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Maruyama et al.

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(54) **DETECTION OF INFLAMMATORY DISEASE AND COMPOSITION FOR PREVENTION OR TREATMENT OF INFLAMMATORY DISEASE**

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C07K 14/705 (2006.01)

(52) **U.S. Cl.** **435/7.1; 435/7.92; 530/350**

(58) **Field of Classification Search** None
See application file for complete search history.

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(57) **ABSTRACT**

A novel method for detection of an inflammatory disease and a novel composition for prevention or treatment of an inflammatory disease are provided. The method for detection of an inflammatory disease comprises using RANKL and/or OPG as a marker in a biological sample. The composition for prevention or treatment of an inflammatory disease comprises RANKL and/or M-CSF as an active ingredient.

4 Claims, 24 Drawing Sheets

Fig. 1

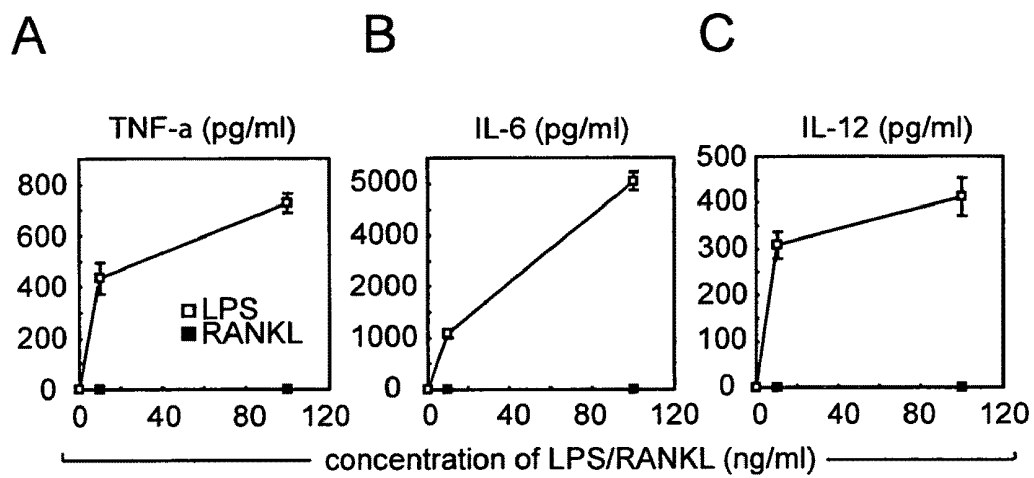


Fig. 2

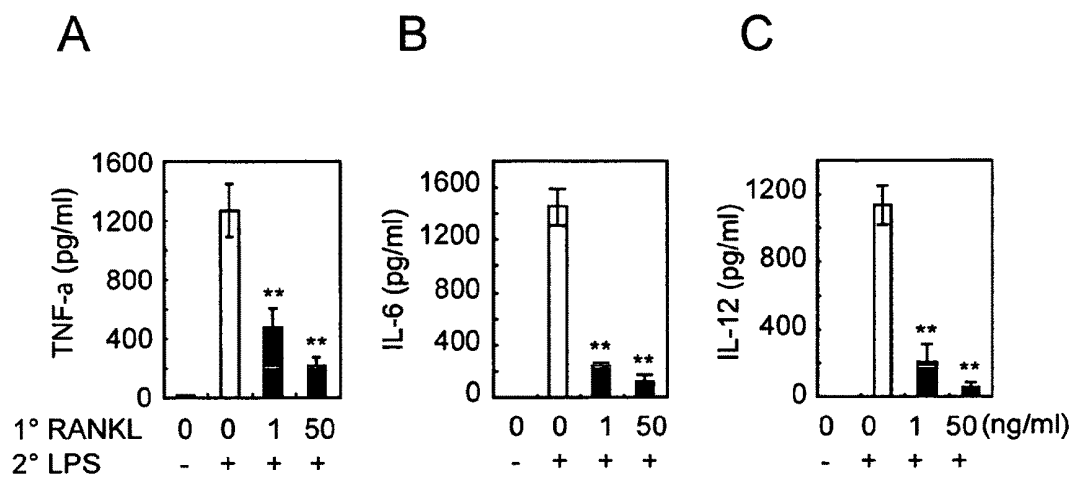


Fig. 3

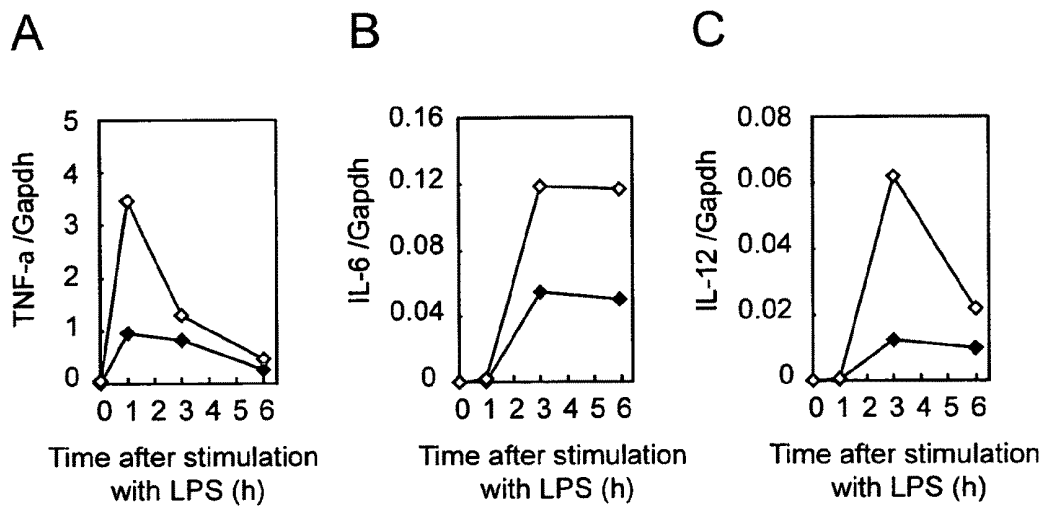


Fig. 4

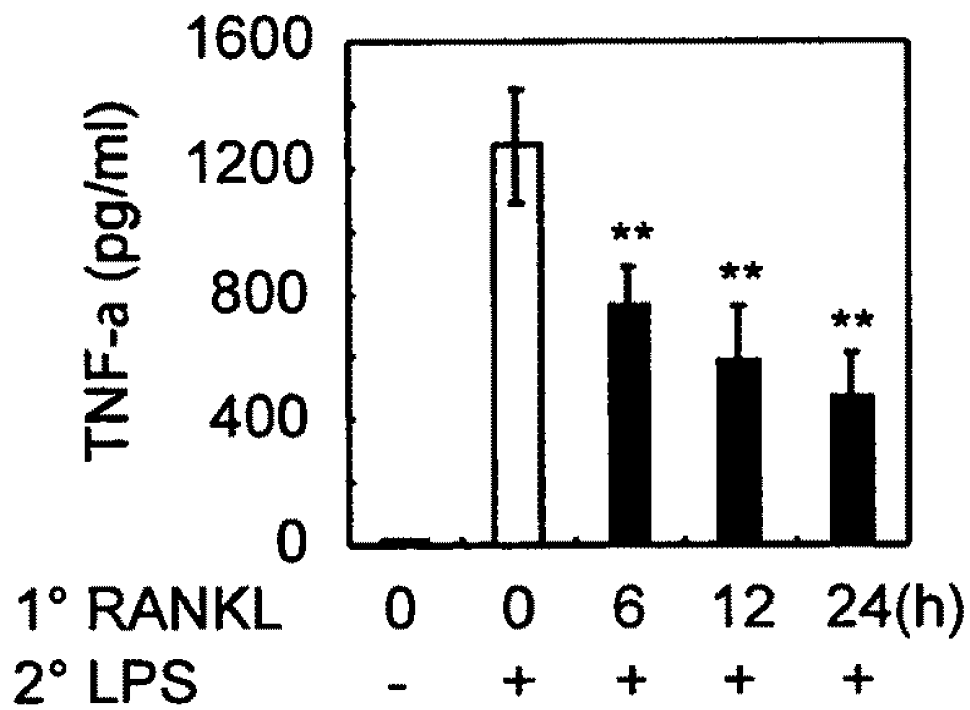


Fig. 5

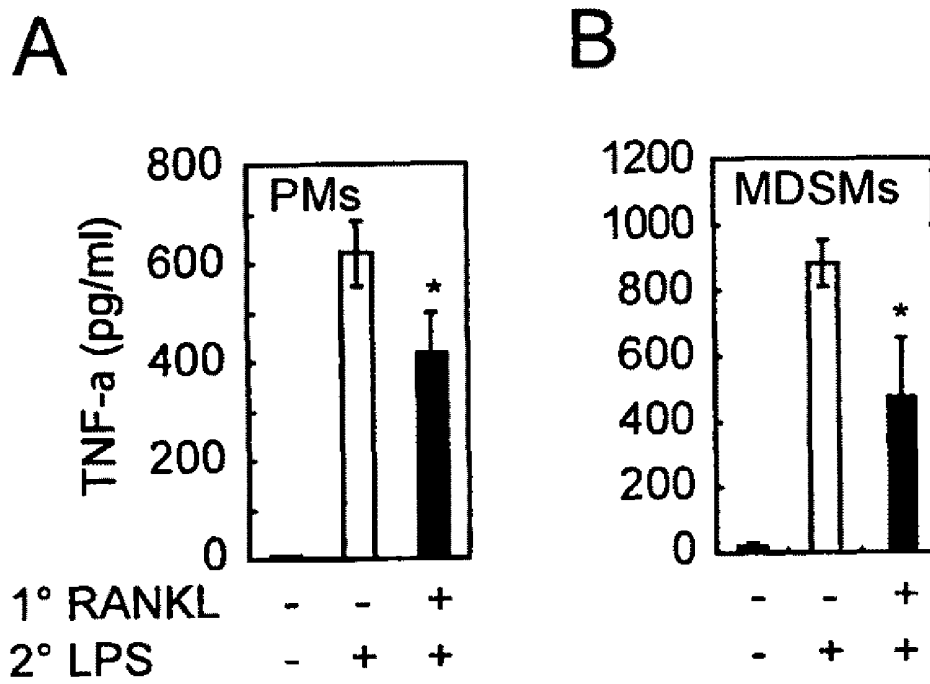


Fig. 6

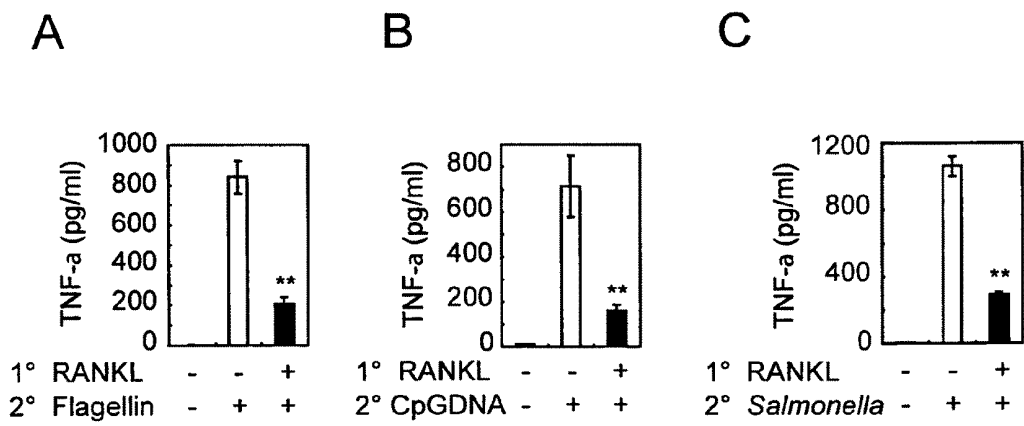


Fig. 7

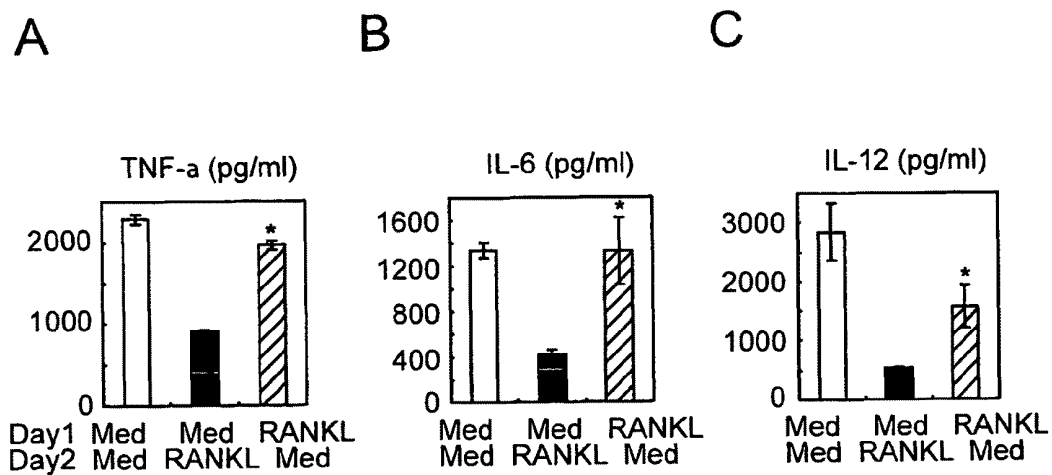


Fig. 8

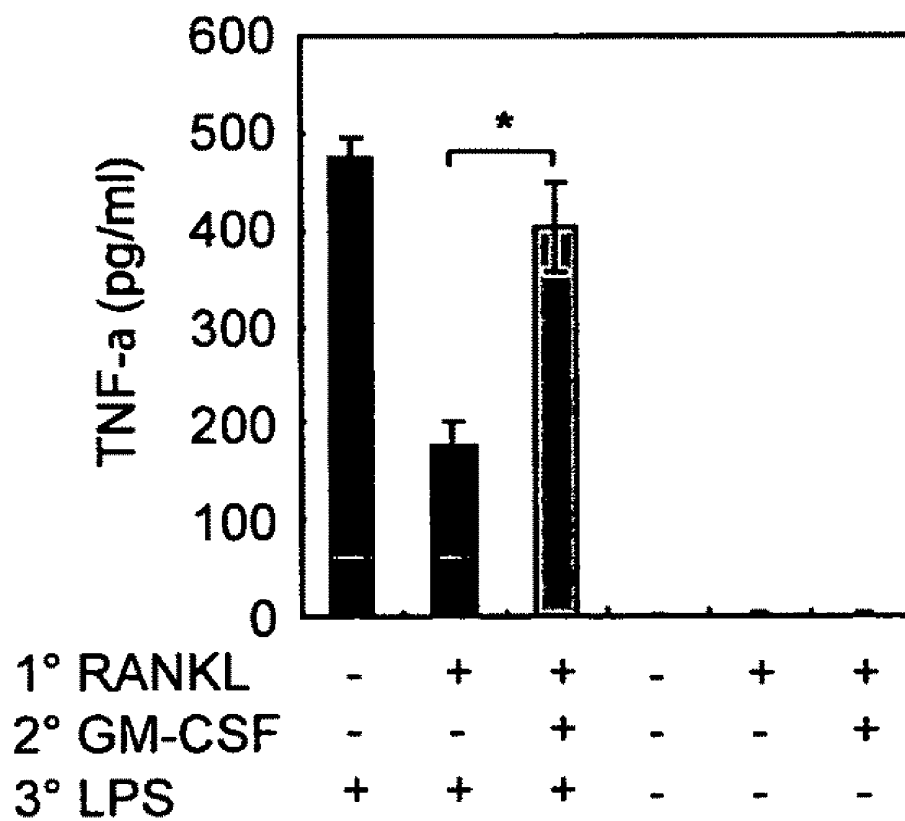
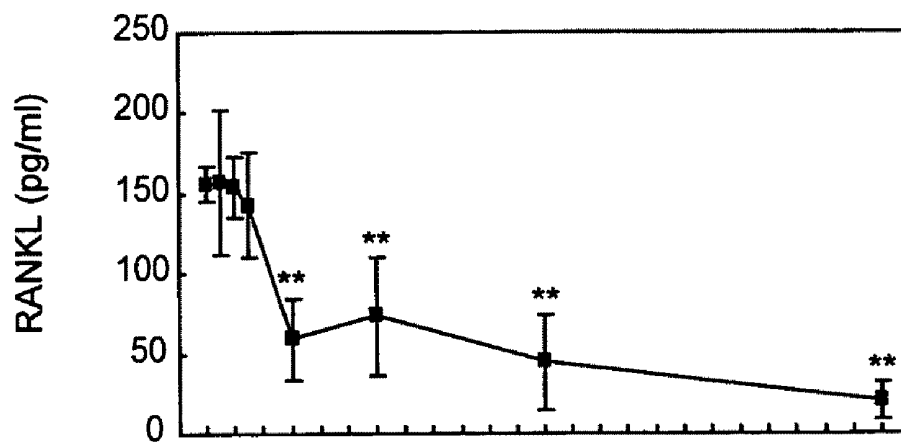


Fig. 9

A



B

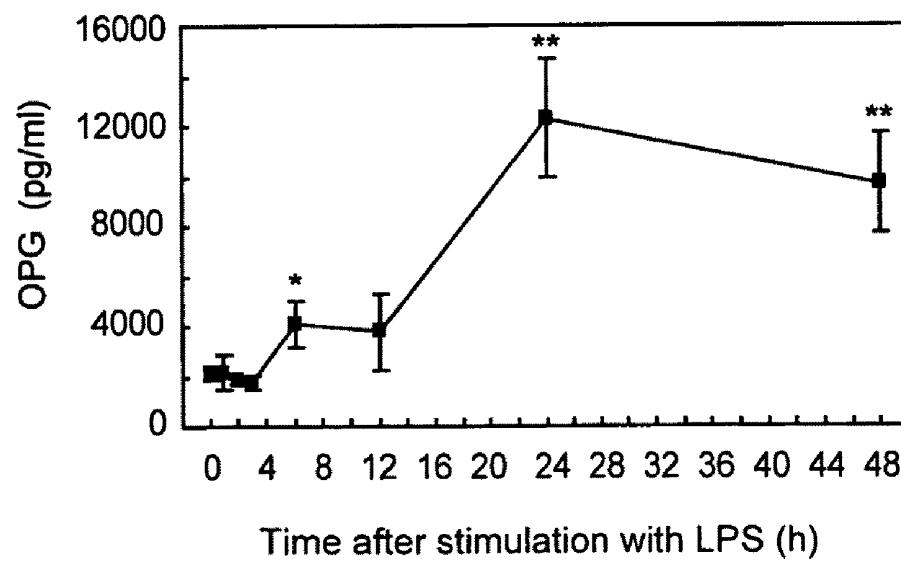
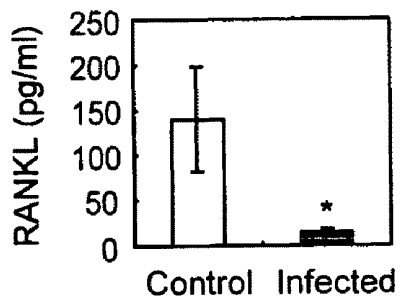


Fig. 10

A



B

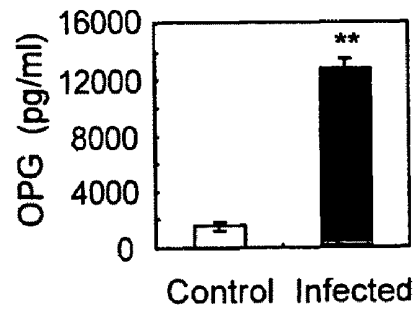
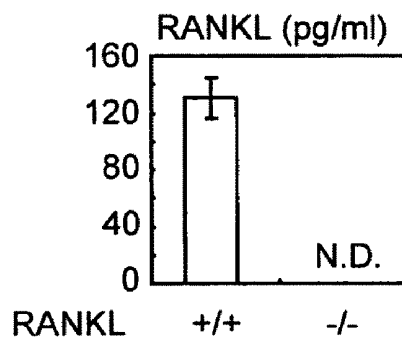


Fig. 11

A



B

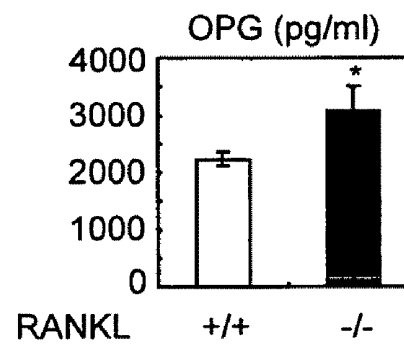


Fig. 12

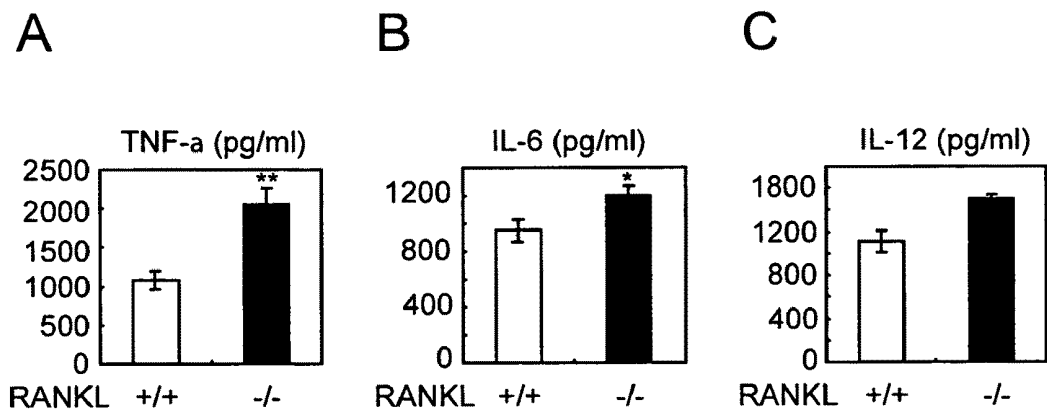
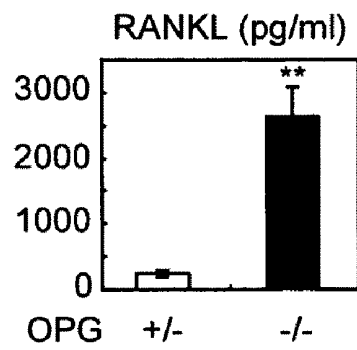


Fig. 13

A



B

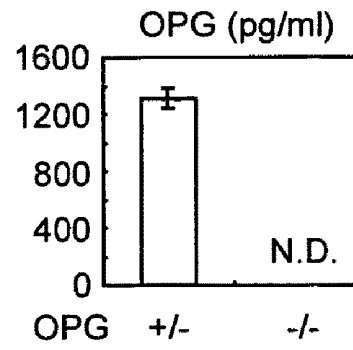


Fig. 14

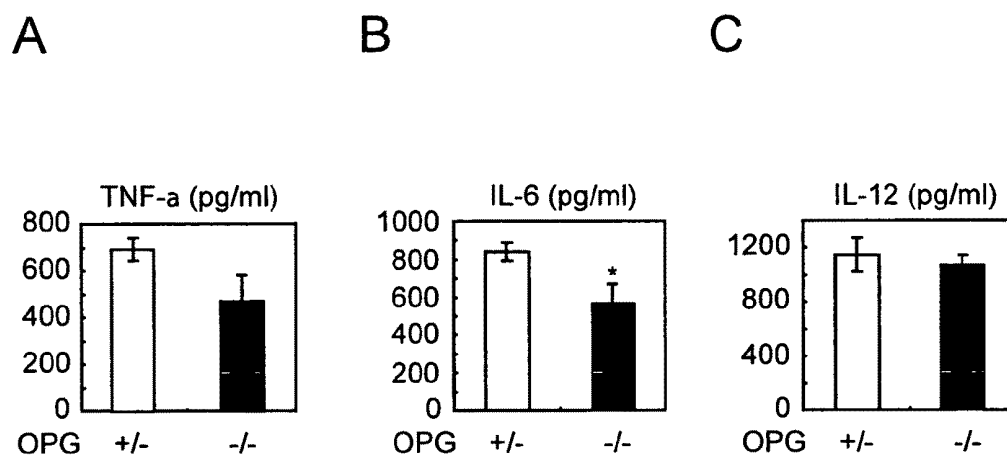


Fig. 15

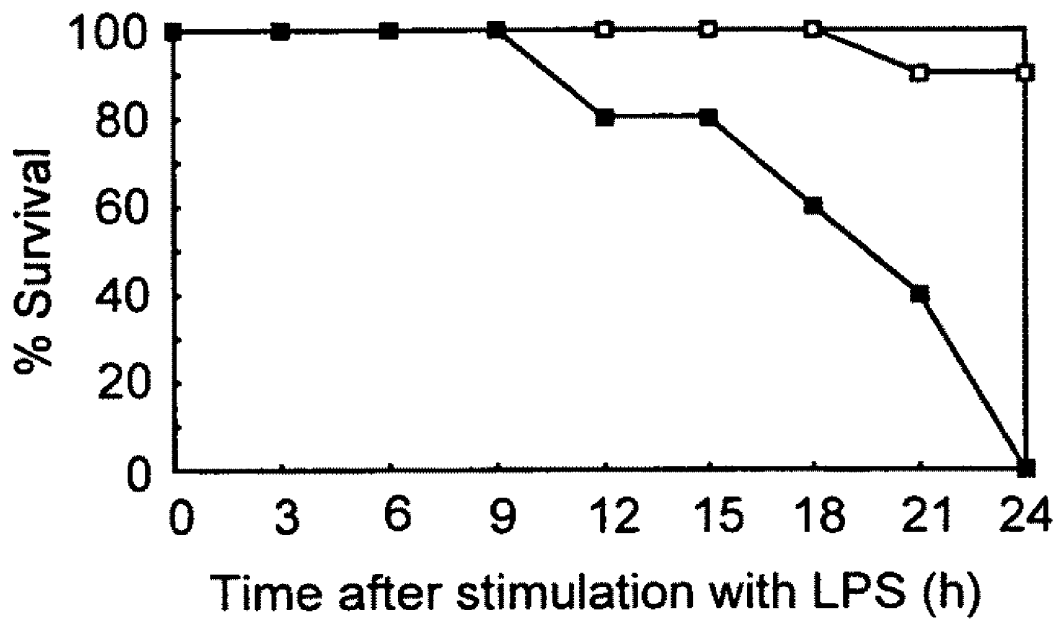
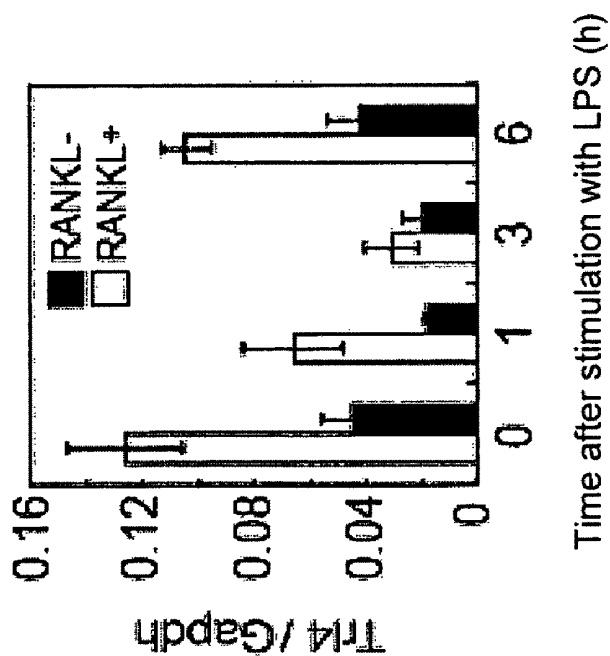


Fig. 16

A



B

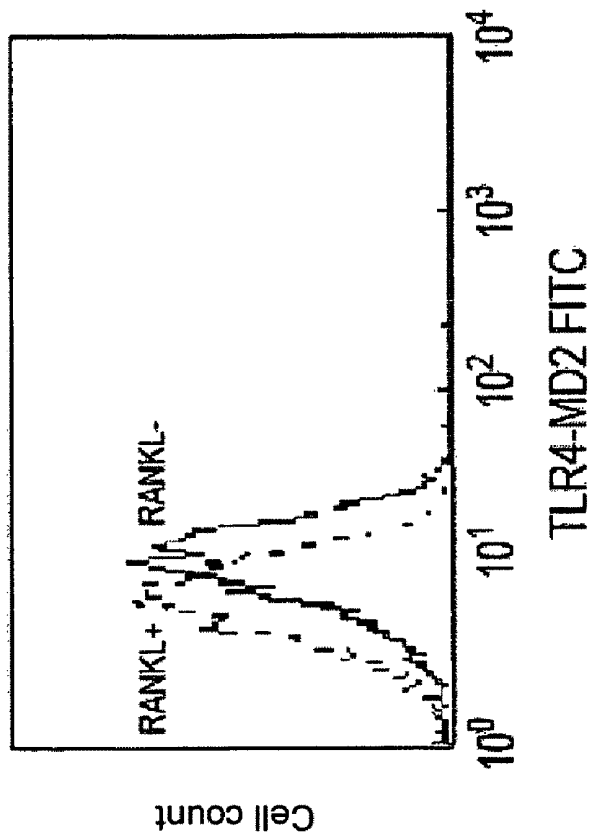


Fig. 17

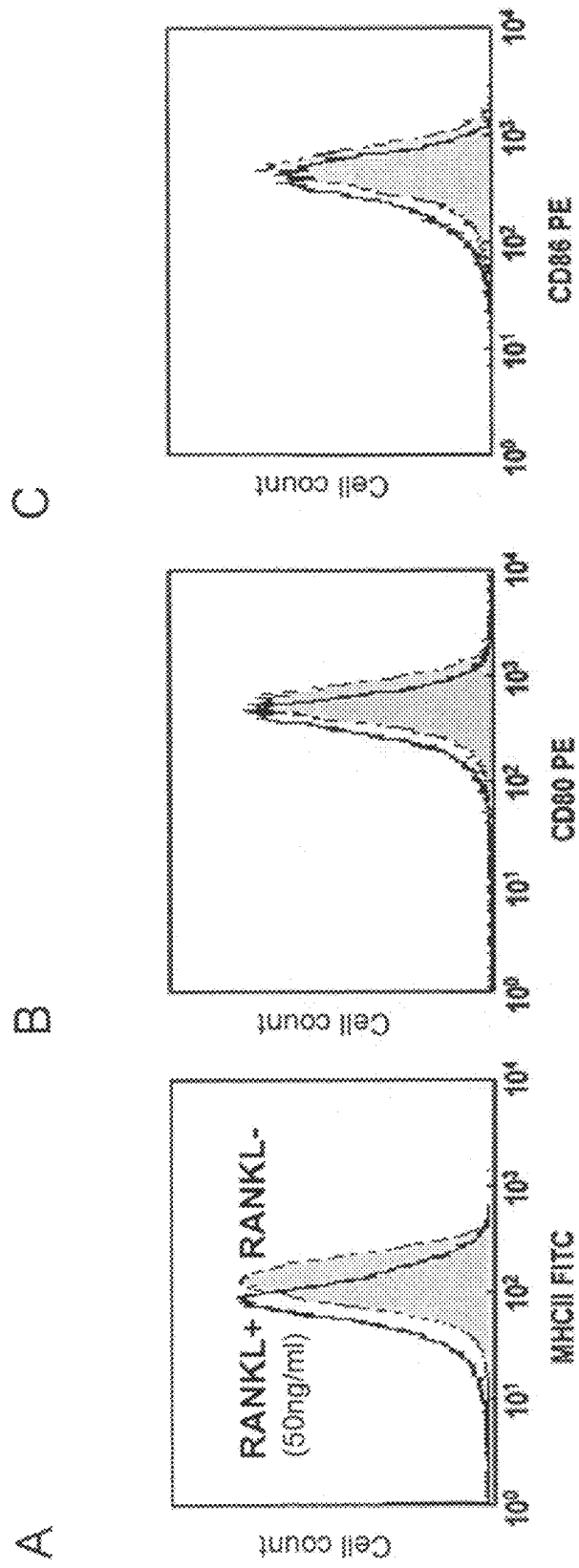


Fig. 18

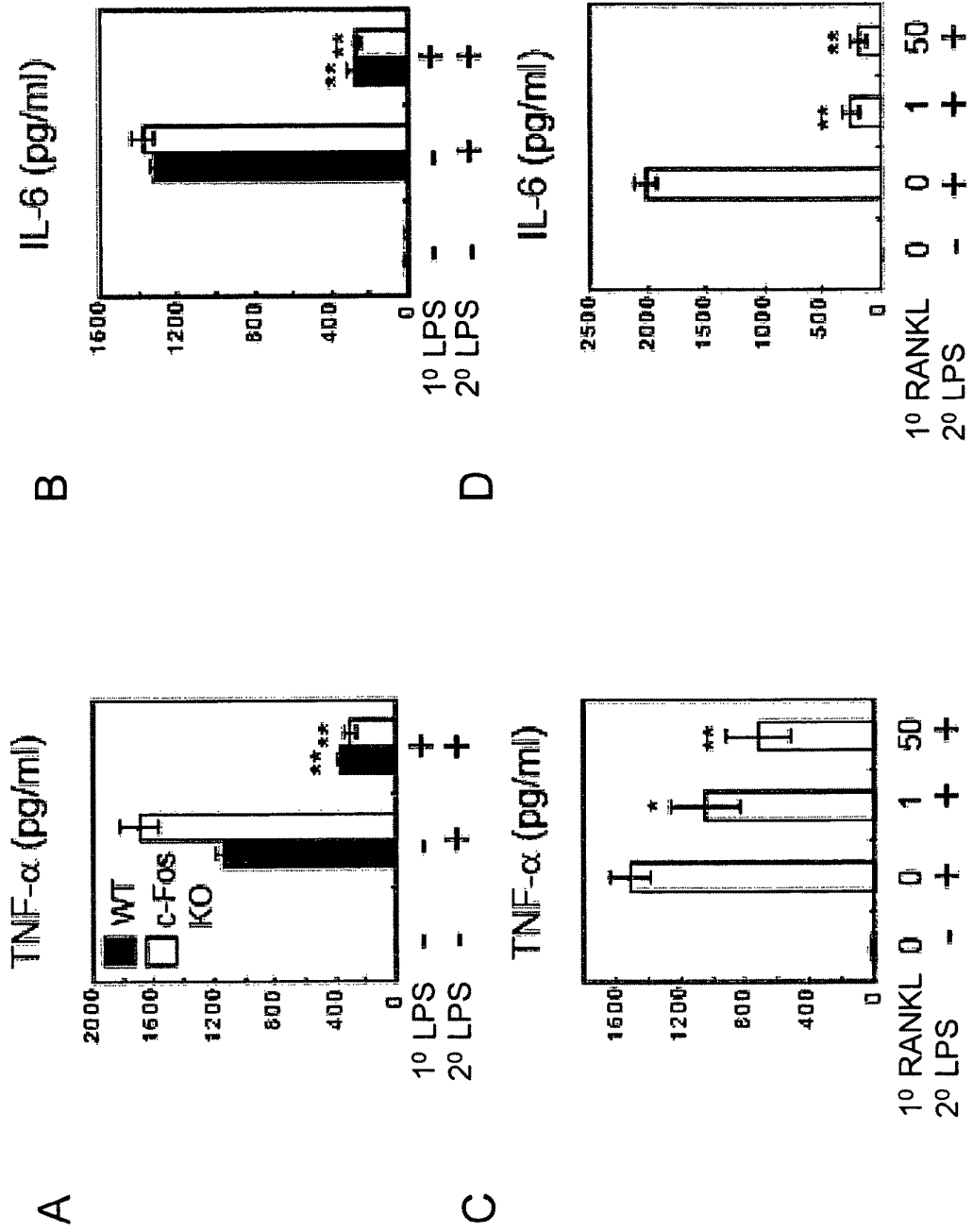


Fig. 19

10 20 30 40 50 60
gtcgactATCAGAGCAGAGAAAAGCGATGGTGGATGGCTCATGGTTAGATCTGGCCAAGAGGAGCAAG
SaII I R A E K A M V D G S W L D L A K R S K

70 80 90 100 110 120
CTTGAAGCTCAGCCTTTTGCTCATCTCACTATTAATGCCACCGACATCCCATCTGGTTCC
L E A Q P F A H L T I N A T D I P S G S

130 140 150 160 170 180
CATAAAGTGAGTCTGTCCTCTTGGTACCATGATCGGGGTTGGGCCAAGATCTCCAACATG
H K V S L S S W Y H D R G W A K I S N M

190 200 210 220 230 240
ACTTTTAGCAATGGAAAATAATAGTTAATCAGGATGGCTTTTATTACCTGTATGCCAAC
T F S N G K L I V N Q D G F Y Y L Y A N

250 260 270 280 290 300
ATTTGCTTTCGACATCATGAACTTCAGGAGACCTAGCTACAGAGTATCTTCAACTAATG
I C F R H H E T S G D L A T E Y L Q L M

310 320 330 340 350 360
GTGTACGTCACTAAAACCAGCATCAAAATCCCAAGTTCTCATACCCTGATGAAAGGAGGA
V Y V T K T S I K I P S S H T L M K G G

370 380 390 400 410 420
AGCACCAAGTATTGGTCAGGGAATTCTGAATTCCATTTTATTCCATAAACGTTGGTGGA
S T K Y W S G N S E F H F Y S I N V G G

430 440 450 460 470 480
TTTTTTAAGTTACGGTCTGGAGAGGAAATCAGCATCGAGGTCTCCAACCCCTCCTTACTG
F F K L R S G E E I S I E V S N P S L L

490 500 510 520 530 540
GATCCGGATCAGGATGCAACATACTTTGGGGCTTTTAAAGTTCGAGATATAGATTGAGCC
D P D Q D A T Y F G A F K V R D I D * (SEQ ID NO: 10)

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NotI

Fig. 20

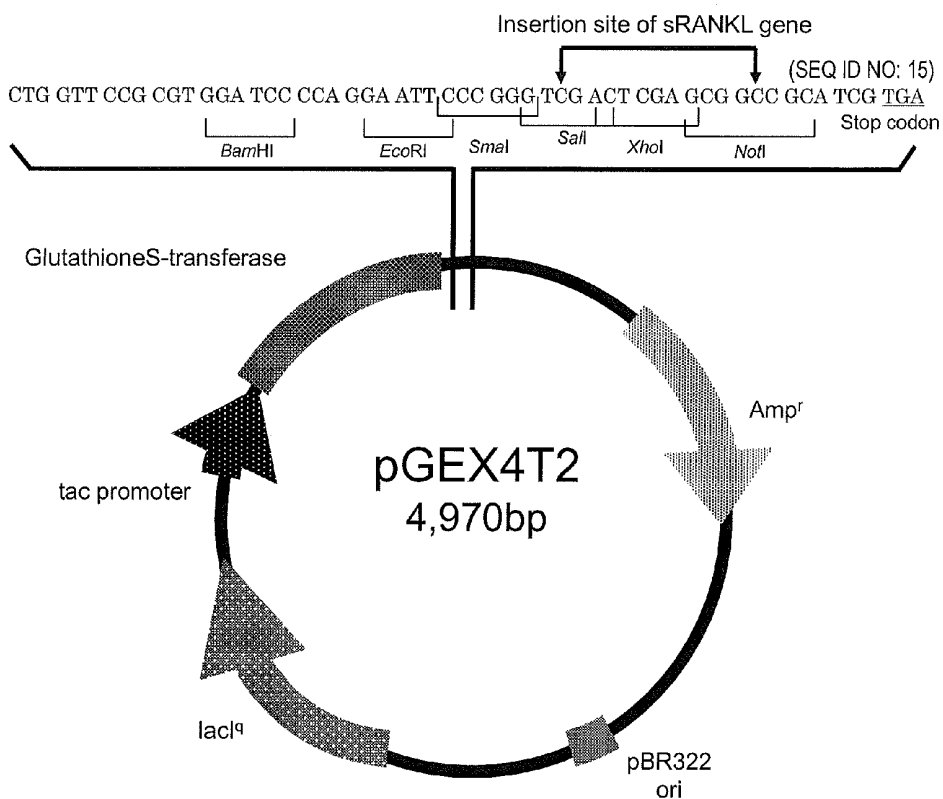


Fig. 21B

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2761 CGAACGACCT ACACCGAACT GAGATACCTA CAGCGTGAGC TATGAGAAAAG CGCCACGCTT
2821 CCCGAAGGGA GAAAGGCGGA CAGGTATCCG GTAAGCGGCA GGGTCGGAAC AGGAGAGCGC
2881 ACGAGGGAGC TTCCAGGGGG AAACGCCTGG TATCTTTATA GTCCTGTCGG GTTTCGCCAC
2941 CTCTGACTTG AGCGTCGATT TTTGTGATGC TCGTCAGGGG GCGGAGCCT ATGGA AAAAC
3001 GCCAGCAACG CGGCCCTTTT ACGGTTCCTG GCCTTTTGCT GGCCTTTTGC TCACATGTTT
3061 TTTCTGCGT TATCCCTGA TTTGTGGAT AACCGTATTA CCGCCTTTGA GTGAGCTGAT
3121 ACCGCTCGCC GCAGCCGAAC GACCGAGCGC AGCGAGTCAG TGAGCGAGGA AGCGGAAGAG
3181 CGCCTGATGC GGTATTTTCT CCTTACGCAT CTGTGCGGTA TTTCACACCG CATAAATTC
3241 GACACCATCG AATGGTGCAA AACCTTTCGC GGTATGGCAT GATAGCGCCC GGAAGAGAGT
3301 CAATTCAGGG TGGTGAATGT GAAACCAGTA ACGTTATACG ATGTCGCAGA GTATGCCGGT

lacIq initiation codon
3361 GTCTCTTATC AGACCGTTTC CCGCGTGGTG AACCAGGCCA GCCACGTTTC TGCGAAAACG
3421 CGGGA AAAAG TGGAAAGCGC GATGGCGGAG CTGAATTACA TTCCAACCG CGTGGCACAA
3481 CAACTGGCGG GCAAACAGTC GTTGTGATT GCGTTGCCA CCTCCAGTCT GGCCCTGCAC
3541 GCGCCGTCGC AAATTGTGCG GCGGATTA AA TCTCGCGCCG ATCAACTGGG TGCCAGCGTG
3601 GTGGTGTGCGA TGGTAGAACG AAGCGCGGTC GAAGCCTGTA AAGCGGCGGT GCACAACTCTT
3661 CTCGCGCAAC GCGTCAGTGG GCTGATCATT AACTATCCGC TGGATGACCA GGATGCCATT
3721 GCTGTGGAAG CTGCCTGCAC TAATGTTCCG GCGTTATTTT TTGATGTCTC TGACCAGACA
3781 CCCATCAACA GTATTATTTT CTCCCATGAA GACGGTACGC GACTGGGCGT GGAGCATCTG
3841 GTCGCATTGG GTCACCAGCA AATCGCGCTG TTAGCGGGCC CATTAAAGTTC TGTCTCGGGC
3901 CGTCTGCGTC TGGCTGGCTG GCATAAATAT CTCACTCGCA ATCAAATTCA GCCGATAGCG
3961 GAACGGGAAG GCGACTGGAG TGCCATGTCC GGTTTTCAAC AAACCATGCA AATGCTGAAT
4021 GAGGGCATCG TTCCCACTGC GATGCTGGTT GCCAACGATC AGATGGCGCT GGGCGCAATG
4081 CGCGCCATTA CCGAGTCCGG GCTGCGGTT GGTGCGGATA TCTCGGTAGT GGGATACGAC
4141 GATACCGAAG ACAGTCATG TTATATCCCG CCGTTAACCA CCATCAAACA GGATTTTCGC
4201 CTGCTGGGGC AAACGAGCGT GGACCGCTTG CTGCAACTCT CTCAGGGCCA GCGGTTGAAG
4261 GGCAATCAGC TGTTGCCCGT CTCACTGGTG AAAAGAAAAA CCACCCTGGC GCCCAATACG
4321 CAAACCGCCT CTCCCCGCGC GTTGGCCGAT TCATTAATGC AGCTGGCACG ACAGGTTTCC
4381 C GACTGGAAA GCGGGCAGTG AGCGCAACGC AATTAATGTG AGTTAGCTCA CTCATTAGGC

lacIq termination codon
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4501 ACAATTTTAC ACAGGAAACA GCTATGACCA TGATTACGGA TTCACTGGCC GTCGTTTTAC
4561 AACGTCGTGA CTGGGAAAAC CCTGGCGTTA CCCAACTTAA TCGCCTTGCA GCACATCCCC
4621 CTTTCGCCAG CTGGCGTAAT AGCGAAGAGG CCCGCACCGA TCGCCCTTCC CAACAGTTGC
4681 GCAGCCTGAA TGGCGAATGG CGCTTTGCCT GGTTCCTGGC ACCAGAAGCG GTGCCGGAAA
4741 GCTGGCTGGA GTGCGATCTT CCTGAGGCCG ATACTGTCTG CGTCCCCTCA AACTGGCAGA
4801 TGCACGGTTA CGATGCGCCC ATCTACACCA ACGTAACCTA TCCCATTACG GTC AATCCGC
4861 CGTTTGTTC CACGGAGAAT CCGACGGGTT GTTACTCGCT CACATTTAAT GTTGATGAAA
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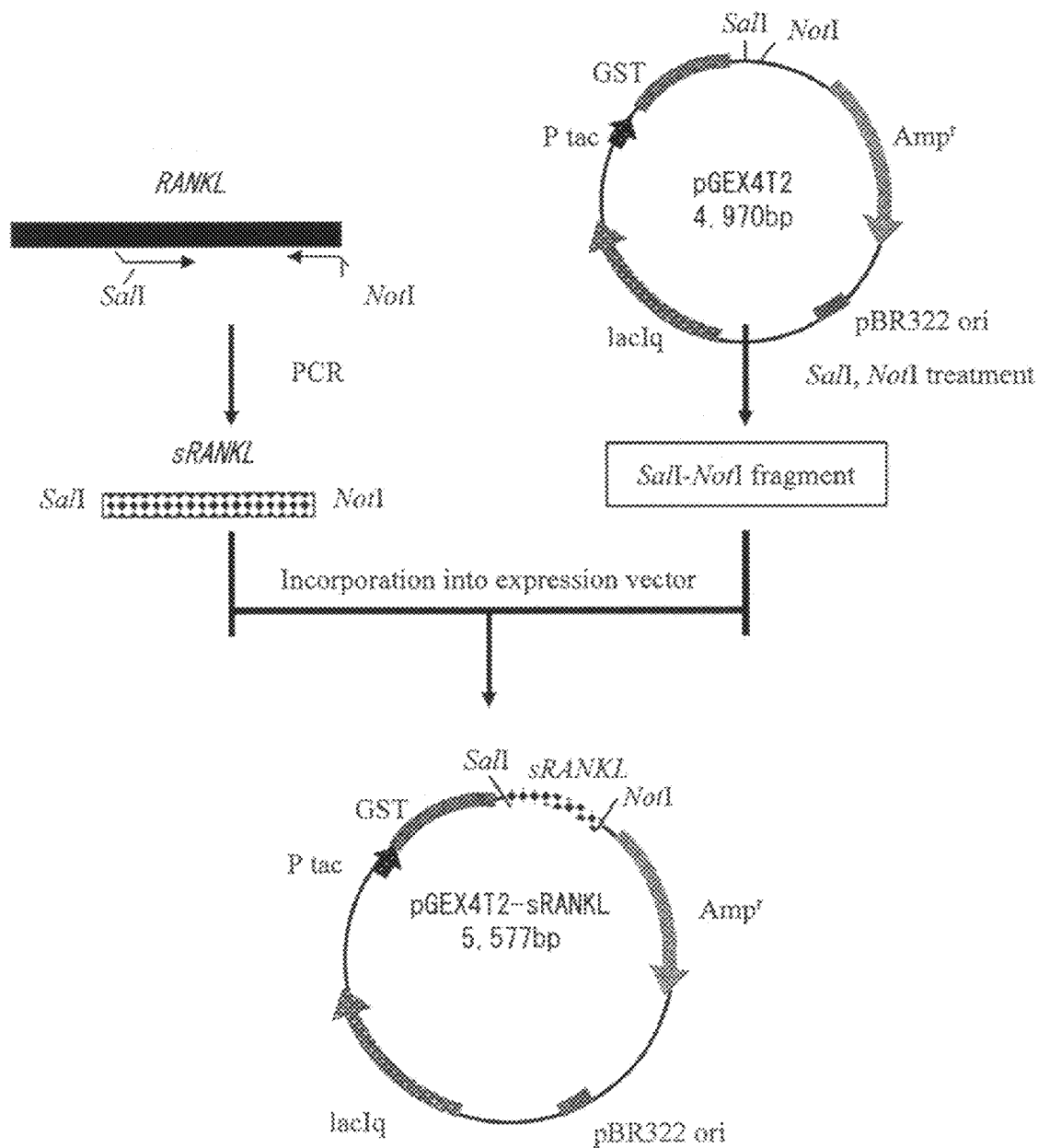
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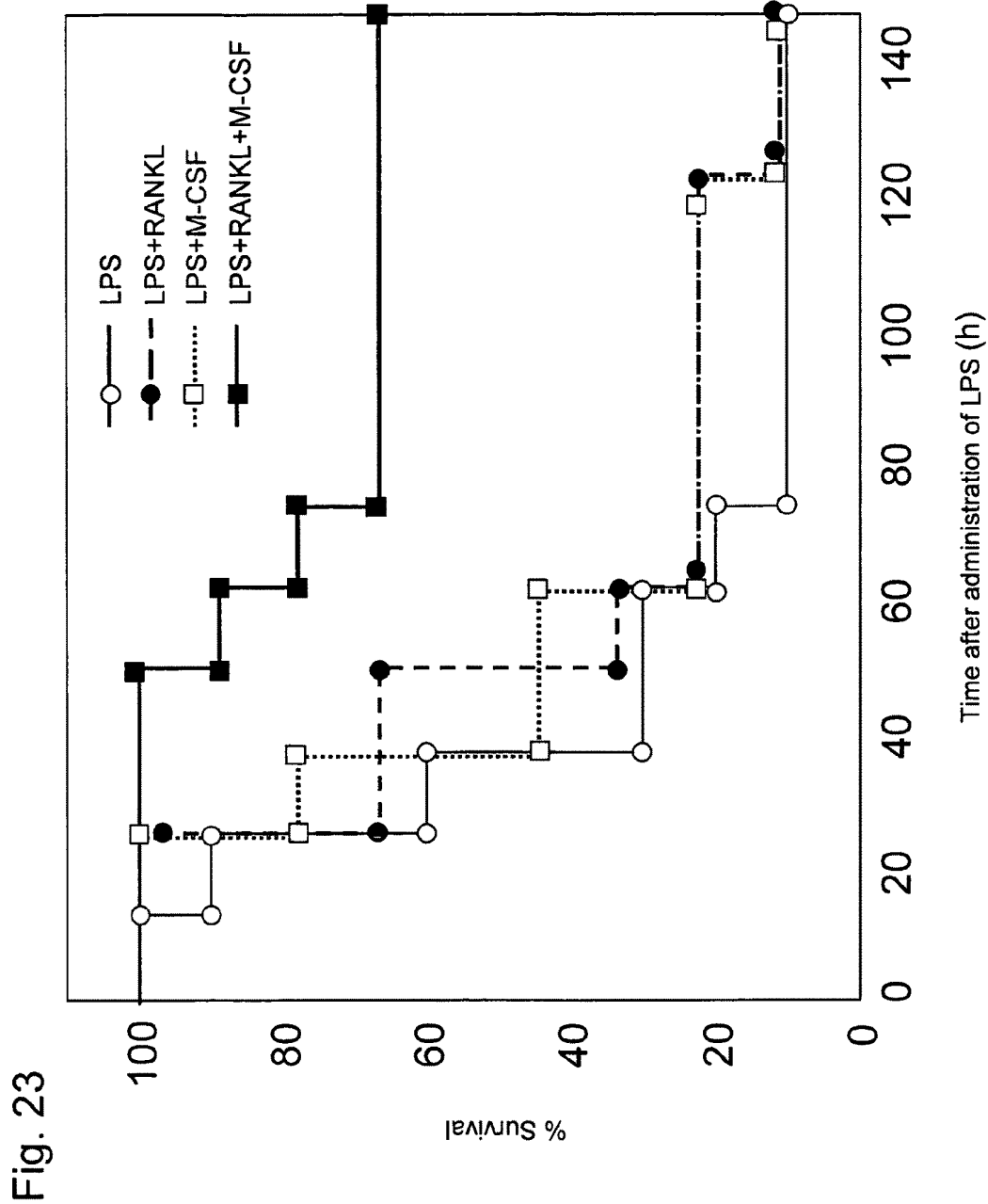
β -lactamase gene:1, 378-2, 238

lacIq gene:3, 319-4, 401

Fig. 22

Method for preparation of recombinant DNA
 Expression of sRANKL in *E. coli*





DETECTION OF INFLAMMATORY DISEASE AND COMPOSITION FOR PREVENTION OR TREATMENT OF INFLAMMATORY DISEASE

TECHNICAL FIELD

The present invention relates to a method for diagnosing or suppressing inflammatory diseases such as sepsis, allergies, and autoimmune diseases with the use of RANKL, which suppresses the production of inflammatory cytokines or improves the survival rate of an inflammatory animal model.

BACKGROUND ART

In recent years, it is known even in surgical fields that humoral factors produced at excessive levels play important roles in formation of pathological conditions during the perioperative period of significantly invasive surgery or the acute phase of a medical emergency such as an infectious disease. It is extremely important to measure various humoral factors in order to gain early, comprehensive knowledge of a pathology that changes every day or every second under such invasion and to apply the understanding clinically. Studies concerning various humoral factors in the process of shifts from sepsis to multiple organ dysfunction syndrome (MODS, by which the functions of a plurality of organs are damaged) have drastically progressed together with the development of molecular biological techniques. Recent studies on MODS have been improved to indicate a study approach that involves analyzing the mechanism of damage at the cellular level and microenvironment or humoral factors, so as to get closer to the pathology. Specifically, based on the understanding that MODS cases are extremely analogous to each other in terms of onset mechanism or pathology even if the causes of MODS cases differ (e.g., an MODS case due to sepsis), the control of factors involved in the shift to pathology that is developed much earlier before the onset of MODS has recently been emphasized. It can be said that clinicians and researchers are currently focusing on elucidation of the pathology of sepsis that occurs at the prestage of MODS and establishment of effective countermeasures against the pathology rather than treatment for MODS itself.

Humoral factors are useful as inflammatory markers. Specifically, CRP (C-reactive protein), TNF- α , IL-1, IL-6, IL-8, IL-10, MIP-1, HMGB-1, MIF, C5a, calcitonin, and the like are known (Marshall et al., *Crit Care Med* 31: 1560, 2003). It has been reported concerning CRP such that CRP is used in combination with the number of platelets or the like for evaluation of the prognosis of severe sepsis (Asayama et al., *Keio J Med* 47: 19 1998). It has also been reported that no significant differences are confirmed between sepsis and trauma (Endo et al., *Journal of Infection* 73: 197 1999). Hence, solid evaluation has not been established for CRP. Similarly, in the case of TNF- α or IL-6, while it has been reported that no significant differences have been confirmed between sepsis and trauma (Endo et al., *Journal of Infection* 73: 197 1999), it has also been reported that IL-6 is used as a prognosis marker for sepsis (Reinhart et al., *Crit Care Med* 29: 765, 2001). Thus, solid evaluation has not been established for TNF- α or IL-6. Moreover, it has been reported that IL-10 is effective (Ono et al., *Am J Surg* 188: 150, 2004), however, it cannot be said that the effects of IL-10 have been sufficiently verified through use. Involvement of MIF in acute respiratory distress syndrome (ARDS), bronchial asthma, or the like has been reported (Donnelly et al., *Nat Med* 3: 320, 1997; Rossi et al., *J Clin Invest* 101: 2869, 1998), but it is unknown whether or

not MIF can be used as a marker. It has not been revealed if the above-mentioned humoral factors cause sepsis or are produced as a result of sepsis.

Inflammatory reactions are biological reactions that limit the spreading of damage to a living body due to invasion and repair such damage. They occur as nonspecific reactions against all injuries or invasions. Inflammatory reactions are actually physiological biological reactions that take place in close association with neuroendocrine reactions, immunoinflammatory reactions, and coagulation-fibrolysis reactions. Inflammatory reactions are expressed locally in the forms of flare, swelling, pain, heat, and the like, and they cause systemic reactions with fever, tachycardia, tachypnea, and increased number of leukocytes when invasion is significant. Such conditions are referred to as systemic inflammatory response syndrome (SIRS). Examples thereof include SIRS not associated with infection or the like, but rather with trauma, burn, pancreatitis, and states after significantly invasive surgery, as well as SIRS associated with infection due to bacteria, fungi, parasites, viruses, or the like. In particular, SIRS caused by infection is referred to as sepsis.

Inflammatory reactions are established by vasodilation, vascular hyperpermeability, leukocyte-vascular endothelial cell activation, or the like. These reactions are induced by complements, amine, kinin, prostanoid, cytokine, and thrombin that are nonspecifically produced as invasion proceeds. Localized inflammatory reactions are induced by local noxious stimuli. However, when biological invasion is significant, systemic escape of these inflammatory mediators (and in particular, cytokine and thrombin) takes place and then systemic vasodilation, hyperpermeability, and leukocyte-vascular endothelial cell activation are observed. Inflammatory reaction has stages of receipt of noxious stimuli, reaction, and repairment. When a systemic inflammatory reaction is sustained, the reaction does not reach the repairment stage, so that biological homeostasis fails. In such case, it is known that MODS is induced to lead the living body to death.

For defense against invasion in the living body, three systems, the nervous system, the endocrine system, and the immune system, undergo reactions while closely interacting with each other. The nervous and endocrine systems are activated as invasion proceeds, resulting in enhanced energy metabolism, gluconeogenesis, increased cardiac output, and the like. Thus, inflammatory reactions are systemically enhanced. Meanwhile, cortisol is known to suppress the immune system and catecholamine is known to suppress the activity of NK cells or killer T cells, which are immunocytes. Bacterial infection or the occurrence of tissue damage activates the complement system or the blood coagulation system, along with which vascular endothelial cells are activated and phagocytic cells including monocytes, macrophages, and neutrophils migrate. Thus, inflammatory cytokines (composed mainly of TNF- α and IL-1) are freed from the damaged sites. With liberation of these inflammatory cytokines, protease or active oxygen, platelet-activating factors, and the like are also freed, forming the pathology of SIRS. Therefore, it is possible to consider that SIRS is also a pathological condition caused by hypercytokinemia (Riedemann et al., *Nat Med* 9: 517, 2003).

It has been reported that in the U.S. that about 750,000 persons are affected with sepsis yearly and 210,000 or more persons lose their lives due to sepsis (Wheeler et al., *N Engl J Med* 340: 207, 1999; Severansky et al., *Sepsis* 3: 11, 1999; Hotchkiss et al., *N Engl J Med* 348: 138, 2003). Furthermore, treatment for sepsis causes significant economic impact because of lengthy ICU hospital stays or increased amounts of resources used. However, although establishment of a

therapeutic method against sepsis that is also referred to as high inflammatory cytokinemia has been attempted as an emergent issue throughout the world, no therapeutic method currently exists by which reduction of sepsis fatalities can be realized (Vincent et al., Clin Infect Dis 34: 1084, 2002). A therapeutic method has been reported recently by which significant improvement in the prognosis of severe sepsis can be achieved via administration of activated protein C (APC) (Bernard et al., N Engl J Med 344: 699, 2001). APC will be approved by the U.S. Food and Drug Administration (FDA) for the first time as a therapeutic agent against severe sepsis. However, the degree of effectiveness of APC is a slight rise in lifesaving rate of only approximately 6%-7%. Thus, sepsis is still considered to be a pathological condition with a very high fatality rate that is extremely difficult to cure. Furthermore, activated protein C is an endogenous protein that activates not only coagulation-suppressing functions, but also fibrinolytic functions, thereby inhibiting thrombus formation or inflammation (DePalo et al., Advances in Sepsis 1: 114, 2001). In addition to the palliative effects of activated protein C, it is inferred that APC increases the risk of bleeding because of its features. In particular, intracranial hemorrhage is a severe adverse event. Thus, administration of APC necessitates sufficient care so that APC is administered in compliance with contraindicated conditions. Attempts that have been made other than the aforementioned attempts are as shown below.

- (1) Large amounts of steroids: Application of large amounts of steroids has succeeded as a pretreatment for animals with endotoxemia or bacillaemia. Based on such successes, the effects of administration of large amounts of steroids to patients with septic shock have been examined in early clinical tests. However, in large-scale double-blind studies, validity has never been reported even in cases of administration of steroids during early development of septic shock (Meduri et al., Sepsis 3: 21, 1999; Bernard et al., N Engl J Med 317: 1565, 1987; Bone et al., Chest 92: 1032, 1987).
- (2) Antiendotoxin antibody: Treatment with a specific anti-endotoxin antibody has been examined using polyclonal human immunoglobulin G against heat-sterilized *E. coli* J5, mouse (E5) and humanized (HA1A) monoclonal antibodies against endotoxin lipid A, and the like. Patients with severe gram-negative bacterial infectious disease have been examined as subjects (Llewelyn et al., Sepsis 3: 39, 1999). Validity has never been confirmed in large-scale tests.
- (3) Anti-TNF treatment: Anti-TNF treatment is neutralization therapy targeting TNF- α (inflammatory cytokine), which uses an anti-TNF monoclonal antibody and a soluble TNF receptor. It has been reported that survival prospects can be improved in many sepsis models via suppression of the effects of TNF- α (Tracey et al., Nature 330: 662, 1987; Pennington et al., Clin Infect Dis 17 (Suppl 2): S515, 1993). Although a plurality of phase II and phase III clinical tests have been conducted, it has never been reported that survival rates have been improved by suppressing the effects of TNF- α with the use of anti-TNF treatment (Abraham et al., JAMA 273: 934, 1995; Reinhart et al., Crit Care Med 24: 733, 1996; Severansky et al., Sepsis 3: 11, 1999; Reinhart et al., Crit Care Med 29: S121, 2001).
- (4) IL-1 receptor antagonist (IL-1Ra): IL-1 is also an inflammatory cytokine, but is known to induce many pathological conditions of sepsis (Ohlsson et al., Nature 348: 550 1990). Such effects can be suppressed with the use of IL-1Ra, which is a natural IL-1 receptor antagonistic substance. However, a lack of differences between

a treatment group (group of treated patients) and a placebo group in terms of survival rate has been demonstrated by three tests (two double-blind tests) conducted for severe sepsis patients (Fisher et al., JAMA 271: 1836, 1994; Opal et al., Crit Care Med 25: 1115, 1997; Severansky et al., Sepsis 3: 11, 1999).

- (5) PAF receptor antagonist (PAFRa): Platelet agglutinating factor (PAF) is a phospholipid involved in cytokine release during sepsis. It has been demonstrated by two double-blind tests that PAF receptor antagonist (BN52021) does not significantly improve survival (Dhainaut et al., Crit Care Med 22: 1720, 1994; Dhainaut et al., Crit Care Med 26: 1963, 1998; Severansky et al., Sepsis 3: 11, 1999). It has been recently demonstrated again by a phase II clinical test using another compound (BB-882) that the compound does not significantly improve the survival of severe sepsis patients.
- (6) Nonsteroidal anti-inflammatory drug: an antiprostaglandin drug, Ibuprofen, has been examined in three double-blind tests, but the usefulness of Ibuprofen has never been demonstrated in any of these cases (Bernard et al., N Engl J Med 336: 912, 1997; Severansky et al., Sepsis 3: 11, 1999).
- (7) Bradykinin antagonist: Bradykinin is a bioactive peptide involved in cytokine release and changes in blood vessels during sepsis. Improvement in lethality with the use of a bradykinin antagonist has never been observed in two double-blind tests (Fein et al., JAMA 277: 482, 1997; Severansky et al., Sepsis 3: 11, 1999).

As described above, although various therapeutic methods for sepsis have been advanced, suppression of the incidence rate has never been confirmed. Sepsis is still a disease for which reduction in the number of deaths therefrom has been impossible to achieve. Novel exploitation of preventive methods or therapeutic methods for sepsis are required.

RANKL, which is a ligand of RANK, is an osteoclastic differentiation-inducing factor and is known to induce osteoclasts from precursor cells of the macrophage system under coexistence with a macrophage colony-stimulating factor (M-CSF) (see Yasuda et al., Proc Natl Acad Sci USA 95: 3597, 1998 and Lacey et al., Cell 93: 165, 1998). Specifically, RANKL is produced by osteoblasts and binds to RANK on precursor cells of the macrophage system, so as to induce the cells to become osteoclasts. Furthermore, at this time, OPG (osteoprotegerin) structurally analogous to RANK suppresses the effects of RANKL, as a decoy receptor. In this manner, bone metabolism is controlled by balancing RANKL and OPG amounts. RANKL is a membrane-associated protein and a part thereof is present in blood in a soluble form. Some bone metabolism diseases confirmed with variation in the concentration of soluble RANKL (sRANKL) have been reported. The usefulness of RANKL as a bone metabolism marker has been suggested, and medical applications of RANKL have been examined (see Rogers et al., J Clin Endocrinol Metab 90: 6323, 2005 and see JP Patent Publication (Kohyo) No. 2004-526748 A; JP Patent Publication (Kohyo) No. 2002-509430 A; and International Publication WO98/46644). Moreover, discussion often takes place concerning RANKL based on the concentration ratio of soluble RANKL to OPG; that is, the ratio of the concentration of soluble RANKL to OPG. As described above, the role of RANKL in the bone metabolism system has been conventionally known; however, the functions of RANKL in the natural immune system have remained unclear and the biological meaning of soluble RANKL existing in blood has also remained unclear.

DISCLOSURE OF THE INVENTION

Objects to be Achieved by the Invention

An object of the present invention is to provide a method for prevention or treatment of symptoms resulting from inflammatory diseases such as infectious diseases, allergies, and autoimmune diseases with the use of RANKL and to provide a composition for prevention or treatment. Particularly, an object of the present invention is to prevent death due to such inflammatory diseases and to treat such diseases. Moreover, an object of the present invention is to provide a method and a reagent for measuring the concentrations of membrane-bound and soluble RANKL and the concentration of OPG, and the like existing in vivo (e.g., in blood or in synovial fluids) as markers for prediction of the degree of inflammation and risks such as the lethality of inflammatory diseases.

Means to Achieve the Objects

As described above, the roles of RANKL in bone metabolism are known. However, the functions of RANKL in the natural immune system have not been elucidated and the biological meanings of RANKL existing in blood have remained unclear.

The present inventors have considered the possible involvement of RANKL in control of the natural immune system and then examined the action of soluble RANKL on macrophages. As a result, the present inventors have discovered that inflammatory cytokine production due to infection is suppressed in various macrophages because of the effects of soluble RANKL. Furthermore, based on the fact that the concentration of blood-soluble RANKL rapidly decreases within several hours after infection, while the concentration of OPG increases, the present inventors have discovered that soluble RANKL alone, OPG alone, or a combination of soluble RANKL and OPG can be a sensitive novel marker for infection. Moreover, the present inventors have discovered that administration of soluble RANKL makes it possible to prevent septic shock due to a drug and that a soluble RANKL can be used as an agent for preventing inflammation or as an anti-inflammatory agent. Thus, the present inventors have completed the present invention.

The present invention is as follows.

- [1] A method for detection of an inflammatory disease with the use of RANKL and/or OPG as a marker in a biological sample.
- [2] The method for detection of an inflammatory disease according to [1], which is a method for detection of an inflammatory disease with the use of RANKL as a marker in a biological sample, comprising determining that a subject is affected with an inflammatory disease when the concentration of soluble RANKL in a biological sample is lower than that of a normal subject.
- [3] The method for detection of an inflammatory disease according to [1], which is a method for detection of an inflammatory disease with the use of OPG as a marker in a biological sample, comprising determining that a subject is affected with an inflammatory disease when the concentration of OPG in a biological sample is higher than that of a normal subject.
- [4] The method for detection of an inflammatory disease according to [1], which is a method for detection of an inflammatory disease with the use of RANKL and OPG as markers in a biological sample, comprising determining that a subject is affected with an inflammatory disease

when the ratio of the concentration of soluble RANKL to the concentration of OPG in a biological sample is lower than that of a normal subject.

- [5] The method for detection of an inflammatory disease according to [1], which is a method for detection of an inflammatory disease with the use of RANKL as a marker in a biological sample, comprising measuring membrane-type RANKL existing on a cell in a biological sample by a flow cytometric method.
- [6] The method for detection of an inflammatory disease according to [1], which is a method for detection of an inflammatory disease with the use of RANKL and OPG as markers in a biological sample, comprising measuring membrane-type RANKL existing on a cell in a biological sample by a flow cytometric method and then determining that a subject is affected with an inflammatory disease when the ratio of the amount of membrane-type RANKL measured to the concentration of OPG is lower than that of a normal subject.
- [7] The method for detection of an inflammatory disease according to [5] or [6], in which the cell in a biological sample is a peripheral blood cell.
- [8] The method for detection of an inflammatory disease according to any one of [1] to [7], in which the inflammatory disease is an infectious disease, an allergic disease, or an autoimmune disease.
- [9] The method for detection of an inflammatory disease according to [8], in which the inflammatory disease is sepsis.
- [10] A detection reagent for detection of an inflammatory disease with the use of RANKL and/or OPG as a marker, comprising an anti-RANKL antibody and/or an anti-OPG antibody.
- [11] A detection reagent for detection of an inflammatory disease with the use of RANKL and/or OPG as a marker, comprising an anti-RANKL antibody and an anti-OPG antibody.
- [12] The detection reagent for detection of an inflammatory disease according to [10] or [11], in which the inflammatory disease is an infectious disease, an allergic disease, or an autoimmune disease.
- [13] The detection reagent for detection of an inflammatory disease according to [12], in which the inflammatory disease is sepsis.
- [14] A composition for prevention or treatment of an inflammatory disease, comprising RANKL and/or M-CSF as an active ingredient.
- [15] The composition for prevention or treatment of an inflammatory disease according to [14], comprising RANKL and M-CSF as active ingredients.
- [16] The composition for prevention or treatment of an inflammatory disease according to [15], in which the composition for prevention or treatment of an inflammatory disease is a combination preparation.
- [17] The composition for prevention or treatment of an inflammatory disease according to [15], in which the composition is a kit comprising a drug that contains RANKL and a drug that contains M-CSF.
- [18] The composition for prevention or treatment of an inflammatory disease according to any one of [15] to [17], which is characterized in that RANKL and/or M-CSF is administered before surgery, so as to prevent a postoperative inflammatory disease.
- [19] The composition for prevention or treatment of an inflammatory disease according to any one of [15] to [18], in which the inflammatory disease is an infectious disease, an allergic disease, or an autoimmune disease.

- [20] The composition for prevention or treatment of an inflammatory disease according to [19], in which the inflammatory disease is sepsis.
- [21] An immunosuppressive agent, comprising RANKL and/or M-CSF as an active ingredient.
- [22] The immunosuppressive agent according to [21], comprising RANKL and M-CSF as active ingredients.
- [23] The immunosuppressive agent according to [22], in which the immunosuppressive agent is a combination preparation.
- [24] The immunosuppressive agent according to [23], in which the immunosuppressive agent is a kit comprising a drug that contains RANKL and a drug that contains M-CSF.

Effects of the Invention

When a subject is affected with an inflammatory disease such as an infectious disease, RANKL expression decreases and OPG expression increases. Therefore, an inflammatory disease can be detected and diagnosed by measurement of the amount of RANKL or OPG. Particularly when the ratio of the amount of RANKL to the amount of OPG is used as an indicator, high-precision detection and diagnosis can be performed.

Furthermore, RANKL and/or M-CSF has an effect of suppressing inflammatory cytokine production by various macrophages. Moreover, an inflammatory disease can be prevented or treated with the use of RANKL and/or M-CSF.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 shows a lack of the induction of cytokine production due to soluble RANKL.

FIG. 2 shows RANKL-induced tolerance in macrophages based on inflammatory cytokine concentrations.

FIG. 3 shows RANKL-induced tolerance in macrophages based on inflammatory cytokine mRNA levels.

FIG. 4 shows RANKL-induced tolerance in macrophages based on the concentrations of TNF- α produced when macrophages were pretreated with soluble RANKL for given periods of time and then stimulated with LPS.

FIG. 5 shows RANKL-induced tolerance in macrophages based on the concentrations of TNF- α produced when peritoneal macrophages (PMs) and M-CSF-dependent spleen-derived macrophages (MDSMs) were pretreated with soluble RANKL for given periods of time and then stimulated with LPS.

FIG. 6 shows RANKL-induced tolerance with respect to various stimulations.

FIG. 7 shows the reversibility of RANKL-induced tolerance.

FIG. 8 shows attenuation of RANKL-induced tolerance by GM-CSF.

FIG. 9 shows changes in serum soluble RANKL and OPG concentrations in response to LPS injection.

FIG. 10 shows changes in serum soluble RANKL and OPG concentrations in response to *Salmonella* infection.

FIG. 11 shows serum soluble RANKL and OPG concentrations in wild-type mice and mice lacking RANKL (Tnfsf11^{-/-}).

FIG. 12 shows serum cytokine concentrations in wild-type mice and mice lacking RANKL (Tnfsf11^{-/-}) into which LPS was injected intraperitoneally.

FIG. 13 shows serum soluble RANKL and OPG concentrations in wild-type mice and mice lacking OPG (Tnfrsf11b^{-/-}).

FIG. 14 shows serum cytokine concentrations in wild-type mice and mice lacking OPG (Tnfrsf11b^{-/-}), into which LPS was injected intraperitoneally.

FIG. 15 shows survival curves after intraperitoneal injection of a high dose of LPS into wild-type mice (n=10) and Tnfsf11^{-/-} mice (n=5).

FIG. 16 shows that TLR4 expression is suppressed by treatment of macrophages with soluble RANKL.

FIG. 17 shows antigen presentation suppressed by soluble RANKL.

FIG. 18 shows that RANKL tolerance is c-Fos-independent.

FIG. 19 shows the nucleotide sequence of cDNA that encodes human-type RANKL residues 140 to 317 (SEQ ID No: 14) and the corresponding amino acid sequence (SEQ ID No: 10).

FIG. 20 shows a restriction enzyme map of a vector containing a RANKL gene (SEQ ID No: 15).

FIG. 21A shows the nucleotide sequence of a vector containing a RANKL gene (1/2, continuing to FIG. 21B) (SEQ ID No: 11).

FIG. 21B shows the nucleotide sequence of a vector containing the RANKL gene (2/2, continued from FIG. 21A) (SEQ ID No: 11).

FIG. 22 shows a method for construction of a GST-RANKL vector.

FIG. 23 shows survival percentage when LPS was administered to the mice to which GST-RANKL and/or M-CSF had been administered.

BEST MODE OF CARRYING OUT THE INVENTION

RANKL (Receptor activator of NF- κ B ligand) is a ligand of RANK (receptor activator of NF- κ B) that is a member of the TNF superfamily and is a type II transmembrane protein having an intracellular domain (the domain comprising amino acids 1 to 48 from the N terminus of RANK), a transmembrane domain, and an extracellular domain (JP Patent Publication (Kohyo) No. 2002-509430 and International Publication WO98/46644 pamphlet). In the extracellular domain, a domain comprising amino acid 152 (from the N terminus) and the following amino acids is a TNF ligand family homologous domain.

OPG (osteoprotegerin) has a structure analogous to that of RANK and can bind to RANKL.

When a subject is affected with an inflammatory disease, RANKL expression decreases, OPG expression increases, blood soluble RANKL concentration decreases, and OPG concentration increases.

The method of the present invention is a method for detection of an inflammatory disease, which comprises measuring RANKL and/or OPG in a biological sample collected from a subject and performing detection with the use of RANKL and/or OPG as a marker. RANKL to be measured in the method of the present invention is soluble RANKL (sRANKL) secreted in a biological sample that is a body fluid such as blood or membrane-type RANKL existing on cells in peripheral blood or the like.

Examples of a biological sample include blood, plasma, serum, tears, urine, amniotic fluids, synovial fluids, spinal fluids, cell extracts, tissue extracts, cells, and tissues. The effects of an inflammatory disease are often expressed throughout the body, so that blood, plasma, and serum are preferable among the above biological samples.

In the method of the present invention, the term "inflammatory disease" refers to a generic term of diseases with the

presentation of inflammatory symptoms, including infectious diseases, allergy diseases, and autoimmune diseases, for example. Examples of such diseases include severely diseased trauma, burn, surgical invasion, acute pancreatitis, peritonitis, malignant tumor, acute abdomen (abdominal disease with an acute abdominal pain as a predominant symptom, which requires emergency surgery), infectious diseases (particularly, nosocomial infections due to Gram-negative bacteria such as *Serratia*, *Pseudomonas aeruginosa*, *Acinetobacter*, *Citrobacter*, and *Enterobacter*), and a severe acute disease (namely, SIRS) requiring treatment in ICU, such as sepsis. Furthermore, allergy diseases such as contact hypersensitivity, allergic rhinitis, food allergy, and asthma are also diseases to be subjected to the method of the present invention. Further examples of the inflammatory disease include inflammatory skin diseases such as atopic dermatitis, contact dermatitis, photosensitive dermatitis, chronic dermatitis of fingers and toes, seborrheic dermatitis, nummular dermatitis, generalized exfoliative dermatitis, stasis dermatitis, local dermatitis due to abrasion, dermatitis medicamentosa, or psoriasis. Further examples of the inflammatory disease include autoimmune diseases such as rheumatoid arthritis, scleroderma, dermatomyositis, autoimmune vasculitis, mixed connective tissue disease, systemic erythematosis, idiopathic thrombocytopenic purpura, Crohn's disease, and human adjuvant disease.

Whether or not a subject is affected with the above inflammatory disease can be detected and diagnosed by the method of the present invention. Furthermore, the severity of the above inflammatory disease can be evaluated and determined. In particular, the severity of sepsis can be evaluated and determined.

A method for measuring RANKL and/or OPG is not limited. For example, RANKL and/or OPG can be measured by immunoassay using an anti-RANKL antibody and/or an anti-OPG antibody, such as Western blotting, EIA, RIA, an agglutination method, immunochromatography, or a flow cytometric method. When membrane-bound RANKL on cells in peripheral blood or the like is measured, a bound RANKL level (amount of RANKL) on cells is measured by a flow cytometric method. Measurement by flow cytometry can be performed using a flow cytometer such as a commercially available FACS. An anti-RANKL antibody and/or anti-OPG antibody can be prepared by a known method. When measurement is performed using an antibody, an antibody to be used herein is labeled adequately with an enzyme such as alkaline phosphatase or a fluorescent dye. In addition, extracellular domains of soluble RANKL and membrane-type RANKL are subjected to measurement in the present invention, an anti-RANKL antibody preferred herein recognizes and binds to the extracellular domains of soluble RANKL and membrane-type RANKL. Soluble RANKL contains no intracellular domain, so that an anti-RANKL antibody to be used in the method of the present invention is an antibody capable of recognizing an extracellular domain other than the intracellular domains of RANKL and preferably recognizing an epitope that exists in a TNF ligand family homologous domain. Furthermore, a commercially available antibody can also be used. Furthermore, RANKL and/or OPG mRNA is detected and then the expression of RANKL and/or OPG may be detected. mRNA can be detected by Northern blotting, an RT-PCR method, a method using DNA chips (DNA microarray), or the like. At this time, a probe or primers comprising partial sequences complementary to the partial sequences of mRNA encoding RANKL and/or OPG are used for measuring specifically mRNA that encodes RANKL and/or OPG. The nucleotide sequences of RANKL and OPG are known.

Probes or primers can be designed based on the known nucleotide sequence information. The number of the nucleotides of a primer or a probe ranges from 5 to 50, preferably 10 to 30, and further preferably 15 to 25. These methods can be performed by known methods.

When a RANKL concentration or amount in a sample collected from a subject is significantly lower than that of a normal subject or when an OPG concentration is significantly higher than that of a normal subject, the subject can be diagnosed as being affected with an inflammatory disease. Furthermore, it can be evaluated and determined that the lower the RANKL concentration or amount or the higher the OPG concentration, the severer the inflammatory disease such as sepsis.

Moreover, when the concentration ratio of soluble RANKL to OPG (soluble RANKL concentration: OPG concentration) in a sample collected from a subject is lower than that of a normal subject, the subject can be diagnosed as being affected with an inflammatory disease. Furthermore, it can be evaluated and determined that the lower the concentration ratio of soluble RANKL to OPG, the severer the inflammatory disease such as sepsis.

The expression of RANKL and/or OPG can fluctuate within several hours after the onset of an inflammatory disease. Hence, an inflammatory disease can be detected at an early phase according to the method of the present invention. Furthermore, several times of measurement of RANKL and/or OPG at appropriate time intervals enables more precise detection.

The present invention encompasses a reagent or a kit for detection of an inflammatory disease with the use of RANKL and/or OPG as a marker. The reagent or the kit comprises an anti-RANKL antibody and/or an anti-OPG antibody. When soluble RANKL and OPG are measured by ELISA or the like, an antibody may be labeled with an enzyme such as alkaline phosphatase or horseradish peroxidase. In addition, when membrane-bound RANKL is measured by a flow cytometric method, an antibody against RANKL may be labeled with a fluorescent dye.

The present invention further encompasses a marker for detection of an inflammatory disease, which comprises RANKL and/or OPG. The present invention further encompasses the use of RANKL and/or OPG as a marker for detection of an inflammatory disease.

The present invention further encompasses a composition (anti-inflammatory agent) or an immunosuppressive agent for treatment or prevention of an inflammatory disease, comprising RANKL and/or M-CSF as an active ingredient. When RANKL alone or a combination of RANKL and M-CSF is administered to a subject, inflammatory cytokine production by macrophages is suppressed in the subject, TLR4 level is suppressed, and the antigen-presenting ability of antigen-presenting cells of the subject is suppressed. Accordingly, RANKL and M-CSF can be used independently or in combination for treatment or prevention of inflammatory diseases. Furthermore, the immunity of a subject can be suppressed by the use of RANKL and M-CSF independently or in combination. In addition, RANKL-induced tolerance (RANKL tolerance) in macrophages is independent from c-Fos required for osteoclast differentiation. Thus, the effects of RANKL on macrophages differ from the "phenomenon of differentiation from macrophages" in the process of differentiation of macrophages into osteoclasts.

The term "inflammatory disease" which is a target of the composition for prevention or treatment of the present invention is a generic term for diseases with presentation of inflammatory symptoms, including infectious diseases, allergy dis-

eases, and autoimmune diseases, for example. Examples of such diseases include severely diseased trauma, burn, surgical invasion, acute pancreatitis, peritonitis, malignant tumor, acute abdomen (abdominal disease with an acute abdominal pain as a predominant symptom, which requires emergency surgery), infectious diseases (particularly, nosocomial infections due to Gram-negative bacteria), and a severe acute disease (namely, SIRS) requiring treatment in ICU, such as sepsis. Furthermore, allergy diseases such as contact hypersensitivity, allergic rhinitis, food allergy, and asthma are also diseases to be subjected to the method of the present invention. Further examples of the inflammatory disease include inflammatory skin diseases such as atopic dermatitis, contact dermatitis, photosensitive dermatitis, chronic dermatitis of fingers and toes, seborrheic dermatitis, nummular dermatitis, generalized exfoliative dermatitis, stasis dermatitis, local dermatitis due to abrasion, dermatitis medicamentosa, or psoriasis. Further examples of the inflammatory disease include autoimmune diseases such as rheumatoid arthritis, scleroderma, dermatomyositis, autoimmune vasculitis, mixed connective tissue disease, systemic erythematosis, idiopathic thrombocytopenic purpura, Crohn's disease, and human adjuvant disease.

Examples of RANKL that can be used for the composition for treatment or the immunosuppressive agent of the present invention include RANKL, soluble RANKL, a soluble RANKL derivative, a soluble RANKL analog, a soluble RANKL fusion protein, or a soluble RANKL mimic. The full-length nucleotide sequence and amino acid sequence of human-derived RANKL are shown in SEQ ID NO: 1 and 2, respectively. A soluble RANKL derivative or a soluble RANKL analog is a protein comprising a partial sequence of the amino acid sequence of RANKL, such as truncated RANKL. A protein having RANKL activity is also included herein. A soluble RANKL derivative preferably contains a TNF ligand family homologous domain that begins from amino acid 152 in the amino acid sequence of SEQ ID NO: 2. Examples of such a soluble RANKL derivative include a protein comprising the amino acid sequence ranging from amino acid 127 to amino acid 317, a protein comprising the amino acid sequence ranging from amino acid 140 to amino acid 317, or a protein comprising the amino acid sequence ranging from amino acid 159 to amino acid 317. Furthermore, examples of a soluble RANKL derivative or a soluble RANKL analog include a protein comprising an amino acid sequence that comprises a deletion, substitution, or addition of one or several amino acids with respect to the amino acid sequence represented by SEQ ID NO: 2 and having RANKL activity, and a protein comprising an amino acid sequence that comprises a deletion, substitution, or addition of one or several amino acids with respect to the amino acid sequence of a protein that comprises a partial sequence of the amino acid sequence of the above RANKL and having RANKL activity. Here, the term "one or several" refers to 1 to 9, preferably 1 to 5, and further preferably 1 or 2. The term "soluble RANKL fusion protein" refers to a fusion protein prepared by fusing another protein to a soluble RANKL protein, a soluble RANKL derivative, or a soluble RANKL analog. An example of such "another protein" is glutathione S-transferase (GST). The nucleotide sequence of DNA encoding a fusion protein prepared by fusing GST to a protein comprising an amino acid sequence ranging from amino acid 127 to amino acid 317 in the amino acid sequence of RANKL and the amino acid sequence of the fusion protein are shown in SEQ ID NOS: 3 and 4, respectively. The nucleotide sequence of DNA encoding a fusion protein prepared by fusing GST to a protein comprising an amino acid sequence ranging from amino acid

140 to amino acid 317 in the amino acid sequence of RANKL and the amino acid sequence of the fusion protein are shown in SEQ ID NOS: 5 and 6, respectively. Furthermore, the nucleotide sequence of DNA encoding a fusion protein prepared by fusing GST to a protein comprising an amino acid sequence ranging from amino acid 159 to amino acid 317 in the amino acid sequence of RANKL and the amino acid sequence of the fusion protein are shown in SEQ ID NOS: 7 and 8, respectively. The soluble RANKL mimic is a compound having a structure analogous to the conformation of RANKL and having RANKL activity. Such RANKL, soluble RANKL, soluble RANKL derivative, soluble RANKL analog, and soluble RANKL fusion protein can be prepared as recombinant proteins by gene-engineering techniques.

Examples of M-CSF to be used for the composition for prevention or treatment or the immunosuppressive agent of the present invention include, similarly to examples of RANKL, M-CSF, an M-CSF derivative, an M-CSF analog, an M-CSF fusion protein, and an M-CSF mimic. Such M-CSF, M-CSF derivative, M-CSF analog, and M-CSF fusion protein can be prepared as recombinant proteins by gene-engineering techniques. Furthermore, a commercially available M-CSF preparation can also be used. An example of such M-CSF is Leukoprol (Trademark, general name: mirimostim).

RANKL alone may be administered to a subject or M-CSF alone may also be administered to a subject for prevention or treatment of an inflammatory disease or immunosuppression. Preferably, both RANKL and M-CSF are administered. When both RANKL and M-CSF are administered, the mixture of the two, that is, a combination preparation prepared by mixing the two can be administered simultaneously. Furthermore, a composition containing RANKL and a composition containing M-CSF are separately formulated, and then the two preparations can be mixed when used and then administered. Furthermore, the preparations may be administered separately in turn. Preferably, both RANKL and M-CSF are administered simultaneously. The composition for prevention or treatment or the immunosuppressive agent of the present invention is also a kit comprising a drug containing RANKL and a drug containing M-CSF for administration of RANKL and M-CSF separately.

Moreover, when the composition for prevention or treatment of an inflammatory disease of the present invention is used for treatment, the composition is administered to a patient affected with an inflammatory disease. Furthermore, when the composition is used for prevention, the composition is administered to a patient suspected of being affected with an inflammatory disease. For example, the composition is administered to a patient affected with a severe infectious disease who may be suspected of developing sepsis or SIRS. Furthermore, for prevention of postoperative infection, the composition of the present invention may be administered before invasive surgery. The immunosuppressive agent of the present invention comprising RANKL and/or M-CSF as an active ingredient is administered to a patient when a tissue or an organ is transplanted in the patient. The immunosuppressive agent of the present invention can suppress graft rejection associated with cell or organ and/or tissue transplantation and graft-versus-host disease.

The composition for prevention or treatment of an inflammatory disease or the immunosuppressive agent of the present invention can be administered in various forms. For example, the composition can be orally administered in the form of tablets, capsules, fine granules, powders, syrups, or the like or can also be parenterally administered in the form of injections, drops, suppositories, sprays, eye drops, intranasal agents, and adhesive preparations, or the like.

The composition for prevention or treatment of an inflammatory disease or the immunosuppressive agent of the present invention contains a carrier, a diluent, and an excipient, which are generally used in the pharmaceutical field. For example, as a carrier and an excipient for tablets, lactose, magnesium stearate, and the like are used. As an aqueous solution for injection, a physiological saline solution, an isotonic solution containing glucose or other adjunctive agents, or the like is used. These examples may also be used in combination with an appropriate solubilizing agent such as alcohol, polyalcohol such as propylene glycol, a nonionic surfactant, and the like. As an oily liquid, sesame oil, soybean oil, or the like is used. As a solubilizing agent, benzyl benzoate, benzylalcohol, or the like may also be used in combination.

The dose is varied depending on symptoms, age, body weights, and the like. The dose in the case of oral administration ranges from approximately 0.001 mg to 1000 mg per day, and administration may be performed once or in divided doses. Furthermore, the dose in the case of parenteral administration ranges from 0.001 mg to 1000 mg per administration, and administration is performed by intravenous injection, intraperitoneal injection, subcutaneous injection, or intramuscular injection or using a suppository, an eye drop, or the like. When RANKL and M-CSF are used in combination, the ratio of the dose of RANKL to the dose of M-CSF is not limited. RANKL and M-CSF may be administered in the same amounts, or the amount of either soluble RANKL or M-CSF to be administered may be greater than the other.

The present invention further encompasses a method for prevention or treatment of an infectious disease or an immunosuppression method, comprising administering soluble RANKL and/or M-CSF to a patient who needs prevention or treatment.

The present invention further encompasses the use of soluble RANKL and/or M-CSF for production of a composition for prevention or treatment of an infectious disease or the use of the same for production of an immunosuppressive agent.

The present invention further encompasses soluble RANKL and/or M-CSF to be used for treatment of an infectious disease or soluble RANKL and/or M-CSF to be used for immunosuppression.

EXAMPLES

The present invention is hereafter described in greater detail with reference to the following examples, although the present invention is not limited thereto.

The following materials and methods are used in the Examples.

Mice

Six to 10-week-old C57BL/6J mice were purchased from Oriental Yeast Co., Ltd. Homozygous mice lacking OPG and control heterozygous mice with a C57BL/6J background were purchased from Clea Japan. Mice lacking RANKL and mice lacking TLR4 with a C57BL/6J background were bred and maintained under specific pathogen-free conditions (SPF). A powder diet was provided to mice lacking RANKL. All experiments were performed in accordance with guidelines for animal use at the Keio University School of Medicine or Oriental Yeast Co., Ltd.

M-CSF-dependent Macrophage

To generate M-CSF-dependent bone marrow-derived macrophages (MDBMs), bone marrow cells were harvested by flushing tibias and femurs with Dulbecco's modified Eagle's medium (DMEM) containing 10% fetal calf serum (FCS) and

antibiotics. After passage through a cell strainer, bone marrow cells were cultured overnight. Non-adherent cells were harvested and cultured in the presence of 10 ng/ml M-CSF. After 3 to 4 days, cells that had adhered to the wells were harvested using a cell scraper and then seeded as M-CSF-dependent bone marrow-derived macrophages at a concentration of 1×10^5 /well in a 24-well plate (Falcon). M-CSF-dependent spleen-derived macrophages (MDSMs) were generated from splenocytes by a method similar to the above method. All macrophages were cultured in the presence of 10 ng/ml M-CSF in all experiments.

Peritoneal Macrophages

Peritoneal cells were harvested by flushing the peritoneal cavity of mice with complete medium (DMEM containing 10% FCS and antibiotics). The thus harvested cells were seeded at a concentration of 1×10^5 /well in a 24-well plate (Falcon). Cells were cultured for 6 hours, the plate was washed with PBS to remove non-adherent cells, and remaining cells were incubated in fresh medium. The thus obtained adherent cells were used as peritoneal macrophages.

In vitro Tolerance Experiments

Macrophages were pretreated with different concentrations of soluble RANKL (<0.10 endotoxin units/mg, R&D) or LPS (also referred to as lipopolysaccharide or endotoxin; LPS used in the Examples is S. Minnesota Re595, Sigma) for given periods of time. Cultured cells were subsequently washed twice with phosphate buffered saline (PBS) and stimulated with LPS, flagellin derived from *Salmonella muenchen* (Calbiochem), CpG oligonucleotide (5'-TCCAT-gACgTTCCTgATgCT-3'; SEQ ID NO: 12), or control GpC oligonucleotides (5'-TCCATgAgCTTCCTgATgCT-3', Proligo; SEQ ID NO: 13) as indicated in the description. In some experiments, 500 U/ml GM-CSF (Pepro Tec) was added 3 hours before stimulation of cells with LPS.

Mouse LPS Administration

LPS (S. Minnesota Re595, Sigma) was administered intraperitoneally. LPS (*E. coli* 055: B5, Sigma) was used in the Example as shown in FIG. 23. Blood was collected by heart puncture at given time points. Blood was allowed to clot for 1 hour and then centrifuged at 15000 rpm at 24° C. for 20 minutes. The serum was stored at -80° C. until cytokine assays.

Bacterial Strain and Infection Experiment

For an infection experiment, an overnight standing culture of *Salmonella enterica* serovar Typhimurium χ 3306 strain (hereinafter, *Salmonella*) in Luria-Bartani broth (LB medium) was diluted and shaken, and mid-log phase bacteria were then collected by centrifugation. *Salmonella* was washed with PBS, diluted with Hanks' salt solution, and used to infect macrophages at a multiplicity of infection (MOI) of 10. After incubation at 37° C. for 1 hour, macrophages were washed with PBS to remove extracellular *Salmonella* and then incubated in complete medium containing 25 μ g/ml gentamycin. After 3 hours, culture supernatants were harvested for cytokine assays. For oral infection with *Salmonella*, mice were subjected to fasting with no water and feed for 12 hours before infection, and then 3.3×10^7 CFU/g body weight of *Salmonella* was administered orally. Four days later, blood was collected by heart puncture.

Enzyme-linked Immunosorbent Assay (ELISA)

TNF- α , IL-6, and IL-12 (p40) concentrations in macrophage culture supernatants were measured using ELISA sets (BD PharMingen). Soluble RANKL and OPG concentrations in mouse serum were measured using ELISA kits (R&D).

PT-PCR Analysis

mRNA was prepared by collecting macrophages and homogenizing the macrophages in Isogen (Nippon gene).

cDNA was synthesized using the Enhanced Avian HS RT-PCR kit (Sigma-Aldrich). Quantitative PCR was performed using an ABI PRISM 7000 apparatus TaqMan Assay-on-demand (Applied Biosystems) and IL-6, TNF- α , IL-12 (p40), and Gapdh primers.

Statistical Analysis

Data are expressed as means \pm SD (standard deviation). All data excluding those of the Example, the results of which are shown in FIG. 23, were analyzed by significant difference tests using the Student's t-test. Only in the Example, the results of which are shown in FIG. 23, significant difference tests were performed using a generalized Wilcoxon test.

The following results were obtained.

(1) Induction of RANKL Tolerance in Macrophages Stimulated with Bacterial Components

First, M-CSF-dependent bone marrow-derived macrophages (MDBMs) were stimulated with soluble RANKL or LPS and then inflammatory cytokine production was measured (FIG. 1A to C). FIG. 1A to C show the lack of the induction of cytokine production because of soluble RANKL. MDBMs were stimulated with LPS or soluble RANKL with a concentration shown in FIG. 1 for 24 hours. Protein concentrations of inflammatory cytokines in culture supernatants were measured by ELISA. Bars in FIG. 1 represent means \pm SD (n=3, culture well). As shown in FIG. 1A to C, it was revealed that while LPS significantly induced the production of cytokines such as TNF- α , IL-6, and IL-12 (p40), soluble RANKL with a concentration as high as 100 ng/ml did not induce detectable levels of cytokine production. Therefore, unlike LPS, soluble RANKL cannot induce inflammatory cytokines.

Next, MDBMs were pretreated for 24 hours with soluble RANKL by increasing the concentration of soluble RANKL in steps and then stimulated with LPS for 6 hours. The concentrations of TNF- α , IL-6, and IL-12 (p40) in the supernatants were measured by ELISA. As a result, it was revealed that pretreatment of soluble RANKL suppresses cytokine production resulting from subsequent stimulation with LPS in a soluble RANKL concentration-dependent manner (FIG. 2A to C). FIG. 2A to C show the protein concentration of each inflammatory cytokine, showing that RANKL-induced tolerance was induced in macrophages. MDBMs were pretreated with soluble RANKL for 24 hours (1 $^\circ$) and then stimulated with 100 ng/ml LPS for 6 hours (2 $^\circ$). As shown in FIG. 2A to C, such induced tolerance was also observed at a low concentration (as low as 1 ng/ml) of soluble RANKL.

The mRNA levels of TNF- α , IL-6, and IL-12 (p40) were measured with time over 6 hours after stimulation with LPS. Compared with macrophages not pretreated with soluble RANKL, macrophages pretreated with soluble RANKL showed significantly suppressed induction of cytokine mRNAs upon LPS stimulation (FIG. 3A to C). FIG. 3A to C show the mRNA level of each inflammatory cytokine, showing that RANKL-induced tolerance was induced in macrophages. MDBMs were pretreated with medium alone (open diamonds) or 10 ng/ml soluble RANKL (close diamonds) for 24 hours and then stimulated with 100 ng/ml LPS for 6 hours. Cells were harvested at each time point as in FIG. 3. Cytokine mRNA levels were measured by quantitative PCR. Values are normalized to Gapdh.

Next, the duration of pretreatment with soluble RANKL was shortened to 24 hours, 12 hours, and 6 hours. MDBMs were pretreated with 1 ng/ml soluble RANKL for given periods of time as in FIG. 4 (1 $^\circ$) and then stimulated with LPS (2 $^\circ$). As a result, it was revealed that even as short as 6 hours of pretreatment significantly suppressed the induction of TNF- α production in response to stimulation with LPS (FIG. 4).

To examine whether or not RANKL-induced tolerance takes place in macrophages other than MDBMs, an experiment was then conducted using peritoneal macrophages (PMs) and M-CSF-dependent spleen-derived macrophages (MDSMs). Peritoneal macrophages (PMs) and M-CSF-dependent spleen-derived macrophages (MDSMs) were pretreated with 50 ng/ml soluble RANKL for 24 hours (1 $^\circ$) and then stimulated with LPS (2 $^\circ$). As a result, the induction of TNF- α production in response to stimulation with LPS was suppressed in these macrophages when the macrophages had been pretreated with soluble RANKL (FIG. 5).

Finally, MDBMs were pretreated with soluble RANKL for 24 hours and then stimulated with flagellin, CpG oligonucleotide, and *Salmonella*. MDBMs were pretreated with 10 ng/ml soluble RANKL for 24 hours (1 $^\circ$) and then stimulated with 1×10^{-11} M flagellin or 3 nM CpG DNA for 6 hours or with *Salmonella* for 3 hours (2 $^\circ$). Cytokines in the culture supernatants were measured by ELISA. As a result, it was revealed that pretreatment with soluble RANKL suppressed the induction of TNF- α production in response to stimulation with the bacterial components or *Salmonella* (FIG. 6A to C). Bars in FIG. 6 represent means \pm SD (n=3, culture well). “*”, P<0.05; “**”, P<0.01” represent the results compared with open bars. These results suggest that soluble RANKL lowers the responsiveness of macrophages to LPS or other bacterial components.

(2) Attenuation of RANKL-induced Tolerance by Soluble RANKL Removal or GM-CSF Treatment

To examine whether or not RANKL-induced tolerance is reversible, MDBMs were treated with soluble RANKL for 24 hours and then cultured in soluble-RANKL-free medium for another 24 hours. MDBMs were treated with medium alone (Med) or 10 ng/ml soluble RANKL for 24 hours (Day 1), treated with medium alone (Med) or 10 ng/ml soluble RANKL for another 24 hours (Day 2), and then stimulated with 100 ng/ml LPS for 6 hours. Culture supernatants (n=3) were harvested for measurement of cytokine concentrations by ELISA. Compared with a control group, production of TNF- α , IL-6, and IL-12 (p40) was sufficiently or at least partially restored, demonstrating that RANKL-induced tolerance is reversible (FIG. 7A to C). Bars in FIG. 7 represent means \pm SD. “*”, P<0.01” represent the results compared with close bars.

GM-CSF is known to inhibit RANKL/RANK signaling. Hence, the effects of GM-CSF on RANKL-induced tolerance were examined. MDSMs were pretreated with soluble RANKL for 24 hours and then treated with soluble RANKL-free GM-CSF-containing medium for 3 hours. Subsequently, cells were stimulated with LPS and then TNF- α concentrations were measured. Specifically, MDBMs were treated with 10 ng/ml soluble RANKL for 24 hours (1 $^\circ$), stimulated with 500 U/ml GM-CSF for 3 hours (2 $^\circ$), and then stimulated with 100 ng/ml LPS for 6 hours (3 $^\circ$). The results are shown in FIG. 8. TNF- α concentrations (n=3) in the culture supernatants were measured by ELISA. Bars in FIG. 8 represent means \pm SD. *, P<0.01. Data representative among three experiments are shown. As shown in FIG. 8, although GM-CSF itself did not induce TNF- α production, brief treatment with GM-CSF significantly increased TNF- α production in macrophages pretreated with soluble RANKL. These results suggest that GM-CSF attenuates RANKL-induced tolerance (FIG. 8). Taken together, it was demonstrated by the Example that short treatment with GM-CSF can restore the responsiveness of macrophages to LPS, as in the case of soluble RANKL removal.

(3) Dynamic Shift of Serum Soluble RANKL and OPG Levels Due to LPS Administration and Bacterial Infection

To compare physiological and pathological soluble RANKL concentrations in serum, LPS was injected intraperitoneally into mice and then serum soluble RANKL and OPG concentrations were measured. LPS was injected intraperitoneally into C57BL/6J mice (n=24) (1 μ g/g body weight). Blood was collected at each time point as in FIG. 9 (n=3, mouse/point). Serum soluble RANKL and OPG concentrations were measured by ELISA. The serum soluble RANKL concentration was approximately 150 pg/ml and OPG concentration was approximately 2000 pg/ml prior to LPS injection. The results are shown in FIGS. 9A and B. Bars in FIG. 9 represent means \pm SD. “*”, P<0.05; “**”, P<0.01” show the results compared with that of 0 h. Surprisingly, soluble RANKL concentration dramatically fell at 6 hours after LPS injection, while OPG concentration was up-regulated on hour 6 and later. Furthermore, C57BL/6J mice (n=6) were orally infected with 3.3×10^7 CFU/g body weight of *Salmonella* (infection). Control mice were caused to drink PBS (control). After 4 days, blood was collected and then serum soluble RANKL and OPG concentrations were measured by ELISA. The results are shown in FIGS. 10A and B. As shown in FIG. 10, down-regulation of serum soluble RANKL and up-regulation of OPG were also observed on day 4 after oral infection with *Salmonella*. Bars in FIG. 10 represent means \pm SD (n=3). “*”, P<0.05; “**”, P<0.01. These results demonstrate that serum soluble RANKL and OPG levels are dynamically regulated in response to LPS and bacterial infection.

(4) Abnormal Cytokine Production in Mice Lacking RANKL and Mice Lacking OPG

To examine the effects of physiological serum soluble RANKL on LPS-induced cytokine production, *Tnfrsf11^{-/-}* mice lacking RANKL were analyzed. FIGS. 11A and B show serum soluble RANKL and OPG concentrations in wild-type mice and mice lacking RANKL (*Tnfrsf11^{-/-}*), as measured by ELISA (n=4 for each genotype). Bars in FIG. 11 represent means \pm SD. “*”, P<0.05; “**”, P<0.01” show the results compared with that of the control. As predicted, serum soluble RANKL was not detectable in *Tnfrsf11^{-/-}* mice, but OPG levels were slightly higher in *Tnfrsf11^{-/-}* mice than in wild-type mice.

Furthermore, LPS (2 μ g/g body weight) was injected intraperitoneally. Ninety minutes later, blood of wild-type mice and *Tnfrsf11^{-/-}* mice were collected. Serum cytokines were measured by ELISA. After LPS injection, significantly elevated TNF- α production and IL-6 production were observed in *Tnfrsf11^{-/-}* mice (FIG. 12A to C). Bars in FIG. 12 represent means \pm SD. “*”, P<0.05; “**”, P<0.01” show the results compared with that of the control. These results are consistent with the notion that the lack of RANKL-induced tolerance potentiates production of inflammatory cytokines in *Tnfrsf11^{-/-}* mice.

To further examine the role of RANKL-induced tolerance in mice, *Tnfrsf11b^{-/-}* mice lacking OPG were analyzed. FIGS. 13A and B show serum soluble RANKL and OPG concentrations in wild-type mice and mice lacking OPG (*Tnfrsf11b^{-/-}*), as measured by ELISA (n=5 for each genotype). Bars in FIG. 13 represent means \pm SD. “*”, P<0.05; “**”, P<0.01” show the results compared with that of the control. Compared with control heterozygous mice lacking OPG, physiological serum soluble RANKL concentration was approximately 10 times higher in *Tnfrsf11b^{-/-}* mice. Therefore, it can be said that mice lacking OPG are always exposed to high soluble RANKL concentrations.

Furthermore, LPS (2 μ g/g body weight) was injected intraperitoneally. Ninety (90) minutes later, blood was collected

from wild-type mice and *Tnfrsf11b^{-/-}* mice and then serum cytokines were measured by ELISA. The results are shown in FIG. 14A to C. Bars in FIG. 14 represent means \pm SD. “*”, P<0.05; “**”, P<0.01” show the results compared with that of the control. It was revealed that after LPS injection, IL-6 production significantly decreased in *Tnfrsf11b^{-/-}* mice. Therefore, it was demonstrated that chronic exposure to high soluble RANKL concentrations suppresses inflammatory cytokine production in mice lacking OPG.

(5) Hypersensitivity of Mice Lacking RANKL to LPS

To analyze the effects of abnormal inflammatory cytokine production in mice lacking RANKL, we intraperitoneally injected a high concentration of LPS into wild-type mice and *Tnfrsf11^{-/-}* mice and then compared the resulting survival percentages. A high dose of LPS (130 μ g/g body weight) was intraperitoneally injected into 6-week-old wild-type mice (n=10) and *Tnfrsf11^{-/-}* mice (n=5), and then survival curves were generated. As a result, while 90% of wild-type mice (n=10, open square) survived at 24 hours after injection, all *Tnfrsf11^{-/-}* mice (n=5, close square) died within 24 hours (FIG. 15). *Tnfrsf11^{-/-}* mice were hypersensitive to LPS. These results suggest that physiological serum soluble RANKL functions to protect mice from endotoxin shock.

(6) Decreased TLR4 by Soluble RANKL

When macrophages were treated with soluble RANKL for 24 hours, the mRNA level of TLR4 was significantly suppressed. Even when macrophages were treated with soluble RANKL, stimulated with LPS, and then the mRNA level of TLR4 at this time was observed with time, the same tendency was observed (FIG. 16A). Bars in FIG. 16 represent means \pm SD. Moreover, it was revealed by flow cytometry analysis that 24 hours of treatment of macrophages with soluble RANKL suppresses the expression of TLR4-MD2 complex on cell surfaces (FIG. 16B). Since TLR4 is an LPS receptor, it was suggested that such a decreased TLR4 level plays a role in the mechanism of RANKL tolerance.

(7) Suppression of Antigen Presentation by Soluble RANKL

It was revealed by flow cytometry analysis that 24 hours of treatment of macrophages with soluble RANKL results in significant decreases in the expression of MHCII, CD80, and CD86 on the surfaces (FIG. 17, continuous lines represent the results of the group treated with soluble RANKL and broken lines (gray) represent the results of the control group). MHCII is a molecule for presenting foreign antigens to lymphocytes and co-stimulatory molecules such as CD80 and CD86 are considered to be essential at this time.

Therefore, decreased expression levels of these molecules suggest that soluble RANKL is capable of suppressing antigen presentation by macrophages.

(8) c-Fos-independent RANKL Tolerance

Transduction of RANKL/RANK signal in osteoclast differentiation requires the c-Fos transcription factor. Mice lacking c-Fos are unable to produce osteoclasts. To examine whether or not c-Fos is required for RANKL tolerance to occur, macrophages derived from mice lacking c-Fos were analyzed. Macrophages derived from mice lacking c-Fos were pretreated with 1 ng/ml LPS for 24 hours and then stimulated with 100 ng/ml LPS to observe LPS tolerance. It was revealed that LPS tolerance occurred similarly to the case of the wild-type mice (FIGS. 18 A and B). Moreover, macrophages derived from the same were pretreated with 10 ng/ml soluble RANKL for 24 hours and then stimulated with 100 ng/ml LPS to observe RANKL tolerance. Thus, it was also revealed that RANKL tolerance occurred similarly to the case of the wild-type mice (FIGS. 18 C and D). Bars in FIG. 18 represent means \pm SD. Therefore, it can be concluded that LPS tolerance or RANKL tolerance is c-Fos-independent.

That is, RANKL tolerance should not be considered to be only the "phenomenon of macrophage differentiation" in the process during which macrophages differentiate into multinucleated osteoclasts.

(9) Mouse Acute Toxicity Test

Mice used herein were 7- to 8-week-old C57BL/6N (female) mice.

LPS used herein was Lipopolysaccharides from *Escherichia coli* O55:B5 (Sigma). Sterile water (1.0 ml) was added to 10 mg each of LPS for dissolution and then the solution was used. M-CSF used herein was Leukoprol (KYOWA HAKKO KOGYO Co., Ltd.). PBS (1.0 ml) was added to 50 µg each of M-CSF for dissolution and then the solution was used.

Preparation of GST-RANKL

Sal I, Not I site was added to cDNA encoding human-type RANKL residues 140 to 317 by PCR. With the use of these endonucleases, the sequence was cloned downstream of Glutathione S-transferase of pGEX-4T-2 (GE healthcare; Genbank Accession Number U13854). The nucleotide sequence of cDNA encoding human-type RANKL residues 140 to 317 (SEQ ID No: 14) and the corresponding amino acid sequence (SEQ ID No: 10) are shown in FIG. 19. Moreover, the restriction enzyme map of a vector containing the RANKL gene (SEQ ID No: 15) is shown in FIG. 20. Furthermore, the nucleotide sequence of the vector (SEQ ID No: 11) is shown in FIGS. 21A and 21B. The nucleotide sequence in FIG. 21B is continued from the nucleotide sequence in FIG. 21A. After induction of IPTG (final concentration: 0.5 mM)-mediated protein expression in BL21 (DE3) *Escherichia coli* (Invitrogen), microbes were suspended in an extraction buffer (50 mM Tris-HCl, pH 8.0, 100 mM NaCl, 1 mM EDTA, 1 mM DTT, and 1% (v/v) TritonX-100) and then disrupted at 4° C.

using an ultrasonicator. After centrifugation at 18000xg for 15 minutes, the supernatants were collected and then subjected to a Glutathione Sepharose (Trademark) column. Subsequently, the resultants were washed with a buffer for washing (50 mM Tris-HCl, pH 8.0, 100 mM NaCl, 1 mM DTT, and 0.1% (v/v) TritonX-100). Subsequently, elution was performed with a Glutathione solution (20 mM reduced glutathione, 50 mM Tris-HCl, and pH 8.0). The molecular weight and the purity of GST-RANKL purified by SDS-PAGE were confirmed, followed by filtration. The molecular weight was 47.0 kDa and the purity was 95% or higher. Furthermore, endotoxin concentrations were measured by limulus ameocyte lysate assay, so that the concentrations were confirmed to be less than 1 EU/µg.

A method for constructing a GST-RANKL expression vector is shown in FIG. 22.

Test of Administration of LPS, RANKL, and M-CSF

Ten (10) µg of GST-RANKL, 2 µg of M-CSF, 2 µg of M-CSF+10 µg of GST-RANKL, and PBS as a control were separately administered intraperitoneally to C57BL/6N mice (9 to 10 mice per group). After 24 hours, LPS (2.5 mg/mouse) was administered intraperitoneally. Subsequent survival was observed until hour 144 after the administration.

The results are shown in FIG. 23. As shown in FIG. 23, survival % was significantly high (P<0.03) when RANKL and M-CSF had been administered.

All publications, patents, and patent applications cited herein are incorporated herein by reference in their entirety. Sequence Listing Free Text

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SEQ ID NOS: 7 and 8: GST-RANKL (aa159-317)

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Ser Gly Ser His Lys Val Ser Leu Ser Ser Trp Tyr His Asp Arg Gly
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Trp Ala Lys Ile Ser Asn Met Thr Phe Ser Asn Gly Lys Leu Ile Val
          195          200          205
Asn Gln Asp Gly Phe Tyr Tyr Leu Tyr Ala Asn Ile Cys Phe Arg His
          210          215          220
His Glu Thr Ser Gly Asp Leu Ala Thr Glu Tyr Leu Gln Leu Met Val
          225          230          235          240
Tyr Val Thr Lys Thr Ser Ile Lys Ile Pro Ser Ser His Thr Leu Met
          245          250          255
Lys Gly Gly Ser Thr Lys Tyr Trp Ser Gly Asn Ser Glu Phe His Phe
          260          265          270
Tyr Ser Ile Asn Val Gly Gly Phe Phe Lys Leu Arg Ser Gly Glu Glu
          275          280          285
    
```

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Ile Ser Ile Glu Val Ser Asn Pro Ser Leu Leu Asp Pro Asp Gln Asp
 290 295 300

Ala Thr Tyr Phe Gly Ala Phe Lys Val Arg Asp Ile Asp
 305 310 315

<210> SEQ ID NO 3
 <211> LENGTH: 1275
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: GST-RANKL
 (aal27-317)

<400> SEQUENCE: 3

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atgtccccta tactaggtta ttggaaaatt aagggccttg tgcaaccac tcgacttctt    60
ttggaatadc ttgaagaaaa atatgaagag catttgatg agcgcgatga aggtgataaa    120
tggcgaaaaca aaaagtttga attgggtttg gagtttccca atcttcctta ttatattgat    180
ggtgatgta aattaacaca gtctatggcc atcatacgtt atatagctga caagcacaac    240
atgttgggtg gttgtccaaa agagcgtgca gagatttcaa tgcttgaagg agcggttttg    300
gatattagat acggtgtttc gagaattgca tatagtaaag accttgaaac tctcaaagtt    360
gattttctta gcaagctacc tgaaatgctg aaaatgttcg aagatcgttt atgtcataaa    420
acatatttaa atgggtgatca tgtaaccocat cctgacttca tgttgatga cgctcttgat    480
gttgttttat acatggacc aatgtgctg gatgcgttcc caaaattagt ttgttttaaa    540
aaacgtattg aagctatccc acaaattgat aagtacttga aatccagcaa gtatatagca    600
tggcctttgc agggctggca agccacgttt ggtggtggcg accatcctcc aaaatcggat    660
ctggttcctg gtggatcccc aggaattccc gggtcgactg tgcaaaagga attacaacat    720
atcgttggat cacagcacat cagagcagag aaagcgatgg tggatggctc atggttagat    780
ctggccaaga ggagcaagct tgaagctcag ccttttgctc atctcactat taatgccacc    840
gacatcccat ctggttccca taaagtgagt ctgtcctctt ggtaccatga tcggggttgg    900
gccaagatct ccaacatgac ttttagcaat ggaaaactaa tagttaatca ggaaggcttt    960
tattacctgt atgccaacat ttgctttoga catcatgaaa cttcaggaga cctagetaca   1020
gagtatcttc aactaatggt gtacgtcact aaaaccagca tcaaaatccc aagttctcat   1080
accctgatga aaggaggaag caccaagtat tggtcagga attctgaatt ccatttttat   1140
tcataaaacg ttggtggatt ttttaagtta cggctctggag aggaaatcag catcgaggtc   1200
tccaaccctc ccttactgga tccggatcag gatgcaacat accttggggc ttttaaagtt   1260
cgagatatag attga                                           1275
    
```

<210> SEQ ID NO 4
 <211> LENGTH: 442
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: GST-RANKL
 (aal27-317)

<400> SEQUENCE: 4

Met Ser Pro Ile Leu Gly Tyr Trp Lys Ile Lys Gly Leu Val Gln Pro
 1 5 10 15

Thr Arg Leu Leu Leu Glu Tyr Leu Glu Glu Lys Tyr Glu Glu His Leu
 20 25 30

Tyr Glu Arg Asp Glu Gly Asp Lys Trp Arg Asn Lys Lys Phe Glu Leu

-continued

35					40					45					
Gly	Leu	Glu	Phe	Pro	Asn	Leu	Pro	Tyr	Tyr	Ile	Asp	Gly	Asp	Val	Lys
50					55					60					
Leu	Thr	Gln	Ser	Met	Ala	Ile	Ile	Arg	Tyr	Ile	Ala	Asp	Lys	His	Asn
65					70					75					80
Met	Leu	Gly	Gly	Cys	Pro	Lys	Glu	Arg	Ala	Glu	Ile	Ser	Met	Leu	Glu
				85					90					95	
Gly	Ala	Val	Leu	Asp	Ile	Arg	Tyr	Gly	Val	Ser	Arg	Ile	Ala	Tyr	Ser
			100					105					110		
Lys	Asp	Phe	Glu	Thr	Leu	Lys	Val	Asp	Phe	Leu	Ser	Lys	Leu	Pro	Glu
		115					120					125			
Met	Leu	Lys	Met	Phe	Glu	Asp	Arg	Leu	Cys	His	Lys	Thr	Tyr	Leu	Asn
130						135					140				
Gly	Asp	His	Val	Thr	His	Pro	Asp	Phe	Met	Leu	Tyr	Asp	Ala	Leu	Asp
145					150					155					160
Val	Val	Leu	Tyr	Met	Asp	Pro	Met	Cys	Leu	Asp	Ala	Phe	Pro	Lys	Leu
				165					170					175	
Val	Cys	Phe	Lys	Lys	Arg	Ile	Glu	Ala	Ile	Pro	Gln	Ile	Asp	Lys	Tyr
			180					185						190	
Leu	Lys	Ser	Ser	Lys	Tyr	Ile	Ala	Trp	Pro	Leu	Gln	Gly	Trp	Gln	Ala
				195				200					205		
Thr	Phe	Gly	Gly	Gly	Asp	His	Pro	Pro	Lys	Ser	Asp	Leu	Val	Pro	Arg
210						215					220				
Gly	Ser	Pro	Gly	Ile	Pro	Gly	Ser	Thr	Arg	Ala	Ala	Ala	Ser	Leu	Val
225						230					235				240
Pro	Arg	Gly	Ser	Pro	Gly	Ile	Pro	Gly	Ser	Thr	Val	Gln	Lys	Glu	Leu
				245					250					255	
Gln	His	Ile	Val	Gly	Ser	Gln	His	Ile	Arg	Ala	Glu	Lys	Ala	Met	Val
			260					265						270	
Asp	Gly	Ser	Trp	Leu	Asp	Leu	Ala	Lys	Arg	Ser	Lys	Leu	Glu	Ala	Gln
			275				280						285		
Pro	Phe	Ala	His	Leu	Thr	Ile	Asn	Ala	Thr	Asp	Ile	Pro	Ser	Gly	Ser
			290				295					300			
His	Lys	Val	Ser	Leu	Ser	Ser	Trp	Tyr	His	Asp	Arg	Gly	Trp	Ala	Lys
305						310					315				320
Ile	Ser	Asn	Met	Thr	Phe	Ser	Asn	Gly	Lys	Leu	Ile	Val	Asn	Gln	Asp
				325					330					335	
Gly	Phe	Tyr	Tyr	Leu	Tyr	Ala	Asn	Ile	Cys	Phe	Arg	His	His	Glu	Thr
				340					345					350	
Ser	Gly	Asp	Leu	Ala	Thr	Glu	Tyr	Leu	Gln	Leu	Met	Val	Tyr	Val	Thr
			355				360					365			
Lys	Thr	Ser	Ile	Lys	Ile	Pro	Ser	Ser	His	Thr	Leu	Met	Lys	Gly	Gly
				370			375					380			
Ser	Thr	Lys	Tyr	Trp	Ser	Gly	Asn	Ser	Glu	Phe	His	Phe	Tyr	Ser	Ile
385						390					395				400
Asn	Val	Gly	Gly	Phe	Phe	Lys	Leu	Arg	Ser	Gly	Glu	Glu	Ile	Ser	Ile
				405					410					415	
Glu	Val	Ser	Asn	Pro	Ser	Leu	Leu	Asp	Pro	Asp	Gln	Asp	Ala	Thr	Tyr
				420					425					430	
Phe	Gly	Ala	Phe	Lys	Val	Arg	Asp	Ile	Asp						
				435				440							

<210> SEQ ID NO 5

<211> LENGTH: 1236

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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: GST-RANKL
(aal140-317)

<400> SEQUENCE: 5
atgtccccta tactaggtta ttggaaaatt aagggccttg tgcaaccac tgcacttctt    60
ttggaatata ttgaagaaaa atatgaagag catttgtatg agcgcgatga aggtgataaa    120
tggcgaaaaca aaaagtgtga attgggtttg gagtttccca atcttcctta ttatattgat    180
ggatgatgta aattaacaca gtctatggcc atcatacgtt atatagctga caagcacaac    240
atgttgggtg gttgtccaaa agagcgtgca gagatttcaa tgcttgaagg agcgggtttg    300
gatattagat acgggtgttc gagaattgca tatagtaaag accttgaaac tctcaaagtt    360
gattttctta gcaagctacc tgaaatgctg aaaatgttcg aagatcgttt atgtcataaa    420
acatatntaa atgggtgatc tgtaacccat cctgacttca tgtgtatga cgtctttgat    480
gttgttttat acatggaccc aatgtgcctg gatgcgttcc caaaattagt ttgttttaa    540
aaacgtattg aagctatccc acaaattgat aagtacttga aatccagcaa gtatatagca    600
tggcctttgc agggctggca agccacgttt ggtggtggcg accatcctcc aaaatcggat    660
ctggttccgc gtggatcccc aggaattccc ggttcgacta tcagagcaga gaaagcgatg    720
gtggatggct catggttaga tctggccaag aggagcaagc ttgaagctca gctttttgct    780
catctcacta ttaatgccac cgacatccca tctggttccc ataaagtgag tctgtcctct    840
tggttaccatg atcgggggtg ggccaagatc tccaacatga cttttagcaa tggaaaacta    900
atagttaatc aggatggctt ttattacctg tatgccaaca tttgctttcg acatcatgaa    960
acttcaggag acctagctac agagtatcct caactaatgg tgtacgtcac taaaaccagc   1020
atcaaaatcc caagttctca taccctgatg aaaggaggaa gcaccaagta ttggtcaggg   1080
aattctgaat tccattttta ttccataaac gttggtggat tttttaagtt acggtctgga   1140
gaggaaatca gcatcgaggt ctccaacccc tctttactgg atccggatca ggatgcaaca   1200
tactttgggg cttttaaagt tcgagatata gattga                               1236
    
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<210> SEQ ID NO 6
<211> LENGTH: 429
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: GST-RANKL
(aal140-317)
    
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<400> SEQUENCE: 6
Met Ser Pro Ile Leu Gly Tyr Trp Lys Ile Lys Gly Leu Val Gln Pro
  1           5           10           15
Thr Arg Leu Leu Leu Glu Tyr Leu Glu Glu Lys Tyr Glu Glu His Leu
  20           25           30
Tyr Glu Arg Asp Glu Gly Asp Lys Trp Arg Asn Lys Lys Phe Glu Leu
  35           40           45
Gly Leu Glu Phe Pro Asn Leu Pro Tyr Tyr Ile Asp Gly Asp Val Lys
  50           55           60
Leu Thr Gln Ser Met Ala Ile Ile Arg Tyr Ile Ala Asp Lys His Asn
  65           70           75           80
Met Leu Gly Gly Cys Pro Lys Glu Arg Ala Glu Ile Ser Met Leu Glu
  85           90           95
Gly Ala Val Leu Asp Ile Arg Tyr Gly Val Ser Arg Ile Ala Tyr Ser
    
```

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	100		105		110	
Lys	Asp	Phe	Glu	Thr	Leu	Lys
	115					120
Val	Asp	Phe	Leu	Ser	Lys	Leu
	120				125	
Met	Leu	Lys	Met	Phe	Glu	Asp
	130					135
Arg	Leu	Cys	His	Lys	Thr	Tyr
	135			140		
Gly	Asp	His	Val	Thr	His	Pro
	145				150	
Asp	Phe	Met	Leu	Tyr	Asp	Ala
	145				155	160
Val	Val	Leu	Tyr	Met	Asp	Pro
				165		
Cys	Leu	Asp	Ala	Phe	Pro	Lys
				170		175
Val	Cys	Phe	Lys	Lys	Arg	Ile
			180			
Glu	Ala	Ile	Pro	Gln	Ile	Asp
			185			190
Lys	Tyr	Lys	Tyr	Ile	Ala	Trp
					200	
Pro	Leu	Gln	Gly	Trp	Gln	Ala
			205			
Thr	Phe	Gly	Gly	Gly	Asp	His
	210				215	
Pro	Pro	Lys	Ser	Asp	Leu	Val
				220		Pro
Gly	Ser	Pro	Gly	Ile	Pro	Gly
	225				230	
Ser	Thr	Arg	Ala	Ala	Ala	Ser
			235			240
Pro	Arg	Gly	Ser	Pro	Gly	Ile
				245		
Ser	Thr	Ile	Arg	Ala	Glu	Lys
				250		255
Ala	Met	Val	Asp	Gly	Ser	Trp
			260			
Leu	Asp	Leu	Ala	Lys	Arg	Ser
			265			270
Glu	Ala	Gln	Pro	Phe	Ala	His
					280	
Leu	Thr	Ile	Asn	Ala	Thr	Asp
					285	
Ser	Gly	Ser	His	Lys	Val	Ser
	290					295
Ser	Ser	Trp	Tyr	His	Asp	Arg
			300			
Trp	Ala	Lys	Ile	Ser	Asn	Met
	305					310
Thr	Phe	Ser	Asn	Gly	Lys	Leu
			315			
Ile	Val	Ile	Val	Ile	Val	Val
						320
Asn	Gln	Asp	Gly	Phe	Tyr	Tyr
				325		
Leu	Tyr	Ala	Asn	Ile	Cys	Phe
				330		
Arg	His	Arg	His	Arg	His	His
				335		
His	Glu	Thr	Ser	Gly	Asp	Leu
				340		
Ala	Thr	Glu	Tyr	Leu	Gln	Leu
				345		
Met	Val	Met	Val	Met	Val	Val
				350		
Tyr	Val	Thr	Lys	Thr	Ser	Ile
				355		
Lys	Ile	Lys	Ile	Pro	Ser	Ser
				360		
His	Thr	Leu	Met	Val	His	Thr
				365		
Lys	Gly	Gly	Ser	Thr	Lys	Tyr
	370					375
Trp	Ser	Gly	Asn	Ser	Glu	Phe
				380		
His	Phe	His	Phe	Lys	Leu	Arg
				385		
Ser	Gly	Glu	Glu	Glu	Glu	Glu
				400		
Ile	Ser	Ile	Glu	Val	Ser	Asn
				405		
Pro	Ser	Leu	Leu	Asp	Pro	Asp
				410		
Gln	Asp	Gln	Asp	Gln	Asp	Gln
				415		
Ala	Thr	Tyr	Phe	Gly	Ala	Phe
				420		
Lys	Val	Arg	Asp	Ile	Asp	
				425		

<210> SEQ ID NO 7
 <211> LENGTH: 1179
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: GST-RANKL
 (aal159-317)

<400> SEQUENCE: 7
 atgtccccta tactaggtta ttggaaaatt aaggccttg tgcaaccac tcgacttctt 60
 ttggaatadc ttgaagaaaa atatgaagag catttgtatg agcgcgatga agtgataaa 120
 tggcgaaaca aaaagtttga attgggtttg gagtttccca atcttctta ttatattgat 180
 ggtgatgtta aattaacaca gtctatggcc atcatagctt atatagctga caagcacaac 240

-continued

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atgttgggtg gttgtccaaa agagcgtgca gagatttcaa tgcttgaagg agcgggttttg 300
gatattagat acgggtgttcc gagaattgca tatagtaaag actttgaaac tctcaaagtt 360
gattttctta gcaagctacc tgaaatgctg aaaatgttcg aagatcgttt atgtcataaa 420
acatatataa atgggtgatca tgtaacccat cctgacttca tgttgatga cgctcttgat 480
gttgttttat acatggaccc aatgtgcctg gatgcgttcc caaaattagt ttgttttaaa 540
aaacgtattg aagctatccc acaaattgat aagtacttga aatccagcaa gtatatagca 600
tggcctttgc agggctggca agccacgttt ggtggtggcg accatcctcc aaaatcggat 660
ctggttccgc gtggatcccc aggaattccc gggtegacta agcttgaagc tcagcctttt 720
gctcatctca ctattaatgc caccgacatc ccatctggtt ccataaaagt gagtctgtcc 780
tcttgggtacc atgatcgggg ttgggccaag atctccaaca tgacttttag caatggaaaa 840
ctaatagtta atcaggatgg cttttattac ctgtatgcca acatttgctt tcgacatcat 900
gaaacttcag gagacctagc tacagagtat cttcaactaa tgggtgacgt cactaaaacc 960
agcatcaaaa tcccagtttc tcataacctg atgaaaggag gaagcaccaa gtattggtca 1020
gggaattctg aattccattt ttattccata aacgttggtg gattttttaa gttacggctc 1080
ggagaggaaa tcagcatcga ggtctccaac cctccttac tggatccgga tcaggatgca 1140
acatactttg gggcttttaa agttcgagat atagattga 1179

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<210> SEQ ID NO 8
<211> LENGTH: 410
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: GST-RANKL
(aal159-317)

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<400> SEQUENCE: 8

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Met Ser Pro Ile Leu Gly Tyr Trp Lys Ile Lys Gly Leu Val Gln Pro
  1           5           10           15
Thr Arg Leu Leu Leu Glu Tyr Leu Glu Glu Lys Tyr Glu Glu His Leu
  20           25           30
Tyr Glu Arg Asp Glu Gly Asp Lys Trp Arg Asn Lys Lys Phe Glu Leu
  35           40           45
Gly Leu Glu Phe Pro Asn Leu Pro Tyr Tyr Ile Asp Gly Asp Val Lys
  50           55           60
Leu Thr Gln Ser Met Ala Ile Ile Arg Tyr Ile Ala Asp Lys His Asn
  65           70           75           80
Met Leu Gly Gly Cys Pro Lys Glu Arg Ala Glu Ile Ser Met Leu Glu
  85           90           95
Gly Ala Val Leu Asp Ile Arg Tyr Gly Val Ser Arg Ile Ala Tyr Ser
 100           105           110
Lys Asp Phe Glu Thr Leu Lys Val Asp Phe Leu Ser Lys Leu Pro Glu
 115           120           125
Met Leu Lys Met Phe Glu Asp Arg Leu Cys His Lys Thr Tyr Leu Asn
 130           135           140
Gly Asp His Val Thr His Pro Asp Phe Met Leu Tyr Asp Ala Leu Asp
 145           150           155           160
Val Val Leu Tyr Met Asp Pro Met Cys Leu Asp Ala Phe Pro Lys Leu
 165           170           175
Val Cys Phe Lys Lys Arg Ile Glu Ala Ile Pro Gln Ile Asp Lys Tyr
 180           185           190

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Leu Lys Ser Ser Lys Tyr Ile Ala Trp Pro Leu Gln Gly Trp Gln Ala
 195 200 205

Thr Phe Gly Gly Gly Asp His Pro Pro Lys Ser Asp Leu Val Pro Arg
 210 215 220

Gly Ser Pro Gly Ile Pro Gly Ser Thr Arg Ala Ala Ala Ser Leu Val
 225 230 235 240

Pro Arg Gly Ser Pro Gly Ile Pro Gly Ser Thr Lys Leu Glu Ala Gln
 245 250 255

Pro Phe Ala His Leu Thr Ile Asn Ala Thr Asp Ile Pro Ser Gly Ser
 260 265 270

His Lys Val Ser Leu Ser Ser Trp Tyr His Asp Arg Gly Trp Ala Lys
 275 280 285

Ile Ser Asn Met Thr Phe Ser Asn Gly Lys Leu Ile Val Asn Gln Asp
 290 295 300

Gly Phe Tyr Tyr Leu Tyr Ala Asn Ile Cys Phe Arg His His Glu Thr
 305 310 315 320

Ser Gly Asp Leu Ala Thr Glu Tyr Leu Gln Leu Met Val Tyr Val Thr
 325 330 335

Lys Thr Ser Ile Lys Ile Pro Ser Ser His Thr Leu Met Lys Gly Gly
 340 345 350

Ser Thr Lys Tyr Trp Ser Gly Asn Ser Glu Phe His Phe Tyr Ser Ile
 355 360 365

Asn Val Gly Gly Phe Phe Lys Leu Arg Ser Gly Glu Glu Ile Ser Ile
 370 375 380

Glu Val Ser Asn Pro Ser Leu Leu Asp Pro Asp Gln Asp Ala Thr Tyr
 385 390 395 400

Phe Gly Ala Phe Lys Val Arg Asp Ile Asp
 405 410

<210> SEQ ID NO 9
 <211> LENGTH: 587
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic

<400> SEQUENCE: 9

gtcgactatc agagcagaga aagcgatggt ggatggctca tggtagatc tggccaagag 60
 gagcaagctt gaagctcagc cttttgctca tctcactatt aatgccaccg acatcccac 120
 tggttcccat aaagtgagtc tgtcctcttg gtaccatgat cggggttggg ccaagatctc 180
 caacatgact tttagcaatg gaaaactaat agttaatcag gatggctttt attacctgta 240
 tgccaacatt tgctttcgac atcatgaaac ttcaggagac ctgctacag agtatcttca 300
 actaatggtg tacgtcacta aaaccagcat caaaatccca agttctcata cctgatgaa 360
 aggaggaagc accaagtatt ggtcagggaa ttctgaatto ctttttatt ccataaacgt 420
 tgggtgattt ttaagttac ggtctggaga ggaaatcagc atcgaggtct ccaaccctc 480
 cttactggat ccggatcagg atgcaacata ctttggggct ttaaagtgc gagatataga 540
 ttgagcccca gtttttgag tgttatgtat ttctggatg cggccgc 587

<210> SEQ ID NO 10
 <211> LENGTH: 178
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic

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<400> SEQUENCE: 10

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Ile Arg Ala Glu Lys Ala Met Val Asp Gly Ser Trp Leu Asp Leu Ala
 1           5           10           15
Lys Arg Ser Lys Leu Glu Ala Gln Pro Phe Ala His Leu Thr Ile Asn
 20           25           30
Ala Thr Asp Ile Pro Ser Gly Ser His Lys Val Ser Leu Ser Ser Trp
 35           40           45
Tyr His Asp Arg Gly Trp Ala Lys Ile Ser Asn Met Thr Phe Ser Asn
 50           55           60
Gly Lys Leu Ile Val Asn Gln Asp Gly Phe Tyr Tyr Leu Tyr Ala Asn
 65           70           75           80
Ile Cys Phe Arg His His Glu Thr Ser Gly Asp Leu Ala Thr Glu Tyr
 85           90           95
Leu Gln Leu Met Val Tyr Val Thr Lys Thr Ser Ile Lys Ile Pro Ser
 100          105          110
Ser His Thr Leu Met Lys Gly Gly Ser Thr Lys Tyr Trp Ser Gly Asn
 115          120          125
Ser Glu Phe His Phe Tyr Ser Ile Asn Val Gly Gly Phe Phe Lys Leu
 130          135          140
Arg Ser Gly Glu Glu Ile Ser Ile Glu Val Ser Asn Pro Ser Leu Leu
 145          150          155          160
Asp Pro Asp Gln Asp Ala Thr Tyr Phe Gly Ala Phe Lys Val Arg Asp
 165          170          175

Ile Asp
    
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<210> SEQ ID NO 11
<211> LENGTH: 4970
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
    
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<400> SEQUENCE: 11

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acgttatcga ctgcacgggtg caccaatgct tctggcgctca ggcagccatc ggaagctgtg      60
gtatggctgt gcaggtcgta aatcactgca taattcgtgt cgctcaaggc gcaactcccgt      120
tctggataat gttttttgcg cgcacatcat aacggttctg gcaaatattc tgaaatgagc      180
tggtgacaat taatcatcgg ctcgataaat gtgtggaatt gtgagcggat aacaatttca      240
cacaggaaac agtattcatg tcccctatac taggttattg gaaaattaag ggccttgtgc      300
aaccactcgc acttcttttg gaatatcttg aagaaaaata tgaagagcat ttgtatgagc      360
gogatgaagg tgataaatgg cgaaacaaaa agtttgaatt gggtttgag tttcccaatc      420
ttccttatta tattgatggt gatgttaaat taacacagtc tatggccatc atacgttata      480
tagctgacaa gcacaacatg ttgggtgggt gtccaaaaga gcgtgcagag atttcaatgc      540
ttgaaggagc ggttttggat attagatcac gtgtttcgag aattgcatat agtaaagact      600
ttgaaactct caaagttgat tttcttagca agctacctga aatgctgaaa atgttcgaag      660
atcgtttatg tcataaaaca tatttaaatg gtgatcatgt aaccatcct gacttcatgt      720
tgtatgacgc tcttgatggt gttttataca tggacceaat gtgcctggat gcgtteccaa      780
aattagtttg ttttaaaaaa cgtattgaag ctatcccaca aattgataag tacttgaat      840
ccagcaagta tatagcatgg cctttgcagg gctggcaagc cacgtttggt ggtggcgacc      900
atcctccaaa atcggatctg gttccgcgtg gatccccagg aattcccggg tcgactcgag      960
cgccgcgcatc gtgactgact gacgatctgc ctgcgcgctt tcggtgatga cggtgaaaac     1020
    
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The invention claimed is:

1. A method for detection of a bacterial infectious disease or sepsis with use of RANKL as a marker in a biological blood sample, the method comprising:
 - (i) measuring by immunoassay a concentration of soluble RANKL in the biological blood sample isolated from a subject; and
 - (ii) determining that the subject is affected with the bacterial infectious disease or sepsis when the concentration of the soluble RANKL in the biological blood sample is lower than that of a normal subject,
 wherein the bacterial infectious disease is a salmonella infectious disease, and the sepsis is lipopolysaccharide or (LPS)-induced sepsis.
2. The method for detection of a bacterial infectious disease or sepsis according to claim 1, wherein the method uses

OPG in addition to RANKL as a marker in the biological blood sample, and the method comprises:

- (i) measuring by immunoassay a concentration of OPG in the biological blood sample isolated from the subject in addition to the concentration of soluble RANKL; and
 - (ii) determining that the subject is affected with bacterial infectious disease or sepsis when a ratio of the concentration of the soluble RANKL relative to the concentration of the OPG in the biological blood sample is lower than the ratio of a normal subject.
3. The method for detection of a bacterial infectious disease or sepsis according to claim 2, the method comprising a step of isolating the blood sample from the subject.
 4. The method for detection of a bacterial infectious disease or sepsis according to claim 1, the method comprising a step of isolating the blood sample from the subject.

* * * * *

专利名称(译)	用于预防或治疗炎性疾病的炎性疾病和组合物的检测		
公开(公告)号	US8257933	公开(公告)日	2012-09-04
申请号	US12/227244	申请日	2006-05-12
[标]申请(专利权)人(译)	学校法人庆应义塾		
申请(专利权)人(译)	庆应大学		
当前申请(专利权)人(译)	庆应大学 东方酵母有限公司.		
[标]发明人	MARUYAMA KENTA MATSUO KOICHI YASUDA HISATAKA		
发明人	MARUYAMA, KENTA MATSUO, KOICHI YASUDA, HISATAKA		
IPC分类号	G01N33/53 C07K14/705 C07K14/51 G01N33/49		
CPC分类号	A61K38/191 A61K38/193 C12Q1/6883 G01N33/566 G01N2333/70575 G01N2800/26 A61K2300/00 C12Q2600/158 A61P29/00 A61P31/00 A61P31/10 Y02A50/481		
其他公开文献	US20090202469A1		
外部链接	Espacenet USPTO		

摘要(译)

提供了一种用于检测炎性疾病的新方法和用于预防或治疗炎性疾病的新组合。检测炎性疾病的方法包括使用RANKL和/或OPG作为生物样品中的标记物。用于预防或治疗炎性疾病的组合物包含RANKL和/或M-CSF作为活性成分。

