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(54) **IDENTIFICATION OF NOVEL SPLICE VARIANTS OF THE HUMAN CATALYTIC SUBUNIT CBETA OF CAMP-DEPENDENT PROTEIN KINASE AND THE USE THEREOF**

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(57) **ABSTRACT**

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The C β gene encodes at least 6 different gene products, designated C β 1, C β 2, C β 3, C β 4, C β 4ab and C β 4abc. As is the case with the murine and bovine splice variants, all the human C β splice variants vary in the N-terminal part preceding the part encoded by exon 2. Homologues to all C β splice variants identified in mouse and bovine were identified in human (C β 1, C β 2, C β 3 and C β 4) in addition to two novel C β splice variants (C β 4ab and C β 4abc), that have previously not been identified in any other species. Genomic DNA- and cDNA sequences encode splice variants and include the nucleotide sequences shown in SEQ ID NO: 1, 2, 3, 4, 5 and 6 respectively. The proteins are new splice variants of the C β protein.

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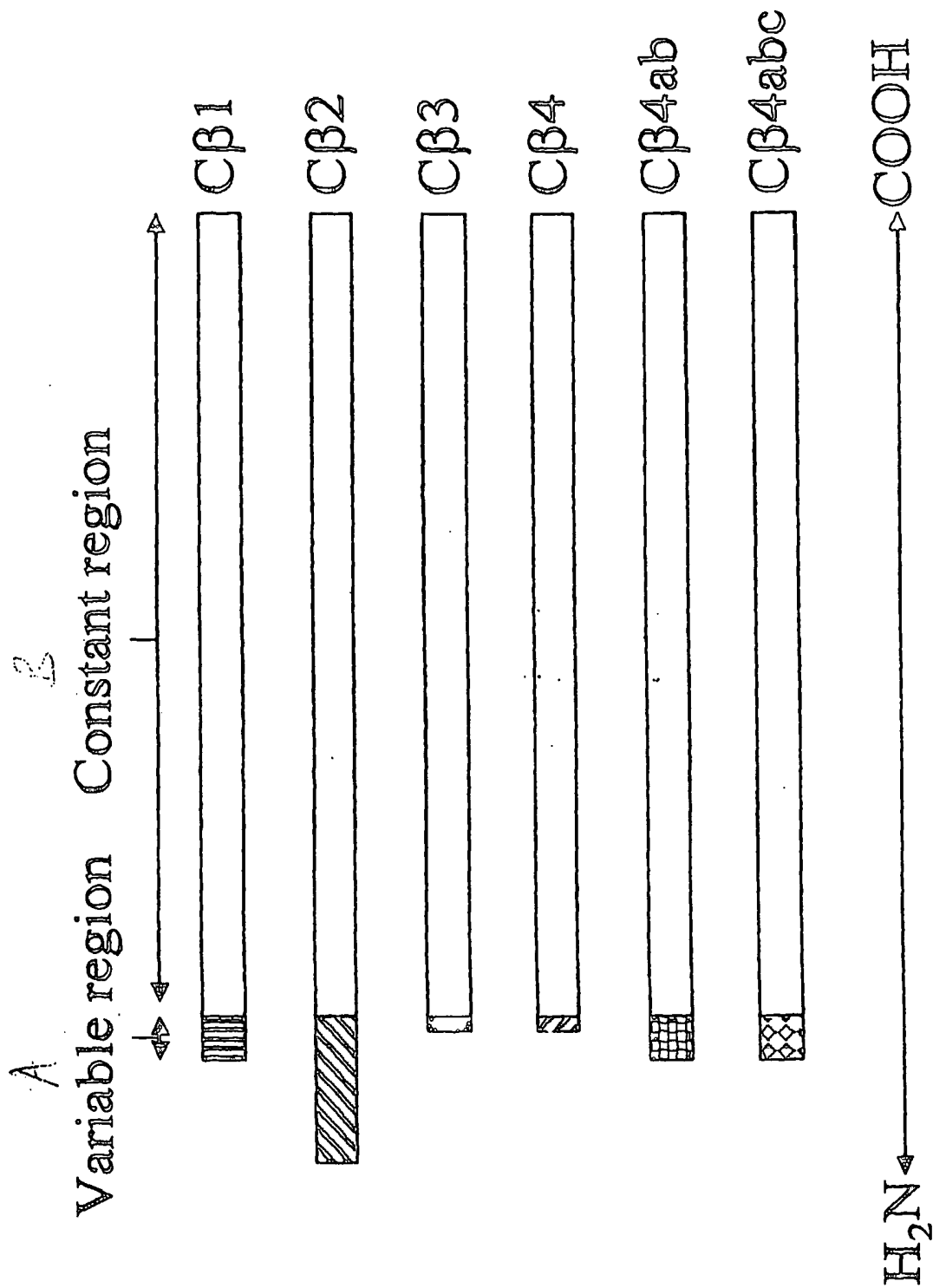
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Dec. 23, 1999 (NO)..... 19996423

FIGURE 1



A

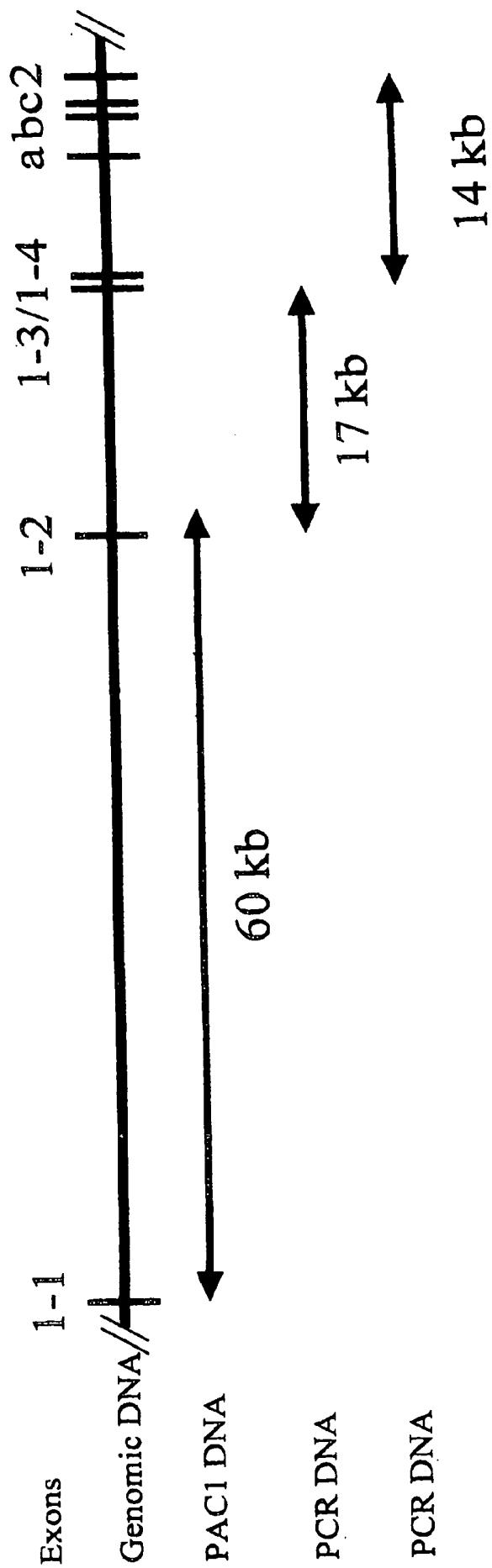


FIGURE 2B

B

Exon 1-1:

ccagcccccttcccttccctgaccccttcttgcctatgccccagacATGGGGAACGCGCGACCG
CCAAGAAAGGCAGCGAGgtggagagcg

Exon 1-2:

agctttatatttaaatgctctcattagcctatatattaatatttaaaccacccaaacataaagccttt
 taggcagatattgcaagtttttaaaatcctcaactctagctgaaaagtgttttgctaagaaaagct
 cagtaatgtgctgttttatattaacaggaaacagaacagcagtagtggtttgaataccctgcaaac
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 ttgagttacattaagtaaaagtgtaaatgcacatgaATGGCAGCTTATAGAGAACCACCTTGTA
CCAGTATACAGGTACAACCTACAGCTCTTCAGAAATTGGAAGGTTTTGCTAGCCGGTTATTCATAG
ACACTCTAAGGTACTGCACATGATCAGAAACAGCTCTGGAAATGACAGCCTTCATTTCTCTGA
ACATACTGCCTTATGGGACAGATCAAgtaagttttg

Exon 1-3 and 1-4:

tgttttaggcagagttcagtggttcgtcacaaataaatgtttcttaatttggtgtttatgactgc
 tcgatttccagagccatgaaatcattgttttgataattctgttcagctcatagttgtttcttag
 gaagatttctgacttacttccacatcacaaaagctcctgcctcacatggcaactgttaaaatggc
 aagttcacgtgctgaagttctacttaacaaggaaccattctatagattctttgtaccattttgta
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 ttgccgtggttaacaactcatgctgatataattgagaacatcttatacatcctggttcgaacat
 ctccctgccattttgagttgttctagtggtatatagaaggaggctgggataactagcttgaaagaaa
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 gttatgctttgcttctagggcttctcttttaaaacaaaagaaagctcttttctgtttctgtgtg
 ctgcatgctccagtggtgtgtgtttacaccatcggttcttctccctctagagattagcataactcc
 tttgctgttgattgtattttgagcaatatgttttgaaagggttggttttctatATGAGTgtaa
 gtatgc

Exon a

cttgatcaag**CACGCAATCATCAGATGCATCTG**gtaggaaaac

Exon b

tggacacaag**CTTGCTCCTCTTCAGAAATATCTG**gtaggcaagt

Exon c

gaacatgtag**ATTCCTTTG**gtatgctcat

Exon 2

atattttcag**TCAAAGAGTT**...

FIGURE 2C

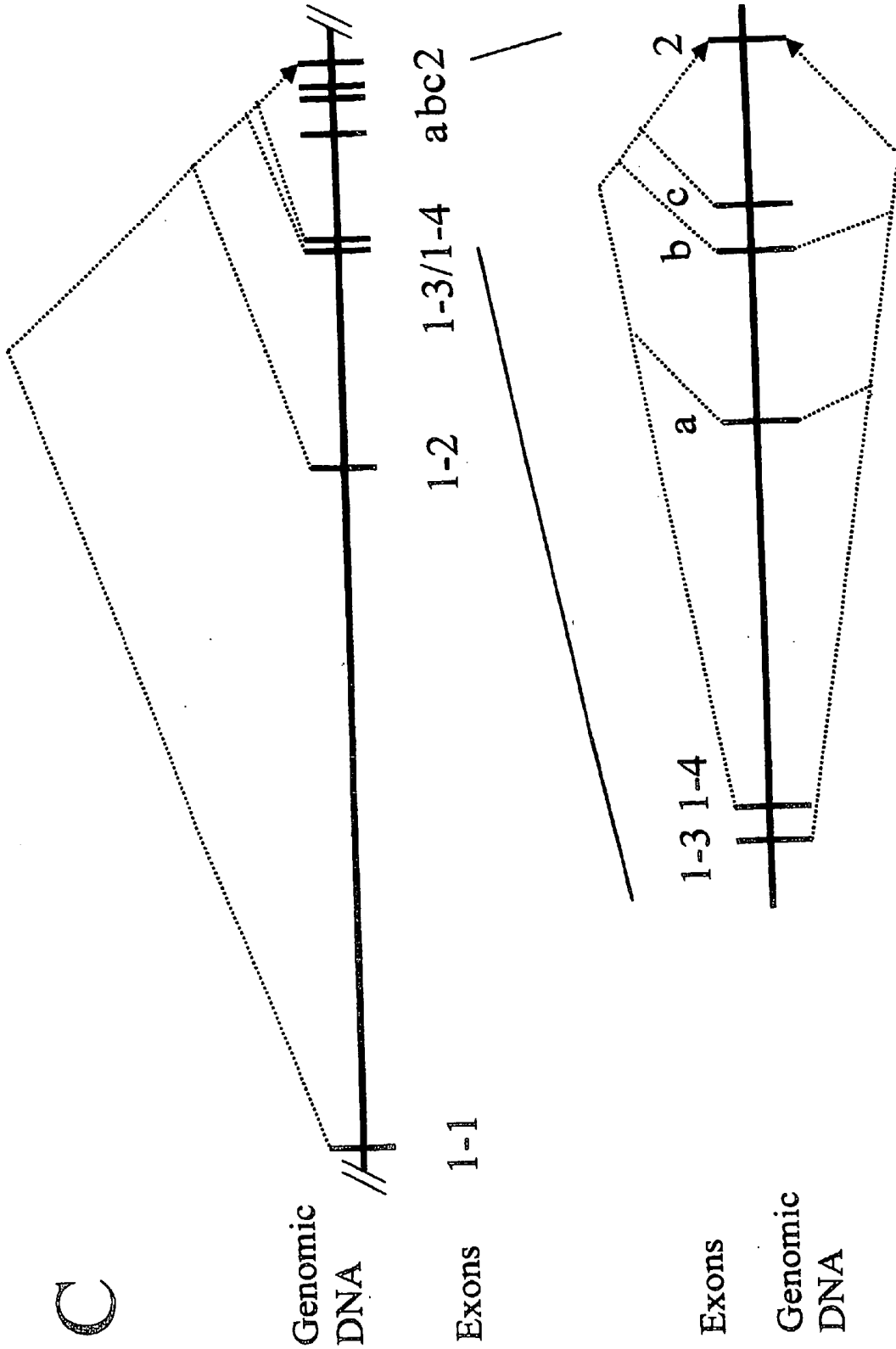
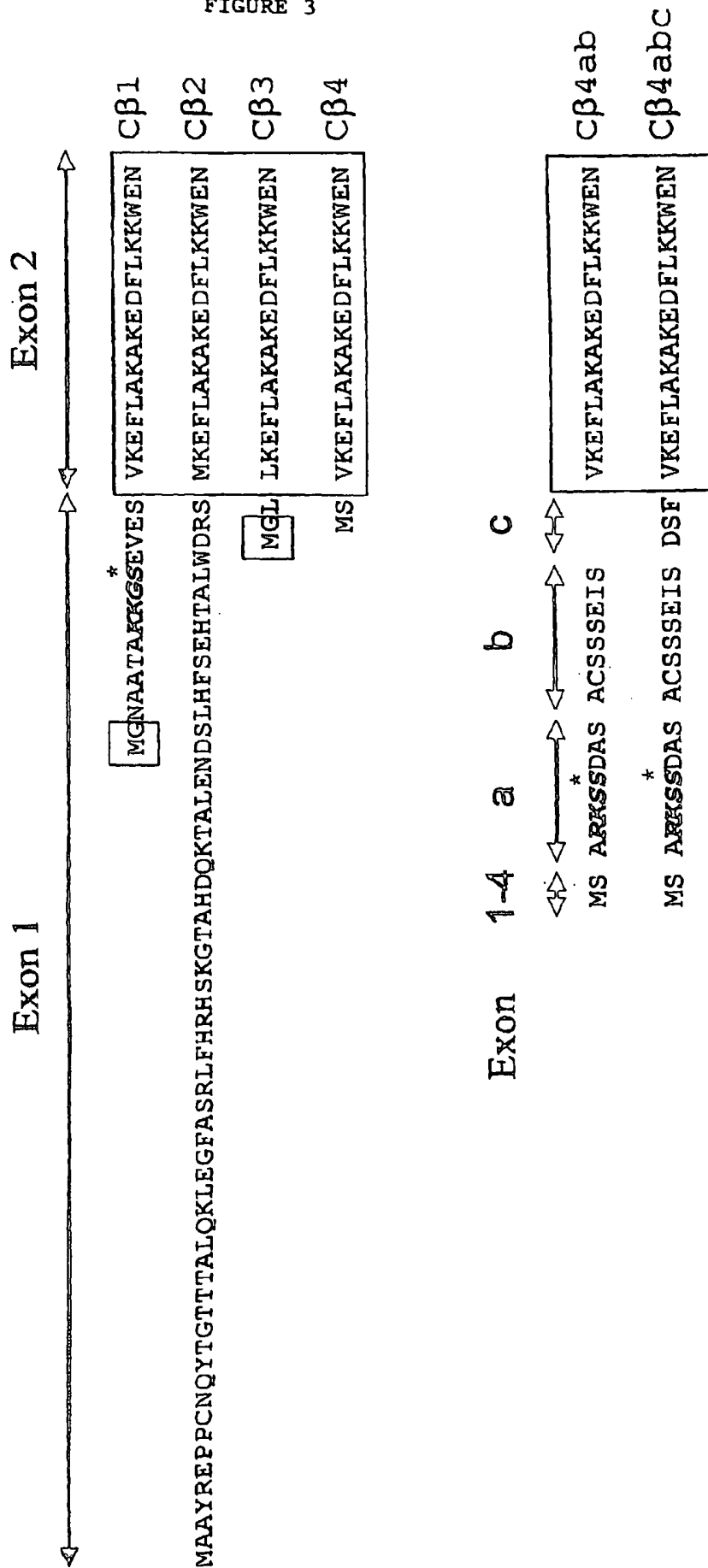
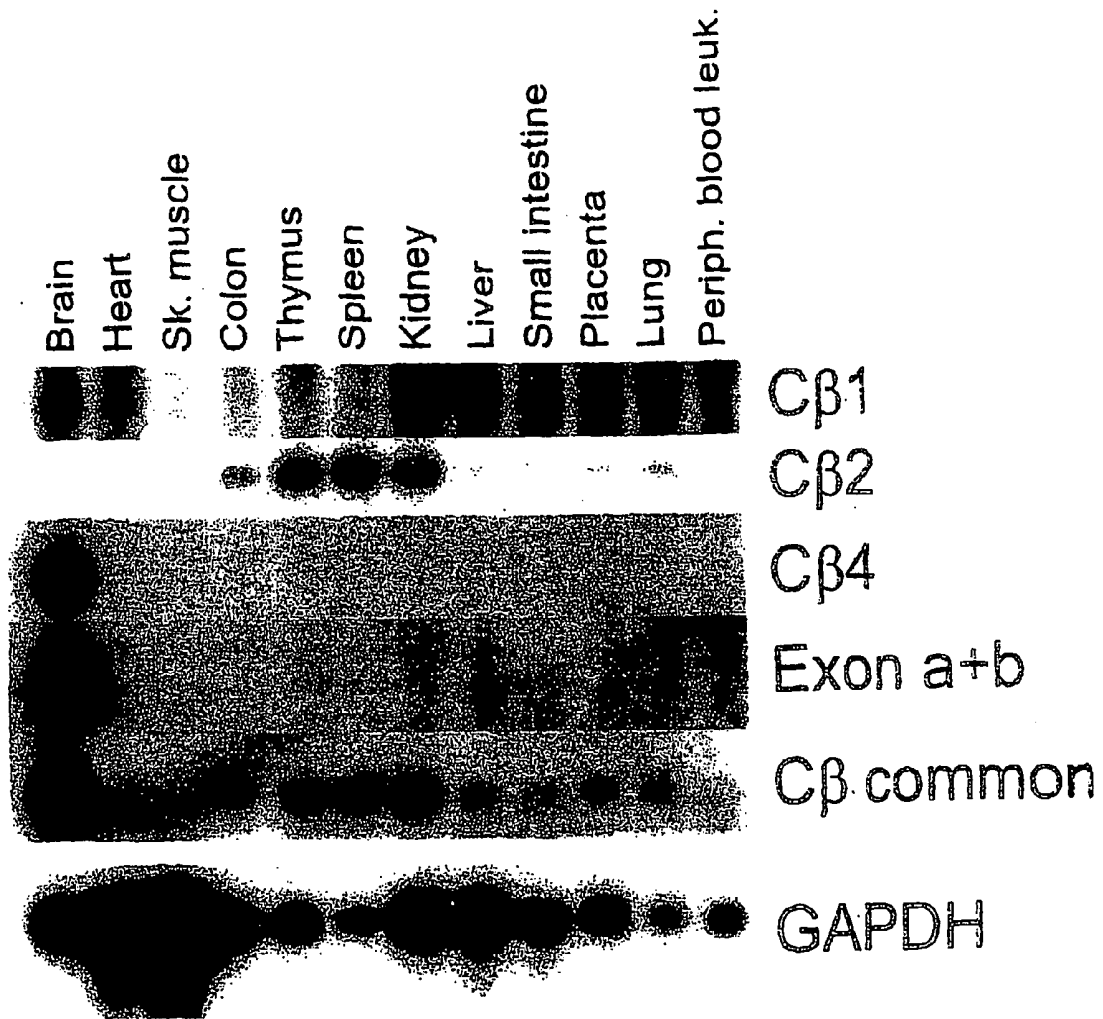


FIGURE 3



FIGUR 4



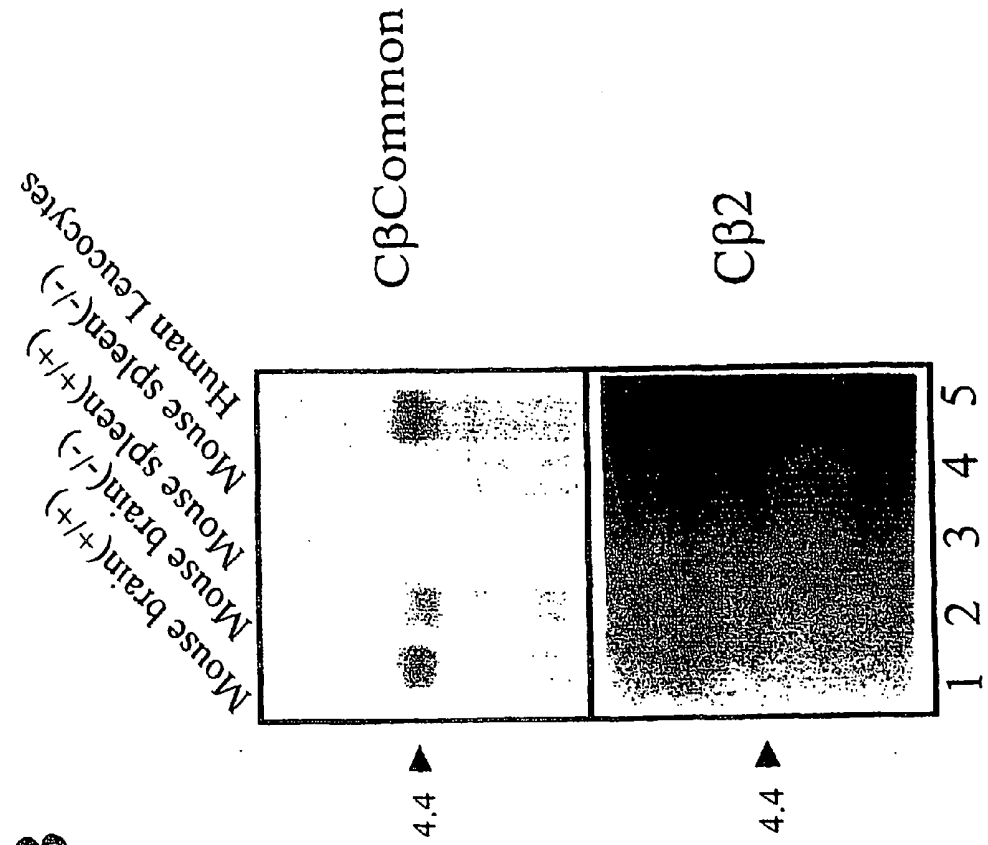


FIGURE 5A

B

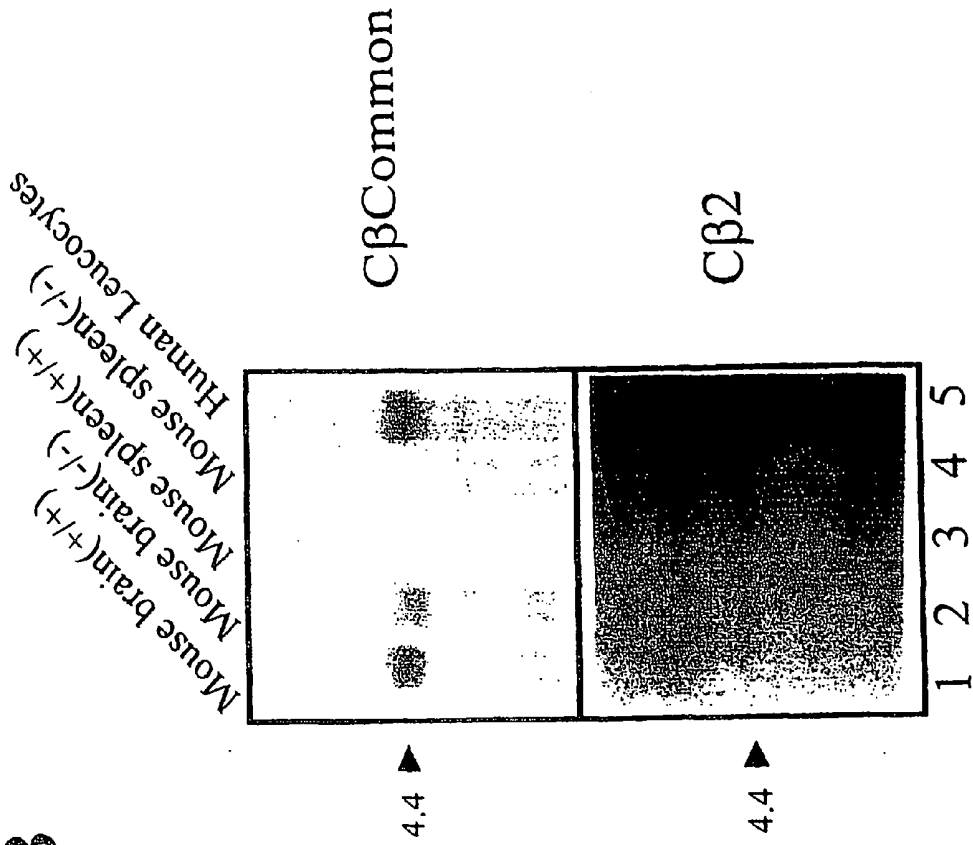


FIGURE 5B

IDENTIFICATION OF NOVEL SPLICE VARIANTS OF THE HUMAN CATALYTIC SUBUNIT CBETA OF CAMP-DEPENDENT PROTEIN KINASE AND THE USE THEREOF

FIELD OF THE INVENTION

[0001] The present invention relates to genomic- and complementary DNA sequences encoding the 6 different gene products, designated C β 1, C β 2, C β 3, C β 4, C β 4ab and C β 4abc which are novel splice variants of C β . The present invention also relates to vectors comprising said DNA sequences and is also directed to said proteins in diagnosis and treatment.

BACKGROUND OF THE INVENTION

[0002] Cyclic 3',5'-adenosine monophosphate (cAMP) is a key intracellular signalling molecule, which main function is to activate the cAMP-dependent protein kinases (PKA) [1]. PKA consists of a heterotetramer, with a regulatory (R) subunit dimer and two catalytic (C) subunits. The holoenzyme is activated when four molecules of cAMP bind to the R subunit dimer, two to each R subunit, releasing two free active C subunits [2]. In man, four different R subunits (RI α , RI β , RII α , RII β), and four different C subunits (C α , C β , C γ and PrKX) have been identified [3]. The C α and C β subunits are expressed in most tissues, while the C γ subunit, which is transcribed from an intron-less gene and represents a retroposon derived from the C α subunit [4], is only expressed in human testis [5]. PrKX is an X chromosome-encoded protein kinase, and was recently identified as a PKA C subunit since it is inhibited by both PKI and RI α and the RI α /PrKX complex is activated by cAMP [6].

[0003] Splice variants of both C α and C β have been identified. The splice variants of C α have been termed C α 1 (previously named C α [7]), C α 2 [8] and C α -s [9]. Originally C α 2 was isolated from interferon-treated cells and identified as a C-terminally truncated C α 1 subunit. However, recently a novel C α 2 splice variant was reported [10]. The novel C α 2 variant was shown to be identical to the previously identified C α splice variant, C α -s. Moreover, C α -s which was originally isolated and characterized from ovine sperm [9], has later been cloned from a human testis cDNA library and identified in human sperm [11]. Both C α -s/C α 2 are encoded with a truncated N-terminal end when compared to C α 1. The variable parts of C α 1 and C α -s are located upstream of exon 2 in the murine C α gene, implying that the variation in the N-terminal end of the C α 1 and C α -s/C α 2 are due to alternative use of different first exons. In bovine, two splice variants of C β have been identified, termed bovine C β 1 [12] and bovine C β 2 [13]. The bovine splice variants contain variable N-terminal ends in which the non-identical sequences are most probably encoded by different forms of exon 1. Bovine C β 2 is expressed at low levels in most tissues with the highest expression in the spleen, thymus, and kidney and to some extent brain. Furthermore, in the mouse, three splice variants of C β have been identified and are designated mouse C β 1, mC β 2 and mouse C β 3 [14]. Whereas mouse C β 1 is ubiquitously expressed, mouse C β 2 and mouse C β 3 have so far only been identified in the brain. The mouse C β 1 and bovine C β 1 are similar in the entire sequence, demonstrating that they represent orthologous protein sequences. However, neither mouse C β 3 nor mouse C β 4 were similar to bovine C β 2 in the N-terminal part,

indicating that their N-terminals are encoded by unrelated exons. Previous to this study, only a single splice variant of human C β had been identified (C β 1), homologous to mouse C β 1 and bovine C β 1.

SUMMARY OF THE INVENTION

[0004] The present invention demonstrate that the C β gene encodes at least 6 different gene products, designated C β 1, C β 2, C β 3, C β 4, C β 4ab and C β 4abc. As is the case with the murine and bovine splice variants, all the human C β splice variants vary in the N-terminal part preceding the part encoded by exon 2. Homologues to all C β splice variants identified in mouse and bovine were identified in human (C β 1, C β 2, C β 3 and C β 4) in addition to two novel C β splice variants (C β 4ab and C β 4abc), that have previously not been identified in any other species. The present invention includes in this respect genomic DNA- and cDNA sequences encoding said splice variants and comprises the nucleotide sequences shown in SEQ ID NO: 1, 2, 3, 4, 5 and 6 respectively. Wherein the said proteins are new splice variants of the C β protein. The present invention is further directed to vectors comprising said cDNA sequences. The invention also includes proteins characterised by the specific amino acid C β splice variant proteins C β 2, C β 4ab and C β 4abc shown in SEQ ID NO: 7, 8 and 9. The invention includes further use of the said C β splice variant proteins and DNA sequences in preparation of pharmaceuticals for diagnostic- and therapeutic purposes.

BRIEF DESCRIPTION OF THE DRAWINGS

[0005] **FIG. 1:** A: Identification of cDNAs encoding human C β splice variants. Schematic representation of the protein-encoding sequences of the various C β splice variants found in human. Human cDNAs from total fetus and brain were amplified using primers complementary to the C β cDNA, subcloned and sequenced. The resulting cDNAs were identical to the previously published C β cDNA (C β 1) downstream of nucleotide 46 (constant region). However, five novel cDNA sequences, designated C β 2, C β 3, C β 4, C β 4ab and C β 4abc, could be identified based on differences in the 5'-ends of the sequences (variable region).

[0006] **FIG. 2:** A: Structure of the human genomic region encoding the novel C β splice variants. Primers were made based on exon 2 and the most 5'-end of the different C β cDNAs, and used to amplify human genomic DNA by PCR. Two overlapping PCR products of 14 and 17 kb, respectively, were identified and mapped by Southern blotting and hybridization to oligonucleotides corresponding to the different cDNAs. As derived from the 14 and 17 kb PCR products, exon 1-2 1-3,1-4 and exon a, b and c are located 31, 14.1, 14, 8.1, 5.4 and 4.4 kb upstream of exon 2. Based on restriction mapping of the PAC clone RPCI-6-228E23, exon 1-1 is located approximately 60 kb upstream of exon 1-2. Exon 1-1 is specific for the splice variant, which encodes C β 1. The exons are indicated as vertical lines. The introns are drawn to scale as indicated. B: Nucleotide sequence of genomic regions encoding novel splice variants of C β . Protein encoding sequences are in capital letters, intron and 5'-untranslated sequences are in lower case letters. Translation initiation codons are underlined. Only the 5'-end of exon 2 is included. C: Schematic representation of how the various human C β exons 5' to exon 2 may be spliced. The upper panel describes a potential model in

which four variants of exon 1 designated exon 1-1,1-2, 1-3 and 14 may alternatively splice with exon 1 to encode the splice variant specific sequence in C β 1, C β 2, C β 3 and C β 4. The lower panel describes a model in which the exons a, b and c may splice with exon 14 and 1-3 upstream of exon 2 to encode the splice variant-specific sequences in C β 4ab, C β 4abc and C β 3ab.

[0007] FIG. 3: Deduced amino acid sequence of C β splice variants. The amino acid sequences of the amino terminal parts of C β 1 and five new splice variants, designated C β 2, C β 3, C β 4, C β 4ab and C β 4abc according to the cDNA clones shown in **FIG. 1A**. The amino acid sequences are shown in the one letter code and demonstrate that six novel C β exons give rise to five different cDNAs as a result of alternative promoter use and alternative splicing. The myristylation motive G-N previously identified in C β 1 is boxed. A PKA autophosphorylation motive that has previously been identified in C β 1, is underlined and Ser10 which is potentially phosphorylated, is labeled by an asterisk. Note that there is a PKA autophosphorylation motif, encoded by exon a, present in C β 4ab and C β 4abc.

[0008] FIG. 4: Tissue distribution of different C β splice variants. Northern blots containing various human tissues were hybridized using probes specific for C β 1, C β 2, C β 4, exon a+b and a probe common to all C β splice variants (C β common). For comparison, the same blots were hybridized using a GAPDH cDNA (GAPDH). All C β 1 mRNAs had the same apparent length (4.4 kb).

[0009] FIG. 5: A: Species distribution of C β 2. A Southern blot containing EcoRI digested genomic DNA from various species was hybridized using a DNA probe corresponding to exon 1-2 (C β 2 specific). A single hybridizing band identifying genomic sequence homologous to human exon 1-2 was identified in mammals such as monkey, dog, rabbit and human except mouse and rat. B: C β 2 is not expressed in the mouse. A Northern blot containing total RNA (20 μ g pr. lane) isolated from wild type (+/+) mouse brain and spleen (lane 1 and 3), brain and spleen of mice ablated (-/-) for C β 1 (lane 2 and 4) and human peripheral blood leukocytes (lane 5) was probed with a C β probe expected to recognize all known C β splice variants (Co Common, upper panel) and a C β probe specific for the C β 2 splice variant (C β 2, lower panel). Messenger RNA recognized by the two probes is indicated as 4.4 kb.

DETAILED DESCRIPTION OF THE INVENTION

[0010] The present invention demonstrate that the human C β gene encodes five novel C β splice variants, designated C β 2, C β 3, C β 4, C β 4ab and C β 4abc, in addition to the previously identified splice variant C β 1 [12]. All the C β splice variants contained a unique N-terminal end, and showed tissue specific expression. As we found no evidence of an additional exon upstream of exon 1-1 and all the cDNA characterized had unique 5'-ends, it is reasonable to assume that the exon 1-1,1-2, 1-3 and 14 each contain a separate promoter, and that the resulting mRNA products are due to alternative use of different promoters. Despite this, we can not rule out the possibility that two or more of these splice variants share a common promoter used to alternatively splice the different exons. Furthermore, we found two C β variants, C β 4ab and C β 4abc, that were the results of alter-

native splicing of either exon a and b, or exon a, b and c, between exon 1-4 and exon 2. The presence of the corresponding mRNA was confirmed by hybridizing a Northern blot with a probe complimentary to the sequences found in exons a and b. This probe and the probe specific for C β 4 bound to an RNA with the same apparent length located in human brain. The location of the exons a, b and c may suggest that they generate splice variants of C β in addition to those demonstrated here. Indeed, a short cDNA from human infant brain have been sequenced and demonstrated to contain a combination of exons 1-3, a, b and 2 (Accession no. AA35 1487, see **FIG. 2C**). We were unable to produce such a cDNA, which could be due to low level expression of C β 3 in adult brain.

[0011] The two splice variants C α 1 and C β 1 are highly conserved in the parts encoded by exon 1, differing in only 2 of the first 16 amino acids [7;12]. It is therefore tempting to suggest that this region serve a specific role in the function of these splice variants. Thus, the fact that we have identified several C β splice variants with variable N-terminal ends could suggest that the N-terminal domain might reflect specific functional features associated with each splice variant. This is supported by studies of the mouse C β 1 KO mouse, which displayed impaired hippocampal plasticity [16]. However, to what extent N-terminal differences influence catalytic activity is not known since it was shown that the N-terminally truncated C β splice variants in mouse, C β 2 and C β 3 were catalytically active, an activity that was inhibited both by PKI and the R subunit in vivo [14]. In addition, a study by Herberg et al [17] showed that deleting amino acids 1-14 in the C β isoform did not influence catalytic activity, demonstrating that the N-terminal specific for the C α 1/C β 1 is not necessary for catalytic activity.

[0012] The N-terminal of C α 1 and C β 1 contain two sites for post-translational modification, a myristylation site and an autophosphorylation site [5;18;19]. In C α 1, C β 1 and C β 3 the N-terminal amino acid is G (Gly) which has been shown as an absolute requirement for myristylation [20]. Despite this, it was previously demonstrated in the mouse that C β 3 does not undergo myristylation in vivo [14]. This phenomena may be explained based on a recent study, demonstrating that the amino acid C-terminal to G must be N if myristylation shall occur. This because deamination of N to yield D is an absolute requirement [21]. Because the amino acid C-terminal to G is L in both mouse and human C β 3, it explains why mouse C β 3 is not myristylated and suggests that the human C β 3 may not be myristylated in vivo.

[0013] The fact that several human C β splice variants (C β 2, C β 3, C β 4, C β 4ab and C β 4abc) lack the ability to become myristylated in vivo, question the role of this post translational modification. Based on the Ca crystal structure it appears that the myristyl group serves to fill and shade a hydrophobic pocket in the large lobe [22], suggesting that this N-terminal modification serves to solubilize the C subunit. This is supported by two independent observations. Firstly, expression of an N-terminally truncated form of C α 1 revealed a C subunit tightly associated with the particulate fraction [23]. Secondly, the C α -s/C α 2 which is a naturally occurring N-terminally truncated splice variant is tightly associate with sub cellular structures in both ovine-[9,24] and human [11] sperm. This taken together with a recent report, which demonstrated that the myristyl group serves to increase the lipophilic properties of the C subunit when

binding the RII- but not the RI subunit [25], suggests that the N-terminal amino acids of C α 1 together with myristylation serves to influence C subunit solubility. Thus, the sequence similarity between C α 1 and C β 1 and the difference in solubility of C α 1 and C α -s/C α 2, may imply comparable difference in solubility between C β 1 and the truncated C β forms.

[0014] Previously a consensus autophosphorylation motif (-KKGS¹⁰-) was identified in C α 1 and C α 1 [12;26], that is phosphorylated when C α 1 is expressed in bacteria [18;23]. In the study by Yonemoto et al. (1993) mutation of S¹⁰ yielded an insoluble enzyme that appeared inactive. Thus, the N-terminal domain may also have implications for catalytic activity by an unknown mechanism. However, like the human C β 2, C β 3, C β 4, the mouse C β 2 and C β 3 lack S¹⁰, yet these splice variants are soluble and catalytically active in vivo [14]. This suggests that the human homologues most probably are active and may imply that S¹⁰ phosphorylation is not crucial for C subunit catalysis. Interestingly, we identified a potential autophosphorylation site (-RKSS⁶-) in C β 4ab and C β 4abc that was encoded by exon a. To what extent this site represents a true autophosphorylation site that will influence C β 4ab and C β 4abc properties, remains to be seen.

[0015] The human C β 2 splice variant was similar to the previously identified bovine C β 2 splice variant, but we have been unable to identify a similar splice variant in mice. Interestingly, the human C β 2 splice variant is expressed only in peripheral tissues, while no detectable C β 2 mRNA signal is found in human brain. However, no C β can be detected outside the brain in mice lacking the C β 1 splice variant [14;16]. In addition, we were unable to detect any signal when hybridizing mouse DNA using a human C β 2 specific probe. Thus, it is likely that mice do not contain a homologue of the human and bovine C β 2 splice variants.

[0016] Interestingly, C β 2 is the most atypical of the C β splice variants. This subunit is encoded with an extended N-terminal domain, which do not resemble any of the other C β splice variants. The unique domain together with the fact that C β 2 lacks the myristylation- as well the autophosphorylation site, and that C β 2 is the only C β splice variant not identified in the brain, may suggest specific and unique features associated with this splice variant in other tissues that will await further studies.

[0017] The inventors suggest that tissue-specific expression of various C β splice variants when complexed with R subunits may imply novel PKA holoenzymes with specific functional features that may be important as mediators of cAMP effects.

[0018] The present invention includes in this respect genomic DNA- and cDNA sequences encoding splice variants C β 1, C β 2, C β 3, C β 4, C β 4ab and C β 4abc and comprises the nucleotide sequence shown in SEQ ID NO: 1, 2, 3, 4, 5 and 6 respectively. Wherein the said proteins are new splice variants of the C β protein. The present invention is further directed to vectors comprising said cDNA sequences. The invention also includes proteins characterised by the specific amino acid C β splice variant proteins; C β 2, C β 4ab and C β 4abc shown in SEQ ID NO: 7, 8 and 9 respectively. The invention includes further use of the said C β splice variant proteins and DNA sequences in preparation of pharmaceuticals for diagnostic- and therapeutic in order to identify, characterize and produce pharmacological compositions.

[0019] C β 2 is an enzyme that is expressed in lymphoid cells, whereby its function is to mediate the regulatory effects of cAMP on T cell activation. Thus, altered levels, location and/or activity of C β 2 will according to the inventors results, have impact on the regulation and normal function of receptors and enzymes which are important for T cell activation and are regulated by cAMP. This knowledge can be used to diagnose hyperreactive and dysfunctional T cells associated with various immune diseases.

[0020] 1) Malfunctioned T cells: It is well known that T cells isolated from patients suffering from T cell-dependent common variable immune deficiency (CVI) and acquired immune deficiency syndrome (AIDS) do not respond to antigen. Furthermore, T cells isolated from patients suffering from certain types of rheumatoid arthritis and other autoimmune diseases are hypersensitive to foreign antigens. In both cases these situations evoke abnormal immune responses that may involve malfunctioned C β 2. This may either be monitored as constitutively activated C β 2, subnormal activity or dislocation of C β 2.

[0021] 1.1) Improving T cell dysfunction: Present invention makes it possible to identify, characterize and produce pharmacological compositions after high throughput screening that specifically will inhibit the enzymatic activity of C β 2. These compositions should be developed such that they can be introduced orally or intravenously to enter the blood system reaching the dysfunctional T cells.

[0022] Furthermore, dislocation of C β 2 protein from the T cell membrane will short cut the regulatory effects of C β 2 on relevant receptors. Thus, the present invention makes it possible to identify, characterize and produce pharmacological composition after high throughput screening that will specifically and irreversibly block C β 2 interaction with the T cell membrane. These compositions should be developed such that they can be introduced orally or intravenously to enter the blood system reaching the T cell.

[0023] 1.2) Down regulation of hyper active T cells: Present invention makes it possible to identify, characterize and produce pharmacological compositions after high throughput screening that specifically will activate the enzymatic activity of C β 2. These compositions should be developed such that they can be introduced orally or intravenously to enter the blood system reaching the dysfunctional T cells.

[0024] 1.3) Kits for diagnosing C β 2 mutations: T cell malfunction caused by mal function or -localization of C β 2 enzyme activity may be caused by mutation(s) in the C β 2 protein. Present invention makes it possible to develop kits, which would diagnostically facilitate if mutated C β 2 is present. Such kits should be developed with C β 2 specific DNA probes.

[0025] Present invention makes it possible to develop a method for inspection and screening of patient T cells for the presence and location of C β 2 comprising:

[0026] a) collection and washing in buffer of isolated peripheral blood T lymphocytes according to [27];

[0027] b) preparing for identification of C β 2 protein by immunofluorescence, T cells are let to settle onto poly L-lysine coated cover slips following detergent-dependent lysis;

- [0028] c) incubation with primary antibody (Ab), either irrelevant Ab or C β 2 specific Ab, Ab overshoot will be removed by washing buffer and T cells incubated with secondary anti-IgG Ab conjugated with a fluorescent;
- [0029] d) inspection of T cells under fluorescent microscopy.
- [0030] Present invention makes it further possible to develop a method of screening patient T cells for membrane associated C β 2 catalytic activity comprising:
- [0031] a) collection and washing in buffer of isolated peripheral blood T lymphocytes according to [27];
- [0032] b) preparation of T cells by lysing in detergent buffer;
- [0033] b) monitoring C β 2 specific catalytic activity by established assay, C β 1 activity is used as an internal control to determine relative activity.
- [0034] Present invention makes it also possible to screen patients for mutations in the C β 2 gene and mRNA comprising:
- [0035] a) collection and washing in buffer of isolated peripheral blood T lymphocytes according to [27];
- [0036] b) isolation of total RNA and genomic DNA according to established methods followed by RT-PCR using C β 2 specific primers according to cDNA sequence of C β 2 specific nucleotides or the C β 2 specific exon, designated exon 1-2.
- [0037] Materials and Methods.
- [0038] General Protocols
- [0039] Complementary DNA probes were radiolabeled using the Megaprime random priming kit and α -[32P]dCTP (Amersham) as instructed by the manufacturers to a specific activity of at least 1×10^9 cpm. Synthetic oligonucleotides were radiolabeled using T4 polynucleotide kinase (Pharmacia) and γ -[32P]ATP as instructed by the manufacturer.
- [0040] DNA was either sequenced manually using Thermo Sequenase radioabeled terminator cycle sequencing kit (Amersham, Buckinghamshire, UK) or by Medigenomix (Martinsried, Germany). Sequences were analyzed using the Wisconsin University GCG program package (UWGC) and the basic local alignment and search tool (BLAST) [5].
- [0041] Identification of cDNAs
- [0042] The 5'-end of human C β cDNA was amplified from human total fetus and brain Marathon RACE-ready cDNAs (Clontech) using the Advantage KlenTaq Polymerase Mix (Clontech) as described by the manufacturer. Amplification was performed using adapter primer 1 (Clontech) and four different primers complementary to the human C β cDNA sequence (5'-CAACCCAAAGAGAAGTAAGAAAGTG-GTCTA-3', 5'-TTGGTTGGTCTGCAAAGAATGGGG-GATAGC-3', 5'-TTTTCTCAITCAAAGTATGCTC-TATTTGC-3' and 5'-AGAATAATGCCGGACTTGAAGATTTTAAAA-3').
- [0043] Five cycles were performed with 45 sec 94° C., 2 min 72° C., five cycles 45 sec 94° C., 2 min 70° C., 25 cycles 45 sec 94° C., 2 min 68° C., and a final extension of 10 min at 72° C. The resulting products were separated by gel electrophoresis, subcloned to pCR2.1TOPO (Invitrogen) as instructed by the manufacturer and sequenced.
- [0044] Amplification of C β gene fragments.
- [0045] A genomic fragment was amplified using an oligonucleotide corresponding to exon 1-3 (5'-GTTTAGGTG-CAATCAITCTGCTGTTTG-3') and a primer complementary to sequences in exon 2 (5'-AAAAAGTCTTCTTTGGCTTTGGCTAGA-3'). Another genomic fragment was amplified using a primer corresponding to exon 1-2 (5'-TGGCAGCTTATAGAGAACCACCTT-3') and a primer complementary to sequence found in exon 1-3 (5'-CAATCCCATGTTGAACCTGGCA-3'). PCR reactions were performed using the Boehringer-Mannheim Expand Long Template PCR kit as instructed by the manufacturer using buffer 2. PCR was performed using human genomic DNA (Boehringer-Mannheim) as template with 1 min at 92° C., 30 cycles of 10 sec 94° C., 30 sec 60° C. and 10 min (extended with 20 sec per cycle from cycle 11 to cycle 30) 68° C., and a final incubation of 7 min at 68° C. Products were separated by agarose gel electrophoresis and analyzed by Southern blotting using radiolabeled cDNAs and synthetic oligonucleotides corresponding to the different exons.
- [0046] Screening of PAC Library and Subcloning of Exon-containing Sequences.
- [0047] The human P1-derived Artificial Chromosome (PAC) library, RPCI-6 was screened and the isolated bacterial clone was grown in liquid culture and plasmid DNA was isolated using ion-exchange columns as described by the manufacturer (Qiagen, Hilden, Germany). Exon-containing DNA restriction fragments were identified by Southern blotting using radio labeled cDNAs and synthetic oligonucleotides. Exon-containing fragments were excised from the gel and subcloned to the pZERO2.1 vector (Invitrogen) as instructed by the manufacturer.
- [0048] Generation of Splice Variant Specific Probes, Northern Blotting and Southern Blotting.
- [0049] DNA fragments corresponding to the splice variant-specific parts of the cDNAs were amplified by PCR. The following primers were used for the different splice variants:
- C β 1: 5'-GCTCTCCACCTCGTCCTTTCTT-3'
and
primer 5'-CCAGCCCCCTTCCCTCCCTGAC-3',
- C β 2: primer 5'-TGGCAGCTTATAGAGAACCACCTT-3'
and
primer 5'-ATTGATCTGTCCATAAGGCAGTAT-3',
- C β 3: primer 5'-TCACAGCTAGCAGTAAGAGCTG-3'
and
primer 5'-CAATCCCATGTTGAACCTGGCA-3',
- C β 4: primer 5'-TCTCCAGTGTGTGTTTACAC-3'
and
primer 5'-ATGATGAAAACCAACCTTTCCA-3'.
- [0050] The primers were used for amplification of the fragments from cloned RACE-products using Taq DNA polymerase (Perkin-Elmer) as described by the manufac-

turer. For generation of a probe specifically recognizing exon a and b, the primers 5'-GATATTTCTGAAGAGGAGCAAGCAGATGCATCTGATGATTTGCGTG-3' and 5'-CACGCAAATCATCAGATGCATCTGCT-TGCTCCTCTTCAGAAATATC-3' were annealed, phosphorylated and ligated. A 1.5 kb fragment of C β cDNA [5] was used for recognizing the parts of the C β mRNA common to all splice variants. Two similar Northern blots containing RNA from various human sources were purchased from Clontech. One blot was hybridized using a probe specific for C β 2, while the other blot was probed in succession with probes specific for C β 3, C β 4, exon a and b, and the 1.5 kb C β cDNA. Both blots were hybridized using GAPDH cDNA as control. As an almost identical pattern of hybridization was obtained using GAPDH on both blots, only one GAPDH blot is shown (**FIG. 4**). All probes were hybridized in ExpressHyb hybridization solution (Clontech) as described by the manufacturer. A Southern blot containing EcoRI-digested DNA from various species (Clontech) and Southern blots containing human and mouse DNA digested with various enzymes were hybridized using the probe specific for C β 2. The filters were prehybridized in 5 \times Denhardt's solution, 5 \times SSC, 50 mM sodium phosphate buffer, pH 6.8, 0.1% SDS, 250 μ g/ml single stranded salmon sperm DNA, and 50% (v/v) formamide at 42 $^{\circ}$ C. for 3 h, and hybridized for 16 h in a similar solution containing the radiolabeled C β common or C β 2 probe. The membranes were washed four times in 2 \times SSC, 0.1% SDS for 5 min at room temperature, followed by two washes using 0.5 \times SSC, 0.1% SDS at 50 $^{\circ}$ C. for 30 min. Autoradiography was performed at -70 $^{\circ}$ C. using Amersham Hyperfilm MP and intensifying screens.

[0051] In order that this invention may be better understood, the following examples are set forth. These examples are for the purposes of illustration only, and are not to be construed as limiting the scope of the invention in any manner.

EXAMPLES

Example 1

[0052] Identification of Exons Encoding Novel Splice Variants of Human C β .

[0053] The 5'-ends of human C β cDNAs were amplified from human brain and total fetus RACE-ready cDNA using four different oligonucleotide primers complementary to the previously published human C β cDNA sequence, in combination with an anchor primer. The resulting PCR products were subcloned, sequenced and compared to the previously published human C β cDNA sequence which is now designated C β 1 (**FIG. 1**). All clones sequenced were shown to lack the 46 first protein-encoding nucleotides in the human C β 1 cDNA sequence. Instead 5 novel stretches of protein encoding sequences were identified (**FIG. 1**, variable region). Each of the clones contained a translation initiation codon and one or more in-frame upstream stop codons. The five novel cDNA sequences were designated C β 2, C β 3, C β 4, C β 4ab and C β 4abc.

[0054] All the C β cDNAs were similar from nucleotide 47 and down stream in the C β 1 cDNA, which corresponds to the start of exon 2 in the murine C β gene. The identification of novel protein-encoding sequences upstream of exon 2,

indicated the presence of several different exons upstream of exon 2. Thus, human genomic DNA was amplified using a combination of primers corresponding to exon 2 (antisense orientation) and the 5'-ends of the different novel cDNAs (sense and antisense orientation) in different combinations. A 17 kb PCR product was the result of an amplification using a primer corresponding to the 5'-end of C β 2 cDNA (sense orientation) and the 5'-end of C β 3 (antisense orientation) Furthermore, a 14 kb PCR product was the result of an amplification using a primer corresponding to the 5'-end of C β 3 cDNA (sense orientation) and a primer corresponding to exon 2 (antisense orientation). These clones enabled us to physically map six novel exons in the C β gene that were designated 1-2,1-3, 1-4, a, b and c, and which were located 31, 14.1, 14, 8.1, 5.4 and 4.4 kb upstream of exon 2, respectively (**FIG. 2A**). Furthermore, a PAC library was screened using the 5' ends of C β 1 and C β 2 cDNAs as probes. One of the clones identified, RPCI-6-228E23, contained both exon 1-2 and an exon containing the entire splice variant-specific part of the C β 1 cDNA, which we termed exon 1-1. This PAC clone was selected for detailed restriction mapping using CpG cutters. The digested PAC DNA was separated by pulsed-field gel electrophoresis (PFGE), transferred to Southern blot membranes and hybridized with exon 1-1 and 1-2, as well as Sp6 and T7 oligonucleotide probes. These results revealed a distance of approximately 60 kb between exon 1-1 and 1-2 (**FIG. 2A**). All nucleotide sequences found in the different C β cDNAs could be identified in a continuous stretch of human genomic DNA, thereby supporting the notion that these cDNAs are products of the same gene. Exon 1-1 was shown to be homologous to the previously identified exon 1A of the murine C β gene. As shown in **FIG. 2B**, exon 1-2 contains the entire C β 2 specific sequence, and exon 1-3 contains the sequence specific for C β 3 which is homologous to the previously identified exon 1B in the mouse C β gene. Finally, exon 1-4 was shown to contain the sequence specific for the human C β 4 splice variant, and to be homologous to the murine exon 1C, which encodes the N-terminal end in the murine C β 2 splice variant. Based on the C β 4ab and C β 4abc cDNA sequences, the exons a, b and c (**FIG. 2B**), were demonstrated to be alternatively spliced in between exon 1-4 and exon 2, with either exons 1-4, a, b and 2 or exons 1-4, a, b, c and 2 (**FIG. 2C**, lower panel). These cDNA sequences represent novel C β splice variants not identified in any other species.

Example 2

[0055] Deduced Amino Acid Sequence of Novel C β 3 Splice Variants.

[0056] The N-terminal parts of the deduced amino acid sequences of the previously published C β 1-sequence and the 5 novel C β splice variants are illustrated in **FIG. 3** (upper and lower panels). The splice variants were identical starting from the sequence encoded by exon 2 (amino acid 17 in C β 1) to the C-terminus, while the N-termini varied both in length and sequence composition. The C β 2 splice variant contains a 63 amino acid sequence substituting the first 16 amino acids in C β 1, and is homologous to the previously identified bovine C β 2 [13]. Furthermore, the human C β 3 splice variant contains four amino acids in the N-terminal substituting the first 16 amino acids in C β 1, and is similar to the previously identified murine C β 3 [14]. The human C β 4 contains three amino acids substituting the first 16 amino acids in C β 1, and is similar to the murine C β 2 [14]. Finally,

the splice variants C β 4ab and C β 4abc contain 18 and 21 amino acids, respectively, that substitute the first 16 amino acids of C β 1. These splice variants show no homology to the N-terminus of any other C subunits identified thus far.

Example 3

[0057] Tissue Distribution of C β Splice Variants.

[0058] To examine the tissue distribution of C β splice variants, exon specific DNA probes and a DNA probe common to all C β splice variants were hybridized to two similar Northern blots containing RNA from various human tissues. For comparison the blots were hybridized to a cDNA encoding glycer-aldehyde 3-phosphate dehydrogenase (GAPDH). In **FIG. 4** (panel C β 1) we show that C β 1 is predominantly expressed in brain and kidney with low level expression in several other tissues as well. C β 2 is expressed at high levels in thymus, spleen and kidney in addition to a weak signal in other tissues (**FIG. 4**, panel C β 2). In contrast to C β 2 the exon 1-4 and exon a and b containing mRNAs appeared to be present exclusively in brain (**FIG. 4**, panels C β 4 and exon a+b). Finally, probing the Northern blot with a probe common to all the C β splice variants, we observed ubiquitous expression of C β with the strongest signal in brain and a somewhat weaker signal in spleen and thymus, when compared to the GAPDH signal (**FIG. 4**, panel C β common). Hybridization using a DNA fragment corresponding to the C β 3 specific cDNA resulted in an almost undetectable signal in the brain and no detectable signals in any other tissues (data not shown).

Example 4

[0059] The Human C β 2 Splice Variant is Not Present in the Mouse.

[0060] Previously we have identified three splice variants of C β in the mouse, C β 1, C β 2 and C β 3 [14]. Based on the present work, it is apparent that mouse C β 2 is not homologous to either bovine or the human C β 2. Instead, mouse C β 2 is homologous to what we now have designated human C β 4. Thus, we investigated whether a C β splice variant similar to human C β 2 was present in the mouse genome. A Zoo-blot containing genomic DNA isolated from human, monkey, rat, mouse, dog, cow, rabbit, chicken and yeast was hybridized using a DNA fragment corresponding to exon 1-2 of human C β . In **FIG. 5** (panel A, lanes 1 to 9) we show that a DNA fragment was detected using C β 2 specific probe in man, monkey, dog, cow, and rabbit. In contrast, the C β 2 specific probe did not recognize any fragments in the rat and mouse suggesting that the C β 2 specific exon is not present in the murine genome. To further substantiate this observation we isolated total RNA from human, wild type mice and mice that are ablated (knockout, KO) for exon 1A of the C β gene [16]. The RNA was isolated from immune tissues and brain since we observed high level expression of C β 2 in human thymus, spleen and peripheral blood leukocytes and high level of the other C β splice variants in the brain (**FIG. 4**). The Northern blots were probed with a C β cDNA probe (expected to recognize all known C β splice variants) and a C β 2 specific probe (see material and methods). In **FIG. 5B** (upper panel) we demonstrate that C β is present in the brain of wild type and C β exon 1 KO (lanes 1 and 2) and in human peripheral blood leukocytes (lane 5). The mouse spleen did not contain C β mRNA (lanes 3 and 4). When probing the

same filter with the C β 2 specific probe (**FIG. 5**, lower panel) C β 2 message was only detected in human peripheral blood leukocytes (lane 5) whereas all the mouse tissues were negative for C β 2 mRNA (lanes 1 to 4).

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<210> SEQ ID NO 9

<211> LENGTH: 2944

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 9

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<210> SEQ ID NO 10

<211> LENGTH: 2973

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 10

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<210> SEQ ID NO 11

<211> LENGTH: 3017

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 11

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<211> LENGTH: 3031

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 12

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agtgccttac cttaaaacaa cacacgagaa gagttaaact ggggtcattt taatgatcaa 2760
tttacctgca tataaaattt atttttaatc aagctgatct taatgtatat aatcatttca 2820
tttgctttat tatcggtgca ggtaggatc taacaccact tcttttcac tgtaccacac 2880
cctggtgaaa cctttgaaga cataaaaaaa acctgtctga gatgttcttt ctaccaatct 2940
atatgtcttt cggttatcaa gtgtttctgc atgtaaatgt catgtaaatg ctgatattga 3000
tttcaactgt ccatctatat ttaaaactgt c 3031

```

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<210> SEQ ID NO 13
<211> LENGTH: 400
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

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<400> SEQUENCE: 13

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Met Ala Ala Tyr Arg Glu Pro Pro Cys Asn Gln Tyr Thr Gly Thr Thr
  1           5           10           15
Thr Ala Leu Gln Lys Leu Glu Gly Phe Ala Ser Arg Leu Phe His Arg
  20           25           30
His Ser Lys Gly Thr Ala His Asp Gln Lys Thr Ala Leu Glu Asn Asp
  35           40           45
Ser Leu His Phe Ser Glu His Thr Ala Leu Trp Asp Arg Ser Met Lys
  50           55           60

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Glu Phe Leu Ala Lys Ala Lys Glu Asp Phe Leu Lys Lys Trp Glu Ser
 65              70              75              80
Pro Ala Gln Asn Thr Ala His Leu Asp Gln Phe Glu Arg Ile Lys Thr
              85              90              95
Leu Gly Thr Gly Ser Phe Gly Arg Val Met Leu Val Lys His Lys Glu
              100              105              110
Thr Gly Asn His Tyr Ala Met Lys Ile Leu Asp Lys Gln Lys Val Val
              115              120              125
Lys Leu Lys Gln Ile Glu His Thr Leu Asn Glu Lys Arg Ile Leu Gln
              130              135              140
Ala Val Asn Phe Pro Phe Leu Val Lys Leu Glu Phe Ser Phe Lys Asp
              145              150              155              160
Asn Ser Asn Leu Tyr Met Val Met Glu Tyr Val Pro Gly Gly Glu Met
              165              170              175
Phe Ser His Leu Arg Arg Ile Gly Arg Phe Ser Glu Pro His Ala Arg
              180              185              190
Phe Tyr Ala Ala Gln Ile Val Leu Thr Phe Glu Tyr Leu His Ser Leu
              195              200              205
Asp Leu Ile Tyr Arg Asp Leu Leu Lys Pro Glu Asn Leu Leu Ile Asp
              210              215              220
Gln Gln Gly Tyr Ile Gln Val Thr Asp Phe Gly Phe Ala Lys Arg Val
              225              230              235              240
Lys Gly Arg Thr Trp Thr Leu Cys Gly Thr Pro Glu Tyr Leu Ala Pro
              245              250              255
Glu Ile Ile Leu Ser Lys Gly Tyr Asn Lys Ala Val Asp Trp Trp Ala
              260              265              270
Leu Gly Val Leu Ile Tyr Glu Met Ala Ala Gly Tyr Pro Pro Phe Phe
              275              280              285
Ala Asp Gln Pro Ile Gln Ile Tyr Glu Lys Ile Val Ser Gly Lys Val
              290              295              300
Arg Phe Pro Ser Ser His Phe Ser Ser Asp Leu Lys Asp Leu Leu Arg
              305              310              315              320
Asn Leu Leu Gln Val Asp Leu Thr Lys Arg Phe Gly Asn Leu Lys Asn
              325              330              335
Gly Val Asn Asp Ile Lys Asn His Lys Trp Phe Ala Thr Thr Asp Trp
              340              345              350
Ile Ala Ile Tyr Gln Arg Lys Val Glu Ala Pro Phe Ile Pro Lys Phe
              355              360              365
Lys Gly Pro Gly Asp Thr Ser Asn Phe Asp Asp Tyr Glu Glu Glu Glu
              370              375              380
Ile Arg Val Ser Ile Asn Glu Lys Cys Gly Lys Glu Phe Ser Glu Phe
              385              390              395              400

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<210> SEQ ID NO 14
<211> LENGTH: 357
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

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<400> SEQUENCE: 14

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Met Ser Ala Arg Lys Ser Ser Asp Ala Ser Ala Cys Ser Ser Ser Glu
  1              5              10              15
Ile Ser Val Met Lys Glu Phe Leu Ala Lys Ala Lys Glu Asp Phe Leu
              20              25              30

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-continued

Lys Lys Trp Glu Ser Pro Ala Gln Asn Thr Ala His Leu Asp Gln Phe
 35 40 45
 Glu Arg Ile Lys Thr Leu Gly Thr Gly Ser Phe Gly Arg Val Met Leu
 50 55 60
 Val Lys His Lys Glu Thr Gly Asn His Tyr Ala Met Lys Ile Leu Asp
 65 70 75 80
 Lys Gln Lys Val Val Lys Leu Lys Gln Ile Glu His Thr Leu Asn Glu
 85 90 95
 Lys Arg Ile Leu Gln Ala Val Asn Phe Pro Phe Leu Val Lys Leu Glu
 100 105 110
 Phe Ser Phe Lys Asp Asn Ser Asn Leu Tyr Met Val Met Glu Tyr Val
 115 120 125
 Pro Gly Gly Glu Met Phe Ser His Leu Arg Arg Ile Gly Arg Phe Ser
 130 135 140
 Glu Pro His Ala Arg Phe Tyr Ala Ala Gln Ile Val Leu Thr Phe Glu
 145 150 155 160
 Tyr Leu His Ser Leu Asp Leu Ile Tyr Arg Asp Leu Leu Lys Pro Glu
 165 170 175
 Asn Leu Leu Ile Asp Gln Gln Gly Tyr Ile Gln Val Thr Asp Phe Gly
 180 185 190
 Phe Ala Lys Arg Val Lys Gly Arg Thr Trp Thr Leu Cys Gly Thr Pro
 195 200 205
 Glu Tyr Leu Ala Pro Glu Ile Ile Leu Ser Lys Gly Tyr Asn Lys Ala
 210 215 220
 Val Asp Trp Trp Ala Leu Gly Val Leu Ile Tyr Glu Met Ala Ala Gly
 225 230 235 240
 Tyr Pro Pro Phe Phe Ala Asp Gln Pro Ile Gln Ile Tyr Glu Lys Ile
 245 250 255
 Val Ser Gly Lys Val Arg Phe Pro Ser Ser His Phe Ser Ser Asp Leu
 260 265 270
 Lys Asp Leu Leu Arg Asn Leu Leu Gln Val Asp Leu Thr Lys Arg Phe
 275 280 285
 Gly Asn Leu Lys Asn Gly Val Asn Asp Ile Lys Asn His Lys Trp Phe
 290 295 300
 Ala Thr Thr Asp Trp Ile Ala Ile Tyr Gln Arg Lys Val Glu Ala Pro
 305 310 315 320
 Phe Ile Pro Lys Phe Lys Gly Pro Gly Asp Thr Ser Asn Phe Asp Asp
 325 330 335
 Tyr Glu Glu Glu Glu Ile Arg Val Ser Ile Asn Glu Lys Cys Gly Lys
 340 345 350
 Glu Phe Ser Glu Phe
 355

<210> SEQ ID NO 15
 <211> LENGTH: 360
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens

<400> SEQUENCE: 15

Met Ser Ala Arg Lys Ser Ser Asp Ala Ser Ala Cys Ser Ser Ser Glu
 1 5 10 15
 Ile Ser Asp Ser Phe Val Met Lys Glu Phe Leu Ala Lys Ala Lys Glu

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Lys Glu Phe Leu Ala Lys Ala Lys Glu Asp Phe Leu Lys Lys Trp Glu
 20 25 30

Asn

<210> SEQ ID NO 17
 <211> LENGTH: 80
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens

<400> SEQUENCE: 17

Met Ala Ala Tyr Arg Glu Pro Pro Cys Asn Gln Tyr Thr Gly Thr Thr
 1 5 10 15

Thr Ala Leu Gln Lys Leu Glu Gly Phe Ala Ser Arg Leu Phe His Arg
 20 25 30

His Ser Lys Gly Thr Ala His Asp Gln Lys Thr Ala Leu Glu Asn Asp
 35 40 45

Ser Leu His Phe Ser Glu His Thr Ala Leu Trp Asp Arg Ser Met Lys
 50 55 60

Glu Phe Leu Ala Lys Ala Lys Glu Asp Phe Leu Lys Lys Trp Glu Asn
 65 70 75 80

<210> SEQ ID NO 18
 <211> LENGTH: 21
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens

<400> SEQUENCE: 18

Met Gly Leu Leu Lys Glu Phe Leu Ala Lys Ala Lys Glu Asp Phe Leu
 1 5 10 15

Lys Lys Trp Glu Asn
 20

<210> SEQ ID NO 19
 <211> LENGTH: 20
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens

<400> SEQUENCE: 19

Met Ser Val Lys Glu Phe Leu Ala Lys Ala Lys Glu Asp Phe Leu Lys
 1 5 10 15

Lys Trp Glu Asn
 20

<210> SEQ ID NO 20
 <211> LENGTH: 36
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens

<400> SEQUENCE: 20

Met Ser Ala Arg Lys Ser Ser Asp Ala Ser Ala Cys Ser Ser Ser Glu
 1 5 10 15

Ile Ser Val Lys Glu Phe Leu Ala Lys Ala Lys Glu Asp Phe Leu Lys
 20 25 30

Lys Trp Glu Asn
 35

<210> SEQ ID NO 21
 <211> LENGTH: 39

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<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 21

Met Ser Ala Arg Lys Ser Ser Asp Ala Ser Ala Cys Ser Ser Ser Glu
 1             5             10             15

Ile Ser Asp Ser Phe Val Lys Glu Phe Leu Ala Lys Ala Lys Glu Asp
             20             25             30

Phe Leu Lys Lys Trp Glu Asn
             35

<210> SEQ ID NO 22
<211> LENGTH: 4
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 22

Lys Lys Gly Ser
 1

<210> SEQ ID NO 23
<211> LENGTH: 4
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 23

Arg Lys Ser Ser
 1

<210> SEQ ID NO 24
<211> LENGTH: 30
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
      primer

<400> SEQUENCE: 24

caacccaaag agaagtaaga aagtgtcta                               30

<210> SEQ ID NO 25
<211> LENGTH: 30
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
      primer

<400> SEQUENCE: 25

ttggttggtc tgcaagaat ggggatagc                               30

<210> SEQ ID NO 26
<211> LENGTH: 29
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
      primer

<400> SEQUENCE: 26

ttttctcatt caaagtatgc tctatttgc                               29

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<210> SEQ ID NO 27
<211> LENGTH: 30
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic primer

<400> SEQUENCE: 27
agaataatgc cggacttgaa gattttgaaa 30

<210> SEQ ID NO 28
<211> LENGTH: 27
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic primer

<400> SEQUENCE: 28
gttttagtgc aatcattctg ctgtttg 27

<210> SEQ ID NO 29
<211> LENGTH: 27
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic primer

<400> SEQUENCE: 29
aaaaagtctt ctttgcttt ggctaga 27

<210> SEQ ID NO 30
<211> LENGTH: 24
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic primer

<400> SEQUENCE: 30
tggcagctta tagagaacca cctt 24

<210> SEQ ID NO 31
<211> LENGTH: 22
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic primer

<400> SEQUENCE: 31
caatcccatg ttgaacctgg ca 22

<210> SEQ ID NO 32
<211> LENGTH: 24
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic primer

<400> SEQUENCE: 32
gctctccacc tcgctgcctt tctt 24

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<210> SEQ ID NO 33
<211> LENGTH: 24
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
primer

<400> SEQUENCE: 33

ccagccccc ttcccttccc tgac 24

<210> SEQ ID NO 34
<211> LENGTH: 24
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
primer

<400> SEQUENCE: 34

attgatctgt ccataaggca gtat 24

<210> SEQ ID NO 35
<211> LENGTH: 22
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
primer

<400> SEQUENCE: 35

tcacagctag cagtaagagc tg 22

<210> SEQ ID NO 36
<211> LENGTH: 22
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
primer

<400> SEQUENCE: 36

tctccagtgt gtgtgtttac ac 22

<210> SEQ ID NO 37
<211> LENGTH: 22
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
primer

<400> SEQUENCE: 37

atgatgaaaa ccaacctttc ca 22

<210> SEQ ID NO 38
<211> LENGTH: 46
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
primer

<400> SEQUENCE: 38

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gatatttctg aagaggagca agcagatgca tctgatgatt tgcgtg

46

<210> SEQ ID NO 39

<211> LENGTH: 46

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic primer

<400> SEQUENCE: 39

cacgcaaatc atcagatgca tctgcttgcct cctcttcaga aatata

46

1. The genomic DNA sequence encoding novel human catalytic subunits C β 2, C β 4ab and C β 4abc variants of c-AMP dependent protein kinase termed C β 3, comprising the nucleotide sequence of SEQ ID NO:1.

2. The cDNA sequence encoding novel human catalytic subunits C β 2, C β 4ab and C β 4abc variants of cAMP dependent protein kinase termed C β , comprising the nucleotide sequences of SEQ ID NOS: 2, 5 and 6, respectively.

3. The vectors comprising the DNA sequences according to claims 1 or 2.

4. The specific amino acid sequences of SEQ ID NOS 7, 8 and 9 of C β 2, C β 4ab and C β 4abc, respectively.

5. A protein encoded by the nucleotide sequences according to claims 1 or 2.

6. A protein encoded by the specific DNA sequences according to claims 1 or 2 comprising the specific amino acid sequence of SEQ ID NOS: 7, 8 and 9.

7. A kit comprising C β 2 specific DNA probes of claims 1 or 2.

8. The use of the C β 2, C β 4ab and C β 4abc proteins of claims 1-6, for the preparation of pharmaceuticals.

9. The use of the C β 2 protein of claims 1-6, for the preparation of a medicament for inhibition of the enzymatic activity of C β 2.

10. The use of the C β 2 protein of claims 1-6, for the preparation of a medicament that will specifically and irreversibly block C β 2 interaction.

11. The use of the C β 2 protein of claims 1-6, for the preparation of a medicament that will activate the enzymatic activity of C β 2.

12. The use of the DNA sequences which is complementary to the C β 2, C β 4ab and C β 4abc DNA according to claims 1 or 2 for the preparation of an anti sense drug.

13. A method for inspection and screening of patient T cells for the presence and location of the C β 2 of claims 1-6, comprising:

- a) collecting and washing in buffer of isolated peripheral blood T lymphocytes;

b) preparing for identification of C β 2 protein by immunofluorescence, T cells are let to settle onto poly L-lysine coated cover slips following detergent-dependent lysis;

c) incubating with primary antibody (Ab), either irrelevant Ab or C β 2 specific Ab, Ab overshoot will be removed by washing buffer and T cells incubated with secondary anti-IgG Ab conjugated with a fluorescent; and

d) inspection of T cells under fluorescent microscopy.

14. A method of screening patient T cells for membrane associated of the C β 2 of claims 1-6, catalytic activity comprising:

a) collecting and washing in buffer of isolated peripheral blood T lymphocytes;

b) preparing of T cells by lysing in detergent buffer;

c) monitoring C β 2 specific catalytic activity by established assay, C β 1 activity is used as an internal control to determine relative activity.

15. A method for screening of patients for mutations in the C β 2 gene of claims 1 or 2 and mRNA comprising:

a) collecting and washing in buffer of isolated peripheral blood T lymphocytes;

b) isolating of total RNA and genomic DNA according to established methods followed by RT-PCR using C β 2 specific primers according to cDNA sequence of C β 2 specific nucleotides or the C β 2 specific exon, designated exon 1-2.

16. A product produced by the method according to claims 13, 14 and 15.

17. A test system for screening for inhibitory- or activating molecules of the C β 2 protein of claims 1-6.

18. The product from the screening method according to claim 17.

* * * * *

专利名称(译)	鉴定营养依赖性蛋白激酶的人催化亚基cbeta的新剪接变体及其用途		
公开(公告)号	US20040152626A1	公开(公告)日	2004-08-05
申请号	US10/168804	申请日	2000-12-22
[标]申请(专利权)人(译)	ORSTAVIK SIGURD REINTON NILS FRENGEN的Eirik LANGELAND BJORN TORE JAHNSEN TORE SKALHEGG BJORN小号		
申请(专利权)人(译)	ORSTAVIK SIGURD REINTON NILS FRENGEN的Eirik LANGELAND BJORN TORE JAHNSEN TORE SKALHEGG BJORN小号		
当前申请(专利权)人(译)	ORSTAVIK SIGURD REINTON NILS FRENGEN的Eirik LANGELAND BJORN TORE JAHNSEN TORE SKALHEGG BJORN小号		
[标]发明人	ORSTAVIK SIGURD REINTON NILS FRENGEN EIRIK LANGELAND BJORN TORE JAHNSEN TORE SKALHEGG BJORN S		
发明人	ORSTAVIK, SIGURD REINTON, NILS FRENGEN, EIRIK LANGELAND, BJORN TORE JAHNSEN, TORE SKALHEGG, BJORN S		
IPC分类号	G01N33/48 A61K31/7088 A61K38/00 A61K38/45 A61K45/00 A61K48/00 A61P37/04 A61P43/00 C12N9/00 C12N9/12 C12N9/99 C12N15/09 C12Q1/02 C12Q1/48 C12Q1/68 G01N21/78 G01N33/15 G01N33/50 G01N33/53 G01N33/569 C07H21/04		
CPC分类号	A61K38/00 C12N9/1205 G01N33/56972 C12Y207/11011 C12Q1/485		
优先权	19996423 1999-12-23 NO		
外部链接	Espacenet USPTO		

摘要(译)

Cbeta基因编码至少6种不同的基因产物，称为Cbeta1，Cbeta2，Cbeta3，Cbeta4，Cbeta4ab和Cbeta4abc。与鼠和牛剪接变体的情况一样，所有人Cbeta剪接变体在由外显子2编码的部分之前的N末端部分中变化。在人类中鉴定了在小鼠和牛中鉴定的所有

Cbeta剪接变体的同源物 (Cbeta1)。 , Cbeta2 , Cbeta3和Cbeta4) 以及之前尚未在任何其他物种中鉴定的两种新型Cbeta剪接变体 (Cbeta4ab和Cbeta4abc)。基因组DNA和cDNA序列编码剪接变体，并包括分别在SEQ ID NO : 1,2,3,4,5和6中显示的核苷酸序列。蛋白质是Cbeta蛋白质的新剪接变体。

