENCE: 10

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Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Ser Ala Val Val Leu
1           5           10           15
Ala Gln Gly Val Met Asp Leu Phe Gly Glu Glu Gly Arg Glu Glu His
20           25           30
Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val
35           40           45
Ser Cys Val Ala Lys Trp Glu Tyr Phe Lys Ile Val Gly Asn Arg Ser
50           55           60
Leu Thr Phe Ala Glu Lys Lys Glu Glu Ile Ser Gln Trp Ala Lys Lys
65           70           75           80
Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
85           90           95
Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Ala Leu
100          105          110
Pro Asn Ala Val Lys Lys Val Asn Asn Leu Leu Asp Asn Glu Asn Gln
115          120          125
Thr Leu Arg Gln Leu Tyr Val Ala Leu Arg Glu Leu Gly Arg Gln Asn

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85					90					95					
Leu	Lys	Gln	Glu	His	Arg	Lys	Asn	Val	Ser	Glu	Leu	Val	Ser	Ala	Leu
			100					105						110	
Pro	Asn	Ala	Val	Lys	Lys	Val	Asn	Asp	Leu	Leu	Asp	Asn	Glu	Asn	Gln
		115					120						125		
Thr	Pro	Arg	Gln	Leu	Tyr	Val	Ala	Leu	Arg	Glu	Leu	Gly	Arg	Gln	Asn
		130					135						140		
Pro	Ala	Leu	Tyr	Arg	Ile	Val	Glu	Tyr	Ile	Asn	Val	Ala	Val	Arg	Leu
		145				150					155				160
Arg	Ser	Glu	Glu	Val	Asp	Glu	Gln	Glu	Gln	Arg	Arg	Arg	Leu	Ser	Ala
			165						170						175
Leu	Pro	Phe	Gly	Asp	His	Asn	Asp	Asn	Met	Glu	Glu	Gln	Asp	Phe	Gly
			180						185					190	
Asp	Gln	Asp	Phe	Arg	Tyr	Val	Tyr	Gly	Phe	Glu	Cys	Ala	Arg	Phe	Leu
		195					200						205		
Leu	Gln	Asn	Gly	Arg	Met	Phe	Gly	Leu	Asn	Thr	Asp	Glu	Arg	Tyr	
		210					215						220		

<210> SEQ ID NO 13
 <211> LENGTH: 769
 <212> TYPE: DNA
 <213> ORGANISM: *Cylicostephanus goldi*

<400> SEQUENCE: 13

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atgaacaaaa cgtaaactt tctcacagtc gttagtgcgc tagtctctggc tcaaggtgtc      60
gtggaccttt ttggtgaaga gggctcgtgaa gaacatcgcc gtcaccatcg tcattcactc      120
ttaccacat  atctccacaa cgctcagctgt gtggcctaaat gggataactt caaaattgtg      180
gggaatagga gtttgacatt tgctgagaaa aagaaagaaa ttagcgagtg ggctaaaaaa      240
tacaatgtag tggtaagctt ttttgacttg atgtaaatgc actcgtatgc cggccctttt      300
aggatgaagt tgcaaggtac aatgcttata gagaaaaact taagcaggaa cacaggaaaa      360
acgtcagcga acttgtttct gatcttccca acgcagtaaa gaaagtgaat gatctcctgg      420
acaatgagaa tcaactctct aggcaacttt acattgccct cagagaactt ggtagacaaa      480
atccagaagt aagttgaaag tgctgcaatt ttaggcttag ataaaacagt tgtttaagct      540
ttaccgtggt gtgcagttta tcaatgtggc tgtaagaata agacgtgaag atttgatga      600
gcaagaacaa cgaacaaggc tgccaacttt accttttggc gacaacaacg acaatttcga      660
agagcaagac ttcggtgaac aagactttcg ctatgtctat ggctttgagt gtgcaagatt      720
tctccttcaa aatggaagaa tgtttggact taacacggat agaagatac      769
    
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<210> SEQ ID NO 14
 <211> LENGTH: 223
 <212> TYPE: PRT
 <213> ORGANISM: *Cylicostephanus goldi*

<400> SEQUENCE: 14

Met	Asn	Lys	Thr	Leu	Thr	Phe	Leu	Thr	Val	Val	Ser	Ala	Val	Val	Leu
			1			5			10					15	
Ala	Gln	Gly	Val	Val	Asp	Leu	Phe	Gly	Glu	Glu	Gly	Arg	Glu	Glu	His
			20					25						30	
Arg	Arg	His	His	Arg	His	Ser	Leu	Leu	Pro	Pro	Tyr	Leu	His	Asn	Val
			35				40						45		
Ser	Cys	Val	Ala	Lys	Trp	Glu	Tyr	Phe	Lys	Ile	Val	Gly	Asn	Arg	Ser
		50					55						60		

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Leu Thr Phe Ala Glu Lys Lys Lys Glu Ile Ser Glu Trp Ala Lys Lys
 65 70 75 80
 Tyr Asn Val Val Asp Glu Val Ala Arg Tyr Asn Ala Tyr Arg Glu Lys
 85 90 95
 Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Asp Leu
 100 105 110
 Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln
 115 120 125
 Thr Pro Arg Gln Leu Tyr Ile Ala Leu Arg Glu Leu Gly Arg Gln Asn
 130 135 140
 Pro Glu Leu Tyr Arg Val Val Glu Phe Ile Asn Val Ala Val Arg Ile
 145 150 155 160
 Arg Arg Glu Asp Leu Asp Glu Gln Glu Gln Arg Thr Arg Leu Ser Thr
 165 170 175
 Leu Pro Phe Gly Asp Asn Asn Asp Asn Phe Glu Glu Gln Asp Phe Gly
 180 185 190
 Glu Gln Asp Phe Arg Tyr Val Tyr Gly Phe Glu Cys Ala Arg Phe Leu
 195 200 205
 Leu Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr
 210 215 220

<210> SEQ ID NO 15
 <211> LENGTH: 847
 <212> TYPE: DNA
 <213> ORGANISM: *Cylicostephanus goldi*

<400> SEQUENCE: 15

atgaacaaaa cgtaacatt tctcacagtc gttagtgcgc tagtctctggc ccaaggtgtc 60
 atggaccttc ttgatgaaga ggctcgtgga gagcatcgcc gtcaccatcg tcattcactc 120
 ttaccacat atctccacaa cgtgagctgt gtggctaaat gggaatactt caaaattgtg 180
 gggaacagga gtttgacgtt tgctgagaaa aagaaagaaa ttagcgagtg ggcaaaaaaa 240
 tacaacgctt tggttaagctt ttgtgactcg atgtagatac cccagatatt ctagataccc 300
 atgctggcct ttttaggatg aagttgcaag ctacaatgct tatagagaaa aactcaagca 360
 ggaacacagg aaaaacgtta gcgaacttgt atctgatctt cccaatgcag tgaagaaagt 420
 gaatgatctc ctggacaatg agaatcaaac tcctaggcaa ctttacgttg ccctcagaga 480
 acttggtaga caaaatccag cagtaagttg aaagtgctgc aatttcaggc ttagataaaa 540
 cagttgttta agctttaccg tggtgtcgag ctcatcaatg tggctgtaag attaagacgt 600
 gaagatttgg atgagcaaga acaacgaaca aggctgtcaa ccttaccttt tggcgacaac 660
 aacaacaatt tcgatgagca ggacttcggt gaacaagact ttcgctatgt ctatggcttt 720
 gagtgtgcaa gatttctcct tcaaaatgga agaattgttg gacttaacac ggatagaaga 780
 tactagtaag agtcaactgt agctcaaaagt ggttcgagct acgaacagca tgccatcatc 840
 acctcct 847

<210> SEQ ID NO 16
 <211> LENGTH: 223
 <212> TYPE: PRT
 <213> ORGANISM: *Cylicostephanus goldi*

<400> SEQUENCE: 16

Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Ser Ala Val Val Leu
 1 5 10 15

-continued

Ala Gln Gly Val Met Asp Leu Leu Asp Glu Glu Ala Arg Gly Glu His
 20 25 30

Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val
 35 40 45

Ser Cys Val Ala Lys Trp Glu Tyr Phe Lys Ile Val Gly Asn Arg Ser
 50 55 60

Leu Thr Phe Ala Glu Lys Lys Lys Glu Ile Ser Glu Trp Ala Lys Lys
 65 70 75 80

Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
 85 90 95

Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Asp Leu
 100 105 110

Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln
 115 120 125

Thr Pro Arg Gln Leu Tyr Val Ala Leu Arg Glu Leu Gly Arg Gln Asn
 130 135 140

Pro Ala Leu Tyr Arg Val Val Glu Leu Ile Asn Val Ala Val Arg Leu
 145 150 155 160

Arg Arg Glu Asp Leu Asp Glu Gln Glu Gln Arg Thr Arg Leu Ser Thr
 165 170 175

Leu Pro Phe Gly Asp Asn Asn Asn Asn Phe Asp Glu Gln Asp Phe Gly
 180 185 190

Glu Gln Asp Phe Arg Tyr Val Tyr Gly Phe Glu Cys Ala Arg Phe Leu
 195 200 205

Leu Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr
 210 215 220

<210> SEQ ID NO 17

<211> LENGTH: 835

<212> TYPE: DNA

<213> ORGANISM: *Cylicostephanus goldi*

<400> SEQUENCE: 17

```

atgaacaaaa cgtaacatt tctcacagtc gttagtgcg ttgtcctggc gcaaggtgtc 60
atggccctat ttggtgaaga gagtcgtgaa gaacaccgcc gtcaccatcg tcattcactc 120
ttaccacat atctccacaa cgtgagctgt gtggctaaat gggagtactt caaaattgtg 180
gggaacagga gtttgacgtt tgctgagaaa aagaaagaaa tcagcgagtg ggctaaaaaa 240
tacaatggtg tgtaagctt ttttgacttg atgtaaatgc actcgcatgc cggcctttat 300
aggatgaagt tgcaagctac aatgcttata gagaaaaact caagcaggaa cacaggaaaa 360
acgtagcga acttgtttct gatcttccca acgcagtaaa gaaagtcagc gatcttttgg 420
acaacgaaaa tcagacttct aggcaacttt atgttgact cagagaactt ggtagacaaa 480
atccggcagt aagttgaaga ggctccaatt ttgggctcaa gcaaaaaataa ttattttagc 540
tataccgtgt cgtcagtat atcaatgtgg ctgtgagatt aagacgaaaa gaacaggatg 600
aacaagaacg acaaggaacg ctgtcagctc taccttttgg cgagaataac gacaatttgg 660
aagagcagga ctttggtgaa caagactttc gctatgtcta tggctttgag tgtgcaagat 720
ttctccttca aaatggaaga atgtttggac tcaacacgga tagaagatac cagtaagagt 780
caactgtagc tcaaagtggg tttgagctac gaacagcatg ccatcatcac ctct 835

```

<210> SEQ ID NO 18

<211> LENGTH: 223

-continued

<212> TYPE: PRT

<213> ORGANISM: *Cylicostephanus goldi*

<400> SEQUENCE: 18

```

Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Ser Ala Val Val Leu
 1          5          10          15
Ala Gln Gly Val Met Ala Leu Phe Gly Glu Glu Ser Arg Glu Glu His
 20          25          30
Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val
 35          40          45
Ser Cys Val Ala Lys Trp Glu Tyr Phe Lys Ile Val Gly Asn Arg Ser
 50          55          60
Leu Thr Phe Ala Glu Lys Lys Lys Glu Ile Ser Glu Trp Ala Lys Lys
 65          70          75          80
Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
 85          90          95
Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Asp Leu
 100          105          110
Pro Asn Ala Val Lys Lys Val Ser Asp Leu Leu Asp Asn Glu Asn Gln
 115          120          125
Thr Ser Arg Gln Leu Tyr Val Ala Leu Arg Glu Leu Gly Arg Gln Asn
 130          135          140
Pro Ala Val Tyr Arg Val Val Glu Tyr Ile Asn Val Ala Val Arg Leu
 145          150          155          160
Arg Arg Lys Glu Gln Asp Glu Gln Glu Arg Gln Gly Thr Leu Ser Ala
 165          170          175
Leu Pro Phe Gly Glu Asn Asn Asp Asn Leu Glu Glu Gln Asp Phe Gly
 180          185          190
Glu Gln Asp Phe Arg Tyr Val Tyr Gly Phe Glu Cys Ala Arg Phe Leu
 195          200          205
Leu Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr
 210          215          220

```

<210> SEQ ID NO 19

<211> LENGTH: 830

<212> TYPE: DNA

<213> ORGANISM: *Cylicostephanus longibursatus*

<400> SEQUENCE: 19

```

atgaacaaaa cgtaaaccatt tctcaccgtc gtctatgccg tagtctggc ccaaggtgtc    60
atggaccttt ttggtgaaga gggctgtgaa gaacatcgcc gtcaccatcg tcattcactc    120
ttaccacccat atctccacaa gtgtagctgt gtggctaaat gggaaactt caaaattgtg    180
gggaacagga gtttgacgtt tgctgagaaa aaggaagaaa ttagcaagtg ggcaaaaaaa    240
tacaatgttg tggtagcgtt ttgtaacccc gtataatata ctctcgcata ctggccggtt    300
caggatgaag ttgcaagcta cagtgttgc agggaaaagc ttaagcagga acacaggaaa    360
aacgttagcg aaattgtttc taatcttccc aatgcagtga agaaagtaaa cgatcttttg    420
gacaatgaaa atcagacccc caggcaactt tacgttgctc tcagaaaaact tggtaaacaa    480
aatccggcag taagttgaaa gagctgcaat tttgggtttg aggagaaaaa actattttag    540
ctttatcgtg ttgtcgagta tatcaatgtg cttgtgagac taagacgtga agaatttgat    600
gaagatcagc gaagatcgct gtcagcttta ccttttggcg acaataacga cgatttgaa    660
gagcaggact ttggtgaaca ggactttogc tatatctatg gotttgagtg tgcaagattt    720

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-continued

 atccttcaaa atggaagaat gttcggactc aacacggata gaagatatta gtaagagtca 780

actgtagctc gagggtttga gctacgaact gcatgccatc atcacctect 830

<210> SEQ ID NO 20

<211> LENGTH: 222

<212> TYPE: PRT

<213> ORGANISM: *Cylicostephanus longibursatus*

<400> SEQUENCE: 20

 Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Tyr Ala Val Val Leu
 1 5 10 15

 Ala Gln Gly Val Met Asp Leu Phe Gly Glu Glu Gly Arg Glu Glu His
 20 25 30

 Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val
 35 40 45

 Ser Cys Val Ala Lys Trp Glu Tyr Phe Lys Ile Val Gly Asn Arg Ser
 50 55 60

 Leu Thr Phe Ala Glu Lys Lys Glu Glu Ile Ser Lys Trp Ala Lys Lys
 65 70 75 80

 Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Ser Ala Cys Arg Glu Lys
 85 90 95

 Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Ile Val Ser Asn Leu
 100 105 110

 Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln
 115 120 125

 Thr Pro Arg Gln Leu Tyr Val Ala Phe Arg Lys Leu Gly Lys Gln Asn
 130 135 140

 Pro Ala Leu Tyr Arg Val Val Glu Tyr Ile Asn Val Leu Val Arg Leu
 145 150 155 160

 Arg Arg Glu Glu Phe Asp Glu Asp Gln Arg Arg Ser Leu Ser Ala Leu
 165 170 175

 Pro Phe Gly Asp Asn Asn Asp Asp Leu Glu Glu Gln Asp Phe Gly Glu
 180 185 190

 Gln Asp Phe Arg Tyr Ile Tyr Gly Phe Glu Cys Ala Arg Phe Ile Leu
 195 200 205

 Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr
 210 215 220

<210> SEQ ID NO 21

<211> LENGTH: 829

<212> TYPE: DNA

<213> ORGANISM: *Cylicocyclus insigne*

<400> SEQUENCE: 21

atgaacaaaa cgtaacatt tctcaccgtc gtctgtgccc tagtctggc ccaaggtgtc 60

atggaccttt ttggtaaga aggtcgtgaa gaacatcgcc gtcaccatcg tcattcactc 120

ttaccacccat atctccacaa tgtgagctgt gtggctaaat gggaaactt caaaaattctg 180

gggaacagaa gtttgacgtt tgctgagaaa aaggaaaaaa tcagcgagtg ggcaaaaaag 240

tacaatggtt tggtagctt ttgtaactcc gtataatata ccctcgcatg ctggccggtt 300

caggatgaag ttgcaagcta caatgcttgc agggaaaaag ttaagcagga acacaggaaa 360

aacgttagcg aaattgttcc taatcttccc aatgcagtaa agaaagtaaa cgatcttttg 420

gacaatgaaa atcagactcc caggcaactt tacgttgccc tcagaaaaact cggtaaacaa 480

aatccgccag taagttgaaa gactgcaact ttgggtttaa gggaaaaaaa ctatttttagc 540

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tttaccgctg tgcgagat atcaatgtgg ttgtgagact aagacgtgaa gaatctgatg 600
aagaacaacg aagaacgctg tcagctttac cttttggcga caataacgac aacttgaag 660
agcaagactt tggggaagaa gactttcgct atatttatgg ctttgagtgt gcaagattta 720
tccttcaaaa tgggagaatg ttcggactca acacggatag aagatatcag taagagtcaa 780
ctgtagctta aaagtttgag ctacgaacag catgccatca tcacctct 829

```

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<210> SEQ ID NO 22
<211> LENGTH: 222
<212> TYPE: PRT
<213> ORGANISM: Cylicocyclus insigne

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<400> SEQUENCE: 22

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```

Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Cys Ala Val Val Leu
1          5          10          15
Ala Gln Gly Val Met Asp Leu Phe Gly Glu Glu Gly Arg Glu Glu His
20          25          30
Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val
35          40          45
Ser Cys Val Ala Lys Trp Glu Tyr Phe Lys Ile Leu Gly Asn Arg Ser
50          55          60
Leu Thr Phe Ala Glu Lys Lys Glu Lys Ile Ser Glu Trp Ala Lys Lys
65          70          75          80
Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Cys Arg Glu Lys
85          90          95
Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Ile Val Ser Asn Leu
100         105         110
Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln
115         120         125
Thr Pro Arg Gln Leu Tyr Val Ala Leu Arg Lys Leu Gly Lys Gln Asn
130         135         140
Pro Pro Leu Tyr Arg Val Val Glu Tyr Ile Asn Val Val Val Arg Leu
145         150         155         160
Arg Arg Glu Glu Ser Asp Glu Glu Gln Arg Arg Thr Leu Ser Ala Leu
165         170         175
Pro Phe Gly Asp Asn Asn Asp Asn Leu Glu Glu Gln Asp Phe Gly Glu
180         185         190
Glu Asp Phe Arg Tyr Ile Tyr Gly Phe Glu Cys Ala Arg Phe Ile Leu
195         200         205
Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr
210         215         220

```

```

<210> SEQ ID NO 23
<211> LENGTH: 830
<212> TYPE: DNA
<213> ORGANISM: Cylicostephanus longibursatus

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<400> SEQUENCE: 23

```

```

atgaacaaaa cgtaaacatt tctcaccgtc gtctatgccg tagtctctggc ccaaggtgtc 60
atggaccttt ttggtgaaga gggctctttaa gaacatcgcc gtcaccatcg tcattcactc 120
ttaccacat atctccacaa tgtgagctgt gtggctaaat gggaaactt caaaattctg 180
gggaacagga gtttgacgtt tgctgagaaa aaggaaaaaa tcagcgagtg ggcaaaaaag 240
tacaatgttg tggtagcctt ttgtaactca gtataatata tctctgcata ctggccgttt 300

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caggatgaag ttgcaagcta caatgcttgc agggaaaagc ttaagcagga acacagaaaa 360
aacgttagcg aaattgtttc taatcttccc aatgcagtga agaaagtaaa cgatcttttg 420
gacaaatgaaa atcagacccc caggcaactt tacgttgccc tcagaaaact tggtaaacaa 480
aatccggcag taagttgaaa gagctgcaat tttgggttg aggaaaaaaa actattttag 540
ctttatcgty ttgtcgagta tatcaatgtg cttgtgagac taagacgtga agaatttgat 600
gaagatcagc gaagatcgct gtcagcttta ccttttggcg acaataacga cgatttggaa 660
gagcaggact ttggtgaaca ggactttcgc tatatctatg gctttgagtg tgcaagattt 720
atccttcaaa atggaagaat gttcggactc aacacggata gaagatatta gtaagagtca 780
actgtagctc aagggtttga gctacgaact gcatgccatc atcaactcct 830

```

<210> SEQ ID NO 24

<211> LENGTH: 222

<212> TYPE: PRT

<213> ORGANISM: *Cylicostephanus longibursatus*

<400> SEQUENCE: 24

```

Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Tyr Ala Val Val Leu
1          5          10          15
Ala Gln Gly Val Met Asp Leu Phe Gly Glu Glu Gly Leu Glu Glu His
20        25        30
Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val
35        40        45
Ser Cys Val Ala Lys Trp Glu Tyr Phe Lys Ile Leu Gly Asn Arg Ser
50        55        60
Leu Thr Phe Ala Glu Lys Lys Glu Lys Ile Ser Glu Trp Ala Lys Lys
65        70        75        80
Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Cys Arg Glu Lys
85        90        95
Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Ile Val Ser Asn Leu
100       105       110
Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln
115       120       125
Thr Pro Arg Gln Leu Tyr Val Ala Leu Arg Lys Leu Gly Lys Gln Asn
130       135       140
Pro Ala Leu Tyr Arg Val Val Glu Tyr Ile Asn Val Leu Val Arg Leu
145       150       155       160
Arg Arg Glu Glu Phe Asp Glu Asp Gln Arg Arg Ser Leu Ser Ala Leu
165       170       175
Pro Phe Gly Asp Asn Asn Asp Asp Leu Glu Glu Gln Asp Phe Gly Glu
180       185       190
Gln Asp Phe Arg Tyr Ile Tyr Gly Phe Glu Cys Ala Arg Phe Ile Leu
195       200       205
Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr
210       215       220

```

<210> SEQ ID NO 25

<211> LENGTH: 832

<212> TYPE: DNA

<213> ORGANISM: *Cylicocyclus nassatus*

<400> SEQUENCE: 25

```

atgaacaaaa cgtaaacatt tctcatcgtc gttagtgcgc tagtctgac ccaaagtgtt 60
atggactttt tcgatgaaga cggctcgtgaa gaacatcgcc gtcacatcgc tcattccctt 120

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ttaccaccgt atotccacaa tatgagctgc gtggccaaat gggaatactt cgagattgtg 180
ggggacagga gtctgacgtt tgctgaaaag aaggaaaaaa tcggcgagtg ggctaataaaa 240
tacaatggtt tggtaagatt ttgtaactct atgtaaagat acccccgtac gtgcacctgt 300
ttaggatgaa gttgcaagct acaatgctta tagagaaaaa ctaaagcagg agcacaggaa 360
aaacgttagc gagcttgtct ctggtcttcc caatgctgtg aagaaaataa acgaactttt 420
agacaatgaa aatcagactg ttaggcaact ttatgttgct ttaagagaac ttggtaaaca 480
aaatccagca gtaagttaaa agaagtgcaa ttttgggctt aactaatgag acaattttag 540
ctctaccgtg ttgtcgagta tatcaatgtg gttgtgagac ttagacgtga agatttggat 600
gagcaggaac aacagagaac gctgtcaacc ccacctttcg gcgagaataa cgaagagcaa 660
gactttgggtg aacaagactt tcaactatc tatggttttg agtgtgccag attcatcctt 720
caaaatggaa gaatgtttgg acttaacacg gatagaagat attagtaaga gttaactgca 780
gctcaatgtg atagagattg agccacaacc caacatgcca tcatcaoctc ct 832

```

<210> SEQ ID NO 26

<211> LENGTH: 220

<212> TYPE: PRT

<213> ORGANISM: *Cylicocyclus nassatus*

<400> SEQUENCE: 26

```

Met Asn Lys Thr Leu Thr Phe Leu Ile Val Val Ser Ala Val Val Leu
1           5           10           15
Thr Gln Ser Val Met Asp Phe Phe Asp Glu Asp Gly Arg Glu Glu His
20           25           30
Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Met
35           40           45
Ser Cys Val Ala Lys Trp Glu Tyr Phe Glu Ile Val Gly Asp Arg Ser
50           55           60
Leu Thr Phe Ala Glu Lys Lys Glu Lys Ile Gly Glu Trp Ala Lys Lys
65           70           75           80
Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
85           90           95
Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Gly Leu
100          105          110
Pro Asn Ala Val Lys Lys Ile Asn Glu Leu Leu Asp Asn Glu Asn Gln
115          120          125
Thr Val Arg Gln Leu Tyr Val Ala Leu Arg Glu Leu Gly Lys Gln Asn
130          135          140
Pro Ala Leu Tyr Arg Val Val Glu Tyr Ile Asn Val Val Val Arg Leu
145          150          155          160
Arg Arg Glu Asp Leu Asp Glu Gln Glu Gln Gln Arg Thr Leu Ser Thr
165          170          175
Pro Pro Phe Gly Glu Asn Asn Glu Glu Gln Asp Phe Gly Glu Gln Asp
180          185          190
Phe His Tyr Ile Tyr Gly Phe Glu Cys Ala Arg Phe Ile Leu Gln Asn
195          200          205
Gly Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr
210          215          220

```

<210> SEQ ID NO 27

<211> LENGTH: 762

<212> TYPE: DNA

-continued

<213> ORGANISM: *Cylicocyclus nassatus*

<400> SEQUENCE: 27

```

atgaacaaaa cgtaacatt tctcatcgtc gttagtcca tagtctggc ccaaagtgtt    60
atggactttt tcgatgaaga aggtcgtgag ggacatcgcc gtcacatcgc tcattcactt    120
ttaccaccat atctccacaa tatgagctgc gtggcctaat ggaataactt cgagattgtg    180
ggggacagga gtctgacgtt ttgctgaaaa aaggaaaaaa tcggcgagtg ggctaaaaaa    240
tacaatggtg tgtaagatt ttgtaactcc atgttaggat acctccgcac gtcgcctgt    300
ttaggatgaa gttgcaagct acaatgctta tagagaaaaa ctaaagcagg agcacaggaa    360
aaacgttagc gagcttgtct ctggtcttcc caatgctgtg aagaaagtaa acgaactttt    420
agacaatgaa aatcagactg ttaggcaact ttatgttgct ttaagagaac ttggtaaaca    480
aatccagca gtaagttaa aagaagtacaa ttttgagctc aactaatgag acaattttag    540
ctctaccgtg ttgtcgagta tatcaatggt gttgtgagac ttagacgtga agattcggat    600
gagcaggaac aacgaagaac tctgtcaacc tcacctttcg gcgagaataa cgaagagcaa    660
gattttggtg aacaagattt tcaatatatc tatggttttg agtgtgcaag attcatcctt    720
caaaatggaa gaatgtttgg actcaatcgc gatagaagat at                          762

```

<210> SEQ ID NO 28

<211> LENGTH: 220

<212> TYPE: PRT

<213> ORGANISM: *Cylicocyclus nassatus*

<400> SEQUENCE: 28

```

Met Asn Lys Thr Leu Thr Phe Leu Ile Val Val Ser Ala Ile Val Leu
1           5           10          15
Ala Gln Ser Val Met Asp Phe Phe Asp Glu Glu Gly Arg Glu Gly His
                20           25           30
Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Met
                35           40           45
Ser Cys Val Ala Lys Trp Glu Tyr Phe Glu Ile Val Gly Asp Arg Ser
                50           55           60
Leu Thr Phe Ala Glu Lys Lys Glu Lys Ile Gly Glu Trp Ala Lys Lys
65           70           75
Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
                85           90           95
Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Gly Leu
                100          105          110
Pro Asn Ala Val Lys Lys Val Asn Glu Leu Leu Asp Asn Glu Asn Gln
                115          120          125
Thr Val Arg Gln Leu Tyr Val Ala Leu Arg Glu Leu Gly Lys Gln Asn
                130          135          140
Pro Ala Leu Tyr Arg Val Val Glu Tyr Ile Asn Val Val Val Arg Leu
145          150          155          160
Arg Arg Glu Asp Ser Asp Glu Gln Glu Gln Arg Arg Thr Leu Ser Thr
                165          170          175
Ser Pro Phe Gly Glu Asn Asn Glu Glu Gln Asp Phe Gly Glu Gln Asp
                180          185          190
Phe His Tyr Ile Tyr Gly Phe Glu Cys Ala Arg Phe Ile Leu Gln Asn
                195          200          205
Gly Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr
                210          215          220

```

-continued

<210> SEQ ID NO 29
 <211> LENGTH: 829
 <212> TYPE: DNA
 <213> ORGANISM: *Cylicocyclus nassatus*

<400> SEQUENCE: 29

```

atgaacaaaa cgtaacatt tctcatcgcc gttagtgcc tagtcctggc ccaaagtatg      60
gacttttttcg atgaagacgg tcgtgaagaa catcgccgctc atcatcgta ttcactttta      120
ccaccatatac tccacaatat gagctgcgcg gccaaatggg aatacttcga gattgtaggg      180
gacaggagtc tgacgtttgc tgaaaagaag gaaaaaatcg gcgagtgggc taaaaaatac      240
aatgttgtagg taagattttg taactccatg taaagatacc cctccatgct gtccecgttta      300
ggatgaagtt gcaagctaca atgcttgcaag aaaaaaactg aagcaagagc acaggaaaaa      360
cgtcagcgag cttgtctctg gtcttcccaa tgctgtgaag aaagtaaagc aactttttaga      420
caatgaaaat cagactgtta ggcaacttta tgttgcttta agagaacttg gtaaacaaaa      480
tccagcagta agttgaaaga agtgcatttt gggcttaact aacgagacaa ttttagctct      540
accgtgttgt cgagtatac aatgtggctg tgagacttag acgtgaagat teggatgagc      600
aggaaaaacg aagaacgctg tcaacctcac ctttcggcga gaataacgaa gagcaggacc      660
ttggtgaaca agatttcac tatactatg gctttgagtg tgcaagattc atccttcaaa      720
atggaagaat gtttgactt aacacggata gaagatatta gtaaaatttg actgcagctc      780
aaagtggtag agattgagct accaacccaa catgccaatca tcacctcct      829
  
```

<210> SEQ ID NO 30
 <211> LENGTH: 219
 <212> TYPE: PRT
 <213> ORGANISM: *Cylicocyclus nassatus*

<400> SEQUENCE: 30

```

Met Asn Lys Thr Leu Thr Phe Leu Ile Ala Val Ser Ala Ile Val Leu
1           5           10          15
Ala Gln Ser Met Asp Phe Phe Asp Glu Asp Gly Arg Glu Glu His Arg
20          25          30
Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Met Ser
35          40          45
Cys Ala Ala Lys Trp Glu Tyr Phe Glu Ile Val Gly Asp Arg Ser Leu
50          55          60
Thr Phe Ala Glu Lys Lys Glu Lys Ile Gly Glu Trp Ala Lys Lys Tyr
65          70          75          80
Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Cys Arg Glu Lys Leu
85          90          95
Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Gly Leu Pro
100         105         110
Asn Ala Val Lys Lys Val Asn Glu Leu Leu Asp Asn Glu Asn Gln Thr
115         120         125
Val Arg Gln Leu Tyr Val Ala Leu Arg Glu Leu Gly Lys Gln Asn Pro
130         135         140
Ala Leu Tyr Arg Val Val Glu Tyr Ile Asn Val Ala Val Arg Leu Arg
145         150         155         160
Arg Glu Asp Ser Asp Glu Gln Glu Lys Arg Arg Thr Leu Ser Thr Ser
165         170         175
Pro Phe Gly Glu Asn Asn Glu Glu Gln Asp Leu Gly Glu Gln Asp Phe
  
```

-continued

	180	185	190	
His Tyr Ile Tyr Gly Phe Glu Cys Ala Arg Phe Ile Leu Gln Asn Gly	195	200	205	
Arg Met Phe Gly Leu Asn Thr Asp Arg Arg Tyr	210	215		

<210> SEQ ID NO 31
 <211> LENGTH: 840
 <212> TYPE: DNA
 <213> ORGANISM: Cyathostomum pateratum

<400> SEQUENCE: 31

atgaacaaaa cgtaacatt tctcacagtc gttagtgccg tagttctggc ccaaggtgtc	60
atggaccttt ttggtaaga gggctgtgaa gaacatcgtc gtcaccatcg tcattcactc	120
ttaccacat atctccacaa tgtgagctgt gaggctaaat gggagtactt caaaattgtg	180
gggaacagga gtttgacggt tgctgagaaa aaggagaaaa ttagcgagtg ggcaaaaaa	240
tacaatggtg tgtaagctt ttttgaattg atgtaaatc actcgcatgc tggcctttt	300
aggatgaagt tgcaagctac aatgcttaca gggaaaaact caagcaggag cacagaaaa	360
acgttagcga acttgtttct gctcttccaa acgcagtaaa gaaagtcaac gatcttctag	420
acaatgaaaa tcagactctt aggcaacttt acgttgccct tagaaaaact ggtagacaaa	480
atccggcagt aagtcgaaa agctgcgtcc ttggacttaa gcggaaaaat tatttcagct	540
ttaccgtatt gtgcagtaca ttaatgtggc tgtaagacta agaagtgaag aagtggatga	600
gcaagaacaa cgaagaagc tgtcagctct accttttggc gaccataacg ataatttga	660
agagcaggac ttoggtgaac aagactttcg ctatgtctat ggctttgagt gtgcaagatt	720
tctcctcaa aatggaagaa tggctggact caacacggat ggaagatatt agtaagaaac	780
aagtgtagct caaagtggta gagtttgagc tacgaactca acatgccatc atcacctcct	840

<210> SEQ ID NO 32
 <211> LENGTH: 223
 <212> TYPE: PRT
 <213> ORGANISM: Cyathostomum pateratum

<400> SEQUENCE: 32

Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Ser Ala Val Val Leu			
1	5	10	15
Ala Gln Gly Val Met Asp Leu Phe Gly Glu Glu Gly Arg Glu Glu His	20	25	30
Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val	35	40	45
Ser Cys Glu Ala Lys Trp Glu Tyr Phe Lys Ile Val Gly Asn Arg Ser	50	55	60
Leu Thr Phe Ala Glu Lys Lys Glu Lys Ile Ser Glu Trp Ala Lys Lys	65	70	75
Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys	85	90	95
Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Ala Leu	100	105	110
Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln	115	120	125
Thr Leu Arg Gln Leu Tyr Val Ala Leu Arg Lys Leu Gly Arg Gln Asn	130	135	140

-continued

Pro Ala Leu Tyr Arg Ile Val Glu Tyr Ile Asn Val Ala Val Arg Leu
 145 150 155 160

Arg Ser Glu Glu Val Asp Glu Gln Glu Gln Arg Arg Arg Leu Ser Ala
 165 170 175

Leu Pro Phe Gly Asp His Asn Asp Asn Leu Glu Glu Gln Asp Phe Gly
 180 185 190

Glu Gln Asp Phe Arg Tyr Val Tyr Gly Phe Glu Cys Ala Arg Phe Leu
 195 200 205

Leu Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Gly Arg Tyr
 210 215 220

<210> SEQ ID NO 33
 <211> LENGTH: 734
 <212> TYPE: DNA
 <213> ORGANISM: *Cylicostephanus goldi*
 <400> SEQUENCE: 33

```

atgaacaaaa cgtaaacatt tctcacagtc gttagtgccg ttgtcctggc ccaaggtgtc    60
atggccctat ttgggtaaga gagtcgtgaa gaacaccgcc gtcaccatcg tcattcactc    120
ttaccacccat atctccacaa cgtgagctgt gtggctaaat gggagtactt caaaattgtg    180
gggaacagga gtttgacgtt tgctgagaaa aagaaagaaa tcagcgagtg ggctaaaaaa    240
tacaatgttg tggatgaagt tgcaagctac aatgcttata gagaaaaact caagcaggaa    300
cacaggaaaa acgtttagca acttgtttct gatcttccca acgcagtaaa gaaagtcaac    360
gatcttttgg acaacgaaaa tcagacttct aggcaacttt atgttgcaact cagagaactt    420
ggtagacaaa atccggcact ataccgtgtc gtcgagtata tcaatgtggc tgtgagatta    480
agacgaaaag aacaggatga acaagaacga caaggaacgc tgcagctct accttttggc    540
gagaataacg acaatttga agagcaggac tttggtgaac aagactttcg ctatgtctat    600
ggctttgagt gtgcaagatt tctccttcaa aatggaagaa tgtttggact caacacggat    660
agaagatacc agtaagagtc aactgtagct caaagtgggt ttgagctacg aacagcatgc    720
catcatcacc tect                                                734
    
```

<210> SEQ ID NO 34
 <211> LENGTH: 223
 <212> TYPE: PRT
 <213> ORGANISM: *Cylicostephanus goldi*
 <400> SEQUENCE: 34

Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Ser Ala Val Val Leu
 1 5 10 15

Ala Gln Gly Val Met Ala Leu Phe Gly Glu Glu Ser Arg Glu Glu His
 20 25 30

Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val
 35 40 45

Ser Cys Val Ala Lys Trp Glu Tyr Phe Lys Ile Val Gly Asn Arg Ser
 50 55 60

Leu Thr Phe Ala Glu Lys Lys Lys Glu Ile Ser Glu Trp Ala Lys Lys
 65 70 75 80

Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
 85 90 95

Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Asp Leu
 100 105 110

Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln

-continued

Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Ile Val Ser Asn Leu
 100 105 110
 Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln
 115 120 125
 Thr Pro Arg Gln Leu Tyr Val Ala Leu Arg Lys Leu Gly Lys Gln Asn
 130 135 140
 Pro Ala Leu Tyr Arg Val Val Glu Tyr Ile Asn Val Leu Val Arg Leu
 145 150 155 160
 Arg Arg Glu Glu Phe Asp Glu Asp Gln Arg Arg Ser Leu Ser Ala Leu
 165 170 175
 Pro Phe Gly Asp Asn Asn Asp Asp Leu Glu Glu Gln Asp Phe Gly Glu
 180 185 190
 Gln Asp Phe Arg Tyr Ile Tyr Gly Phe Glu Cys Ala Arg Phe Ile Leu
 195 200 205
 Gln Asn Gly Arg Met Phe Gly Ile Asn Thr Asp Arg Arg Tyr
 210 215 220

<210> SEQ ID NO 37
 <211> LENGTH: 132
 <212> TYPE: PRT
 <213> ORGANISM: Cyathostomum sp.

<400> SEQUENCE: 37

Arg Glu Lys Ala Arg Ile Ile Gln Asp Glu Tyr Thr Lys Arg Met Gln
 1 5 10 15
 Gln Val Thr Pro Gln Ala Gln Glu Phe Leu Ala Lys Trp Glu Lys Thr
 20 25 30
 Trp Phe Thr Asn Val Gln Gln Tyr Ser Gly Asp Lys Lys Ala Phe Phe
 35 40 45
 Lys Gln Met Ile Glu Leu Ile Pro Gln Leu Met Glu Glu Val His Gly
 50 55 60
 Phe Ser Glu Glu Thr Trp Lys Ser Leu Glu Glu Gln Phe Pro Glu Gln
 65 70 75 80
 Thr Ala Ala Trp Lys Asp Asn Glu Asp Arg Leu Lys Gln Phe Tyr Glu
 85 90 95
 Phe Ile Lys Ser Leu Pro Lys Gln Asp Leu Ala Glu Asp Pro Glu Ala
 100 105 110
 Phe Arg Lys Phe Ala His Leu Gly Leu Gln Lys Leu Leu Pro Ile Glu
 115 120 125
 Ala Leu Arg Ala
 130

<210> SEQ ID NO 38
 <211> LENGTH: 396
 <212> TYPE: DNA
 <213> ORGANISM: Cyathostomum sp.

<400> SEQUENCE: 38

agggagaagg ctgaattat tcaagacgaa tacactaaac gtatgcagca ggtcacacca 60
 caagctcagg aattcctggc aaaatgggag aagacatggt tcacgaatgt gcagcaatat 120
 agcggagata agaaagcttt cttcaagcag atgattgagc taatccctca actaatggag 180
 gaggttcatt ggttctcgga agagacttgg aagagccttg aggagcaatt cccagagcag 240
 acagccgcat ggaagataa tgaggatcgc ctaaagcaat tttatgagtt tatcaagagc 300
 ctaccaagc aggacttagc tgaggatcgc gaagcattca gaaagtgcgc tcacctcgga 360

-continued

ctccagaaac ttcttccaat tgaagctctc agagct 396

<210> SEQ ID NO 39
 <211> LENGTH: 565
 <212> TYPE: DNA
 <213> ORGANISM: *Cyathostomum catinatum*.

<400> SEQUENCE: 39

tggtcacacc acaagctcag gagttcctgg ccaaggttaag ctattacctt accaggggtga 60
 ggggaaagaa gttggcagcg gtcggaaacc cggtaatcta ctgactttac caattathtt 120
 cagtgggaga agacatggtt cacgaatata cagcaataca gtggagacaa gcaagccttc 180
 tttaaagcaga tgattgaact aattcctcaa cttatggagg aggttcaggt aagttagccg 240
 caaaaathtt taaccaatgg ttgagctoga cattttttca gggattcaca gaggagactt 300
 ggaatagcct gagggagcaa ttcccgagc agacagccgc atggaaggat cgtgagtatc 360
 tttcataatt actgtacttg gaattatact ttacaatcat aatcctactc ttagacgagg 420
 atcgccgtgaa gcaattctat gagttcatta agagcctacc caaacaacaa ttagctgagg 480
 tgattttcat tgatttttcg aaaaatatat ttttgataca ttctttttca ggatccggaa 540
 gctttcagaa agttcgctca cctcg 565

<210> SEQ ID NO 40
 <211> LENGTH: 569
 <212> TYPE: DNA
 <213> ORGANISM: *Cyathostomum catinatum*.

<400> SEQUENCE: 40

ttgtcacacc acaagctcag gagttcctgg ctaaggttaag ctattacctt accaggggtga 60
 gggggaagaa gttgggagcg gtcggaaacc cggtaatcta ctgactttac caattathtt 120
 cagtgggaga ggacatggtt cacgaatata cagcaataca gtggagacaa gcaagccttc 180
 tttaaagcaga tgattgaact aattcctcaa cttatggagg aggttcaggt aagttggccg 240
 caaaaathtt taaccaatgg ttgagctoga cattttttca gggattcaca gaggagactt 300
 ggaatagcct gagggagcaa ttcccgagc agacagccgc atggaaggat cgtaagtatc 360
 tttcataatt actgtacttg gaattatact ttacaatcat aatcctactc ttagacgagg 420
 atcgccgtgaa gcaattctat gagttcatta agagcctacc caaacaacaa ttagctgagg 480
 tgattttcat tgatttttcg tacgaaaaat atatttttga tacattcttt ttcaggatcc 540
 ggaagctttc agaaagttcg ctcacctcg 569

<210> SEQ ID NO 41
 <211> LENGTH: 561
 <212> TYPE: DNA
 <213> ORGANISM: *Cylicostephanus longibursatus*

<400> SEQUENCE: 41

aggtcacacc acaagctcag gaattcctgg caaaggttaag ctatcacctt accaggggtga 60
 ggggtagaag ttaggagcga gggaaaccgg tgatctctta taccattac ttcagtggga 120
 gaagatatgg ttcacgaatg tacagcaata tagtgagac aagcaagcct tcttcaagca 180
 gatgattgaa ctaattcctc aacttatgga ggaggtacag gtaagtcagc taaagtgatt 240
 ttaagaaaaa attaagcctg atttccttt cagggattct cagaggagac ttggaatagc 300
 ctaaaggagc aattcctgga gcagacagcc gcatggaagg atagttagta tttttcataa 360

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ttactgtact tggaattata ctttacaatc ataatectac cctcagacga ggagcgctg 420
aagcaattct atgagttcat taagagccta cccaaacaac aaatagctga ggtgatttct 480
attgattttt cgtacgaaaa gtatatTTTT aatacattct tttgcaggat ccggaagcct 540
tcagaaagtt cgctcacctc g 561

```

```

<210> SEQ ID NO 42
<211> LENGTH: 589
<212> TYPE: DNA
<213> ORGANISM: Cylicocyclus nassatus

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```

<400> SEQUENCE: 42

```

```

aggtcacacc acaagctcag gaattcctgg caaaggttaag ctaccatatt tcgaggggga 60
gggcaatttt ggagcgaggg aggagagaa agggagagaa aactgggttg ggatcactaa 120
ctctaccgcg cacttcaggt gggagaagac atgggttcacg aatgtgcagc aatatacgcg 180
agataagaaa gcctttttca aacagatgat tgagctaate cctcaactaa tggaagaggt 240
tcattgtaagt caaccaaaat ggctttttaag cggagattaa actcgaattt ttcttcaggg 300
gttctcggag gagacttggg agagccttga ggagcaattc ccagagcaga cagcccgatg 360
gaaggatagt aagcattctt catagctccc gcctttatca tttatcttca cgatagtaat 420
cttattttta gatgaggatc gcctgaagca attttatgag ttcatcaaga gcctacccaa 480
gcaggactta gctgaggtaa ctttcatggt tttttcctga gctgtaaaaa tgcttgcaac 540
taacaacttt tetaggatcc ggaagctttc agaaagtctg ctcacctcg 589

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<210> SEQ ID NO 43
<211> LENGTH: 195
<212> TYPE: PRT
<213> ORGANISM: Cyathostomum sp.

```

```

<400> SEQUENCE: 43

```

```

Lys Lys Glu Ser Gln Gly Phe Phe Ser Ile Pro Val Asp Asn Leu Arg
1           5           10           15
Ala Ser Pro Phe Leu Leu Gln Tyr Ile Lys Glu Tyr Ile Pro Asp Tyr
          20           25           30
Lys Asn Ala Met Glu Lys Phe Glu Asp Ile Pro Lys Gln Tyr Arg Asp
          35           40           45
Leu Ile Pro Glu Glu Val Ala Thr His Leu Lys Ala Ile Thr Ala Glu
          50           55           60
Glu Lys Ala Val Leu Lys Glu Val Met Lys Asp Tyr Ala Lys Tyr Lys
          65           70           75           80
Asp Glu Glu Glu Phe Leu Lys Ala Leu Lys Glu Lys Ser Glu Gly Leu
          85           90           95
His Glu Lys Ala Ser Lys Leu His Asn Phe Ile Lys Gly Lys Val Asp
          100          105          110
Ala Leu Gly Asp Glu Ala Lys Ala Phe Val Lys Lys Val Ile Ala Ala
          115          120          125
Ala Arg Glu Val His Ala Lys Leu Leu Ala Gly Asp Lys Pro Ser Leu
          130          135          140
Glu Asp Ile Lys Lys Lys Ala Lys Glu His Met Gly Glu Phe Glu Lys
          145          150          155          160
Leu Ser Asp Asp Ala Lys Glu Asp Leu Lys Lys Asn Phe Pro Ile Leu
          165          170          175
Thr Ser Val Trp Thr Asn Glu Lys Thr Arg Ala Leu Ile Asp Lys Tyr
          180          185          190

```

-continued

Val Glu Asn
195

<210> SEQ ID NO 44
<211> LENGTH: 573
<212> TYPE: DNA
<213> ORGANISM: Cyathostomum sp.

<400> SEQUENCE: 44

```
atgcttcgaa taactttctt ccttgccttc tttggtgtct acactttttc tgcacctctt    60
ggaccgctg aagagaagat agatgtggaa aaaatggaaa aatttgaaga tattccaaag    120
caatatcgag accttattcc ggaagaggta gctacacacc tcaaagccat caccgctgaa    180
gagaaagctg ttctaaaaga ggtaatgaag aattatgcaa agtacaagaa cgaggaggag    240
tttttgaag cgttgaaga aaaatcagag agtttgcatt gaaaagccag caaacttcac    300
aattttatca aaggaaggt tgacgcactt ggagatgaag caaaggcatt tgtgaagaag    360
gttatcgag ctgctcgaga agtgcactgc aaacttcttg ccggggacaa accatcgctt    420
gaagatatca agaagaaagc caaggagcat atggctgaat tcgagaaact aagcgatgat    480
gccaaaggag atctcaaaaa gaatttccca atccttactt ccgtctggac aaatgagaaa    540
acaagagcgt tgattgacaa atatgtggag aac                                573
```

<210> SEQ ID NO 45
<211> LENGTH: 145
<212> TYPE: PRT
<213> ORGANISM: Cyathostomum sp.

<400> SEQUENCE: 45

```
Gly Lys Met Ser Asp Leu Trp Thr Ala Ile Ser Glu Thr Asn Lys Val
1           5           10           15
Arg Leu Phe Asn Thr Leu Ser Leu Gly Ile Ala Gly Val Leu Cys Ile
20          25          30
Thr Thr Ala Phe Ile Pro Val Glu Asn Gln Val Val Cys Ala Val Leu
35          40          45
Ile Thr Leu Leu Gln Gly Val Ile Gly Phe Asn Ser Ala Gly Tyr Asn
50          55          60
Lys Ala Ala Val Ile Val Ala Arg Gln His Ala His Leu Leu Leu Thr
65          70          75          80
Cys Phe Gly Leu Ile Val Thr Phe Val Pro Leu Val Gln Pro Phe Ile
85          90          95
Val Gln Leu Val Ala Pro Asp His Ser Trp Asp Gln Trp Phe Tyr Leu
100         105         110
Phe Val Gly His Gly Leu Val Leu Val Ile Ala Asn Leu Phe Phe Cys
115         120         125
Leu Thr Ile Glu Ala Lys Pro Ala Ala Phe Thr Gln Lys Thr Asp Ser
130         135         140
Ser
145
```

<210> SEQ ID NO 46
<211> LENGTH: 435
<212> TYPE: DNA
<213> ORGANISM: Cyathostomum sp.

<400> SEQUENCE: 46

```
ggtaaaatgt cagatttatg gacggcaata agcgaacaaa ataaagtccg cttgttcaac    60
```

-continued

```

accttgctgc tgggaattgc tggcgtactg tgtataacta ctgctttcat tcctgtggaa 120
aatcaggttg tttgcgctgt tttaatcacg ttattgcaag gagttatcgg attcaattca 180
gctggatata acaaaagctgc agtcattggt gctaggcagc atgetcatct tctggtgacc 240
tgetttgggc tcattgtcac ttttgtcccc ttggtgcagc cattcatagt tcaacttgtg 300
gcccctgacc atagctggga ccaatggttt tatctgtttg ttgggcatgg tctcgtactt 360
gttatagcga atttattctt ttgtctcact atcgaggcga aaccggcagc gttcacacag 420
aaaactgatt catca 435

```

```

<210> SEQ ID NO 47
<211> LENGTH: 173
<212> TYPE: DNA
<213> ORGANISM: Cyathostomum sp.

```

```

<400> SEQUENCE: 47
ggtttaatta cccaagtttg aggtactttc taaatctgac cggatcaact gatttgtgtc 60
tgattaaatt ttgaaaatct ctccctgaat agggagagta caagagtgca tatccaaaaa 120
aaaaaaaaaa aaaaaaaaaa aaaaacatgt cggccgcctc ggctctaga ata 173

```

```

<210> SEQ ID NO 48
<211> LENGTH: 287
<212> TYPE: DNA
<213> ORGANISM: Cyathostomum sp.

```

```

<400> SEQUENCE: 48
ggtttaatta cccaagtttg agtgtcatga agcttgcctg aaaaaagcag agaaccaag 60
aggagatagt ttcacagttc cgccagacag gaaatgcgtg ccaagatggt ttgcggaaga 120
ggagaaacgt cgttcaacta gaatgagaag gcattgattc tgtttagtcg ttgagatatt 180
taaaaattct ttgcagaaaa ccttttcaaa tcataaagtc gaagaccaca aaaaaaaaaa 240
aaaaaaaaaa aaaaaaaaaa atgtcggccc cctcggcctc tagaata 287

```

```

<210> SEQ ID NO 49
<211> LENGTH: 620
<212> TYPE: DNA
<213> ORGANISM: Cyathostomum sp.

```

```

<400> SEQUENCE: 49
ggtttaatta cccaagtttg aggctgcttc aacagtaggt ttagaaatga catcgcggat 60
atggcgcgcg acccagagcc ctccattatt gctactcctg ttgttgatca gtctaccagt 120
agctgagtggt agtattcgac tatgtggagt gcgactaaca cgaactctta tggctatctg 180
caggaatcaa ttatcgggtt attcgcgaaag taaaagatct gctatgtggg aagagcctcg 240
actggaaacc gtgcactcaa caatgaaacg atcagggatc gccaccgaat gctgcgagaa 300
tcggtgtctca ttagctact taaagacata ctgctgcagc acttagcctt ggcactctaa 360
gccgctttta tctctctccc atgatctctc ttcgttatct gtataaccga atatagtcac 420
tccgaaaatg cggatgctta ggccaatttg ttgacgtttg cgcgatgaat catttgetgt 480
tcgtcattat ctcacagacg tgtaaaagat ctctttttat gaaagtctat tttggttgag 540
ctgcaccatt aaaccgttca caaaaaaaaa aaaaaaaaaa aaaaaaaaaa acatgtcggc 600
cgctcggccc tctagaataa 620

```

```

<210> SEQ ID NO 50

```

-continued

<211> LENGTH: 172
 <212> TYPE: DNA
 <213> ORGANISM: *Cyathostomum* sp.
 <400> SEQUENCE: 50
 ggtttaatta cccaagtttg aggtactttc tagatctgac cggatcaact gattgtggtc 60
 tgattaaatt ttggaatct cttcctgaac agggagagta caagagtgta tatgaaaaaa 120
 aaaaaaaaaa aaaaaaaaaa aaaacatgtc ggccgcctcg gcctctagaa ta 172

<210> SEQ ID NO 51
 <211> LENGTH: 327
 <212> TYPE: DNA
 <213> ORGANISM: *Cyathostomum* sp.
 <400> SEQUENCE: 51
 ggtttaatta cccaagtttg aggatgctta gtttcaagct cgtttcttc ttcgtacttc 60
 tcacagcttg tgtgctaaca gatccaagag tgtaatccg agaaaagcga atggactgga 120
 gacgttacta tagcagatgg ggtcgcggaa gctctaattg gggaaaccgc ggaggtacct 180
 tcggcggacg aaaatggagt taccgcactt ttggacaatg gggacattaa catctgatgt 240
 atgaaaagat ctaatgaaat aaagcttcga aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa 300
 atgtcggccg cctcggcctc tagaata 327

<210> SEQ ID NO 52
 <211> LENGTH: 324
 <212> TYPE: DNA
 <213> ORGANISM: *Cyathostomum* sp.
 <400> SEQUENCE: 52
 ggtttaatta cccaagtttg agaatgttcg aaaaattcct tctgctactg atcgttggtga 60
 tcgcccctcat ttctttggcg tctgcagatt tttcatgctt cttcgggtgat accatctgca 120
 agagcattac atgcaggggc tgcaccgtcg ccacttgctt taatggagac tgtatgtgca 180
 cactatgtaa ctgatgatct tcacatgtcg cattaccatt tgtaacaaat acattttctc 240
 ttgttcataa taaatttttc actcaaaaaa aaaaaaaaaa aaaaaaaaaa aaaaaaacatg 300
 tcggccgcct cggcctctag aata 324

<210> SEQ ID NO 53
 <211> LENGTH: 328
 <212> TYPE: DNA
 <213> ORGANISM: *Cyathostomum* sp.
 <400> SEQUENCE: 53
 ggccgcggga ttttctagag gccgaggcgg gtttttaggt gttcctcaaa cttgggtaat 60
 taaaccacga ggccgaggcg ggttttaggt tgttctcaaa cttgggtaat taaaccacga 120
 tggcgaggcg ggttttaggt tgttctcaaa cttgggtaat taaaccacga tggcgaggcg 180
 ggttttaggt tgttctcaa acttgggtaa ttaaaccaag aggcgaggcg ggttttaggt 240
 ttgttctca aacttgggta attaaaccac gatggcgagg cgggttttag gttgttctca 300
 aacttgggta attaaaccaa tcactagt 328

<210> SEQ ID NO 54
 <211> LENGTH: 154
 <212> TYPE: DNA
 <213> ORGANISM: *Cyathostomum* sp.
 <400> SEQUENCE: 54

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```

ggccgcggga ttattctaga ggccgaggca gtggtatcaa cgcagagtgg ccattacggc 60
cggggagagg gaaaagtctt tttctctcgc gataccacaaa aaaaaaaaaa aaaaaaaaaa 120
aaaaaaaaacatg tcggccgcct cggcctctag aata 154

```

```

<210> SEQ ID NO 55
<211> LENGTH: 387
<212> TYPE: DNA
<213> ORGANISM: Cyathostomum sp.

```

```

<400> SEQUENCE: 55

```

```

ggccgcggga ttttctagag gccgaggcgt cttacttggg tggetcaata actgaaagct 60
tagaattcat taaccttaa cccacagggg ttatttgaca tgcttgactt gaaatgatg 120
ctcttctgct tgtagttggt ttattatgct agctgtaagt atactctggt agaccagaac 180
atcaatgtgc tagttgaatg tatcatgtta tcactttgtc aactctata cgaatctagg 240
tgtggcaggc cacaccctc tcctgacctt gttcaccatc aattagcttt tagctgttat 300
ttaataacat cacactgatt gcaaaaaaaaa aaaaaaaaaa aaaaaaaaaa acatgtcggc 360
cgctcgggcc tctaaaaaat cactagt 387

```

```

<210> SEQ ID NO 56
<211> LENGTH: 292
<212> TYPE: DNA
<213> ORGANISM: Cyathostomum sp.

```

```

<400> SEQUENCE: 56

```

```

ggccgcggga ttattctaga ggccgaggca gtggtatcaa cgcagagtgg ccattacggc 60
cgaagcagtg gtatcaacgc agagtggcca ttacggccgg gtggtgacca cgggtgacgg 120
ggaattaggg ttcgattccg gagagggagc ctgagaaacg gctaccacat ccaaggaagg 180
cagcagggcg gcaaattacc cactcccgcg ccggggaggt agtgacgaaa aaaaaaaaaa 240
aaaaaaaaaa aaaaacatgt cggccgcctc ggctctaga ataactacta gt 292

```

```

<210> SEQ ID NO 57
<211> LENGTH: 199
<212> TYPE: DNA
<213> ORGANISM: Cyathostomum sp.

```

```

<400> SEQUENCE: 57

```

```

ggccgcggga ttttctagag gccgaggcgg gttttagctc aaacttgggt aattaaccg 60
gtaggatggc gaggcggggt tctcaaaact ggtaattaa accagtagga tggcgaggcg 120
ggtttctcaa acttgggtaa ttaaacgggt aggaggccga ggcgggtctc aaacttgggt 180
aattaacca atcactagt 199

```

```

<210> SEQ ID NO 58
<211> LENGTH: 167
<212> TYPE: DNA
<213> ORGANISM: Cyathostomum sp.

```

```

<400> SEQUENCE: 58

```

```

caagtttgag gtactttcta gatctgacct gatcaactga ttgtggtctg attaaatctt 60
ggaatctct tctgaacag ggagagtaca agagtgtata ttaagaaaaa aaaaaaaaaa 120
aaaaaaaaaa catgtcggcc gcctcggcct ctagaataat cactagt 167

```

```

<210> SEQ ID NO 59

```

-continued

<211> LENGTH: 22
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 59

ggtttaatta cccaagtttg ag 22

<210> SEQ ID NO 60
<211> LENGTH: 17
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 60

attctagagg ccgaggc 17

<210> SEQ ID NO 61
<211> LENGTH: 17
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 61

ttctagaggc cgaggcg 17

<210> SEQ ID NO 62
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 62

aaaaaggagg tgtttggttc 20

<210> SEQ ID NO 63
<211> LENGTH: 23
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 63

cttgaatttg ataaaactac acc 23

<210> SEQ ID NO 64
<211> LENGTH: 18
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 64

aattgtgggg aacaggag 18

<210> SEQ ID NO 65
<211> LENGTH: 21
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 65

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aatgaaaatc agactcctag g 21

<210> SEQ ID NO 66
 <211> LENGTH: 34
 <212> TYPE: DNA
 <213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 66

aattcggatc cgcaaggtgt catggacctt tttg 34

<210> SEQ ID NO 67
 <211> LENGTH: 38
 <212> TYPE: DNA
 <213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 67

ccgcaagctt atatctttca tctgtgttga gtccaaac 38

<210> SEQ ID NO 68
 <211> LENGTH: 60
 <212> TYPE: PRT
 <213> ORGANISM: *Cyclicoclyclus ashworthi*
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <222> LOCATION: (43)..(43)
 <223> OTHER INFORMATION: X is E or V
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <222> LOCATION: (50)..(50)
 <223> OTHER INFORMATION: X is G or D
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <222> LOCATION: (57)..(57)
 <223> OTHER INFORMATION: X is E or V

<400> SEQUENCE: 68

Leu Thr Phe Ala Glu Lys Lys Gly Lys Ile Ser Glu Trp Ala Lys Lys
 1 5 10 15

Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
 20 25 30

Leu Lys Gln Glu His Arg Lys Asn Val Ser Xaa Leu Val Ser Gly Leu
 35 40 45

Pro Xaa Ala Val Lys Lys Val Asn Xaa Leu Leu Asp
 50 55 60

<210> SEQ ID NO 69
 <211> LENGTH: 60
 <212> TYPE: PRT
 <213> ORGANISM: *Cyathostomum catinatum*
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <222> LOCATION: (8)..(8)
 <223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid

<400> SEQUENCE: 69

Leu Thr Phe Ala Glu Lys Lys Xaa Glu Ile Ser Glu Trp Ala Lys Lys
 1 5 10 15

Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
 20 25 30

Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Ala Leu
 35 40 45

-continued

Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp
50 55 60

<210> SEQ ID NO 70
<211> LENGTH: 60
<212> TYPE: PRT
<213> ORGANISM: Coronocyclus coronatus

<400> SEQUENCE: 70

Leu Thr Phe Ala Glu Lys Lys Glu Lys Ile Ser Glu Trp Ala Lys Lys
1 5 10 15

Tyr Lys Val Glu Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
20 25 30

Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Ala Leu
35 40 45

Pro Gly Ala Val Lys Lys Val Asn Glu Leu Leu Asp
50 55 60

<210> SEQ ID NO 71
<211> LENGTH: 60
<212> TYPE: PRT
<213> ORGANISM: Cyclicostephanus goldi

<400> SEQUENCE: 71

Leu Thr Phe Ala Glu Lys Lys Lys Glu Ile Ser Glu Trp Ala Lys Lys
1 5 10 15

Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
20 25 30

Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Asp Leu
35 40 45

Pro Ser Ala Val Lys Lys Val Asn Asp Leu Leu Asp
50 55 60

<210> SEQ ID NO 72
<211> LENGTH: 60
<212> TYPE: PRT
<213> ORGANISM: Coronocyclus labiatus

<400> SEQUENCE: 72

Leu Thr Phe Ala Glu Lys Lys Glu Lys Ile Ser Glu Trp Ala Lys Lys
1 5 10 15

Tyr Asn Val Val Asp Glu Val Ala Arg Tyr Asn Ala Tyr Arg Glu Lys
20 25 30

Leu Lys Gln Glu Tyr Arg Lys Asn Val Ser Glu Leu Val Ser Gly Leu
35 40 45

Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp
50 55 60

<210> SEQ ID NO 73
<211> LENGTH: 60
<212> TYPE: PRT
<213> ORGANISM: Cyclicocyclus leptostomum

<400> SEQUENCE: 73

Leu Thr Phe Ala Glu Lys Lys Gly Lys Ile Ser Glu Trp Ala Lys Lys
1 5 10 15

Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
20 25 30

Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Gly Leu

-continued

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      35              40              45
Pro Gly Ala Val Lys Lys Val Asn Glu Leu Leu Asp
   50              55              60

```

```

<210> SEQ ID NO 74
<211> LENGTH: 60
<212> TYPE: PRT
<213> ORGANISM: Cylicostephanus longibursatus
<400> SEQUENCE: 74

```

```

Leu Thr Phe Ala Glu Lys Lys Glu Glu Ile Ser Lys Trp Ala Lys Lys
 1              5              10              15
Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
              20              25              30
Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Ile Val Ser Asp Leu
   35              40              45
Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp
   50              55              60

```

```

<210> SEQ ID NO 75
<211> LENGTH: 60
<212> TYPE: PRT
<213> ORGANISM: Cylicostephanus minutus
<400> SEQUENCE: 75

```

```

Leu Thr Phe Ala Glu Lys Lys Glu Lys Ile Ser Glu Trp Ala Lys Lys
 1              5              10              15
Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
              20              25              30
Leu Lys Gln Glu His Arg Lys Asn Val Ser Gln Leu Val Ser Ala Leu
   35              40              45
Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp
   50              55              60

```

```

<210> SEQ ID NO 76
<211> LENGTH: 60
<212> TYPE: PRT
<213> ORGANISM: Cylicocyclus nassatus
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (25)..(25)
<223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid
<400> SEQUENCE: 76

```

```

Leu Thr Phe Ala Glu Lys Lys Glu Lys Ile Gly Glu Trp Ala Lys Lys
 1              5              10              15
Tyr Asn Val Val Asp Glu Val Ala Xaa Tyr Asn Ala Tyr Arg Glu Lys
              20              25              30
Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Gly Leu
   35              40              45
Pro Asn Ala Val Lys Lys Val Asn Glu Leu Leu Asp
   50              55              60

```

```

<210> SEQ ID NO 77
<211> LENGTH: 60
<212> TYPE: PRT
<213> ORGANISM: Cyathostomum pateratum
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<222> LOCATION: (8)..(8)
<223> OTHER INFORMATION: X is K or E

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<400> SEQUENCE: 77

Leu Thr Phe Ala Glu Lys Lys Xaa Glu Ile Ser Glu Trp Ala Lys Lys
 1 5 10 15
 Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
 20 25 30
 Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Ala Leu
 35 40 45
 Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp
 50 55 60

<210> SEQ ID NO 78

<211> LENGTH: 223

<212> TYPE: PRT

<213> ORGANISM: Cyathostomum pateratum

<400> SEQUENCE: 78

Met Asn Lys Thr Leu Thr Phe Leu Thr Val Val Ser Ala Val Ala Leu
 1 5 10 15
 Ala Gln Gly Val Met Asp Leu Phe Gly Glu Glu Gly Arg Glu Glu His
 20 25 30
 Arg Arg His His Arg His Ser Leu Leu Pro Pro Tyr Leu His Asn Val
 35 40 45
 Ser Cys Glu Ala Lys Trp Glu Tyr Phe Lys Ile Val Gly Asn Arg Ser
 50 55 60
 Leu Thr Phe Ala Glu Lys Arg Lys Glu Ile Ser Glu Trp Ala Lys Lys
 65 70 75 80
 Tyr Asn Val Val Asp Glu Val Ala Ser Tyr Asn Ala Tyr Arg Glu Lys
 85 90 95
 Leu Lys Gln Glu His Arg Lys Asn Val Ser Glu Leu Val Ser Ala Leu
 100 105 110
 Pro Asn Ala Val Lys Lys Val Asn Asp Leu Leu Asp Asn Glu Asn Gln
 115 120 125
 Thr Pro Arg Gln Leu Tyr Val Ala Leu Arg Lys Leu Gly Arg Gln Asn
 130 135 140
 Pro Ala Leu Tyr Arg Ile Val Glu Tyr Ile Asn Val Ala Val Arg Leu
 145 150 155 160
 Arg Ser Glu Glu Val Asp Glu Gln Glu Gln Arg Arg Arg Leu Ser Ala
 165 170 175
 Leu Pro Phe Gly Asp His Asn Asp Asn Leu Glu Glu Gln Asp Phe Gly
 180 185 190
 Glu Gln Asp Phe Arg Tyr Val Tyr Gly Phe Glu Cys Ala Arg Phe Leu
 195 200 205
 Leu Gln Asn Gly Arg Met Phe Gly Leu Asn Thr Asp Glu Arg Tyr
 210 215 220

<210> SEQ ID NO 79

<211> LENGTH: 203

<212> TYPE: PRT

<213> ORGANISM: Nippostrongylus brasiliensis

<400> SEQUENCE: 79

Met Lys Ala Leu Leu Ile Ala Val Leu Ala Leu Thr Ala Ala Ala His
 1 5 10 15
 Tyr Arg Gly Lys Glu Ser Glu Thr Gly His Gly Arg His His His His
 20 25 30
 Pro Pro Pro Pro Phe Leu Lys Asp Val Asp Lys Ser Ala Arg Lys

-continued

Thr Cys Asn Asn
210

<210> SEQ ID NO 81
<211> LENGTH: 201
<212> TYPE: PRT
<213> ORGANISM: Ostertagia ostertagi
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (4)..(4)
<223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid

<400> SEQUENCE: 81

Lys Ser Ala Xaa Phe Leu Leu Leu Ile Gly Ala Thr Val Leu Gly His
1 5 10 15
Ala Arg His Asn Glu Gly Cys Pro Arg Glu Glu Ser His Arg Ala Gln
20 25 30
Pro Arg Pro Lys Phe Leu His His Val Gly Ile Lys Ala Arg Arg Glu
35 40 45
Tyr Phe His Ile Val Arg Ser Gly Glu Ile Ile Ala Lys Gln Asp Glu
50 55 60
Gln Ile Leu Asp Trp Ala Lys Lys Tyr Gly Val Glu Glu Glu Val Glu
65 70 75 80
Glu Phe Asn Asn Lys Thr Ala Ser Tyr Val Glu Glu Leu Val Gln Asn
85 90 95
Val Thr Asn Leu Ile Ala Glu Leu Pro Thr Ala Leu Glu Ala Phe Leu
100 105 110
Asn Ile Thr Gln Asn Lys Asp Gln Thr Arg Met Glu Met Lys Lys Ala
115 120 125
Leu Arg Glu Met Arg Thr Glu Glu Phe Glu Val Phe Asp Ala Leu Lys
130 135 140
Ala Ala Phe Lys Val Phe Lys Pro Asn His Cys Leu Tyr His Arg Cys
145 150 155 160
Thr Asp Ser Gln Ser Ser Glu Glu Leu Val Asp Asp Trp Met Asp Phe
165 170 175
Gln Glu Lys Asp Asp Gln Ile Ser Lys Met Leu Asp Pro His Asp Glu
180 185 190
Phe Thr Met Met Gln Lys Ser Gly Val
195 200

<210> SEQ ID NO 82
<211> LENGTH: 241
<212> TYPE: PRT
<213> ORGANISM: Caenorhabditis elegans

<400> SEQUENCE: 82

Met Ala Leu Ser Tyr Ser Phe Ile Phe Thr Leu Phe Ala Phe Ser Ala
1 5 10 15
Val Val Leu Ala Gly Pro Gly Gly Arg His Gly His Gly Gly Gly Gly
20 25 30
Phe Gly Gly Ala Pro Gln Leu Pro Pro Phe Leu Gln Asn Val Thr Ala
35 40 45
Glu Gly Arg Gln Ala Phe Phe Ala Ile Val Ser Asn Thr Ser Leu Thr
50 55 60
Ile Ser Glu Thr Glu Ser Gln Ile Ser Ser Trp Ala Gln Thr Tyr Gly
65 70 75 80
Val Ser Ser Gln Val Thr Glu Phe Gln Thr Lys Val Glu Glu Lys Leu
85 90 95

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Asn Glu Ile Lys Gln Asn Val Thr Ala Val Ile Asn Asn Leu Ser Thr
 100 105 110

 Val Glu Thr Gln Leu Glu Ala Ile Phe Ala Asn Lys Ser Gln Thr Ile
 115 120 125

 Arg Glu Gln Phe Gln Ala Leu Gly Gln Leu Lys Asp Gln Tyr Pro Gln
 130 135 140

 Glu Val Gly Val Leu Leu Phe Leu Ala Lys Pro Lys Gly Glu His Gly
 145 150 155 160

 Gly Gln Gly Pro Phe Gly Gly Phe Pro Gly Gly His Gln Gly Gly Phe
 165 170 175

 Pro Gly Gly Asn Gln Gly Gly Phe Gly Gly Asn Gln Gly Gly Phe Gly
 180 185 190

 Gly Asn Gln Gly Gly Phe Pro Phe Gly Asn Gln Gly Gly Asn Gln Gly
 195 200 205

 Gly Phe Pro Phe Gly Asn Pro Gly Asn Gln Gly Gly Phe Gly Gly Asn
 210 215 220

 Gln Gly Gly Asn Gln Gly Gly Phe Gly Gly Asn Arg Gly Gly Arg Gly
 225 230 235 240

 Phe

<210> SEQ ID NO 83

<211> LENGTH: 235

<212> TYPE: PRT

<213> ORGANISM: Caenorhabditis elegans

<400> SEQUENCE: 83

Met Ser Tyr Tyr Ser Thr Ser Leu Tyr Ile Phe Ala Ile Thr Met Ala
 1 5 10 15

 Thr Met Val Leu Ala Gly Pro Arg Gly Gly Phe Gly Gly Gly Pro Gly
 20 25 30

 Gly Pro Gly Gly Arg Gly Arg His Gly Pro Pro Met Pro Pro Phe Leu
 35 40 45

 Gln Asn Val Thr Asp Glu Gly Arg Arg Ala Phe Phe Asp Ile Ala Arg
 50 55 60

 Asn Gln Asn Leu Thr Ile Ala Glu Met Glu Ser Gln Thr Ser Thr Trp
 65 70 75 80

 Ala Gln Thr Tyr Gly Val Ser Asp Val Tyr Ser Glu Phe Glu Ala Asn
 85 90 95

 Ile Thr Ala His Arg Asn Glu Val Gln Gln Asn Val Thr Gln Val Val
 100 105 110

 Ser Gln Leu Ser Ala Ala Gln Thr Ala Leu Glu Ala Val Met Asn Asn
 115 120 125

 Lys Asn Gln Thr Arg Gln Gln Met Lys Glu Ala Ile Asp Asn Leu Lys
 130 135 140

 Thr Gln Tyr Pro Gln Glu Ile Pro Ala Leu Phe Phe Ile Ser Gly Ser
 145 150 155 160

 Phe Arg Arg Gly Pro Gly Gly Arg His Gly Gly Pro Gly Gly Pro Gly
 165 170 175

 Gly Arg Arg Met Gly Pro Gly Gly Arg Gly Gly Asp Ser Arg Glu Gly
 180 185 190

 Pro Met Met Gly Gly Met Gly Arg Gly Gly Phe Gly Gly Gln Gly Met
 195 200 205

-continued

Gly	Gly	Met	Gly	Ala	Gly	Leu	Gly	Gln	Gly	Arg	Arg	Gly	Gly	Pro	Asp
	210					215					220				

Ser	Met	Asn	Glu	Ser	Ser	Asp	Val	Asn	Asp	Phe
225				230						235

The invention claimed is:

1. A method of diagnosing a cyathostomin infection in an animal, said method comprising the steps of:
 - contacting a sample with a cyathostomin larval antigen, wherein the cyathostomin larval antigen is at least 60% identical to the sequence of SEQ ID NO: 37; and identifying a level of anti-cyathostomin larval antigen antibodies in the sample, wherein the anti-cyathostomin larval antigen antibodies bind to antigen comprising an amino acid sequence at least 60% identical to the sequence of SEQ ID NO:37; wherein a level of anti-cyathostomin larval antigen antibodies is indicative of the cyathostomin infection.
 2. The method of claim 1, wherein the level of anti-cyathostomin larval antigen antibodies is evaluated relative to the level of anti-cyathostomin larval antigen antibodies present in a reference or control sample obtained from a healthy animal, an animal without a moderate or high mucosal burden of cyathostomin parasites and/or an animal without larval cyathostominosis.
 3. The method of claim 1, wherein the sample is contacted with a cyathostomin larval antigen comprising SEQ ID NO: 37.
 4. The method of claim 1, wherein the cyathostomin larval antigen is bound, conjugated or immobilised on or to a suitable substrate.
 5. The method of claim 3, wherein the sample is contacted with one or more agent(s) capable of binding:
 - (a) a cyathostomin larval antigen comprising SEQ ID NO: 37; or
 - (b) a cyathostomin larval antigen comprising an amino acid sequence at least 60% identical to SEQ ID NO: 37.
 6. The method of claim 5, wherein the binding agent(s) are bound, conjugated or immobilized on or to a suitable substrate.
 7. The method of claim 1, wherein the animal is a member of the Equidae family.
 8. The method of claim 1, wherein the animal is a horse.
 9. The method of claim 1, wherein the sample is a biological sample selected from the group consisting of: whole

- blood; serum; plasma; saliva; sweat; semen; tissue biopsy; tissue scraping; tissue/organ wash/lavage; and fecal preparation.
10. The method of claim 3, wherein the cyathostomin larval antigen is bound, conjugated or immobilised on or to a suitable substrate.
11. The method of claim 1, wherein the level of anti-cyathostomin larval antigen antibody is identified using an immunological detection technique.
12. The method of claim 11, wherein the immunological detection technique is selected from the group consisting of enzyme-linked immunosorbent assay (ELISA), Western blot and dot blot.
13. The method of claim 1, wherein the method further comprises contacting a sample with a second cyathostomin larval antigen, wherein the second cyathostomin larval antigen is at least 60% identical to the sequence of SEQ ID NO: 1; and identifying a second level of second anti-cyathostomin larval antigen antibodies in the sample, wherein the anti-cyathostomin larval antigen antibodies bind to antigen comprising an amino acid sequence at least 60% identical to the sequence of SEQ ID NO: 1, wherein the level of anti-cyathostomin larval antigen antibodies to SEQ ID NO: 37 or the second level of second anti-cyathostomin larval antigen antibodies to SEQ ID NO: 1 is indicative of the cyathostomin infection.
14. The method of claim 13, wherein the second cyathostomin larval antigen comprises SEQ ID NO: 1.
15. The method of claim 3, wherein the method further comprises contacting a sample with a cyathostomin larval antigen at least 60% identical to the sequence of SEQ ID NO: 1; and identifying a level of anti-cyathostomin larval antigen antibodies in the sample, wherein the anti-cyathostomin larval antigen antibodies bind to antigen comprising an amino acid sequence at least 60% identical to the sequence of SEQ ID NO: 1.
16. The method of claim 15, wherein the sample is further contacted with a cyathostomin larval antigen comprising SEQ ID NO: 1.

* * * * *

专利名称(译)	马寄生虫检测		
公开(公告)号	US9239330	公开(公告)日	2016-01-19
申请号	US14/193380	申请日	2014-02-28
申请(专利权)人(译)	MOREDUN研究所		
当前申请(专利权)人(译)	MOREDUN研究所		
[标]发明人	MATTHEWS JACQUI HODGKINSON JANE PROUDMAN CHRISTOPHER		
发明人	MATTHEWS, JACQUI HODGKINSON, JANE PROUDMAN, CHRISTOPHER		
IPC分类号	G01N33/569 G01N33/53		
CPC分类号	G01N33/5308 G01N33/569 G01N2333/4353		
优先权	2009005511 2009-03-31 GB PCT/GB2010/000616 2010-03-31 WO		
其他公开文献	US20140242619A1		
外部链接	Espacenet USPTO		

摘要(译)

本发明提供了一种诊断cyathostomin感染的方法，所述方法包括鉴定样品中抗cyathostomin幼虫抗原抗体水平的步骤，其中抗cyathostomin幼虫抗原抗体水平指示cyathostomin感染。

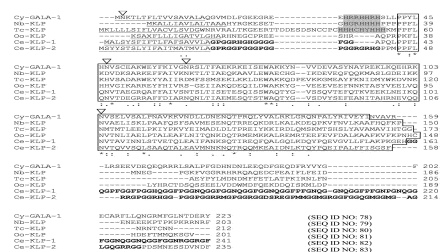


Figure 1

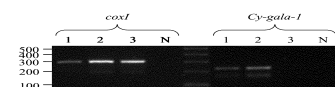


Figure 2