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Venge

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(54) **METHOD FOR ESTIMATION OF THE AMOUNT OF SPECIFIC CELL TYPES**

5,747,265 A * 5/1998 Parsons et al. 435/7.2

(75) Inventor: **Per Venge**, Uppsala (SE)

FOREIGN PATENT DOCUMENTS

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(73) Assignee: **Phadia AB**, Uppsala (SE)

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(*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 504 days.

Moshfegh et al., Methods of Simultaneous Quantitative Analysis of Eosinophil and Neutrophil Adhesion and Transmigration, *Scand. J. Immunol.* 50: 262-269 (1999).*

(21) Appl. No.: **10/178,211**

Carlson et al., Human Eosinophil Peroxidase: purification and characterization (*The Journal of Immunology*, 134 (3): 1875-1879 (Mar. 1985).*

(22) Filed: **Jun. 24, 2002**

McEuen et al., Mass, charge, and subcellular localization of a unique secretory product identified by the basophil-specific antibody BB1 (*J. Allergy Clin Immunol.* 107 : 842-848 (May 2001).*

(65) **Prior Publication Data**

US 2003/0077666 A1 Apr. 24, 2003

Cella et al., Platelet factor 4 (PF4) and heparin released platelet factor 4 (HR-PF4) in diabetes mellitus. Effect of the duration of the disease. *Folia haematologica (Leipzig, Germany:1928)*, 113 (5): 646-654 (1986) (Abstract).*

Related U.S. Application Data

(60) Provisional application No. 60/300,759, filed on Jun. 25, 2001.

Xu et al. The development of an assay for human neutrophil lipocalin (HNL) to be used as a specific marker of neutrophil activity in vivo and vitro, *Journal of Immunological Methods* 171: 245-252 (1994).*

(30) **Foreign Application Priority Data**

Jun. 25, 2001 (SE) 0102220

(Continued)

(51) **Int. Cl.**
G01N 33/53 (2006.01)

Primary Examiner—Gailene R Gabel

(52) **U.S. Cl.** 435/7.1; 435/7.23; 435/7.24; 435/287.2; 436/517; 436/17; 436/172; 436/175; 436/177

(74) *Attorney, Agent, or Firm*—Porter Wright Morris & Arthur LLP

(58) **Field of Classification Search** 435/2, 435/7.1, 7.2, 7.23, 7.24, 7.92, 326, 332, 372.2, 435/372.3, 287.2; 436/517, 519, 10, 17, 436/63, 64, 172, 175, 177

(57) **ABSTRACT**

See application file for complete search history.

The present invention relates to the estimation of the amount of subtypes of specific cells, for example the number of certain subtypes of leukocytes, by measurements of unique proteins in extracts of blood and other biological material. The knowledge of the number or amount of specific subtypes of white cells is important in the clinical diagnosis and surveillance of subjects with inflammatory disease including infectious disease, cancer, allergy/asthma etc.

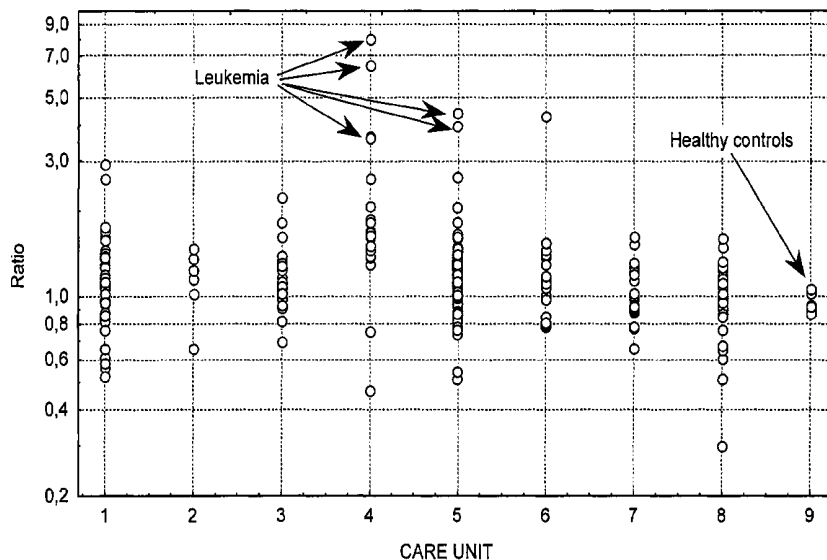
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20 Claims, 3 Drawing Sheets

MPO/LF ratio in a hospitalized population



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Cella et al., Platelet factor 4 (PF4) and heparin released platelet factor 4 (HR-PF4) in diabetes mellitus. Effect of the duration of the disease. *Folia haematologica* (Leipzig, Germany:1928), 113 (5): 646-654 (1986).*

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Peterson et al, *Eur. J. Haematol.*, 40:415-423 (1988).

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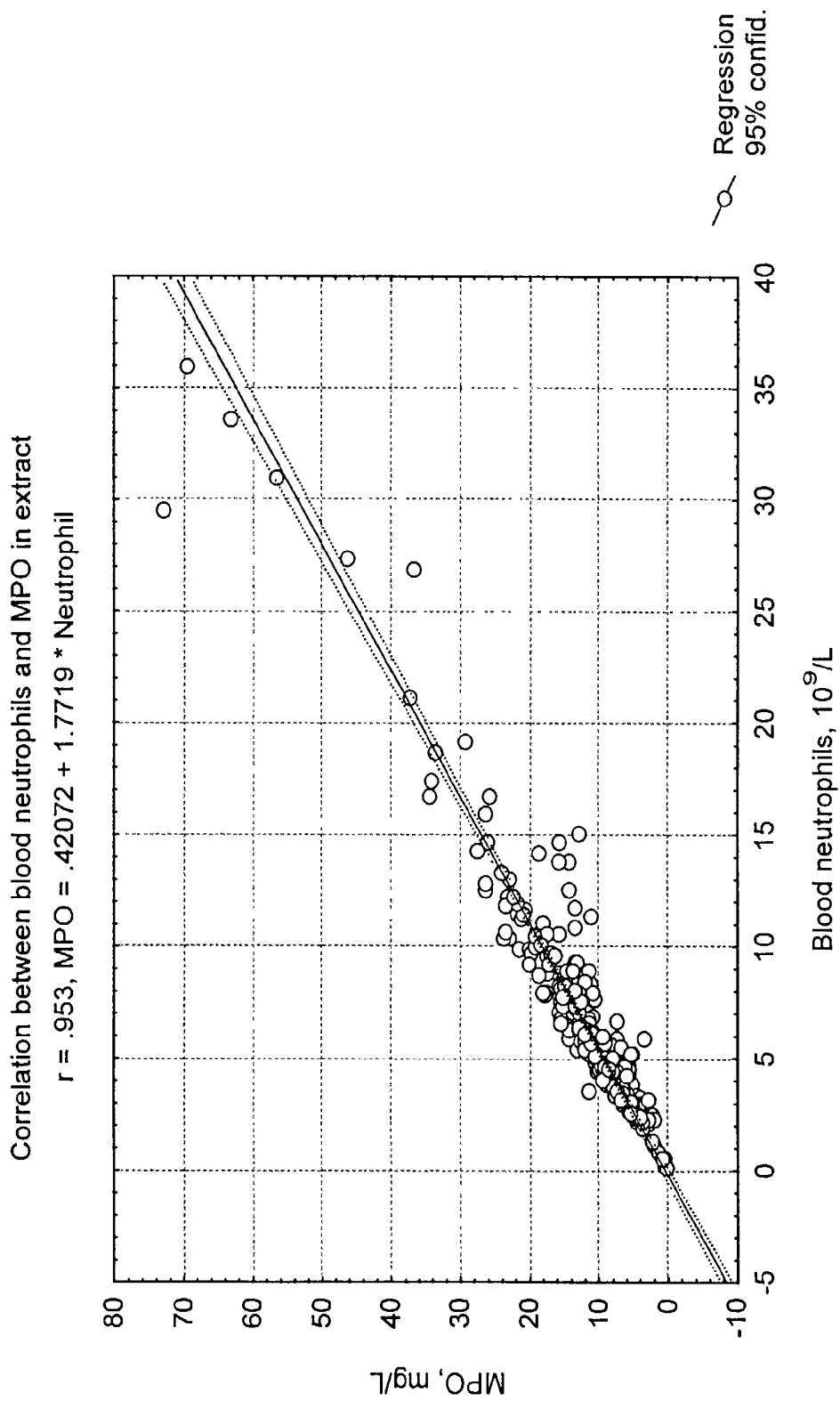


Figure 1

Correlation between blood eosinophil counts and EPO-concentration in extract

$$r=0.954, \text{ EPO} = -1.151 + 210.98 * \text{EOS}$$

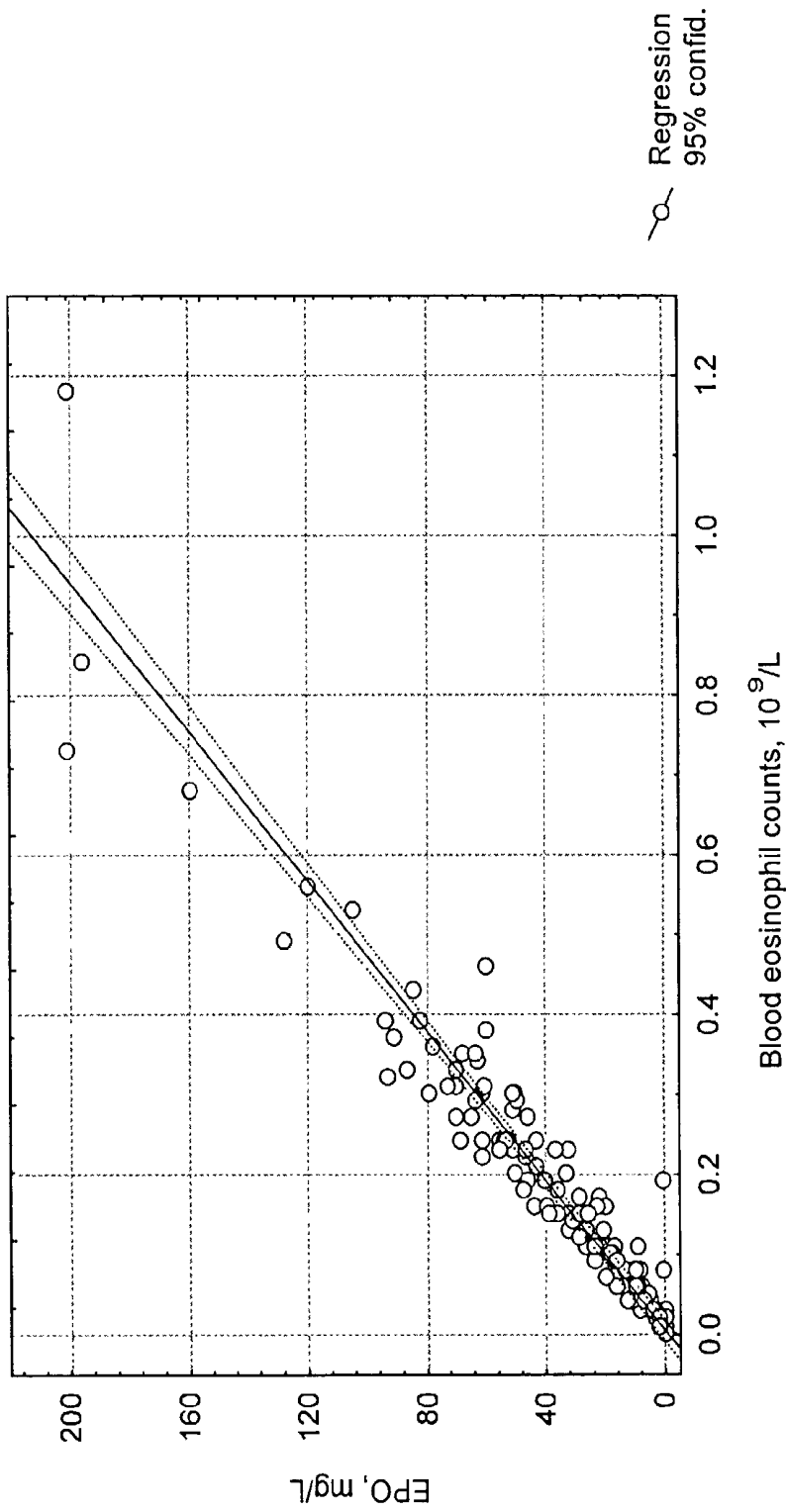


Figure 3

METHOD FOR ESTIMATION OF THE AMOUNT OF SPECIFIC CELL TYPES

RELATED APPLICATION

The present application claims priority under 35 U.S.C. §119 of U.S. application Ser. No. 60/300,759 filed Jun. 25, 2001.

FIELD OF THE INVENTION

The present invention relates to the estimation of the amount of subtypes of specific cells, for example the number of certain subtypes of leukocytes, by measurements of unique proteins in extracts of blood and other biological material. The knowledge of the number or amount of specific subtypes of white cells is important in the clinical diagnosis and surveillance of subjects with inflammatory disease including infectious disease, cancer, allergy/asthma etc.

BACKGROUND OF THE INVENTION

The estimation of the number of various leukocytes in blood and other body fluids is one of the most widely used tools in medicine. The traditional way of obtaining this information is the counting and differentiating of the cells under the light microscope. This technique is complemented by the automated counting in cell counters based on the principle of counting the number of particles in the fluid and the measurement of various physical parameters such as size, forward and side scatter, but also my histochemical staining of cells. An extension of these techniques is the flow cytometer principle in which antibodies are used to identify individual cells based on their cell surface antigens or by means of their content of intracellular antigens after permeabilisation of the cells.

In WO 00/58726 there is described a method for quantitative leukocyte count in whole blood. However, this method does not quantitate different specific subtypes of leukocytes in respect of number or ratio.

SUMMARY OF THE INVENTION

There exists a need of easy-to-use, inexpensive and reliable tests to estimate the number of various white blood cells such as neutrophils and eosinophils, in blood and other body fluids, applicable in the point-of-care situation, thus supporting the medical doctor in his/hers immediate decision-making.

The present inventor has found that the extraction of whole blood with for example detergents such as CTAB (N-cetyl-N,N,N-trimethylammonium bromide), and the subsequent measurement by specific immunoassays of the neutrophil proteins, MPO (myeloperoxidase), HNL (human neutrophil lipocalin) or lactoferrin, or the specific measurements of eosinophil proteins such as EPX (eosinophil protein x) or EPO (eosinophil peroxidase), will accurately identify the number of neutrophils or eosinophils present in the blood. The estimation of the numbers of neutrophils is useful in the diagnosis and monitoring of subjects with inflammatory diseases such as infections, rheumatoid diseases, but also in conjunction with medical treatment, in particular cytostatic treatment, where the reduced production of neutrophils i.e. neutropenia, may occur as a serious adverse effect of treatment. The estimation of eosinophil numbers is useful in patients with allergic disease, chronic inflammatory diseases, parasitic disease, certain cancers such as Hodgkin's disease, but also as a general indicator of disease, since elevated numbers of eosinophils may occur in a number of diseases for unknown reasons.

In the broadest sense the invention means that the number of any given cell population in a body fluid may be possible to estimate, given the availability of immunoassays for molecules that are unique for the cell population to be estimated.

Thus, the invention could be adopted to the estimation of lymphocyte populations in e.g. patients with HIV-infections, cancer, autoimmune disease, but also for the estimation of populations at various maturation stages of myeloid cells, since some intracellular proteins are produced primarily by immature cells and other proteins primarily by more mature cells.

Thus, the invention relates to a method to in vitro estimate the amount, wherein the amount refers to either the number or ratio of specific cell subtypes in a patient sample, comprising

- a) extracting an aliquot of said sample; and
- b) measuring the concentration of cell specific molecule(s) in said extracted sample.

The specific cell subtypes may be subtypes of leukocytes.

The cell specific molecules may for example be extra- and intracellular proteins or antigens, cell surface markers etc.

The sample is preferably blood or other body fluid. A very small amount of sample is extracted, such as 1-10 µl of sample but larger volumes might be considered.

The extraction is preferably with cationic detergent. The extraction time is very short, for example 1 minute. Preferably, the detergent is CTAB.

Preferably, the measuring in step b) is by an immunoassay, such as ELISA (enzyme-linked immunosorbent assay), EIA (enzyme immunoassay), FEIA (fluoroenzyme immunoassay), or RIA (radio immunoassay).

In the method of the invention, the said concentration of said cell specific molecule(s) may be correlated with the number of the respective cell type(s).

In one embodiment, two cell specific molecules are measured in step b) and a ratio between the concentrations of said molecules is determined.

The cell specific molecule(s) may be a neutrophil protein, such as MPO (myeloperoxidase), HNL (human neutrophil lipocalin) or lactoferrin for measuring neutrophils.

The cell specific molecule may also be an eosinophil protein, such as EPX (eosinophil protein x), EPO (eosinophil peroxidase) for measuring eosinophils.

Alternatively, the cell specific molecule is/are a basophil protein, such as BB 1, or a thrombocytic protein, such as β-thromboglobulin, for measuring thrombocytes.

In another alternative the cell specific molecule is/are cell surface markers such as CD20 for measuring B-lymphocytes and CD3 T-lymphocytes or CD4 and CD8 for measuring different lymphocyte populations.

In a further alternative, the cell specific molecule is/are CD14 or lysozyme for measuring monocytes.

DETAILED DESCRIPTION OF THE INVENTION

The present invention will be described more closely below in association with the accompanying drawings in which

FIG. 1 shows a correlation between the number of blood neutrophils and the concentration of MPO protein in detergent extracted whole blood.

FIG. 2 shows ratio between MPO and lactoferrin in detergent extracted whole blood.

FIG. 3 shows a correlation between the number of eosinophils and the concentration of EPO protein in detergent extracted whole blood.

EXPERIMENTAL SECTION

Isolation and Purification of Human Neutrophil and Eosinophil Granule Proteins

Granules were prepared from the buffy coat of granulocytes obtained from healthy blood donors using a modification of the procedure described by Peterson et al (*Eur. J. Haematol.* 40 (1988) 415-423). In brief, the red blood cells were allowed to sediment using Dextran T-500 before collection of the leukocyte rich plasma. Leucocytes were washed twice in 0.34 M Sucrose and the suspended in 5 volumes of 0.34 M Sucrose. The leukocytes were cavitated using N₂ at a pressure of 750 psi for 30 min at +4° C. (Klempner et al., *J. Cell. Biol.* 86 (1980) 21-28; and Borregaard et al., *J. Cell. Biol.* 97 (1983) 52-61). The cavitate was suspended in 0.34 M Sucrose, 0.17 M NaCl and centrifuged for 20 min at 450×g at +4° C.

The supernatant was centrifuged for 20 min at 10,000×g at +4° C. to sediment the granules. Myeloperoxidase (MPO) was purified from granule extracts according Olsson et al (*Scand. J. Haematol.* 9 (1972) 483-491) and Cooray et al (*Vet. Immunol. Immunopathol.* 38 (1993) 261-272). The final preparation was completely homogenous according to the absorbance ratio A430 nm/A280 m which was 0.80 (Agner Acta. Chem. Scand. 12 (1958) 89-94. Human neutrophil lipocalin (HNL) was purified as described (Xu et al., *Scand J Clin Lab Invest* 54 (1994) 365-376. HNL was purified to homogeneity according SDS-PAGE electrophoresis and silver staining and the antigen did not react with antibodies against the other neutrophil proteins, MPO; Lactoferrin, Cathepsin G, Elastase and Lysozyme. Lactoferrin was purified as described (Reiter *Int. J. Tissue React.* 5 (1983) 87-96. Eosinophil Peroxidase (EPO) was purified as described (Carlson et al *J. Immunol.* 134 (1985) 1875-1879) and the final preparation was homogenous according to the absorbance ratio A415 nm/A280 nm that was 1.15.

Eosinophil protein (EPX) was purified to homogeneity as described (Peterson et al., *Immunol.* 50 (1983) 19-26. The final preparation appeared as one band on SDS-PAGE electrophoresis and did not react with antibodies against eosinophil cationic protein (ECP), elastase, cathepsin G, MPO and EPO.

Production of Antibodies

Polyclonal Antibodies

Antibodies against MPO, HNL, EPO and EPX was raised in rabbits by multiple site intracutaneous injections into the rabbits of total 50-100 µg of the purified proteins suspended in Freund's complete and incomplete adjuvant. The specificity of the antibodies was evaluated by double immuno diffusion (Ouchterlony *Acta Pathol. Microbiol. Scand* 26 (1949) 507-) in agarose and tested against extracts of neutrophils and eosinophil granules and the following purified proteins: cathepsin G, elastase, MPO, lysozyme, lactoferrin, ECP, EPX; EPO.

Monoclonal Antibodies

Female Balb/c mice were immunized subcutaneously with purified protein. Priming was done by injecting 50 µg of pure protein mixed with Freund's complete adjuvant. Three boosters were done with approximately 50 µg of pure protein in PBS (phosphate buffered saline). Spleen cells were fused as described (Galfré et al., *Nature* 266 (1977) 550-552) with Sp2/0 myeloma cells. Supernatants from the cell cultures were screened for antibodies using ELISA technique with antigen-coated wells. Antibodies in supernatants were also

screened for specificity to respective granule protein and mapped for epitopes in BIAcore® (BIAcore, Uppsala, Sweden). Hybridomas were selected according to the ELISA and BIAcore experiments and cloned, expanded and purified. All selected antibodies were of IgG1 subtype.

Immunoassays

HNL was assessed using a radioimmunoassay as described (Xu et al., *J. Immunol. Methods* 171 (1994) 245-252. Inter- and intra assay variations were less than 10% and detection limit was less than 4 µg/l.

EPX and Myeloperoxidase was measured using commercially available radioimmunoassays (Pharmacia Diagnostics AB, Uppsala, Sweden). Inter- and intra assay variations were less than 10% and detection limit was less than 3 and 8 µg/l, respectively

EPO was measured using a prototype immunofluorometric assay utilising the Pharmacia CAP system® as described (Nielsen et al., *Allergy* 53 (1998) 778-785. Inter- and intra assay variations were less than 8% and detection limit was less than 0.5 µg/l. Lactoferrin was estimated as described (Olofsson et al., *Scand J Haematol* 18 (1977) 73-80). Inter- and intra assay variations were less than 8% and detection limit was less than 2 µg/l.

Blood Samples

EDTA-containing blood samples drawn from patients were randomly collected at the University Hospital, Uppsala, Sweden. Blood cell counts were performed on each sample by means of a Coulter STKS (Beckman Coulter, Inc.) cell counter.

Granule proteins were extracted from granulocytes by means of adding CTAB (N-cetyl-N,N,N-trimethyl-ammonium bromide) at a final concentration of 0.05-0.5% to a small aliquot of blood, 1-10 µl. The mixture was then incubated for at least 1 minute and then stored frozen at -20° C. before analysis.

Statistical Evaluation

Regression analysis was performed using the statistical package, Statistica (Statsoft, Tulsa, USA).

EXAMPLE 1

Estimation of the Number of Blood Neutrophils

The present invention shows that extraction of a small aliquot of blood, 1-10 µl, with CTAB, final concentration 0.05-0.5%, for at least 1 minute and subsequent measurement of the neutrophil protein MPO by means of a specific immunoassay accurately estimates the numbers of neutrophils in the blood.

As shown in FIG. 1, the concentration of MPO in the extract was significantly and linearly correlated (r=0.96) to the number of neutrophils in the extracted blood as estimated by means of a Coulter STKS (Beckman Coulter, Inc.) cell counter. From the equation of the regression line it is apparent that the deviation from origo was minimal, indicating the cell specificity of the measurement. The results were obtained from a mixed population of hospitalized patients (n=275) having both elevated and reduced levels of neutrophils in their blood. Thus, some patients had highly elevated levels due to acute bacterial infections and others had seriously reduced levels due to leukemia or cytostatic drug treatment. In spite of the inclusion of these extremes in the calculation, the relationship between number of neutrophils and the concentration of MPO was linear over the entire range measured. When HNL was measured the corresponding correlation was

$r=0.93$ and also with a linear relationship to the number of neutrophils over the entire range. Lactoferrin measurement also showed a linear relationship over the entire range and a correlation coefficient of $r=0.82$.

EXAMPLE 2

Estimation of the Degree of Maturation of the Neutrophil Population

It is well known that MPO is stored in the primary granules of neutrophils, whereas lactoferrin and HNL are stored in secondary granules. This is because the production of MPO primarily takes place during the early maturation steps i.e. by myeloblasts and promyelocytes, whereas lactoferrin and HNL primarily are produced during later maturation steps i.e. by myelocytes. It is also known that production of MPO is less affected by an increased requirement of neutrophils in the circulation, such as in acute infections, than the production of lactoferrin and HNL. The ratio between the content of either of the secondary granule proteins and MPO would therefore provide us with an estimate of the relative size of the various maturation stages of neutrophils in the blood and an indication of the bone marrow turnover of neutrophils.

It is shown in FIG. 2 that a ratio between MPO concentration and lactoferrin concentration in extracted whole blood varies about 20-fold between patients, with myeloid leukemia patients having the highest ratios.

EXAMPLE 3

Estimation of the Number of Blood Eosinophils

In this example it is shown that extraction of a small aliquot of blood, 1-10 μ l, with CTAB, 0.05-0.5%, for at least 1 minute and the subsequent measurement of the eosinophil protein EPO by means of a specific immunoassay accurately estimates the numbers of eosinophils in the blood.

As shown in FIG. 3, the concentration of EPO in the extract was significantly and linearly correlated ($r=0.95$) to the number of eosinophils in the extracted blood as estimated by means of a Coulter STKS (Beckman Coulter, Inc.) cell counter. From the equation of the regression line it is apparent that the deviation from origo was minimal, indicating the cell specificity of the measurement. The results were obtained from a mixed population of hospitalized patients ($n=275$) having both elevated and reduced levels of eosinophils in their blood. Thus, some patients had elevated numbers because of allergy and asthma, chronic inflammatory diseases, cancer etc. and some had reduced numbers because of, among other things, acute infections. In spite of the inclusion of these extremes in the calculation, the relationship between number of eosinophils and the concentration of EPO was linear over the entire range measured. When EPX was measured the corresponding correlation was $r=0.93$ and also with a linear relationship to the number of eosinophils over the entire range.

The above examples 1-3 describe neutrophils, different maturation forms of neutrophils, and eosinophils. However the invention is not to be construed as limited to these cell types.

For example the basophil protein BB1 may measure basophils.

Cell surface markers such as CD20 may measure B-lymphocytes and CD3 T-lymphocytes.

The cell surface markers CD4 and CD8 may be used to measure different lymphocyte populations.

Monocytes may be measured by CD14 or lysozyme.

Thrombocytes may be measured by β -tromboglobulin

Determination of ratios is especially interesting for myeloid cells as described in Example 2, but also for various subpopulations of lymphocytes.

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- Xu SY, Peterson CGB, Carlson M et al. The development of an assay for human neutrophil lipocalin (HNL)—to be used as a specific marker of neutrophil activity in vivo and vitro. *J Immunol Methods* 1994; 171:245-52.

The invention claimed is:

1. A method for in vitro estimating an amount of specific cells in a patient sample using cell specific molecules, the method comprising
 - a) extracting cell specific molecules from cells contained in an aliquot of the patient sample;
 - b) measuring the concentration of at least two cell specific molecules in the extracted aliquot by immunological assay; and

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- c) estimating the amount of specific cells that contained the at least two cell-specific molecules from a ratio between the measured concentrations of the cell specific molecules.
2. The method according to claim 1, wherein the extraction is conducted with cationic detergent.
3. The method according to claim 2, wherein the detergent is CTAB (N-cetyl-N,N,N-trimethylammonium bromide).
4. The method according to claim 1, wherein the patient sample is whole blood.
5. The method according to claim 1, wherein the cell specific molecules are intracellular and extracellular proteins or cell surface markers.
6. The method according to claim 1, wherein the concentration of each cell specific molecule is correlated with a number of a subtype of cells.
7. The method according to claim 1, wherein the cell specific molecules comprise a neutrophil protein.
8. The method according to claim 7, wherein the neutrophil protein comprises MPO (myeloperoxidase), HNL (human neutrophil lipocalin) or lactoferrin.
9. The method according to claim 1, wherein the cell specific molecules comprise an eosinophil protein.
10. The method according to claim 9, wherein the eosinophil protein comprises EPX (eosinophil protein x) or EPO (eosinophil peroxidase).

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11. The method according to claim 1, wherein the cell specific molecules comprise a basophil protein.
12. The method according to claim 11, wherein the basophil protein is BB1.
13. The method according to claim 1, wherein the cell specific molecules comprise a thrombocytic protein for measuring thrombocytes.
14. The method according to claim 13, wherein the thrombocytic protein is β -thromboglobulin.
15. The method according to claim 1, wherein the cell specific molecules comprise cell surface markers for measuring B-lymphocytes and CD3 T-lymphocytes.
16. The method according to claim 15, wherein the cell surface markers are CD20.
17. The method according to claim 1, wherein the cell specific molecules comprise cell surface markers for measuring different lymphocyte populations.
18. The method according to claim 17, wherein the cell surface markers are CD4 or CD8.
19. The method according to claim 1, wherein the cell specific molecules are CD14 or lysozyme for measuring monocytes.
20. The method according to claim 1, wherein the cell specific molecules comprise myeloperoxidase and lactoefin.

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UNITED STATES PATENT AND TRADEMARK OFFICE
CERTIFICATE OF CORRECTION

PATENT NO. : 7,405,047 B2
APPLICATION NO. : 10/178211
DATED : June 24, 2002
INVENTOR(S) : Per Venge

Page 1 of 1

It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

Claim 20, column 8, line 24, change to "lactoeffin" to -- lactoferrin --.

Signed and Sealed this

Twenty-third Day of September, 2008

A handwritten signature in black ink, appearing to read "Jon W. Dudas". The signature is stylized with a large, sweeping initial "J" and a circular flourish at the end.

JON W. DUDAS
Director of the United States Patent and Trademark Office

UNITED STATES PATENT AND TRADEMARK OFFICE
CERTIFICATE OF CORRECTION

PATENT NO. : 7,405,047 B2
APPLICATION NO. : 10/178211
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Page 1 of 1

It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

Claim 20, column 8, line 24, change to "lactoeffin" to -- lactoferrin --.

This certificate supersedes the Certificate of Correction issued September 23, 2008.

Signed and Sealed this

Fourteenth Day of October, 2008

A handwritten signature in black ink, reading "Jon W. Dudas". The signature is written in a cursive style with a large, looped initial "J".

JON W. DUDAS
Director of the United States Patent and Trademark Office

专利名称(译)	估计特定细胞类型的量的方法		
公开(公告)号	US7405047	公开(公告)日	2008-07-29
申请号	US10/178211	申请日	2002-06-24
[标]申请(专利权)人(译)	VENGE PER		
申请(专利权)人(译)	VENGE PER		
当前申请(专利权)人(译)	PER VENGE		
[标]发明人	VENGE PER		
发明人	VENGE, PER		
IPC分类号	G01N33/53 G01N33/50 G01N33/566 G01N33/569		
CPC分类号	G01N33/50 G01N33/53 G01N33/566 G01N33/56972 G01N2333/705 Y10T436/25375 G01N2333/70514 G01N2333/70517 G01N2333/7452 Y10T436/25125 Y10T436/107497 G01N2333/7051		
优先权	0102220 2001-06-25 SE 60/300759 2001-06-25 US		
其他公开文献	US20030077666A1		
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摘要(译)

本发明涉及通过测量血液和其他生物材料的提取物中的独特蛋白质来估计特定细胞的亚型数量，例如白细胞的某些亚型的数量。了解白细胞特定亚型的数量或数量对于炎症性疾病（包括传染病，癌症，过敏/哮喘等）的受试者的临床诊断和监测非常重要。

