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(54) **METHOD FOR INVESTIGATION OF LIVER DAMAGE TYPE**

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(57) **ABSTRACT**

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Provided is a test method for discriminating a liver injury type using as an indicator a level of lithocholic acid (hereinafter abbreviated as LCA) in a biological sample of a subject. That is, provided is a test method comprising measuring a level of LCA in a biological sample collected from a subject, and discriminating a liver injury type as hepatocellular injury type, cholestasis type, or a mixed type thereof using the measured level of LCA as an indicator.

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Figure 1

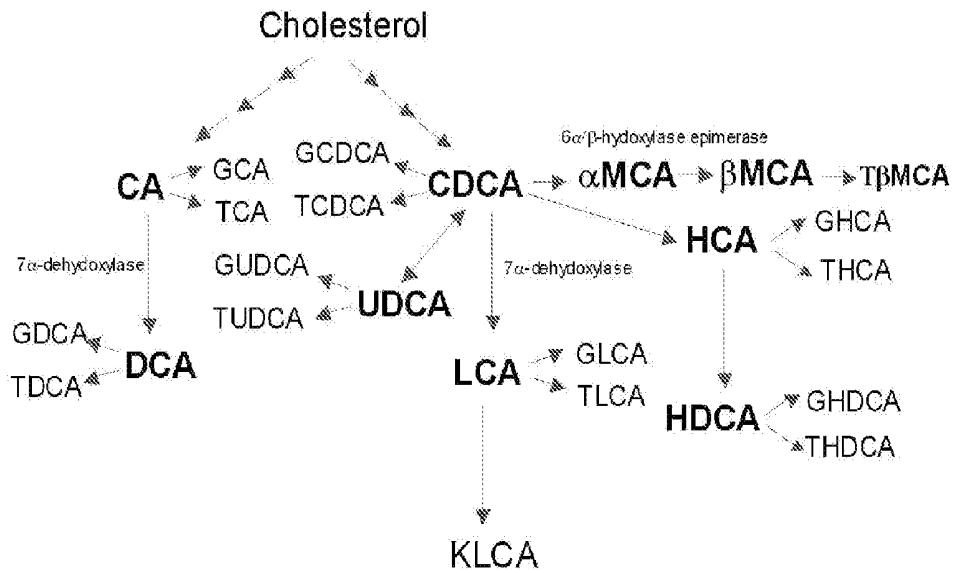


Figure 2

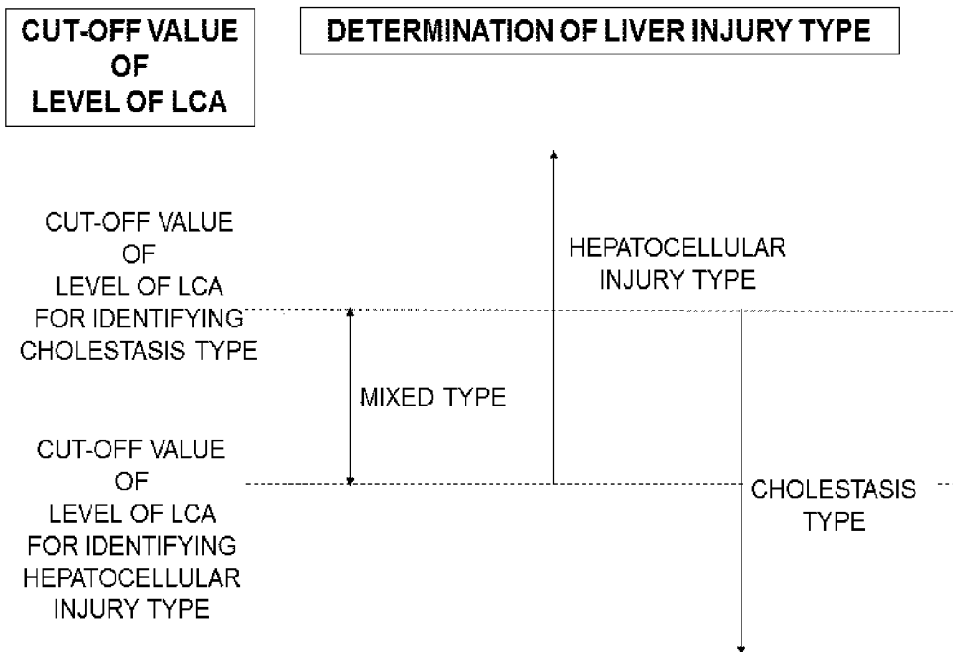


Figure 3

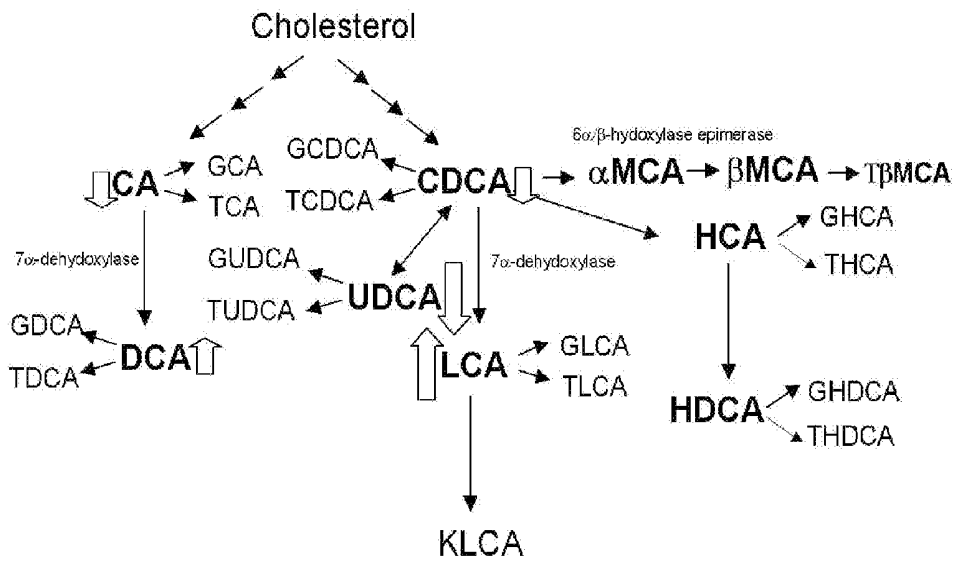
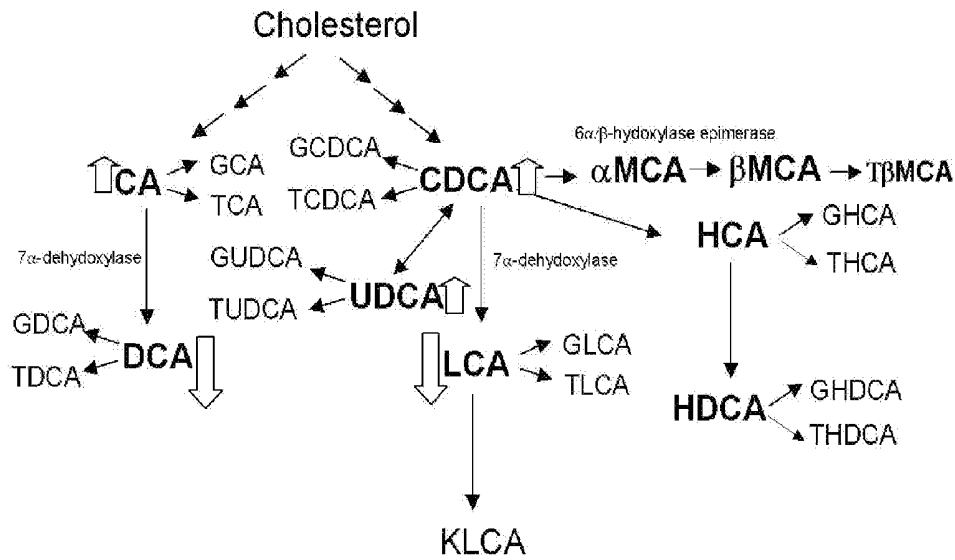


Figure 4



## METHOD FOR INVESTIGATION OF LIVER DAMAGE TYPE

### TECHNICAL FIELD

[0001] The present invention relates to a test method for liver injury, and more particularly, to a test method for discriminating a liver injury type. More particularly, the present invention relates to a test method comprising measuring a level of lithocholic acid (hereinafter sometimes abbreviated as LCA) in a biological sample collected from a subject, and discriminating a liver injury type using the measured level of LCA as an indicator.

### BACKGROUND ART

[0002] Liver is a glandular tissue connected to digestive tract and has various important functions for living organisms, such as production and secretion of bile, a detoxification action, carbohydrate metabolism, protein metabolism, production of a blood coagulation factor, a hormone regulation action, and storage of various components of living organisms, including fat, glycogen, a protein, and a vitamin. Therefore, when those functions are acutely or chronically damaged due to viral infection, a drug or a toxic agent, or excess alcohol intake, the maintenance of homeostasis of liver functions is disrupted, resulting in a critical health problem. Among various diseases found in comprehensive medical examination or the like, liver dysfunction accounts for a high proportion of the diseases, and about 30% of Japanese adults are estimated to have liver dysfunction.

[0003] Diseases included in liver disorders are classified based on their etiologies or clinical symptoms. For example, the diseases may be classified based on their etiologies into viral hepatitis, drug-induced liver injury, alcoholic liver injury, autoimmune liver injury, metabolic liver injury, and the like. In addition, the diseases may be classified based on their clinical symptoms into hepatocellular injury type and cholestasis type. Hepatocellular injury is found in viral hepatitis, toxic liver injury, fatty liver, cirrhosis, and the like. Hepatocellular injury is caused by, for example, necrosis, fatty metamorphosis, multinucleation, or nuclear vacuolar degeneration of a hepatocyte. The hepatocyte, which is also referred to as hepatic parenchymal cell, is one of the cells constituting liver and accounts for most major parts of liver. The hepatocyte is an exocrine cell which secretes bile, while it is also an endocrine cell which secretes plasma proteins into an apical membrane side and stores glycogen to regulate blood glucose. Therefore, when the hepatocyte is damaged, a significant reduction in liver function is caused. Cholestasis is caused by blockade of bile excretion from liver due to some cause, with the result that abnormal excretion of bile is brought about over a part or whole of intrahepatic or extrahepatic bile duct system, and thereby the bile accumulates in liver and blood. Then, various symptoms such as jaundice and hepatitis are induced.

[0004] At present, diagnosis and treatment of liver diseases are performed according to an approach in which a physician empirically diagnoses a liver injury type using a symptom of a patient and values of serum biochemical parameters as indicators and then treatment is started (Non Patent Literature 1). Enzymes which leak from the hepatocyte by hepatocellular injury are used as the biochemical parameters. Specifically, there are used, for example, aspartate aminotransferase (hereinafter abbreviated as AST; sometimes referred to as

glutamate-oxaloacetate transaminase (GOT)), alanine aminotransferase (hereinafter abbreviated as ALT; sometimes referred to as glutamate-pyruvate transaminase (GPT)),  $\gamma$ -glutamyl transpeptidase (hereinafter abbreviated as  $\gamma$ -GTP), and alkaline phosphatase (hereinafter abbreviated as ALP).

[0005] In addition, in non-clinical and clinical practices, measurement of total bile acids (hereinafter abbreviated as TBA) is generally performed. Bile acids is a general term for compounds that are steroid derivatives having cholanic acid skeletons widely found in bile of mammals and are major components of bile which plays a key role in digestion and absorption of fat. The bile acids are produced from metabolism of cholesterol in hepatocytes of liver (FIG. 1), followed by conjugating with glycine and taurine (aminoethylsulfonic acid) or the like, and then excreted as bile. The bile acids are roughly classified into primary bile acids and secondary bile acids. The primary bile acids are bile acids which are synthesized in liver. Examples of the primary bile acids may include cholic acid (hereinafter sometimes abbreviated as CA), chenodeoxycholic acid (hereinafter sometimes abbreviated as CDCA), and ursodeoxycholic acid (hereinafter sometimes abbreviated as UDCA) which has an isomeric relationship with CDCA. The secondary bile acids are bile acids which are produced from the primary bile acids excreted via bile duct to intestinal tract in a dehydroxylation reaction, a dehydrogenation reaction, a hydrogenation reaction, or a deconjugation reaction by enterobacteria. Examples of the secondary bile acids may include deoxycholic acid (hereinafter sometimes abbreviated as DCA) and LCA. Those five kinds of bile acid components given as examples each have three forms which are a free form, a glycine-conjugated form, and a taurine-conjugated form. In addition, some bile acid components each have a sulfated form or a glucuronidated form. In many animal species including humans, CA and CDCA having hydroxyl groups at position 3, 7, and 12 of a cholanic acid skeleton are produced as the primary bile acids. Meanwhile, some animals produce bile acids inherent in the species. For example,  $\alpha$ -muricholic acid (hereinafter sometimes abbreviated as  $\alpha$ MCA) and  $\beta$ -muricholic acid (hereinafter sometimes abbreviated as  $\beta$ MCA) are found in mice and hyocholic acid (hereinafter sometimes abbreviated as HCA) is found in swine, and these bile acids are rare in humans and are characteristic of each of the animal species.  $\alpha$ MCA and pmCA are primary bile acids and taurine-conjugated forms thereof are also known. HCA is a primary bile acid and produces hyodeoxycholic acid (hereinafter abbreviated as HDCA) by its 7 $\alpha$ -dehydroxylation in intestine. Those bile acid components also have glycine-conjugated forms and taurine-conjugated forms.

[0006] In treatment of liver diseases, discrimination of a liver injury type is important for selection of treatment suitable for each liver injury type. Hitherto, liver injury types have been determined based on biochemical parameters. For example, a liver injury type is determined using, as indicators, increases in ALT and AST in hepatocellular injury type and increases in ALP and  $\gamma$ -GTP in cholestasis type. However, a determination method based on biochemical parameters sometimes leads to misjudgment of a liver injury type. Therefore, an attempt has been made to measure levels of many components constituting bile acids in serum and discriminate a liver injury type based on the results to further determine a therapeutic strategy.

[0007] There have been previous reports on various relationships between classification of liver injury types based on etiologies and levels of bile acid components (Non Patent Literatures 2 to 6). However, no consensus has been reached and no clinical application has been achieved.

[0008] In addition, there have been some reports on the association between classification of liver injury types based on clinical symptoms and levels of bile acid components (Non Patent Literatures 6 to 8). However, similarly to the classification of liver injury types based on etiologies, a variety of results have been shown and no consensus has been reached. Meanwhile, for classification of types of drug-induced liver injury based on clinical symptoms, there has been proposed an assessment method using biochemical parameters rather than bile acid components (Non Patent Literature 9). Based on this proposal, the Japan Society of Hepatology either proposes scoring of drug-induced liver injury (Non Patent Literature 10). Further, the inventors of the present invention have reported that a liver disease caused by administration of a drug to a rat was compared to a typical cholestasis model obtained by ligation of bile duct, or administration of 1-naphtyl isothiocyanate (ANIT), or the like, and that injury types of liver diseases may be discriminated by measuring levels of bile acid components (Non Patent Literature 11). However, at present, there is no report on a definitive biomarker that clinically diagnoses an early stage of cholestasis in liver at a capillary bile duct level. Thus, the decision of a therapeutic strategy is difficult in many cases.

#### CITATION LIST

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#### SUMMARY OF INVENTION

##### Technical Problem to be Solved by the Invention

[0021] Liver injuries can be classified into hepatocellular injury type and cholestasis type based on clinical symptoms. There is no established technique capable of determining those types, although discrimination of the types is important for selection of treatment suitable for each of the types. Therefore, treatment such as medication is currently performed according to a physicians' comprehensive decision based on their clinical experience during common treatment. Hitherto, in the case of determining a liver injury type based on a biochemical parameter, increased ALT and AST have been used as an indicator of hepatocellular injury type, while increased ALP and  $\gamma$ -GTP have been used as an indicator of cholestasis type. However, determination based on only those indicators leads to misjudgment in some cases. Accordingly, a marker capable of more definitely discriminating a liver injury type at the initial stage of the onset of the liver disease enables early determination of a therapeutic strategy.

[0022] An object of the present invention is to find a useful biomarker which enables early diagnosis of a liver injury type, and provide a method of discriminating a liver injury type or a method of assisting discrimination of a liver injury type, in both of which the biomarker is used as an indicator.

##### Solution to Problem

[0023] The inventors of the present invention have made intensive investigations to achieve the object, and found the association of a level of lithocholic acid with a liver injury type by measuring levels of 24 kinds of bile acid components, a hepatic fibrosis marker, and an oxidative stress marker in clinical serum samples and subjecting the data obtained by the measurement to multivariable analysis. Specifically, the level of lithocholic acid showed a tendency to be high in hepatocellular injury-type liver injury in a population of patients with liver injury, and in contrast, the level showed a tendency to be low in cholestasis-type liver injury. In addition, it is found that in hepatocellular injury-type liver injury, a level of ursodeoxycholic acid showed a tendency to be low and a level of type IV collagen, which was a hepatic fibrosis marker, showed a tendency to be high. Meanwhile, in cholestasis-type liver injury, a level of deoxycholic acid showed a tendency to be low, and levels of serum sulfated bile acid (hereinafter abbreviated as SSBA), type IV collagen and hyaluronic acid (hereinafter abbreviated as HA), which were hepatic fibrosis markers, and reactive oxygen species (hereinafter abbreviated as ROS), which was an oxidative stress marker, showed a tendency to be high. Further, levels of taurine conjugates and glycine conjugates of various bile acids showed similar tendencies to levels of the components in a free form. Thus, the inventors of the present invention have revealed that a liver injury type can be discriminated by

using a biomarker including those bile acid components that show characteristic tendencies depending on liver injury types, and achieved the present invention.

**[0024]** That is, the present invention relates to the following.

1. A test method for discriminating a liver injury type, comprising measuring a level of LCA in a biological sample collected from a subject to use the level of LCA as an indicator.

2. The test method according to the above-mentioned item 1., further comprising comparing the measured level of LCA to a predetermined cut-off value of the level of LCA.

3. The test method according to the above-mentioned item 1., further comprising determining the liver injury type as hepatocellular injury type when the measured level of LCA is equal to or higher than a predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury.

4. The test method according to the above-mentioned item 1., further comprising determining the liver injury type as cholestasis type when the measured level of LCA is equal to or lower than a predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury.

5. The test method according to the above-mentioned item 1., further comprising determining the liver injury type as a mixed type of hepatocellular injury type and cholestasis type when the measured level of LCA is higher than a predetermined cut-off value for identifying hepatocellular injury-type liver injury and is lower than a predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury.

6. The test method according to the above-mentioned item 1., further comprising:

**[0025]** measuring any one or both of levels of UDCA and type IV collagen, to use as indicators the any one or both of levels of UDCA and type IV collagen in addition to the level of LCA; and

**[0026]** determining the liver injury type as hepatocellular injury type when the measured level of LCA is equal to or higher than a predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury and the measured level of UDCA and/or type IV collagen satisfies any one of the following:

**[0027]** (1) the level of UDCA is equal to or lower than a predetermined cut-off value of the level of UDCA;

**[0028]** (2) the level of type IV collagen is equal to or higher than a predetermined cut-off value of the level of type IV collagen for identifying hepatocellular injury-type liver injury; and

**[0029]** (3) the level of UDCA is equal to or lower than a predetermined cut-off value of the level of UDCA for identifying hepatocellular injury-type liver injury and the level of type IV collagen is equal to or higher than a predetermined cut-off value of the level of type IV collagen for identifying hepatocellular injury-type liver injury.

7. The test method according to the above-mentioned item 1., further comprising:

**[0030]** measuring any one or more of levels of DCA, SSBA, type IV collagen, HA, and ROS, to use as indicators the levels of DCA, SSBA, type IV collagen, HA, and ROS in addition to the level of LCA; and

**[0031]** determining the liver injury type as cholestasis type when the measured level of LCA is equal to or lower than a predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury and the measured levels of

DCA, SSBA, type IV collagen, HA, and ROS satisfy any one or two or more of the following:

**[0032]** (4) the level of DCA is equal to or lower than a predetermined cut-off value of the level of DCA for identifying cholestasis-type liver injury;

**[0033]** (5) the level of SSBA is equal to or higher than a predetermined cut-off value of the level of SSBA for identifying cholestasis-type liver injury;

**[0034]** (6) the level of type IV collagen is equal to or higher than a predetermined cut-off value of the level of type IV collagen for identifying cholestasis-type liver injury;

**[0035]** (7) the level of HA is equal to or higher than a predetermined cut-off value of the level of HA for identifying cholestasis-type liver injury; and

**[0036]** (8) the level of ROS is equal to or higher than a predetermined cut-off value of the level of ROS for identifying cholestasis-type liver injury.

8. The test method according to the above-mentioned item 1., further comprising:

**[0037]** measuring a level of type IV collagen, to use as an indicator the level of type IV collagen in addition to the level of LCA; and

**[0038]** determining the liver injury type as a mixed type of hepatocellular injury type and cholestasis type when the measured level of LCA is higher than a predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury and is lower than a predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury and when the measured level of type IV collagen is equal to or higher than a predetermined cut-off value of the level of type IV collagen for identifying cholestasis-type liver injury.

9. The test method according to the above-mentioned item 2., in which the predetermined cut-off value of the level of LCA is calculated from a receiver operating characteristic curve, which is hereinafter referred to as ROC curve, of the level of LCA.

10. The test method according to the above-mentioned item 3., 5., 6., or 8., in which the predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury is calculated from a predetermined ROC curve of the level of LCA for hepatocellular injury-type liver injury.

11. The test method according to the above-mentioned item 4., 5., 6., or 8., in which the predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury is calculated from a predetermined ROC curve of the level of LCA for cholestasis-type liver injury.

12. The test method according to the above-mentioned item 6., in which the predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury, the predetermined cut-off value of the level of UDCA for identifying hepatocellular injury-type liver injury, and the predetermined cut-off value of the level of type IV collagen for identifying hepatocellular injury-type liver injury are calculated from a predetermined ROC of the level of LCA for hepatocellular injury-type liver injury, a predetermined ROC of the level of UDCA for hepatocellular injury-type liver injury, and a predetermined ROC of the level of type IV collagen for hepatocellular injury-type liver disease, respectively.

13. The test method according to the above-mentioned item 7., in which the predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury, the predetermined cut-off value of the level of DCA for identifying

cholestasis-type liver injury, the predetermined cut-off value of the level of SSBA for identifying cholestasis-type liver injury, the predetermined cut-off value of the level of type IV collagen for identifying cholestasis-type liver injury, the predetermined cut-off value of the level of HA for identifying cholestasis-type liver injury, and the predetermined cut-off value of the level of ROS for identifying cholestasis-type liver injury are calculated from a predetermined ROC of the level of LCA for cholestasis-type liver injury, a predetermined ROC of the level of DCA for cholestasis-type liver injury, a predetermined ROC of the level of SSBA for cholestasis-type liver injury, a predetermined ROC of the level of type IV collagen for cholestasis-type liver injury, a predetermined ROC of the level of HA for cholestasis-type liver injury, and a predetermined ROC of the level of ROS for cholestasis-type liver injury, respectively.

14. The test method according to any one of the above-mentioned items 1. to 13., in which the biological sample is a blood sample.

15. The test method according to any one of the above-mentioned items 1. to 14., in which the biological sample is a serum sample.

16. A selection method of a therapeutic agent for liver injury according to a liver injury type, wherein the method is applied for a subject whose liver injury type is determined by the test method of any one of the above-mentioned items 1 to 15.

#### Advantageous Effects of Invention

**[0039]** According to the present invention, the test method comprising measuring a level of LCA in a biological sample collected from a subject and discriminating a liver injury type using the measured level of LCA as an indicator can be provided.

**[0040]** The test method according to the present invention is used alone or in combination with measurement of biochemical parameters which has hitherto been performed, and thereby enables early diagnosis of liver injury types based on clinical symptoms, specifically, hepatocellular injury type, cholestasis type, and a mixed type thereof, and early determination of a therapeutic strategy. The test method according to the present invention can be used in an examination for liver disease and is extremely useful as a method of assisting diagnosis and treatment of liver disease.

#### BRIEF DESCRIPTION OF DRAWINGS

**[0041]** FIG. 1 is a view for illustrating a metabolic pathway of bile acids.

**[0042]** FIG. 2 is a view for illustrating a method of discriminating a liver injury type using a cut-off value of a level of LCA.

**[0043]** FIG. 3 is a schematic view for illustrating that in hepatocellular injury-type liver injury, the level of LCA showed a tendency to increase significantly and a level of UDCA showed a tendency to decrease significantly in bile acid components. Levels of CA and CDCA, which had an isomeric relationship with UDCA, tended to decrease with no significant difference and a level of DCA tended to increase with no significant difference.

**[0044]** FIG. 4 is a schematic view for illustrating that in cholestasis-type liver injury, both of the levels of LCA and DCA showed a tendency to increase significantly in bile acid components. All of the levels of CDCA, UDCA, and CA showed a tendency to decrease with no significant difference.

#### DESCRIPTION OF EMBODIMENTS

**[0045]** Abbreviations of bile acid components used in this description are shown in Table 1. In this description, the names of bile acid components are sometimes described as abbreviations shown in Table 1.

TABLE 1

Bile acid components	Abbreviations
Cholic acid	CA
Glycocholic acid	GCA
Taurocholic acid	TCA
Deoxycholic acid	DCA
Glycodeoxycholic acid	GDCA
Taurodeoxycholic acid	TDCA
Chenodeoxycholic acid	CDCA
Glycochenodeoxycholic acid	GCDCA
Taurochenodeoxycholic acid	TCDCa
Ursodeoxycholic acid	UDCA
Glycoursodeoxycholic acid	GUDCA
Tauroursodeoxycholic acid	TUDCA
Lithocholic acid	LCA
Glycolithocholic acid	GLCA
Taurolithocholic acid	TLCA
12-Ketolithocholic acid	12_KLCA
$\alpha$ -Muricholic acid	$\alpha$ MCA
$\beta$ -Muricholic acid	$\beta$ MCA
Tauro- $\beta$ -muricholic acid	T $\beta$ MCA
Hyochoolic acid	HCA
Glychoyochoolic acid	GHCA
Taurohyochoolic acid	THCA
Hyodeoxycholic acid	HDCA

**[0046]** The present invention relates to a test method for liver injury, and more particularly, to a test method for discriminating a liver injury type. The test method according to the present invention comprises measuring a level of LCA in a biological sample collected from a subject, and discriminating a liver injury type using the measured level of LCA as an indicator. The discrimination of a liver injury type using the level of LCA as an indicator may be performed by comparing the measured level of LCA to a predetermined cut-off value of the level of LCA. An example of the test method according to the present invention may be a test method for discriminating a liver injury type, comprising measuring a level of LCA in a biological sample collected from a subject, and further comprising comparing the measured level of LCA to a predetermined cut-off value of the level of LCA.

**[0047]** In the test method according to the present invention, a level of a different biomarker from LCA, the biomarker being associated with a liver injury type, may be measured in addition to the level of LCA, and the liver injury type can be discriminated using the level of LCA and the level of the biomarker as indicators.

**[0048]** Examples of the different biomarker from LCA, the biomarker being associated with a liver injury type, may include UDCA and type IV collagen, which are associated with hepatocellular injury-type liver injury. In addition, as a biomarker associated with cholestasis-type liver injury, there may be given, for example, DCA, SSBA, type IV collagen, HA, and ROS.

**[0049]** The test method according to the present invention may be performed alone or in combination with various test methods, such as measurement of biochemical parameters, which have hitherto been performed.

**[0050]** The "liver injury type" as used herein means a type to be classified based on liver injury symptoms. The "liver

injury symptoms” refer to various complaints and examination findings which are exhibited by a patient with liver injury. Specific examples of the “liver injury type” may include hepatocellular injury type and cholestasis type. In the hepatocellular injury type, a hepatic parenchymal cell, which is one of the cells constituting liver and accounts for most major parts of liver, is damaged, and a remarkable reduction in liver function is caused. The cholestasis type is caused by blockade of excretion of bile from liver due to some cause, with the result that abnormal excretion of bile is brought about over a part or whole of intrahepatic or extrahepatic bile duct system, and thereby the bile accumulates in liver and blood. Then, various symptoms such as jaundice and hepatitis are induced.

**[0051]** The test method according to the present invention can discriminate hepatocellular injury type and cholestasis type, and can also discriminate a type classified as both of the hepatocellular injury type and the cholestasis type, that is, a mixed type.

**[0052]** The “subject” to be subjected to the test method according to the present invention means an individual who has been diagnosed as having liver dysfunction and/or who is suspected of having liver dysfunction based on some testing results. The “subject” encompasses an individual who has a liver disease classified as viral hepatitis, drug-induced liver injury, alcoholic liver injury, autoimmune liver injury, metabolic liver injury, or the like or/and who is suspected as having the liver disease. Preferred is an individual who has been diagnosed as having liver dysfunction.

**[0053]** The “biological sample” as used herein is not particularly limited as long as the sample contains bile acid components, and a preferred example thereof may be a sample which may contain a hepatic fibrosis marker and/or an oxidative stress marker in addition to the bile acid components. Specific examples thereof may include blood isolated from a subject, and serum and plasma prepared from the blood, preferably serum.

**[0054]** A “therapeutic agent for liver injury” and a “treatment method of liver injury” as used herein include examples such as therapeutic agents for liver injury and treatment methods of liver injury which are described in “Manual for handling disorders due to adverse drug reactions, Drug-induced liver injury,” and medications and treatment methods which are performed in medical facilities. In the case of the hepatocellular injury type, for example, intravenous injection of stronger Neo-Minophagen C formulation which is a glycyrrhizin formulation and has an anti-allergic action is performed as well as oral administration of UDCA having a hepatocyte membrane protecting action. When hepatocellular injury is increased in severity to induce fulminant hepatitis, for example, intravenous hyperalimentation (IVH) or artificial liver support is used. In the case of the cholestasis type, for example, vitamin A or vitamin K is administered for covering lack of a liposoluble vitamin and then UDCA, prednisolone, phenobarbital, taurine, colestimide, or the like is used as a medication.

**[0055]** LCA is one of the bile acids. The bile acids are produced from cholesterol metabolism in hepatocytes of liver, followed by conjugating with glycine and taurine (aminoethylsulfonic acid) or the like, and then excreted as bile, which play a key role in digestion and absorption of fat. LCA is a secondary bile acid produced in such a way that CDCA, which is one of the primary bile acids synthesized in liver, is excreted via bile duct into intestinal tract and subjected to a dehydration and oxidation reaction by 7- $\alpha$  dehydroxylase.

**[0056]** In a population of patients with liver injury, when levels of LCA in serum were compared between patients with hepatocellular injury-type liver injury and patients with cholestasis-type liver injury, the level of LCA showed a tendency to be high in the patients with hepatocellular injury-type liver injury, and in contrast, the level of LCA showed a tendency to be low in the patients with cholestasis-type liver injury. Further, in the cholestasis-type liver injury, a level of DCA in serum showed a tendency to be low and a level of SSBA showed a tendency to be high. Meanwhile, in the hepatocellular injury-type liver injury, a level of UDCA in serum showed a tendency to be low. Levels of taurine conjugates and glycine conjugates of various bile acids also showed a tendency to be similar to the levels of the components in a free form. It can be considered that: in the cholestasis-type liver injury, production of DCA and LCA is suppressed by cholestasis, which is accompanied by an increase in the levels of CA, CDCA, and UDCA; and in contrast, in the hepatocellular injury-type liver injury, production of DCA and LCA, which have high lipophilicity, is enhanced, resulting in suppression of production of CA, CDCA, and UDCA.

**[0057]** In the cholestasis-type liver injury, in addition to the level of LCA, levels of type IV collagen and HA, which are hepatic fibrosis markers, and ROS, which is an oxidative stress marker, in serum also showed a tendency to be high. Meanwhile, in the hepatocellular injury-type liver injury, the level of type IV collagen in serum showed a tendency to be high.

**[0058]** The discrimination of a liver injury type may be performed by comparing values of measurement items to their predetermined cut-off values. The “cut-off value” means a value which distinguishes between a positive range and a negative range. The cut-off values of measurement items may be individually set for liver injury types. The cut-off value may be set according to a method well known. For example, the cut-off value may be set using ROC analysis, which is generally used as a method of investigating usability of a diagnostic examination. In the ROC analysis, when shifting thresholds, an ROC curve is prepared which plots sensitivity for each threshold on the vertical axis and a false positive fraction (FPF, false-positive rate: 1-specificity) on the horizontal axis. In the case of the examination with no diagnostic capability, the ROC curve forms a diagonal straight line. Meanwhile, along with an improvement of a diagnostic capability of the examination, the diagonal line forms a curve which has an arc left-upward. In the case of the examination with a diagnosis capability of 100%, the diagonal line forms a curve which passes from a left side to an upper side. For the setting of the cut-off value, a method based on the fact that an ROC curve of independent variables having excellent sensitivity and specificity comes to near the upper left corner, is given as an example, in which a point having the minimum distance from the upper left corner is set as a cut-off value. Alternatively, a method may be given in which a point farthest from the diagonal dotted line at an area under the curve (abbreviated as AUC) of 0.500 in an ROC curve is set as a cut-off value, that is, (sensitivity+specificity-1) is calculated and a maximum point among the obtained values, i.e., Youden index may be set as the cut-off value. Herein, the “sensitivity” means a true positive rate. In addition, the “specificity” means a true negative rate. Besides, in an alternative method, the cut-off value may be quantitatively set based on the relationship between each of the measurement items and the frequency of a liver injury type.

**[0059]** The discrimination of a liver injury type by the test method according to the present invention is described with referring to FIG. 2. When the level of LCA is equal to or higher than a predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury, the liver injury type is determined as hepatocellular injury type. When the level of LCA is equal to or lower than a predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury, the liver injury type is determined as cholestasis type. In addition, when the level of LCA is higher than a predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury and is lower than a predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury, the liver injury type is determined as a type containing both of hepatocellular injury-type and cholestasis type, that is, a mixed type. Further, for more detailed discrimination, when the level of LCA is higher than a predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury, the liver injury type is determined as hepatocellular injury type in which the mixed type is not included. In addition, when the level of LCA is lower than a predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury, the liver injury type is determined as cholestasis type in which the mixed type is not included.

**[0060]** In the test method according to the present invention, any one or both of levels of UDCA and type IV collagen may be measured in addition to the level of LCA. The measured level of UDCA and/or type IV collagen is compared with the corresponding predetermined cut-off value. When the measured level of LCA is equal to or higher than a predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury and the measured level of UDCA and/or type IV collagen satisfies any one of the following, the liver injury type can be determined as hepatocellular injury type: (1) the level of UDCA is equal to or lower than a predetermined cut-off value of the level of UDCA for identifying hepatocellular injury-type liver injury; (2) the level of type IV collagen is equal to or higher than a predetermined cut-off value of the level of type IV collagen for identifying hepatocellular injury-type liver injury; and (3) the level of UDCA is equal to or lower than a predetermined cut-off value of the level of UDCA for identifying hepatocellular injury-type liver injury and the level of type IV collagen is equal to or higher than a predetermined cut-off value of the level of type IV collagen for identifying hepatocellular injury-type liver injury.

**[0061]** In the test method according to the present invention, any one or more of levels of DCA, SSBA, type IV collagen, HA, and ROS may be measured in addition to the level of LCA. Each of the measured levels is compared with the corresponding predetermined cut-off value. When the measured level of LCA is equal to or lower than a predetermined cut-off value of the level of LCA and the measured levels of DCA, SSBA, type IV collagen, HA, and ROS satisfy any one or two or more of the following, the liver injury type can be determined as cholestasis type: (4) the level of DCA is equal to or lower than a predetermined cut-off value of the level of DCA; (5) the level of SSBA is equal to or higher than a predetermined cut-off value of the level of SSBA; (6) the level of type IV collagen is equal to or higher than a predetermined cut-off value of the level of type IV collagen; (7) the level of HA is equal to or higher than a predetermined cut-off

value of the level of HA; and (8) the level of ROS is equal to or higher than a predetermined cut-off value of the level of ROS.

**[0062]** In the test method according to the present invention, a level of type IV collagen may be measured in addition to the level of LCA. The measured level is compared with the corresponding predetermined cut-off value. When the measured level of LCA is higher than a predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury and is lower than a predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury and when the measured level of type IV collagen is equal to or higher than a predetermined cut-off value of the level of type IV collagen, the liver injury type can be determined as a mixed type of hepatocellular injury type and cholestasis type.

**[0063]** Specific examples of the cut-off values of the measurement items used in the test method according to the present invention are shown in Table 2-1 and Table 2-2. Table 2-1 shows specific examples of cut-off values of biomarkers for identifying hepatocellular injury-type liver injury. Table 2-2 shows specific examples of cut-off values of biomarkers for identifying cholestasis-type liver injury. Each of the cut-off values shown as examples is a cut-off value predetermined by measuring levels of biomarkers using as a biological sample a serum sample collected from a liver injury patient with no application of any therapeutic agent for liver injury, comparing the measured levels of biomarkers between a liver injury patient group and patients with hepatocellular injury-type liver injury or patients with cholestasis-type liver injury by ROC analysis, and determining a Youden index in Examples described below. The cut-off value may also be set for each measurement facility or population to be measured. The setting may be appropriately changed by storing measured data on a level of a biomarker of interest for the same samples as described above, and performing the analysis according to the same analytical approach as described above. In addition, when performing measurement of a level of a biomarker using a different sample from the serum sample, the cut-off value may be appropriately set according to the same analytical approach as described above by analyzing the measured data on each of levels of biomarkers from the sample to be used.

TABLE 2-1

		Cut-off values
Bile acid components	LCA (nmol/mL)	≥0.01985
	UDCA (nmol/mL)	≤0.903
Hepatic fibrosis marker	Type IV collagen (ng/ml)	≥128

TABLE 2-2

		Cut-off values
Bile acid components	LCA (nmol/mL)	≤0.0241
	DCA (nmol/mL)	≤0.175
	SSBA (nmol/mL)	≥21.1
Oxidative stress marker	ROS (U)	≥216
Hepatic fibrosis markers	Type IV collagen (ng/ml)	≥288
	HA (ng/ml)	≥47

**[0064]** A description is given of a specific example of the case where the test method according to the present invention is performed using specific cut-off values exemplified in Table 2-1 and Table 2-2 above.

**[0065]** A specific example of the test method according to the present invention may be a test method for discriminating a liver injury type, the test method including the steps of:

**[0066]** measuring a level of LCA in a serum sample collected from a subject;

**[0067]** comparing the measured level of LCA to a predetermined cut-off value of the level of LCA; and

**[0068]** determining the liver injury type as hepatocellular injury type when the measured level of LCA is equal to or higher than a predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury,

**[0069]** in which the cut-off value is 0.0195 nmol/L.

**[0070]** In addition, another specific example of the test method may be a test method for discriminating a liver injury type, the test method including the steps of:

**[0071]** measuring a level of LCA in a serum sample collected from a subject;

**[0072]** comparing the measured level of LCA to a predetermined cut-off value of the level of LCA; and

**[0073]** determining the liver injury type as cholestasis type when the measured level of LCA is equal to or lower than a predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury,

**[0074]** in which the cut-off value is 0.0241 nmol/L.

**[0075]** Still another specific example of the test method may be a test method for discriminating a liver injury type, the test method including the steps of:

**[0076]** measuring a level of LCA in a serum sample collected from a subject;

**[0077]** comparing the measured level of LCA to a predetermined cut-off value of the level of LCA; and

**[0078]** determining the liver injury type as a mixed type of hepatocellular injury type and cholestasis type when the measured level of LCA is higher than a predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury and is lower than a predetermined cut-off value for identifying cholestasis-type liver injury,

**[0079]** in which the predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury is 0.0195 nmol/L and the predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury is 0.0241 nmol/L.

**[0080]** In addition, another specific example of the test method may be a test method for discriminating a liver injury type, the test method including the steps of:

**[0081]** measuring a level of LCA in a serum sample collected from a subject;

**[0082]** comparing the measured level of LCA to a predetermined cut-off value of the level of LCA;

**[0083]** further measuring any one or both of levels of UDCA and type IV collagen;

**[0084]** comparing the measured level of UDCA and/or type IV collagen to the predetermined cut-off value of the level of LCA; and

**[0085]** determining the liver injury type as hepatocellular injury type when the measured level of LCA is equal to or higher than a predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury and the measured level of UDCA and/or type IV collagen satisfies any one of the following:

**[0086]** (1) the level of UDCA is equal to or lower than a predetermined cut-off value of the level of UDCA;

**[0087]** (2) the level of type IV collagen is equal to or higher than a predetermined cut-off value of the level of type IV collagen for identifying hepatocellular injury-type liver injury;

**[0088]** (3) the level of UDCA is equal to or lower than a predetermined cut-off value of the level of UDCA for identifying hepatocellular injury-type liver injury and the level of type IV collagen is equal to or higher than the predetermined cut-off value of the level of type IV collagen for identifying hepatocellular injury-type liver injury,

**[0089]** in which the predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury is 0.0195 nmol/L, the predetermined cut-off value of the level of UDCA for identifying hepatocellular injury-type liver injury is 0.903 nmol/L, and the predetermined cut-off value of the level of type IV collagen for identifying hepatocellular injury-type liver injury is 128 ng/ml.

**[0090]** In addition, another specific example of the test method may be a test method for discriminating a liver injury type, the test method including the steps of:

**[0091]** measuring a level of LCA in a serum sample collected from a subject;

**[0092]** comparing the measured level of LCA to a predetermined cut-off value of the level of LCA;

**[0093]** further measuring any one or more of levels of DCA, SSBA, type IV collagen, HA, and ROS,

**[0094]** comparing each of the measured levels obtained by the further measurement to the corresponding predetermined cut-off value; and

**[0095]** determining the liver injury type as cholestasis type when the measured level of LCA is equal to or lower than a predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury and the measured levels of DCA, SSBA, type IV collagen, HA, and ROS satisfy any one or two or more of the following:

**[0096]** (4) the level of DCA is equal to or lower than a predetermined cut-off value of the level of DCA for identifying cholestasis-type liver injury;

**[0097]** (5) the level of SSBA is equal to or higher than a predetermined cut-off value of the level of SSBA for identifying cholestasis-type liver injury;

**[0098]** (6) the level of type IV collagen is equal to or higher than a predetermined cut-off value of the level of type IV collagen for identifying cholestasis-type liver injury;

**[0099]** (7) the level of HA is equal to or higher than a predetermined cut-off value of the level of HA for identifying cholestasis-type liver injury; and

**[0100]** (8) the level of ROS is equal to or higher than a predetermined cut-off value of the level of ROS for identifying cholestasis-type liver injury,

**[0101]** in which: the predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury is 0.0241 nmol/L; the predetermined cut-off value of the level of DCA for identifying cholestasis-type liver injury is 0.175 nmol/L; the predetermined cut-off value of the level of SSBA for identifying cholestasis-type liver injury is 21.1 nmol/L; the predetermined cut-off value of the level of type IV collagen for identifying cholestasis-type liver injury is 288 ng/ml; the predetermined cut-off value of the level of HA for identifying cholestasis-type liver injury is 47 ng/ml; and the predetermined cut-off value of the level of ROS for identifying cholestasis-type liver injury is 216 U.

**[0102]** In addition, another specific example of the test method may be a test method for discriminating a liver injury type, the test method including the steps of:

**[0103]** measuring a level of LCA in a serum sample collected from a subject;

**[0104]** comparing the measured level of LCA to a predetermined cut-off value of the level of LCA;

**[0105]** further measuring a level of type IV collagen in the serum sample;

**[0106]** comparing the measured level of type IV collagen to a predetermined cut-off value of the level of type IV collagen; and

**[0107]** determining the liver injury type as a mixed type of hepatocellular injury type and cholestasis type when the measured level of LCA is higher than a predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury, and is lower than a predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury and when the measured level of type IV collagen is equal to or higher than a predetermined cut-off value of the level of type IV collagen for identifying cholestasis-type liver injury,

**[0108]** in which: the predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury and the predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury are 0.0195 nmol/L and 0.0241 nmol/L, respectively; and the predetermined cut-off value of the level of type IV collagen for identifying cholestasis-type liver injury is 288 ng/ml.

**[0109]** The measurement of levels of bile acid components may be performed by a method which has hitherto been used for the measurement. Examples thereof may include liquid chromatography-tandem mass spectrometry (LC-MS/MS) and gas chromatography-mass spectrometry (GC/MS). In addition, levels of bile acid or sulfated bile acid may be measured with a commercially available bile acid measurement kit. For example, the measurement of the level of sulfated bile acid may be performed with a UBASTEC-AUTO bile acid (USBA) kit (LPS Co., Ltd.). The measurement methods are not limited thereto and any method may be used for the measurement as long as the method can measure bile acids and components thereof.

**[0110]** The measurement of the level of type IV collagen may be performed by a method which has hitherto been used for the measurement. For example, the measurement of the level of type IV collagen may be performed with a commercially available type IV collagen measurement reagent kit. Examples of such method may include enzyme immunoassay (EIA) and enzyme-linked immunosorbent assay (ELISA) in which an antibody to type IV collagen is used. Specifically, a level of type IV collagen.7S may be measured as described below. Anti-human type IV collagen rabbit polyclonal antibody is added to a biological sample to produce a complex (complex 1) of type IV collagen.7S and anti-human type IV collagen rabbit polyclonal antibody, followed by adding with human type IV collagen.7S (labeled antigen) labeled with a radioisotope, such as iodine-125 ( $^{125}\text{I}$ ), to form a complex (complex 2) with anti-human type IV collagen.7S rabbit polyclonal antibody which has failed to bind with type IV collagen.7S in the sample. Subsequently, anti-rabbit  $\gamma$ -globulin goat serum (goat antibody) is added to the complex 2, followed by anti-rabbit  $\gamma$ -globulin goat serum (goat antibody), resulting in a reaction of the goat antibody with the complex 2 to form a complex of labeled antigen, anti-human

type IV collagen.7S, and rabbit polyclonal antibody-goat antibody. The resultant precipitate is collected and its radioactivity is measured to determine the level of type IV collagen.7S.

**[0111]** The measurement of the level of HA may be performed by a method which has hitherto been used for the measurement. For example, the level of HA may be measured with a commercially available HA measurement reagent kit. An example of such method may be enzyme-linked immunosorbent assay (ELISA) in which an antibody to HA is used.

**[0112]** The measurement of the level of ROS may be performed by a method which has hitherto been used for the measurement. For example, the level of ROS may be measured with a commercially available ROS measurement reagent kit. Specifically, a total amount of substances producing free radicals may be measured as described below. Free radicals (ROO. or RO.) are generated from a peroxide (ROOH) by the Fenton reaction in which an ion of a metal, such as iron, is used as a catalyst, and the generated free radicals are allowed to react with N,N-diethyl-1,4-phenylene-diamine sulfate (abbreviated as DEPPD). Then, the DEPPD.+ generated by the reaction is subjected to measurement of absorbance at a wavelength of 505 nm.

**[0113]** The present invention encompasses a selection method of a therapeutic agent for liver injury according to a liver injury type, which is applied for a subject whose liver injury type is determined by the test method according to the present invention.

**[0114]** The present invention also encompasses a treatment method of liver injury according to a liver injury type, which is applied for a subject whose liver injury type is determined by the test method according to the present invention.

**[0115]** Examples of the therapeutic agent for liver injury and the treatment method of liver injury according to a liver injury type include therapeutic agents for liver injury and treatment method of liver injury which are described in "Manual for handling disorders due to adverse drug reactions, Drug-induced liver injury," and medications and treatment methods which are performed in medical facilities. In the case of the hepatocellular injury type, for example, intravenous injection of stronger Neo-Minophagen C formulation which is a glycyrrhizin formulation and has an anti-allergic action is performed as well as oral administration of UDCA. When hepatocellular injury is increased in severity to induce fulminant hepatitis, for example, intravenous hyperalimentation (IVH) or artificial liver support is used. In the case of the cholestasis type, for example, vitamin A or vitamin K is administered for covering lack of a liposoluble vitamin and then UDCA, prednisolone, phenobarbital, taurine, colestimide, or the like is used as a medication.

**[0116]** Now, the present invention is specifically described byway of Examples. However, the present invention is not construed as being limited to these Examples.

#### Example 1

**[0117]** In order to find a useful biomarker which enables early diagnosis of a liver injury type, serum samples obtained from liver injury patients and healthy volunteers were used to measure levels of bile acid components, and an oxidative stress marker and a hepatic fibrosis marker. The collection of serum samples and a serial experiment were performed after obtaining the consents of the sample donors, i.e. the patients and the healthy volunteers.

**[0118]** 1. Subject Sample

**[0119]** Under the approval by the ethics committees of Daiichi Sankyo Co., Ltd. and the Jikei University School of Medicine, 304 serum samples were collected from 150 patients who were suspected of having liver injuries for about 2 years, such as viral, autoimmune, alcoholic, and genetic liver injuries. In addition, serum samples of healthy individuals obtained by blood collection from volunteers were used. The serum samples were stored at  $-80^{\circ}$  C. until biomarkers were measured.

**[0120]** 2. Evaluation Items

**[0121]** Levels of bile acid components, specifically, 24 kinds of bile acid components, i.e., CA, DCA, CDCA, UDCA, LCA, 12\_KLCA, aMCA, PICA, HDCA, GCA, GDCA, GCDCA, GUDCA, GLCA, GHCA, GHCA, TCA, TDCA, TCDCA, TUDCA, TLCA, T $\beta$ MCA, THCA, and THDCA, and serum sulfated bile acid (SSBA) were measured.

**[0122]** In addition, levels of ALT, AST, ALP,  $\gamma$ -GTP, total bilirubin (hereinafter abbreviated as T.BIL), direct bilirubin (hereinafter abbreviated as D.BIL), TBA, and albumin (hereinafter abbreviated as ALB), and prothrombin time (hereinafter abbreviated as PT), which were biochemical parameters, were measured. Further, levels of 8-OHdG and ROS, which were oxidative stress markers, were measured as well as type IV collagen and HA, which were hepatic fibrosis markers.

**[0123]** 3. Measurement Method

**[0124]** A total of 24 kinds of bile acid components were measured by a LC-MS/MS method. The minimum limit of determination was 0.03 nmol/ml only for 12-KLCA and 0.01 nmol/ml for other substances to be measured. The maximum limit of determination is 10 nmol/mL only for 12-KLCA and CA and 30 nmol/mL for other substances to be measured. Naphtalam was used as an internal standard (IS). ACQUITY UPLC BEH C18 of 1.7 m and 2.1 $\times$ 50 mm (manufactured by Waters Corporation) was used for a column. 0.1% formic acid and acetonitrile were used for a mobile phase, the ratio of which was changed according to the measurement time. Electrospray ionization (abbreviated as ESI) negative was selected for ionization and selected ion recording (abbreviated as SIR) was selected for a detection method. Deproteinization was used as pretreatment. A level of sulfated bile acid was measured with BioMajesty JCA-BM2250 (JEOL Ltd.) using 300  $\mu$ L of serum and a UBASTEC-AUTO bile acid (USBA) kit (LPS Co., Ltd.).

**[0125]** 8-OHdG and ROS, which were oxidative stress markers, were measured as described below. 8-OHdG was measured with a reagent kit commercially available from Japan Institute for the Control of Aging. For measurement of ROS, free radicals (ROO $\cdot$  or RO $\cdot$ ) are generated from a peroxide (ROOH) by the Fenton reaction in which an ion of a metal, such as iron, is used as a catalyst, and the generated free radicals were allowed to react with N,N-diethyl-1,4-phenylene-diamine sulfate (abbreviated as DEPPD). The DEPPD.+ generated by the reaction have an absorption at a wavelength of 505 nm. Then, the measurement of the absorbance was performed to determine a total amount of substances producing free radicals.

**[0126]** The biochemical parameters were measured by methods which were generally used in laboratory examinations.

**[0127]** Type IV collagen.7S and HA, which were hepatic fibrosis markers, were measured as described below. Type IV

collagen.7S was allowed to react with anti-human type IV collagen rabbit polyclonal antibody to produce a type IV collagen.7S-anti-human type IV collagen rabbit polyclonal antibody complex (complex 1). Then, Iodine-125 ( $^{125}$ I)-labeled human type IV collagen.7S (labeled antigen) was added to react with anti-human type IV collagen.7S rabbit polyclonal antibody which had failed to bind with type IV collagen.7S in the sample, resulting in forming a labeled antigen-anti-human type IV collagen.7S rabbit polyclonal antibody complex (complex 2). Subsequently, anti-rabbit  $\gamma$ -globulin goat serum (goat antibody) was added to allow a reaction of the goat antibody with the complex 2, resulting in forming a complex of the labeled antigen, the anti-human type IV collagen.7S, and the rabbit polyclonal antibody-goat antibody as a precipitate. After an unreacted labeled antigen was removed, the radioactivity of the precipitate was measured and a level of type IV collagen.7S was determined. HA was measured with an HA ELISA kit (manufactured by Cosmo Bio Co., Ltd.).

**[0128]** 4. Data Analysis Method

**[0129]** The correlation of variations in various biomarkers and liver injury symptoms comprehensively determined was analyzed. 304 serum samples obtained from 150 patients with liver injury were used and the summary statistics of the background information were calculated. The analysis was performed without taking the correlation between samples in the same case into account.

**[0130]** In order to investigate biomarkers associated with identification of types of liver injury based on symptoms, a multivariable logistic regression model adjusted by patient background factors was used to calculate the odds ratio for liver injury symptoms comprehensively determined by biomarker candidate factors (hepatocellular injury or cholestasis) with the 95% confidence interval. As the biomarker candidate factors, levels of bile acid components, oxidative stress markers, and hepatic fibrosis markers were used. Sex, age, EMI, alcohol drinking, therapeutic drugs for liver injury (antivirus agent, liver protection agent, bile acid formulation, amino acid formulation/hypoalbuminemia improving drug, and vitamin K formulation), and complications (dyslipidemia, diabetes, obstructive jaundice, and gallstone) were used as the patient background factors.

**[0131]** The presence or absence of liver injury symptoms was determined based on the assessment results given by two or more investigators adopted from the assessment results given by independent three investigators. Samples were used as a total analysis unit and the analysis was performed without taking the correlation between samples in the same case into account. A two-sided significance level of 5% was adopted when a statistical test was performed.

**[0132]** In the analysis of biomarkers, for indicators with no reference values, 97.5 percentiles of measured values of healthy individuals (corresponding to mean+2SD in the case of normal distribution) were set as reference values, and values higher than the reference values were categorized into high values and values lower than the reference values were categorized into low values.

**[0133]** Then, in order to determine objective assessment scales of biomarkers found to be associated with liver injury symptoms, serum samples of patients with no application of a therapeutic agent for liver injury were used to calculate cut-off values of the biomarkers which identify the presence or absence of liver injury symptoms. Though the calculation of a cut-off value may be performed by various procedures, the cut-off value was set to a value that derived through ROC analysis to maximize the sum of sensitivity and specificity.

**[0134]** 5. Results

**[0135]** Background information on patients who have provided their serum samples is shown in Table 3.

TABLE 3

		Liver injury patients	
Number of assessed samples		304	
Sex		Male	126 (41.4)
		Female	178 (58.6)
Age (years)		Mean (standard deviation)	61.0 (15.2)
		<65	160 (52.6)
		>=65	144 (47.4)
BMI (kg/m <sup>2</sup> )		Mean (standard deviation)	23.6 (3.9)
		<25	198 (65.1)
		>=25	95 (31.3)
		Impossible to calculate	11 (3.6)
Alcohol drinking		No	247 (81.3)
		Yes	50 (16.4)
		Not known	7 (2.3)
Therapeutic drugs for liver injury		Yes	24 (7.9)
Drugs usually (hitherto) taken		Yes	44 (14.5)
		Yes	166 (54.6)
		Yes	69 (22.7)
		Yes	5 (1.6)
		Yes	51 (16.8)
		Yes	35 (11.5)
		Yes	3 (1.0)
		Yes	3 (1.0)
Other combined drugs		Yes	21 (6.9)
Drugs usually (hitherto) taken		Yes	10 (3.3)
		Yes	3 (1.0)
		Yes	31 (10.2)
		Yes	109 (35.9)
		Yes	33 (10.9)
Complications		Yes	57 (18.8)
		Yes	61 (20.1)
		Yes	12 (3.9)
		Yes	58 (19.1)
		Yes	36 (11.8)
Biochemical parameters		Median (interquartile range)	56.0 (30.5, 92.5)
		Median (interquartile range)	55.0 (37.5, 99.5)
		Median (interquartile range)	290.0 (231.0, 420.5)
		Median (interquartile range)	67.0 (32.0, 156.0)
		Median (interquartile range)	1.1 (0.7, 1.7)
		Median (interquartile range)	0.1 (0.1, 0.3)
		Median (interquartile range)	18.0 (7.7, 42.3)
		Median (interquartile range)	3.7 (3.2, 4.1)
		Median (interquartile range)	84.0 (73.0, 96.0)
Levels of bile acid components		Median (interquartile range)	0.0324 (0.0154, 0.0747)
		Median (interquartile range)	0.0393 (0.0199, 0.0832)
		Median (interquartile range)	0.6915 (0.0721, 4.9150)
		Median (interquartile range)	0.0000 (0.0000, 0.0055)
		Median (interquartile range)	0.4340 (0.1070, 1.1700)
		Median (interquartile range)	0.2090 (0.0220, 0.7135)
		Median (interquartile range)	0.0000 (0.0000, 0.0030)
		Median (interquartile range)	0.0000 (0.0000, 0.0040)
		Median (interquartile range)	0.1020 (0.0404, 0.3495)
		Median (interquartile range)	0.0224 (0.0105, 0.0719)
		Median (interquartile range)	2.9600 (0.2165, 12.2000)
		Median (interquartile range)	0.0000 (0.0000, 0.0022)
		Median (interquartile range)	3.7000 (1.6100, 8.6450)
		Median (interquartile range)	0.4890 (0.0366, 1.2650)
		Median (interquartile range)	0.0442 (0.0213, 0.1125)
		Median (interquartile range)	0.9725 (0.4075, 3.0050)
		Median (interquartile range)	0.0092 (0.0021, 0.0311)
		Median (interquartile range)	0.1520 (0.0165, 1.0800)
		Median (interquartile range)	0.0000 (0.0000, 0.0001)
		Median (interquartile range)	0.7400 (0.2945, 2.9950)
		Median (interquartile range)	0.0867 (0.0205, 0.2635)
		Median (interquartile range)	0.0013 (0.0000, 0.0482)
		Median (interquartile range)	0.0076 (0.0012, 0.0254)
		Median (interquartile range)	0.2435 (0.0648, 1.0300)
		Median (interquartile range)	15.80 (8.78, 28.46)

TABLE 3-continued

		Liver injury patients	
Oxidative stress markers	8-OHdG (ng/mL)	Median (interquartile range)	0.16 (0.12, 0.22)
	ROS (U)	Median (interquartile range)	219.5 (182.5, 258.0)
Oxidative stress markers	Type IV collagen (ng/mL)	Median (interquartile range)	186.5 (136.0, 258.0)
	HA (ng/mL)	Median (interquartile range)	136.0 (51.5, 388.0)

Data are represented by the number of patients (%) or median (interquartile range).

[0136] Multivariable analysis with covariate adjustment was carried out for biomarkers which define symptoms of liver injury identified for types, and the results are shown in Table 4 and Table 5.

TABLE 4

		Hepatocellular injury type				Cholestasis type			
		Number of assessed samples	Number of liver injury patient samples (%)	Odds ratio (95% confidence interval)	P value	Number of liver injury patient samples (%)	Odds ratio (95% confidence interval)	P value	
Levels of bile acid components	LCA	<=97.5 pct	144	90 (62.5)	1.00	—	70 (48.6)	1.00	—
		>97.5 pct	160	125 (78.1)	2.06 (1.16, 3.66)	0.014	32 (20.0)	0.29 (0.16, 0.53)	<0.0001
	UDCA	<=97.5 pct	148	109 (73.6)	1.00	—	47 (31.8)	1.00	—
		>97.5 pct	156	106 (67.9)	0.31 (0.12, 0.77)	0.0112	55 (35.3)	1.69 (0.71, 4.01)	0.235
	CDCA	<=97.5 pct	268	194 (72.4)	1.00	—	85 (31.7)	1.00	—
		>97.5 pct	36	21 (58.3)	0.50 (0.22, 1.13)	0.0950	17 (47.2)	1.98 (0.88, 4.48)	0.0991
	DCA	<=97.5 pct	232	160 (69.0)	1.00	—	93 (40.1)	1.00	—
		>97.5 pct	72	55 (76.4)	1.48 (0.74, 2.94)	0.267	9 (12.5)	0.26 (0.11, 0.58)	0.001
	CA	<=97.5 pct	261	187 (71.6)	1.00	—	85 (32.6)	1.00	—
		>97.5 pct	43	28 (65.1)	0.75 (0.34, 1.65)	0.470	17 (39.5)	1.36 (0.63, 2.93)	0.627
SSBA	<=97.5 pct	201	137 (68.2)	1.00	—	50 (24.9)	1.00	—	
	>97.5 pct	103	78 (75.7)	1.02 (0.49, 2.09)	0.967	52 (50.5)	5.05 (2.45, 10.41)	<0.0001	
Oxidative stress markers	8-OHdG	<=97.5 pct	292	210 (71.9)	1.00	—	96 (32.9)	1.00	—
		>97.5 pct	12	5 (41.7)	0.26 (0.07, 0.96)	0.0434	6 (50.0)	1.40 (0.35, 5.57)	0.637
	ROS	<=97.5 pct	157	111 (70.7)	1.00	—	36 (22.9)	1.00	—
		>97.5 pct	147	104 (70.7)	0.84 (0.47, 1.52)	0.570	66 (44.9)	2.68 (1.49, 4.80)	0.001
Hepatic fibrosas markers	Type IV collagen	<=150	80	47 (58.8)	1.00	—	20 (25.0)	1.00	—
		>150	162	128 (79.0)	2.19 (1.05, 4.56)	0.036	62 (38.3)	3.54 (1.62, 7.74)	0.0015
	HA	<=50	58	38 (65.5)	1.00	—	13 (22.4)	1.00	—
		>50	182	130 (71.4)	1.20 (0.57, 2.56)	0.629	70 (38.5)	3.66 (1.28, 8.47)	0.0024

Adjustment factors: sex, age, BMI, Alcohol drinking, therapeutic drugs for liver injury (antiviral agent, liver protection agent, bile acid formulation, amino acid formulation, hypoalbuminemia improving drug, and vitamin K formulation), and complications (dyslipidemia, diabetes, obstructive jaundice, and gallstone)

TABLE 5

		Hepatocellular injury type				Cholestasis type			
		Number of assessed samples	Number of liver injury patient samples (%)	Odds ratio (95% confidence interval)	P value	Number of liver injury patient samples (%)	Odds ratio (95% confidence interval)	P value	
Levels of bile acid components	12_KLCA	<=97.5 pct	235	165 (70.2)	1.00	—	77 (32.8)	1.00	—
		>97.5 pct	69	50 (72.5)	0.88 (0.45, 1.71)	0.70	25 (36.2)	1.03 (0.53, 1.98)	0.939
	GLCA	<=97.5 pct	182	113 (62.1)	1.00	—	74 (40.7)	1.00	—
		>97.5 pct	122	102 (83.6)	2.17 (1.13, 4.15)	0.0195	28 (23.0)	0.56 (0.30, 1.04)	0.0655
	GUDCA	<=97.5 pct	140	104 (74.3)	1.00	—	45 (32.1)	1.00	—
		>97.5 pct	164	111 (67.7)	0.16 (0.05, 0.47)	0.0009	57 (34.8)	2.08 (0.78, 5.55)	0.143
	GCDCA	<=97.5 pct	168	114 (67.9)	1.00	—	36 (21.4)	1.00	—
		>97.5 pct	136	101 (74.3)	1.29 (0.69, 2.42)	0.430	66 (48.5)	4.25 (2.23, 8.10)	<0.0001
	GDCA	<=97.5 pct	255	169 (66.3)	1.00	—	94 (36.9)	1.00	—
		>97.5 pct	49	46 (93.9)	7.34 (2.11, 25.53)	0.0017	8 (16.3)	0.35 (0.15, 0.84)	0.0182
	GCA	<=97.5 pct	220	154 (70.0)	1.00	—	50 (22.7)	1.00	—
		>97.5 pct	84	61 (72.6)	1.09 (0.54, 2.16)	0.824	52 (61.9)	6.11 (3.06, 12.23)	<0.0001
	TLCA	<=97.5 pct	209	132 (63.2)	1.00	—	68 (32.5)	1.00	—
		>97.5 pct	95	83 (87.4)	4.68 (2.01, 10.88)	0.0003	34 (35.8)	1.15 (0.59, 2.24)	0.678
	TUDCA	<=97.5 pct	152	105 (69.1)	1.00	—	46 (30.3)	1.00	—
		>97.5 pct	152	110 (72.4)	0.56 (0.22, 1.42)	0.221	56 (36.8)	3.20 (1.18, 8.64)	0.0221
TCDCA	<=97.5 pct	175	113 (64.6)	1.00	—	41 (23.4)	1.00	—	
	>97.5 pct	129	102 (79.1)	2.31 (1.17, 4.54)	0.0152	61 (47.3)	3.32 (1.76, 6.29)	0.0002	

TABLE 5-continued

		Hepatocellular injury type				Cholestasis type		
		Number of assessed samples	Number of liver injury patient samples (%)	Odds ratio (95% confidence interval)	P value	Number of liver injury patient samples (%)	Odds ratio (95% confidence interval)	P value
TDCA	<=97.5 pct	256	172 (67.2)	1.00	—	83 (34.4)	1.00	—
	>97.5 pct	48	43 (89.6)	3.60 (1.31, 9.89)	0.0131	14 (29.2)	0.85 (0.39, 1.83)	0.675
TCA	<=97.5 pct	222	154 (69.4)	1.00	—	52 (23.4)	1.00	—
	>97.5 pct	82	61 (74.4)	1.17 (0.57, 2.40)	0.670	50 (61.0)	7.54 (3.65, 15.60)	<0.0001

Adjustment factors: sex, age, BMI, alcohol drinking, therapeutic drugs for liver injury (antiviral agent, liver protection agent, bile acid formulation, amino acid formulation, hypoalbuminemia improving drug, and vitamin K formulation), and complications (dyslipidemia, diabetes, obstructive jaundice, and gallstone)

**[0137]** Factors found to be significantly associated with the hepatocellular injury type are LCA (OR=2.06, P=0.0140), UDCA (OR=0.31, P=0.0112), and type IV collagen (OR=2.19, P=0.0357). The levels of LCA and type IV collagen showed a tendency to be high and the level of UDCA showed a tendency to be low (Table 4). The levels of CDCA (OR=0.50, P=0.0950), which had an isomeric relationship with UDCA, and CA (OR=0.75, P=0.4702), which was also a primary bile acid, showed a tendency to be low, although their association with the hepatocellular injury type was weak (FIG. 3). When taurine conjugates and glycine conjugates of bile acids were taken into consideration, the levels of TCDCA (OR=2.31, P=0.0152), GDCA (OR=7.34, P=0.0017), TDCA (OR=3.60, P=0.0131), GLCA (OR=2.17, P=0.0195), and TLCA (OR=4.68, P=0.0003) showed a tendency to be significantly high and the level of GUDCA (OR=0.16, P=0.0009) showed a tendency to be significantly low.

**[0138]** Meanwhile, factors found to be significantly associated with the cholestasis type are LCA (OR=0.29, P<0.0001), DCA (OR=0.26, P=0.0010), SSBA (OR=5.05, P<0.0001), type IV collagen (OR=3.54, P=0.0015), HA (OR=3.66, P=0.0024), and ROS (OR=2.68, P=0.0010). The levels of LCA and DCA showed a tendency to be low and the levels of other factors than LCA and DCA showed a tendency to be high (Table 4). Meanwhile, it was found that the levels of CDCA (OR=1.98, P=0.0991), UDCA (OR=1.69, P=0.2353), and CA (OR=1.36, P=0.4269) showed a tendency to be high, although their association with the cholestasis type was weak. That is, contrasting results to the hepatocellular injury type were shown (FIG. 4). When taurine conjugates and glycine conjugates of bile acids were taken into consideration, the levels of GCDCA (OR=4.25, P<0.0001), TCDCA (OR=3.32, P=0.0002), GCA (OR=6.11, P<0.0001), TCA (OR=7.54, P<0.0001), and TUDCA (OR=3.20, P=0.0221) showed a tendency to be significantly high and the level of GDCA (OR=0.35, P=0.0182) showed a tendency to be low. It was found that the level of type IV collagen, which was an indicator of hepatic fibrosis, also showed a tendency to be high in the cholestasis type similarly to the hepatocellular injury type.

**[0139]** In the biomarkers found to be associated with liver injury symptoms, cut-off values which can be used as references for determination of a liver injury type are calculated from serum samples of patients with no application of a therapeutic drug for liver injury, and the results are shown in Table 2-1 and Table 2-2. Cut-off values of biomarkers for identifying hepatocellular injury-type liver injury are shown in Table 2-1. Cut-off values of biomarkers for identifying cholestasis-type liver injury are shown in Table 2-2.

**[0140]** When comparing the results of the multivariable analysis between the hepatocellular injury type and the

cholestasis type, it was found that LCA increased in the hepatocellular injury type, and in contrast, decreased in the cholestasis type. From the results, it was suggested that LCA was a useful marker for discrimination of liver injury symptoms. It can be considered that in the hepatocellular injury type, LCA, which was a secondary bile acid, showed a high value. With this increased metabolism, UDCA, which is an isomer of CDCA that is a parent compound of LCA, showed a low value. Meanwhile, it can be considered that in the cholestasis type, CDCA and CA, which were primary bile acids, showed a tendency to increase owing to cholestasis, although their association with the cholestasis type was weak. With this increased level, excretion of bile acids to intestinal tract was suppressed, resulting in decreased levels of DCA and LCA. In addition, in the hepatocellular injury type, TCDCA, GDCA, TDCA, GLCA, and TLCA significantly increased, while GUDCA significantly decreased, which may be considered as below. With a significant increase in LCA and a change in DCA found to show a tendency to increase but without significance, production of conjugates thereof was also enhanced. GUDCA showed a tendency to decrease with a decrease in UDCA, but TCDCA, which was a taurine conjugate of CDCA, show a tendency to increase. From the results, it was suggested that in the hepatocellular injury type, metabolism from a primary bile acid to a secondary bile acid having high lipid-solubility was increased. It was found that in the cholestasis type, GCDCA, TCDCA, GCA, TCA, and TUDCA significantly increased, while GDCA decreased. From the results, it can be considered that both conjugated reactions of CDCA and CA was increased due to cholestasis. It has been reported that SSBA increases in biliary diseases (Non Patent Literature 12). Also in this test, SSBA showed a high value in the cholestasis type, but no significant change in SSBA was found in the hepatocellular injury type.

**[0141]** It was found that the measured hepatic fibrosis markers significantly increased in both the hepatocellular injury type and the cholestasis type. Among the measured oxidative stress markers, 8-OHdG tended to significantly decrease in the hepatocellular injury type, but the value was not largely changed compared to that of a healthy population. The number of samples of the patient population having a level of 8-OHdG higher than the cut-off value was small. Thus, the association of 8-OHdG with the hepatocellular injury type could not be clearly determined. In addition, ROS, which was reactive oxygen, showed a high value in the cholestasis type, but no significant change in ROS was found in the hepatocellular injury type.

**[0142]** Biochemical parameters, which have hitherto been used in examination of liver diseases, have low correlations with the biomarkers measured in this examination, and hence,

it is difficult to accurately discriminate liver injury symptoms with only use of these biochemical parameters.

**[0143]** From those results, it was suggested that the level of LCA was specific for each of the hepatocellular injury type and the cholestasis type and showed significant changes, and hence was useful as a marker for discriminating types of liver injury symptoms. In addition, primary bile acids and SSBA are also effective for determination of hepatocellular injury type, cholestasis type, and a mixed type thereof as markers which may assist discrimination of types of liver injury symptoms using the level of LCA.

#### INDUSTRIAL APPLICABILITY

**[0144]** As described above, the present invention provides a test method for discriminating a liver injury type. The present invention enables early diagnosis and early determination of a therapeutic strategy of a liver injury type based on clinical symptoms, specifically, hepatocellular injury type, cholestasis type, and a mixed type thereof. As described above, the present invention is extremely useful in the field of examination of liver diseases.

1. A test method for discriminating a liver injury type, comprising measuring a level of lithocholic acid, which is hereinafter abbreviated as LCA, in a biological sample collected from a subject to use the level of LCA as an indicator.

2. The test method according to claim 1, further comprising comparing the measured level of LCA to a predetermined cut-off value of the level of LCA.

3. The test method according to claim 1, further comprising determining the liver injury type as hepatocellular injury type when the measured level of LCA is equal to or higher than a predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury.

4. The test method according to claim 1, further comprising determining the liver injury type as cholestasis type when the measured level of LCA is equal to or lower than a predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury.

5. The test method according to claim 1, further comprising determining the liver injury type as a mixed type of hepatocellular injury type and cholestasis type when the measured level of LCA is higher than a predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury and is lower than a predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury.

6. The test method according to claim 1, further comprising:

measuring any one or both of levels of ursodeoxycholic acid, which is hereinafter abbreviated as UDCA, and type IV collagen, to use as indicators the any one or both of levels of UDCA and type IV collagen in addition to the level of LCA; and

determining the liver injury type as hepatocellular injury type when the measured level of LCA is equal to or higher than a predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury and the measured level of UDCA and/or type IV collagen satisfies any one of the following:

(1) the level of UDCA is equal to or lower than a predetermined cut-off value of the level of UDCA for identifying hepatocellular injury-type liver injury;

(2) the level of type IV collagen is equal to or higher than a predetermined cut-off value of the level of type IV collagen for identifying hepatocellular injury-type liver injury; and

(3) the level of UDCA is equal to or lower than a predetermined cut-off value of the level of UDCA for identifying hepatocellular injury-type liver injury and the level of type IV collagen is equal to or higher than a predetermined cut-off value of the level of type IV collagen for identifying hepatocellular injury-type liver injury.

7. The test method according to claim 1, further comprising:

measuring any one or more of levels of deoxycholic acid, which is hereinafter abbreviated as DCA, serum sulfated bile acid, which are hereinafter abbreviated as SSBA, type IV collagen, hyaluronic acid, which is hereinafter abbreviated as HA, and reactive oxygen species, which is hereinafter abbreviated as ROS, to use as indicators the levels of DCA, SSBA, type IV collagen, HA, and ROS in addition to the level of LCA; and

determining the liver injury type as cholestasis type when the measured level of LCA is equal to or lower than a predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury and the measured levels of DCA, SSBA, type IV collagen, HA, and ROS satisfy any one or two or more of the following:

(4) the level of DCA is equal to or lower than a predetermined cut-off value of the level of DCA for identifying cholestasis-type liver injury;

(5) the level of SSBA is equal to or higher than a predetermined cut-off value of the level of SSBA for identifying cholestasis-type liver injury;

(6) the level of type IV collagen is equal to or higher than a predetermined cut-off value of the level of type IV collagen for identifying cholestasis-type liver injury;

(7) the level of HA is equal to or higher than a predetermined cut-off value of the level of HA for identifying cholestasis-type liver injury; and

(8) the level of ROS is equal to or higher than a predetermined cut-off value of the level of ROS for identifying cholestasis-type liver injury.

8. The test method according to claim 1, further comprising:

measuring a level of type N collagen, to use as an indicator the level of type N collagen in addition to the level of LCA; and

determining the liver injury type as a mixed type of hepatocellular injury type and cholestasis type when the measured level of LCA is higher than a predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury and is lower than a predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury and when the measured level of type IV collagen is equal to or higher than a predetermined cut-off value of the level of type IV collagen for identifying cholestasis-type liver injury.

9. The test method according to claim 2, wherein the predetermined cut-off value of the level of LCA is calculated from a receiver operating characteristic curve, which is hereinafter referred to as ROC curve, of the level of LCA.

10. The test method according to claim 3, wherein the predetermined cut-off value of the level of LCA for identify-

ing hepatocellular injury-type liver injury is calculated from a predetermined ROC curve of the level of LCA for hepatocellular injury-type liver injury.

11. The test method according to claim 4, wherein the predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury is calculated from a predetermined ROC curve of the level of LCA for cholestasis-type liver injury.

12. The test method according to claim 6, wherein the predetermined cut-off value of the level of LCA for identifying hepatocellular injury-type liver injury, the predetermined cut-off value of the level of UDCA for identifying hepatocellular injury-type liver injury, and the predetermined cut-off value of the level of type IV collagen for identifying hepatocellular injury-type liver injury are calculated from a predetermined ROC of the level of LCA for hepatocellular injury-type liver injury, a predetermined ROC of the level of UDCA for hepatocellular injury-type liver injury, and a predetermined ROC of the level of type IV collagen for hepatocellular injury-type liver disease, respectively.

13. The test method according to claim 7, wherein the predetermined cut-off value of the level of LCA for identifying cholestasis-type liver injury, the predetermined cut-off value of the level of DCA for identifying cholestasis-type liver injury, the predetermined cut-off value of the level of

SSBA for identifying cholestasis-type liver injury, the predetermined cut-off value of the level of type IV collagen for identifying cholestasis-type liver injury, the predetermined cut-off value of the level of HA for identifying cholestasis-type liver injury, and the predetermined cut-off value of the level of ROS for identifying cholestasis-type liver injury are calculated from a predetermined ROC of the level of LCA for cholestasis-type liver injury, a predetermined ROC of the level of DCA for cholestasis-type liver injury, a predetermined ROC of the level of SSBA for cholestasis-type liver injury, a predetermined ROC of the level of type IV collagen for cholestasis-type liver injury, a predetermined ROC of the level of HA for cholestasis-type liver injury, and a predetermined ROC of the level of ROS for cholestasis-type liver injury, respectively.

14. The test method according to claim 1, wherein the biological sample is a blood sample.

15. The test method according to claim 1, wherein the biological sample is a serum sample.

16. A selection method of a therapeutic agent for liver injury according to a liver injury type, wherein the method is applied for a subject whose liver injury type is determined by the test method of claim 1.

\* \* \* \* \*

专利名称(译)	肝损伤类型的调查方法		
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摘要(译)

本发明提供一种使用受试者的生物样品中的石胆酸 ( 以下简称为LCA ) 的水平作为指标来区分肝损伤类型的试验方法。即，提供一种测试方法，包括测量从受试者收集的生物样品中的LCA水平，并使用测量的LCA水平将肝损伤类型区分为肝细胞损伤类型，胆汁淤积型或其混合型。指示符。

