US 20020081629A1

(19) United States

(12) Patent Application Publication (10) Pub. No.: US 2002/0081629 A1

Pauwels et al. (43) Pub. Date: Jun. 27, 2002

(54) APPARATUS FOR THE SIMULTANEOUS TRANSFER OF LIQUID ANALYTES

(75) Inventors: **Rudi Wilfried Jan Pauwels**, Bonheiden (BE); **Christiaan Hubert Simon Roelant**, Leuven (BE); **Koenraad Lodewijk August Van Acker**, Temse (BE)

Correspondence Address:
Patricia A. Kammerer
HOWREY SIMON ARNOLD & WHITE, LLP
750 Bering Drive
Houston, TX 77057-2198 (US)

(73) Assignee: **TIBOTEC N.V.**

(21) Appl. No.: **10/025,391**

(22) Filed: **Dec. 19, 2001**

Related U.S. Application Data

(62) Division of application No. 09/530,907, filed on Jun. 30, 2000, which is a 371 of international application No. PCT/IB98/01399, filed on Sep. 8, 1998.

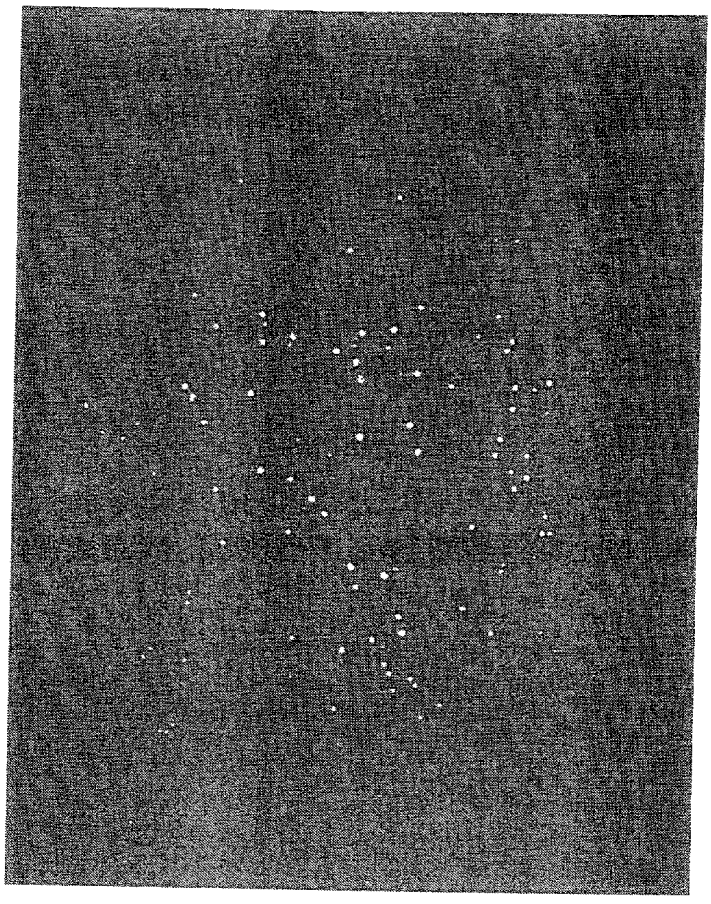
Publication Classification

(51) **Int. Cl.**[7] **G01N 33/53**; C12M 1/34; G01N 33/543

(52) **U.S. Cl.** **435/7.1**; 435/287.2; 436/518

(57) **ABSTRACT**

A method for the rapid screening of analytes, such as potential drug candidates, comprises the steps of applying a plurality of analytes to be screened onto one or more, solid support(s) **(61)** such that the analytes remain isolated from one another; contacting said analyte-carrying solid support(s) **(61)** with targets provided in a semi-solid or liquid medium, whereby said analytes are released from the solid support(s) **(61)** to the targets; and measuring analyte-target interactions. This method allows for the manipulation of thousands of different analytes simultaneously. When the analyte is applied to the solid support **(61)** it can diffuse thereon so as to produce a concentration gradient and serial dilution of analyte if a dose response curve for a candidate drug is required. The method described can be readily automated.



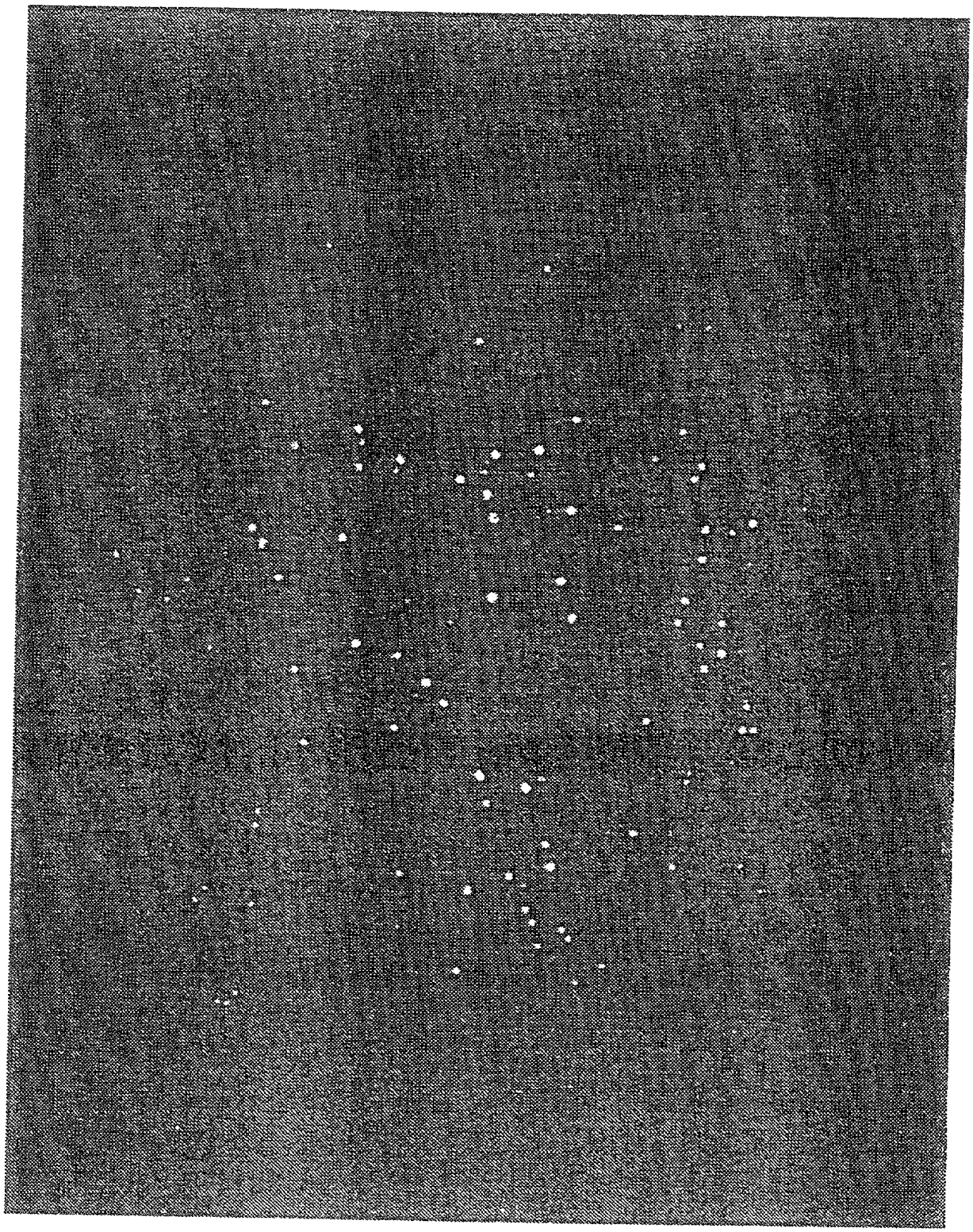


Fig. 1

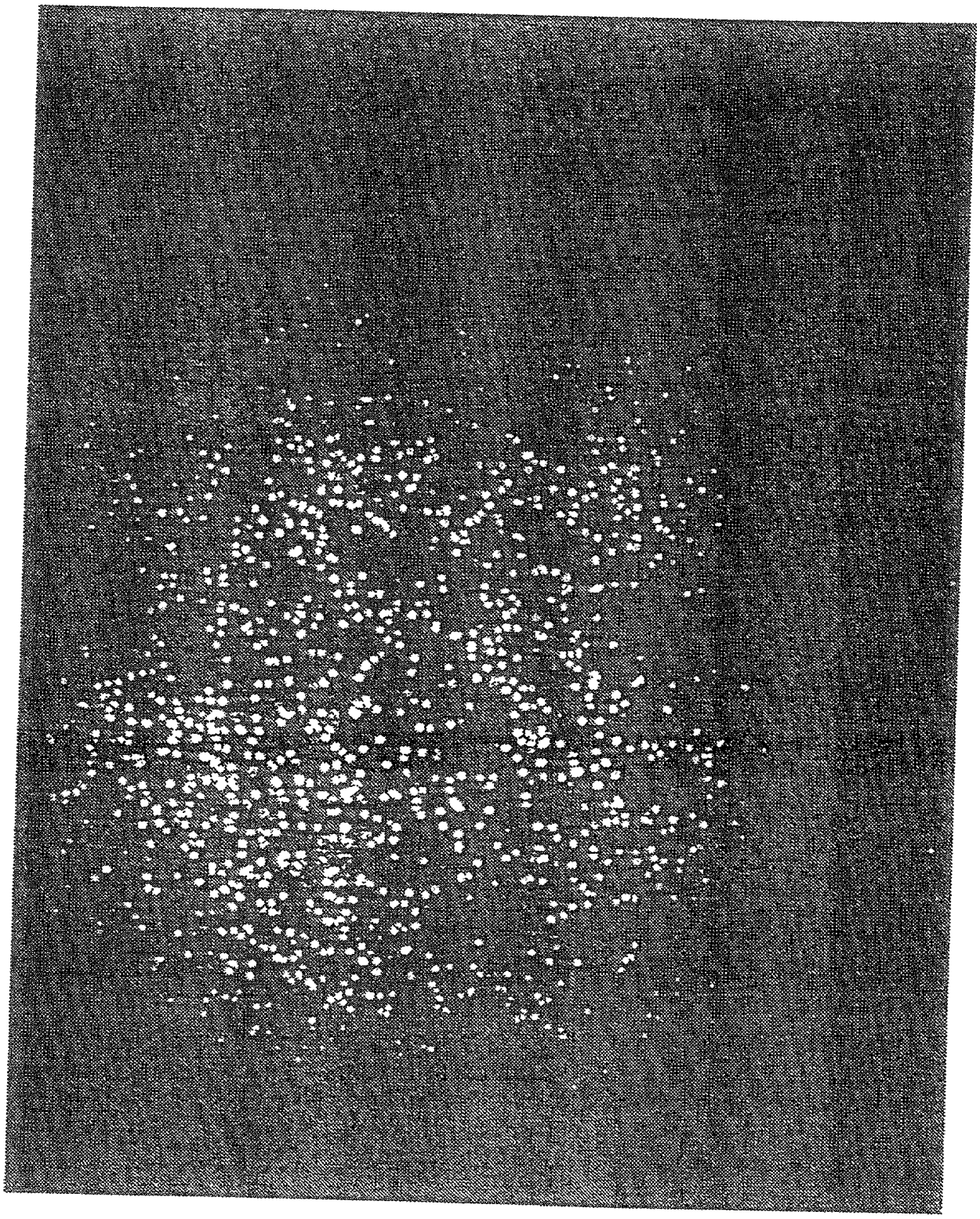


Fig. 2

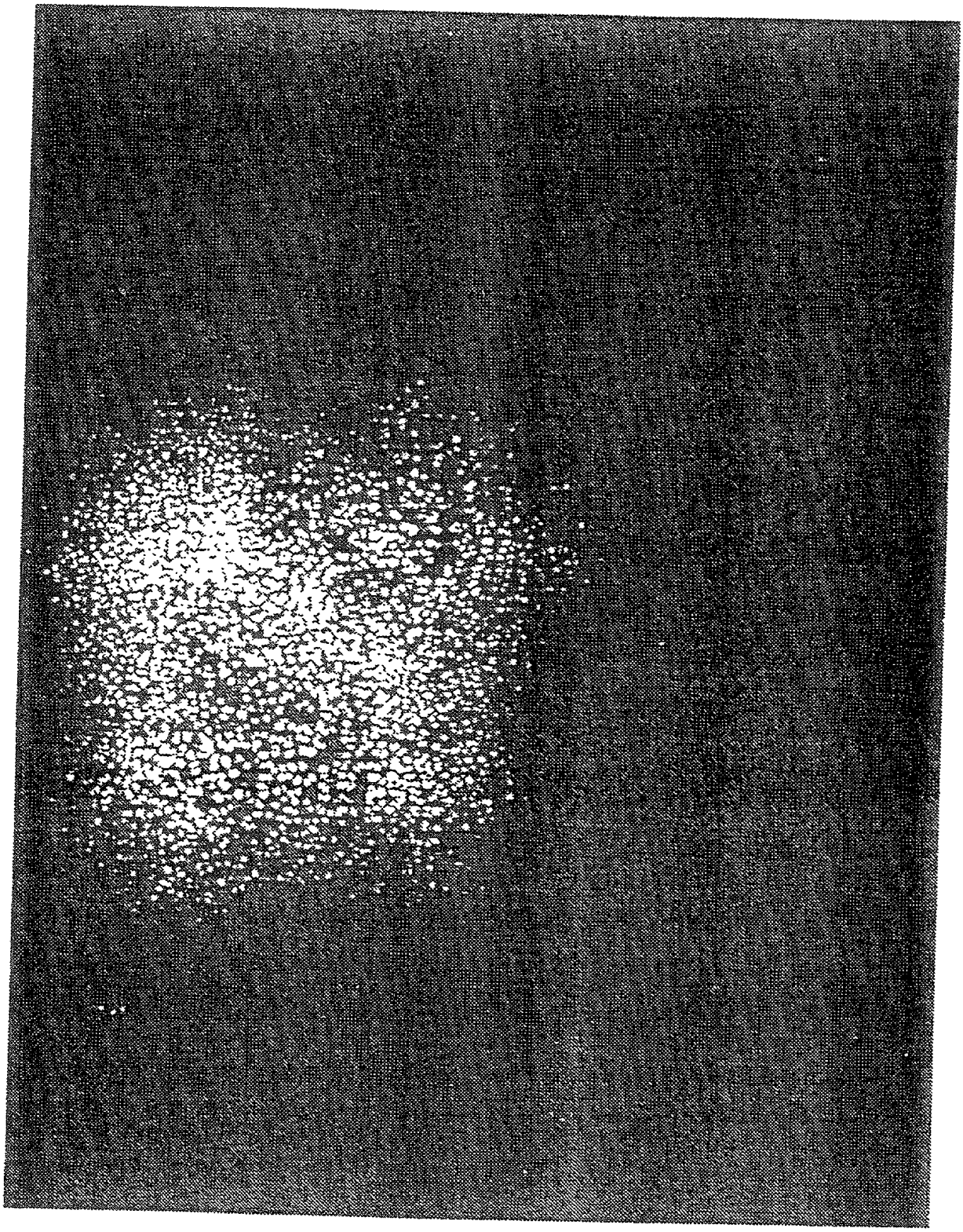


Fig. 3

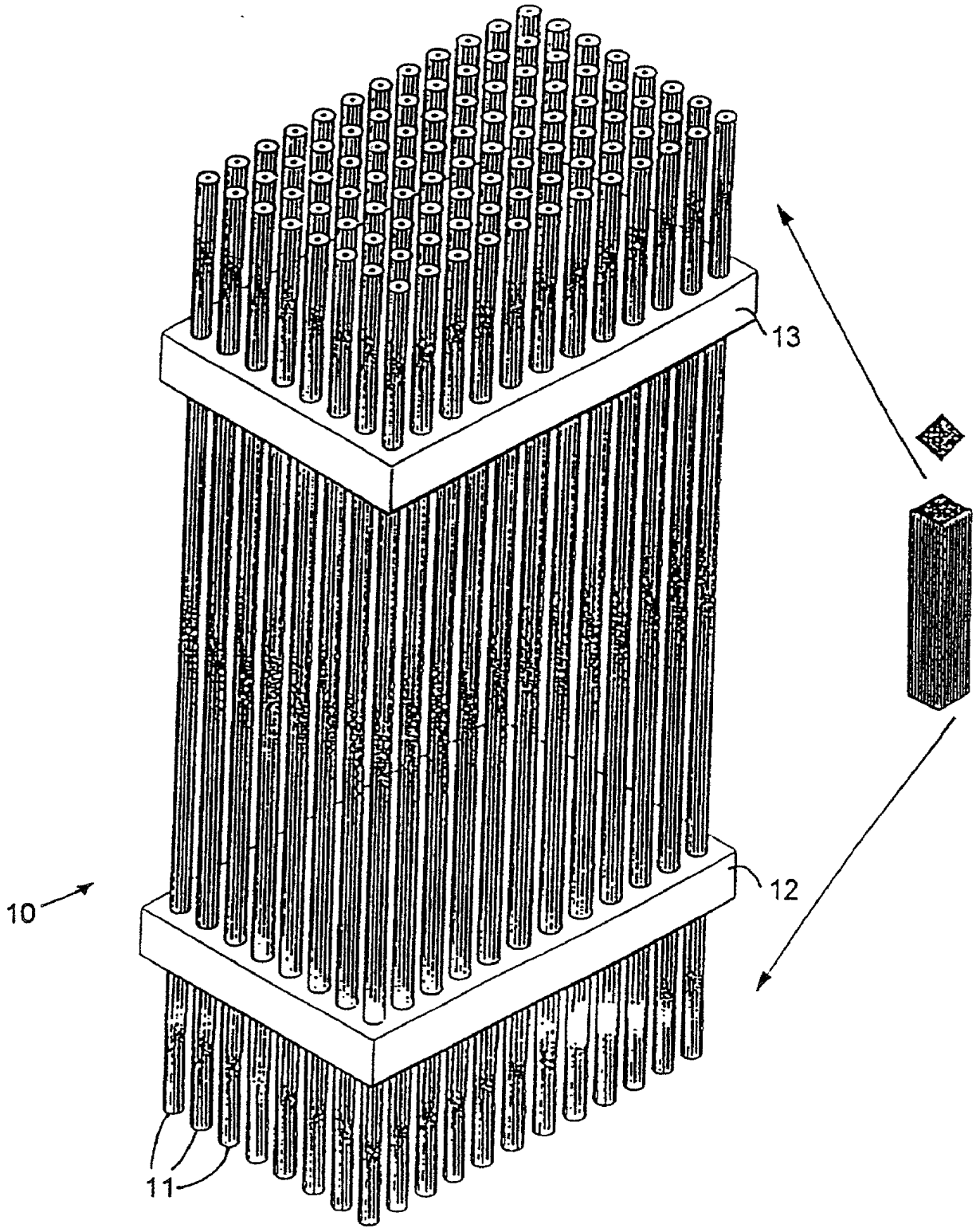


Fig. 4

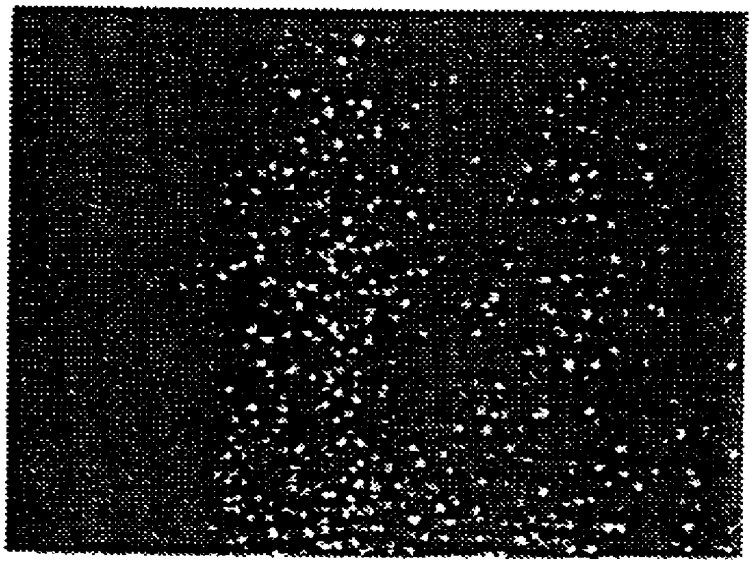


Fig. 5a

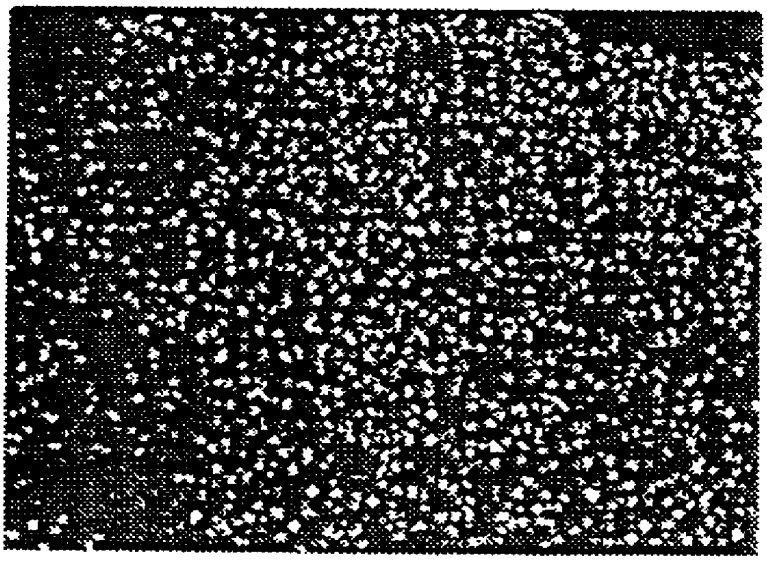


Fig. 5b

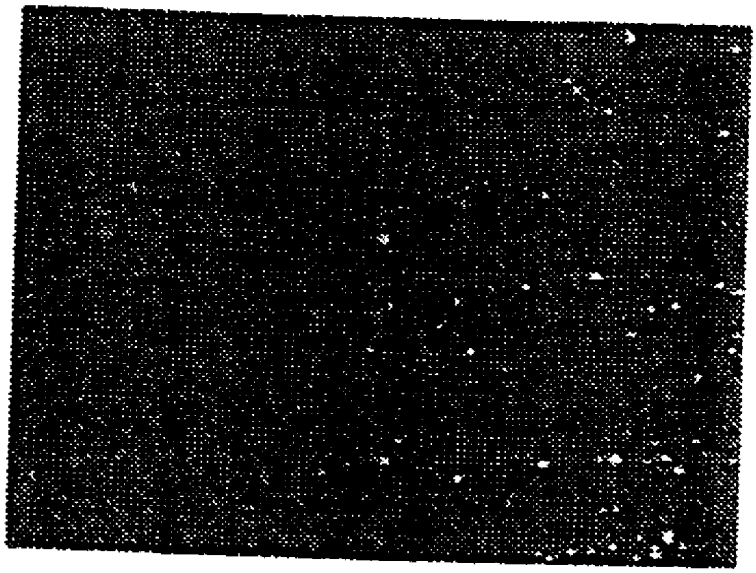


Fig. 6a

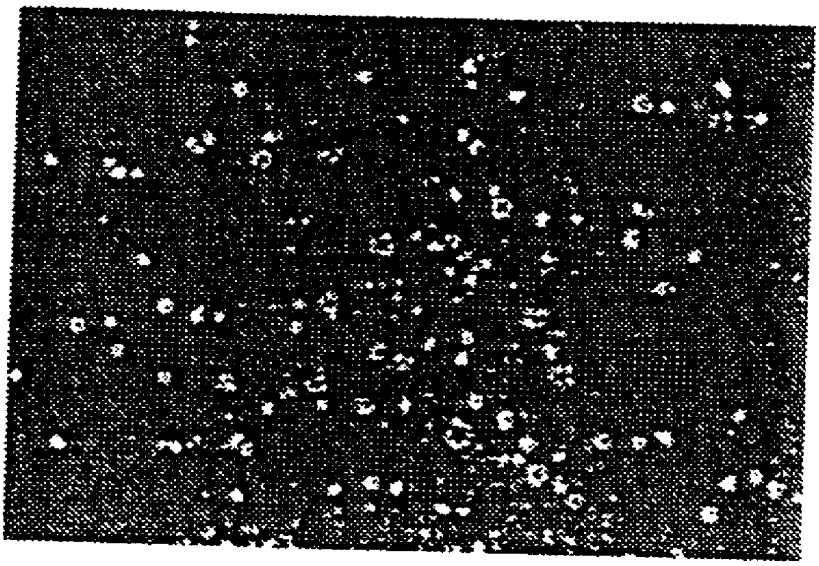


Fig. 6b

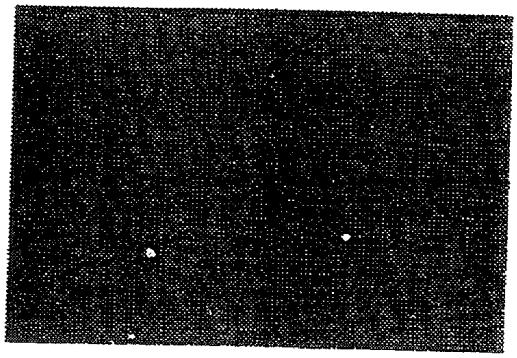


Fig. 7a

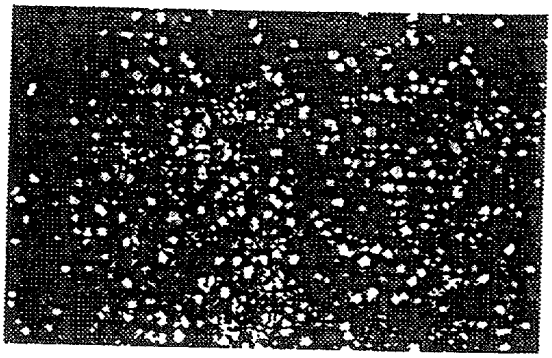


Fig. 7b

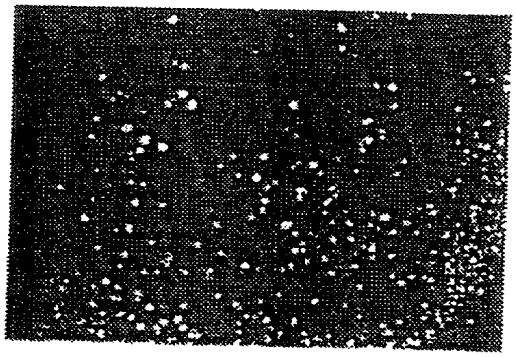


Fig. 7c

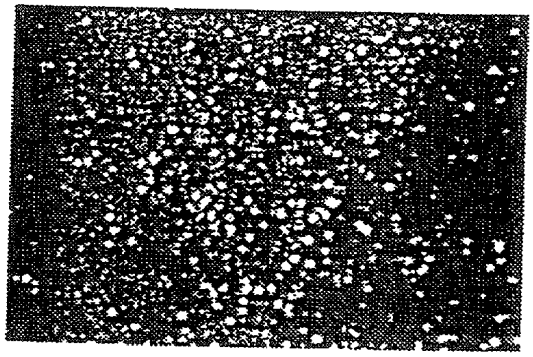


Fig. 7d

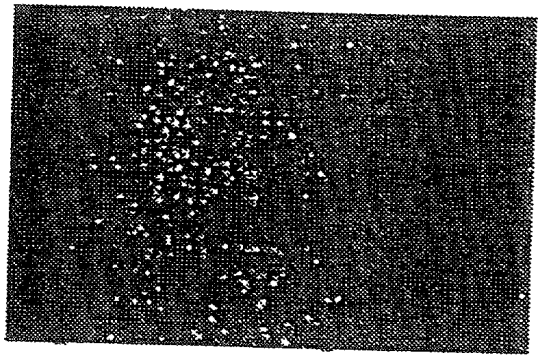


Fig. 7e

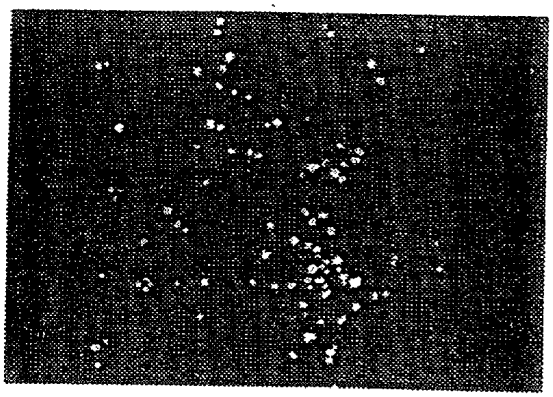


Fig. 8a

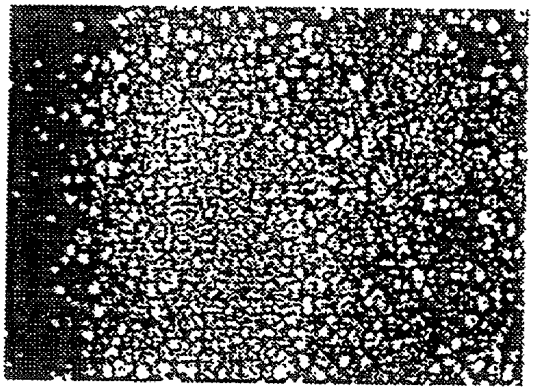


Fig. 8b

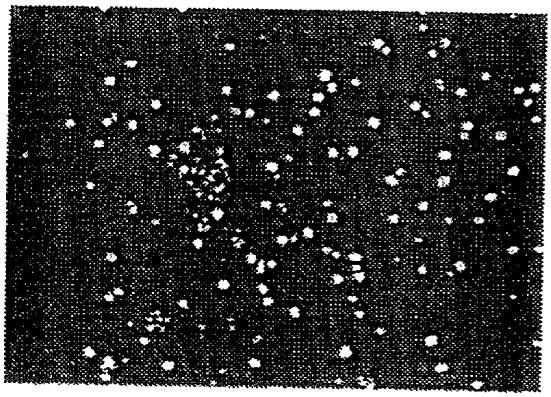


Fig. 8c

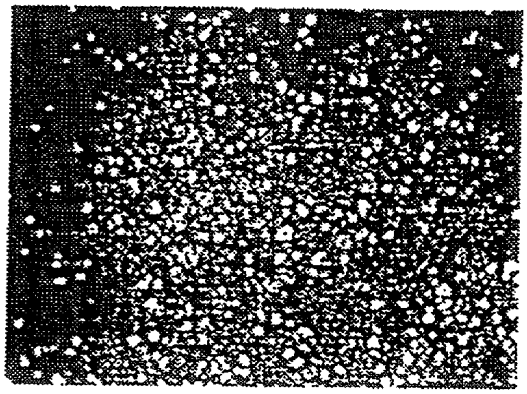


Fig. 8d

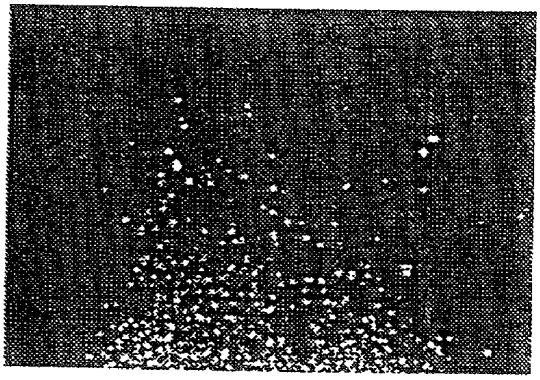


Fig. 8e

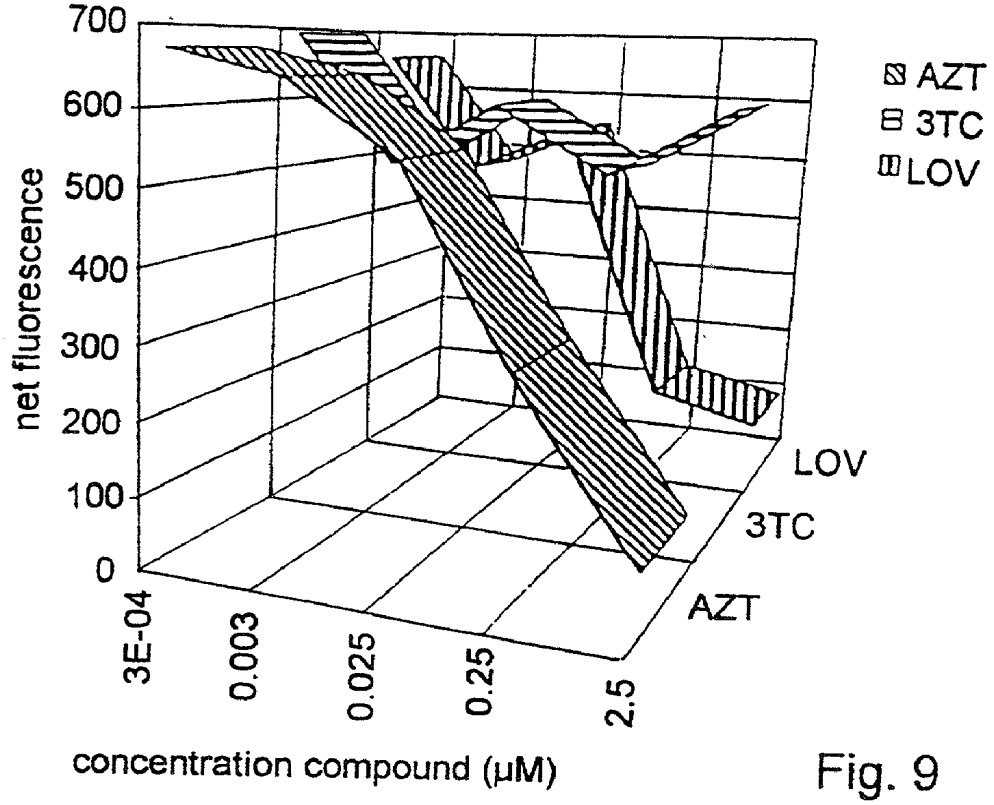

700
600
500
400
300
200
100
0
net fluorescence
AZT
3TC
LOV
LOV
3TC
AZT
3E-04
0.003
0.025
0.25
2.5
concentration compound (µM)

Fig. 9

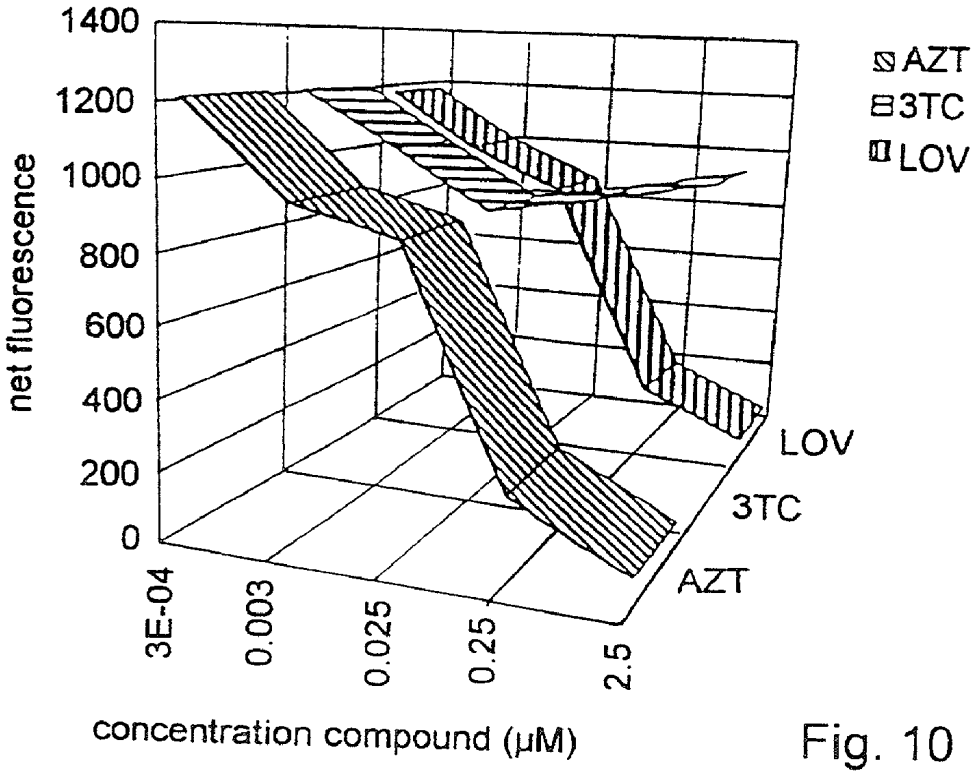

1400
1200
1000
800
600
400
200
0
net fluorescence
AZT
3TC
LOV
LOV
3TC
AZT
3E-04
0.003
0.025
0.25
2.5
concentration compound (µM)

Fig. 10

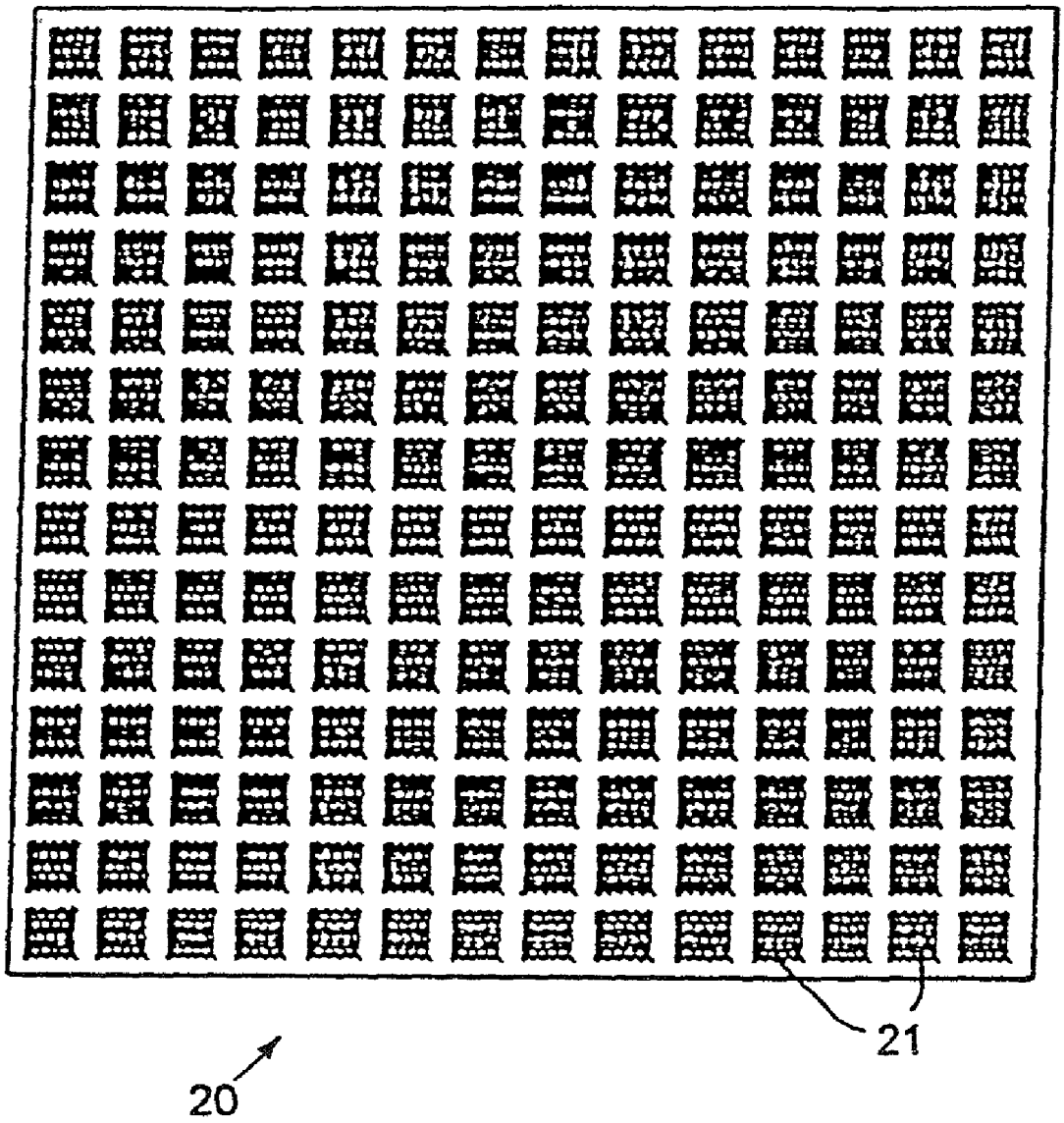


Fig. 11

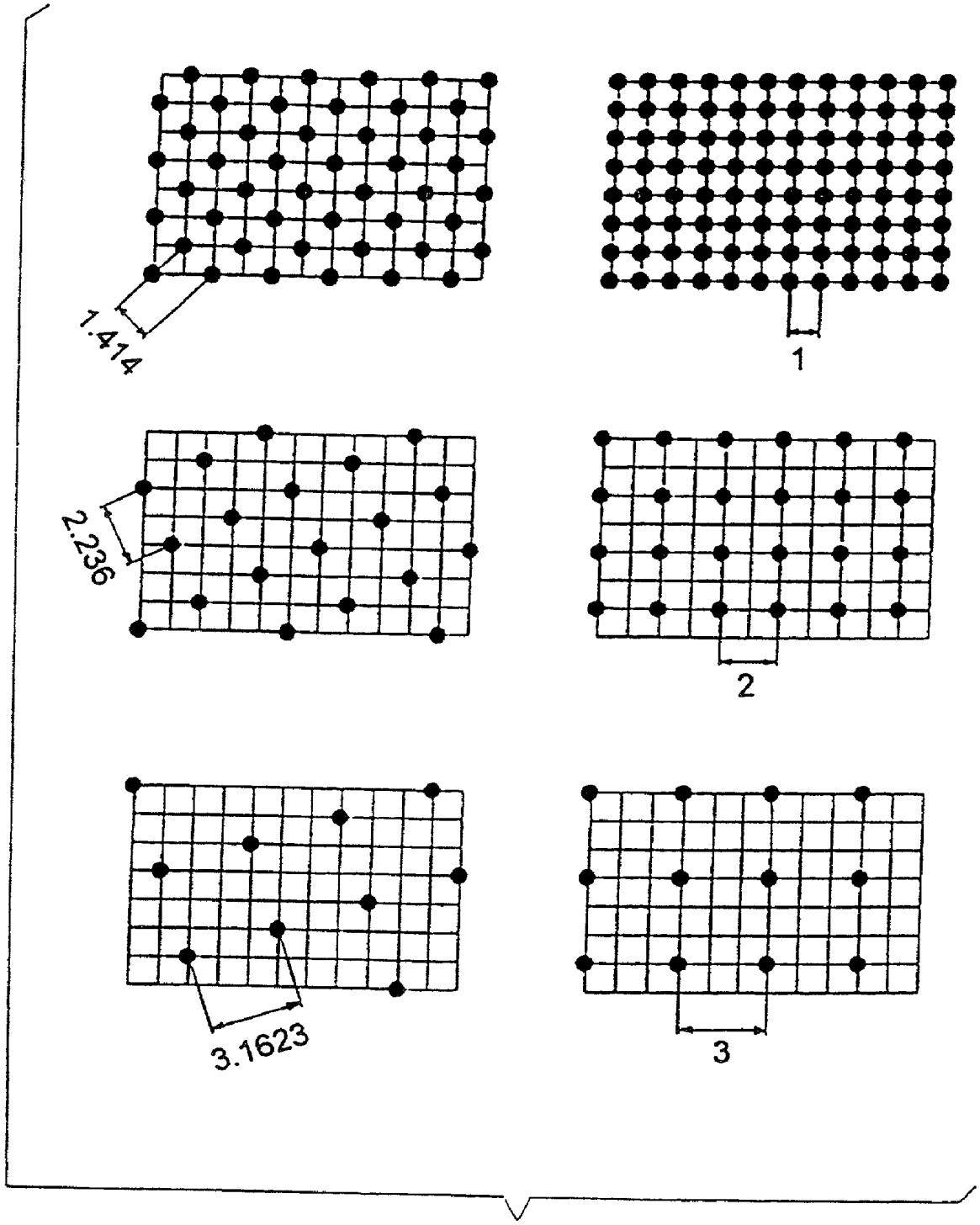


Fig. 12

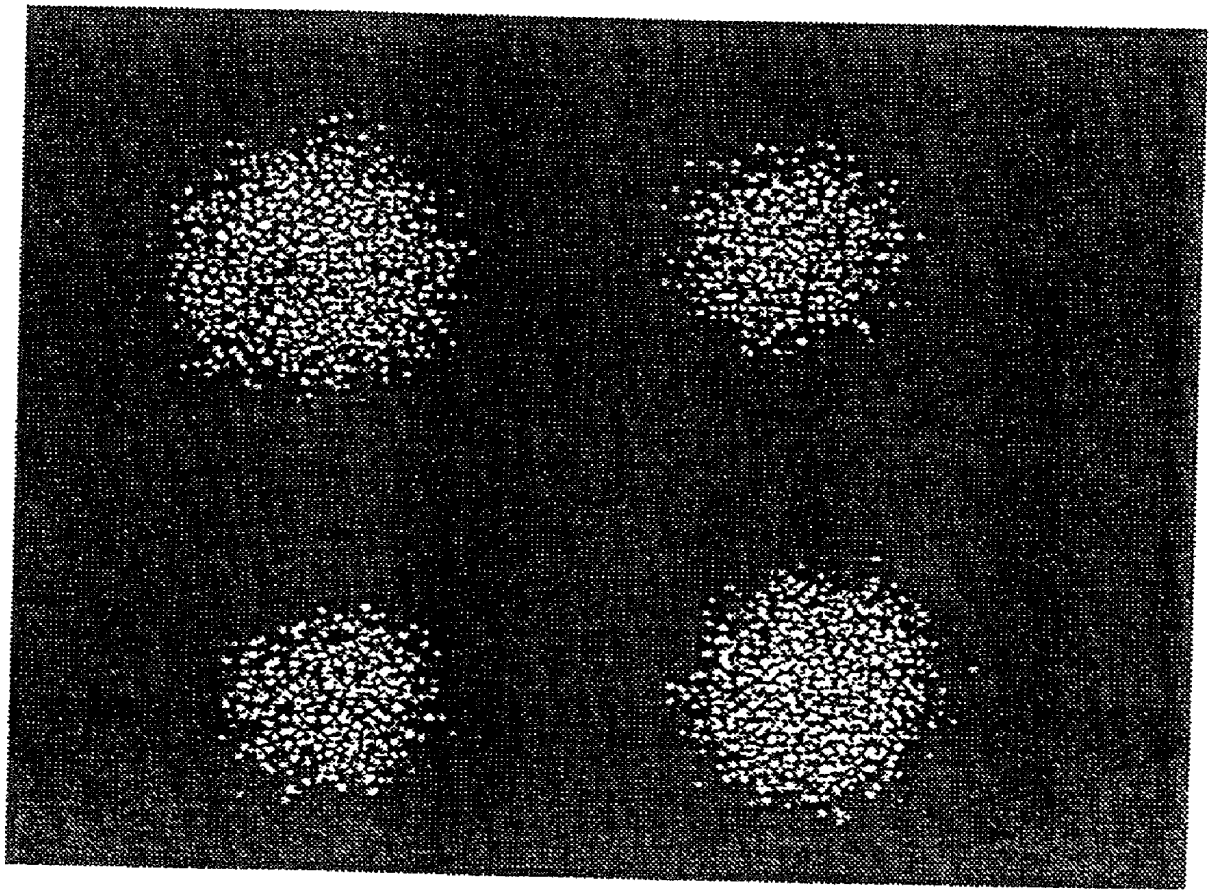


Fig. 13

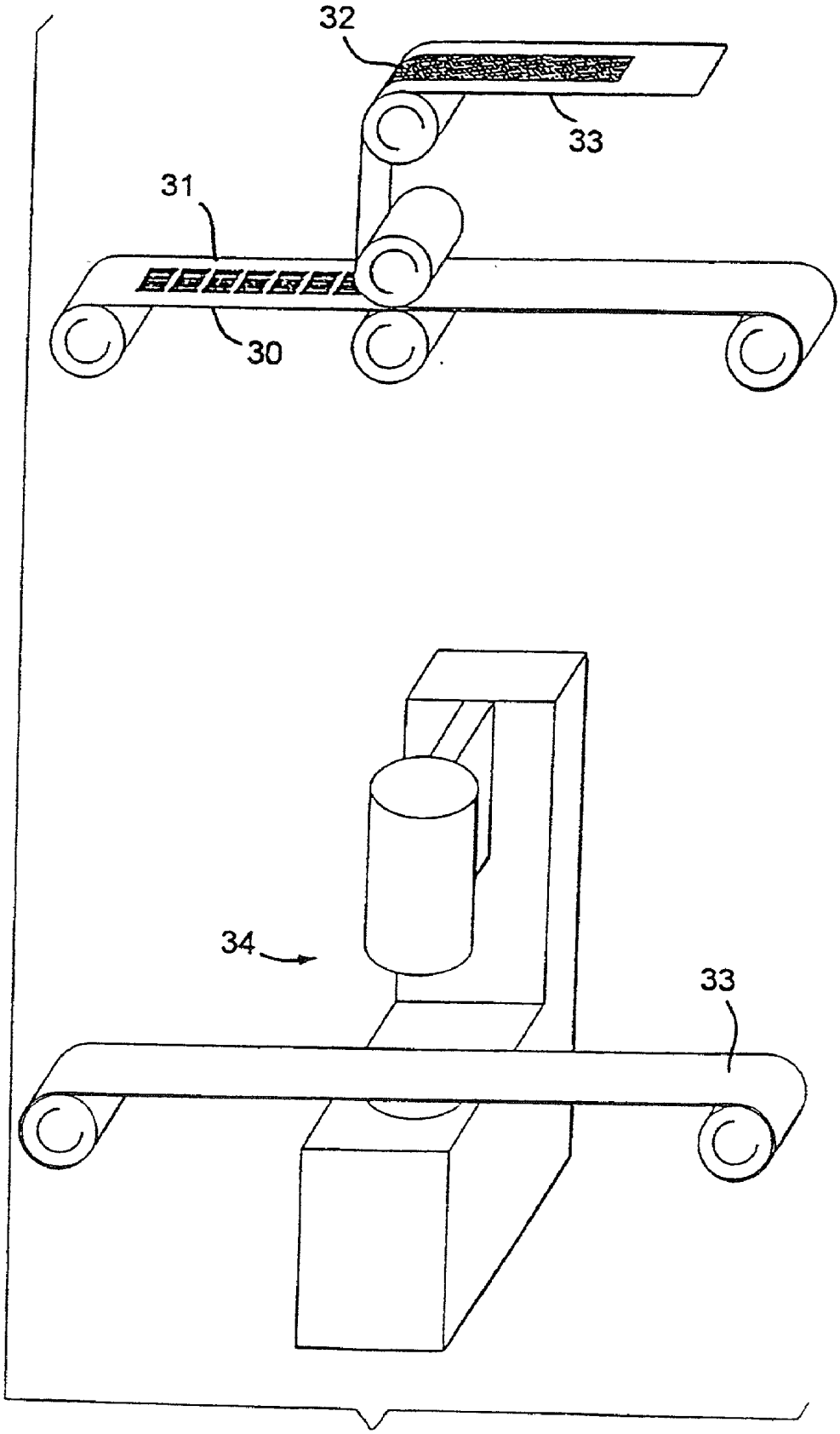


Fig. 14

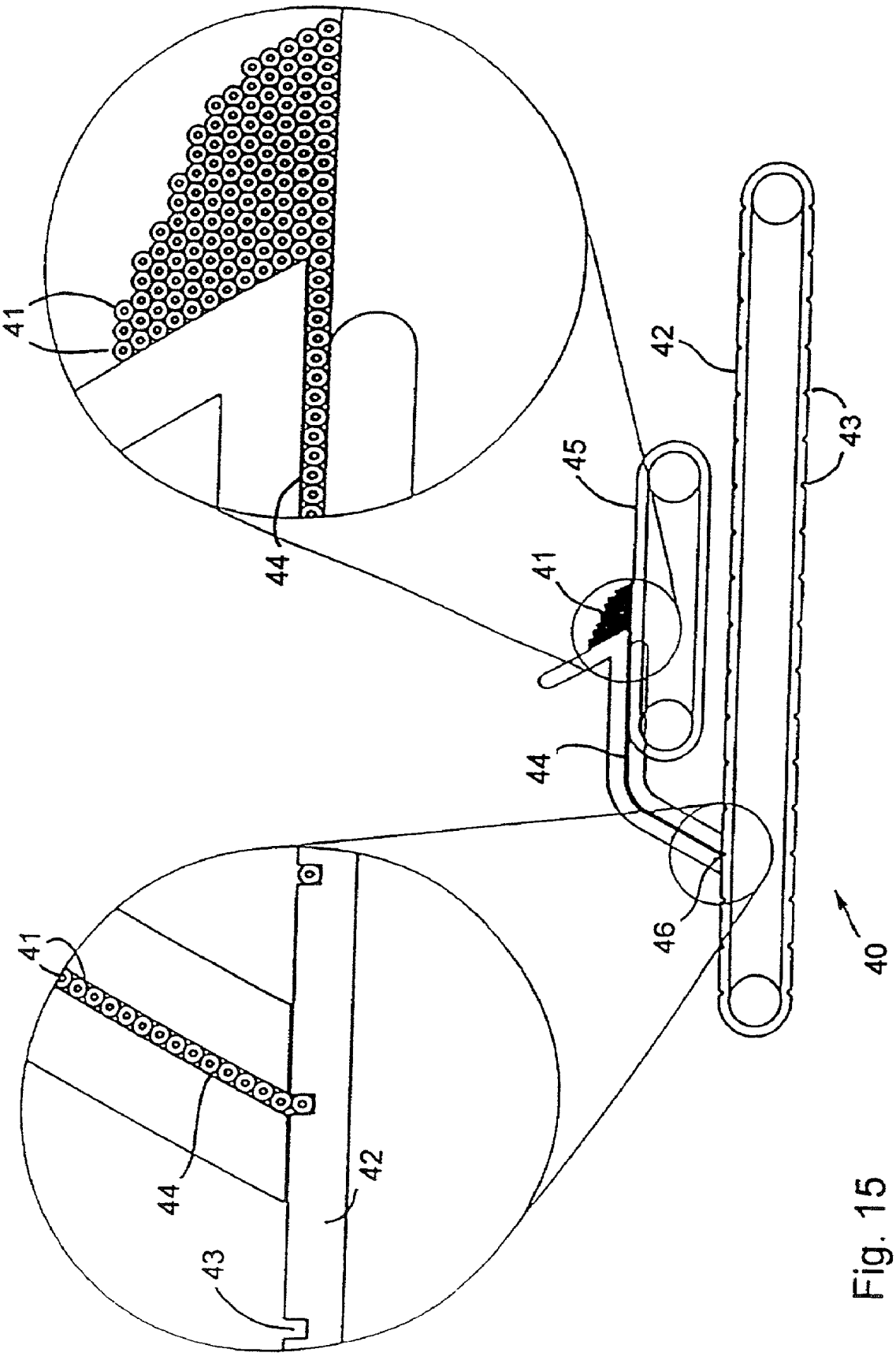


Fig. 15

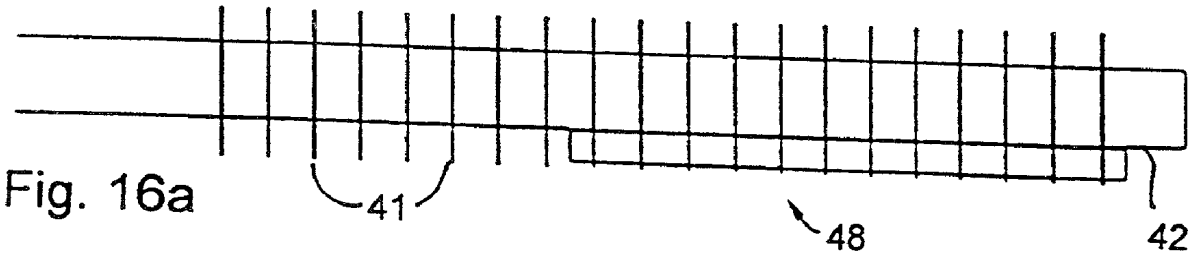


Fig. 16a

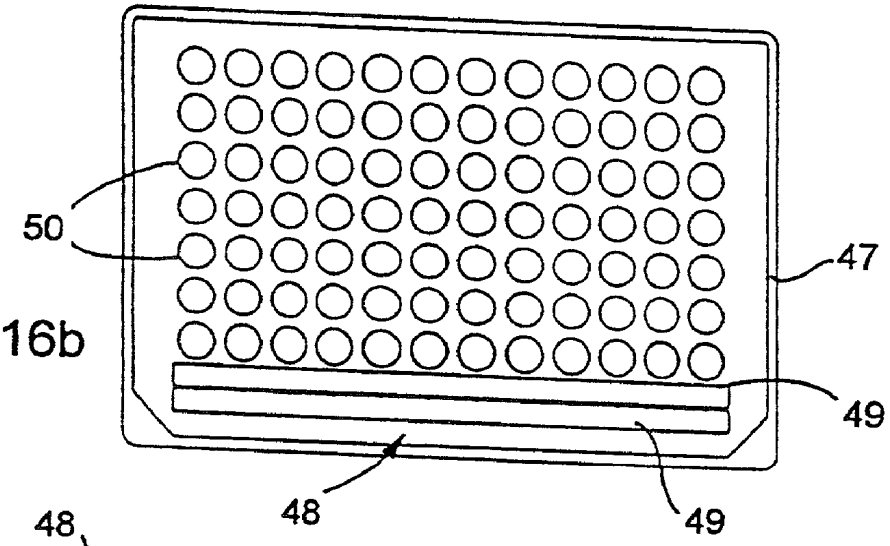


Fig. 16b

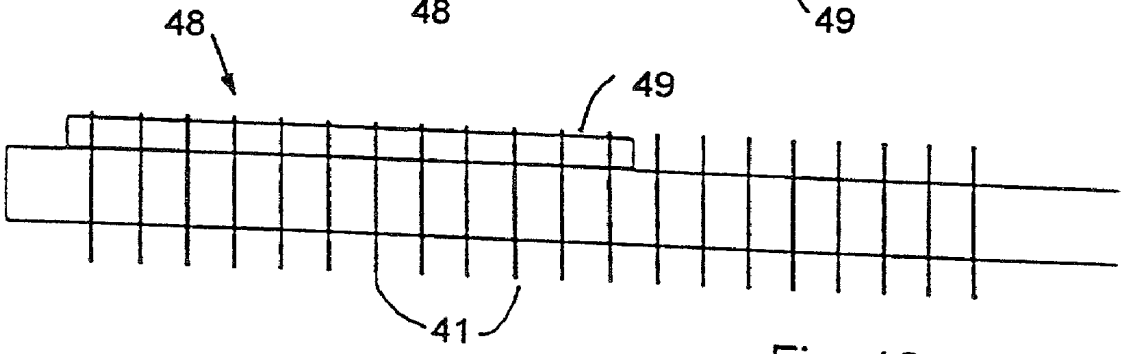


Fig. 16c

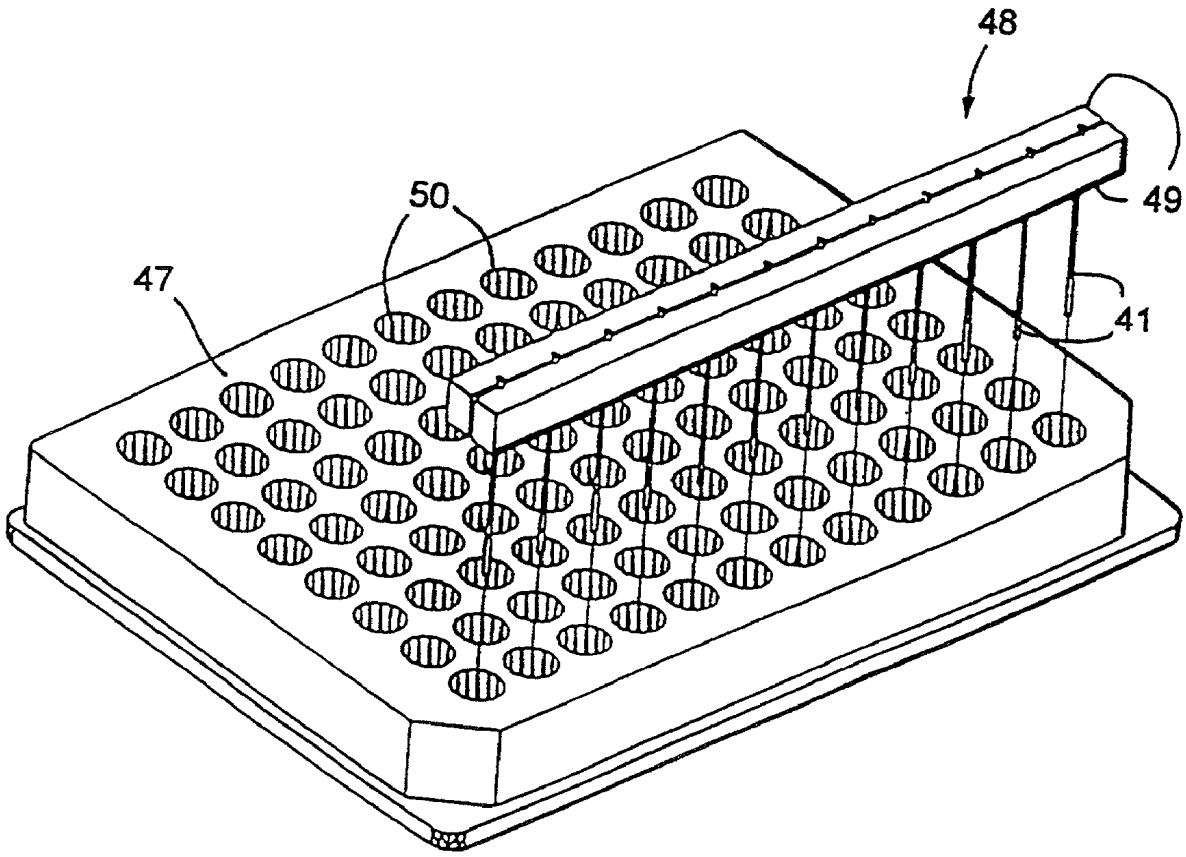


Fig. 17

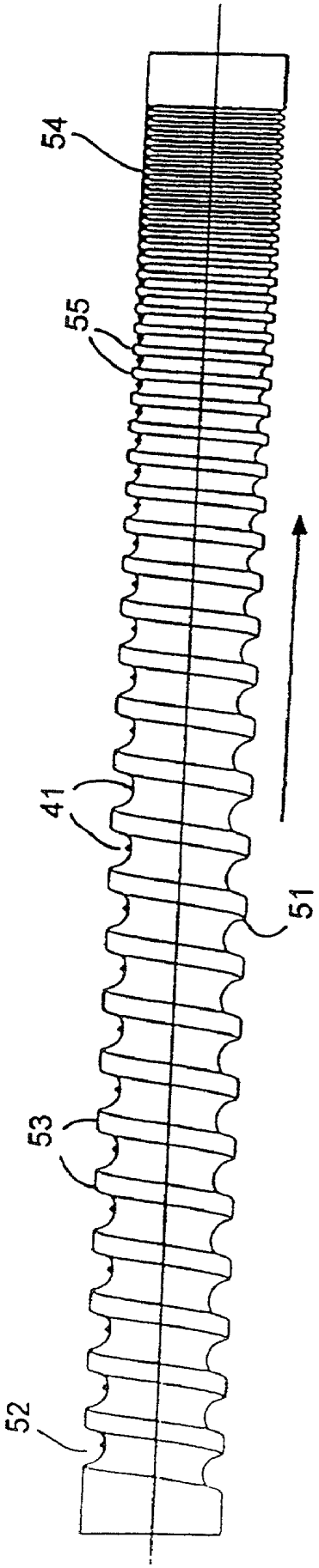


Fig. 18

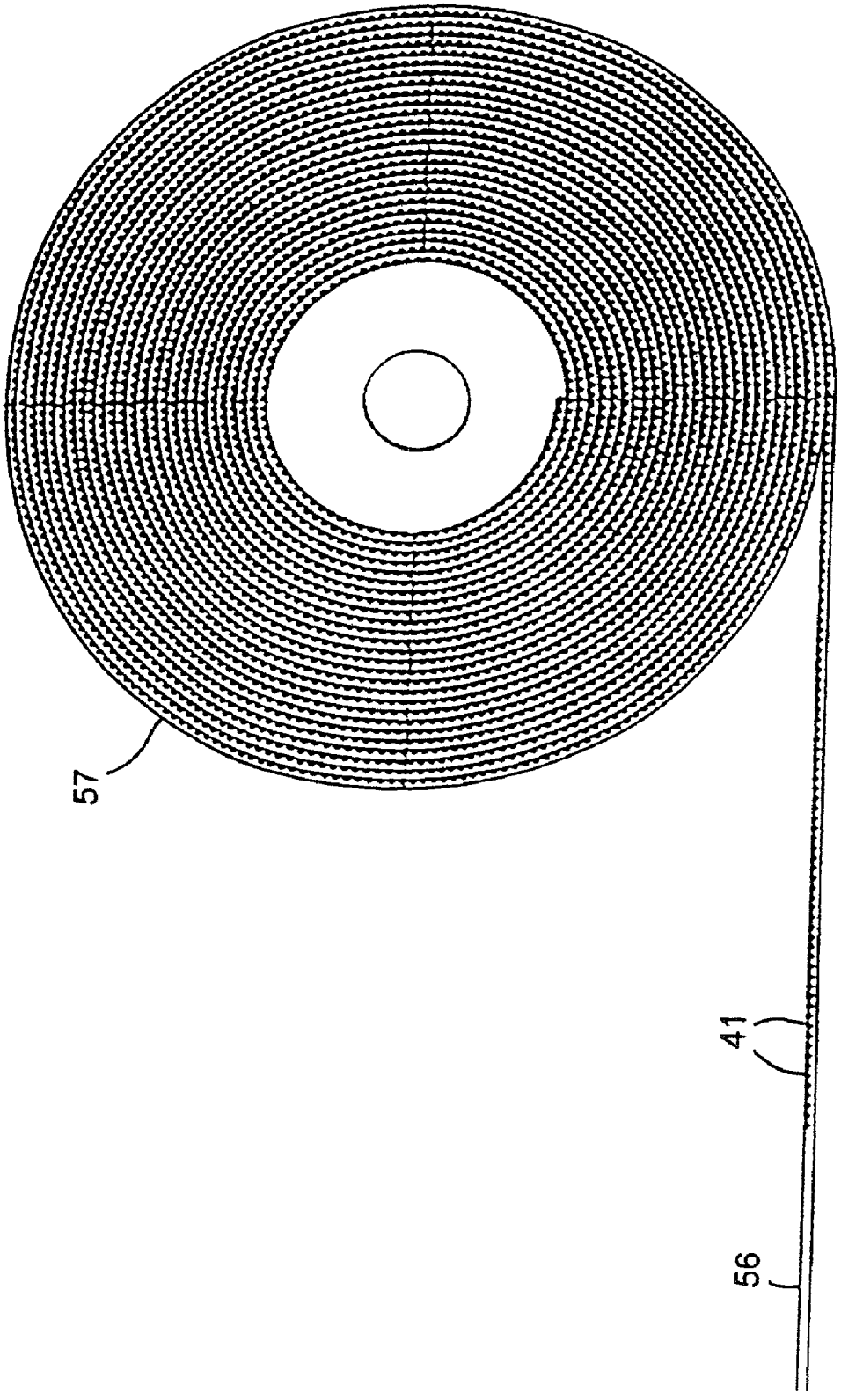


Fig. 19

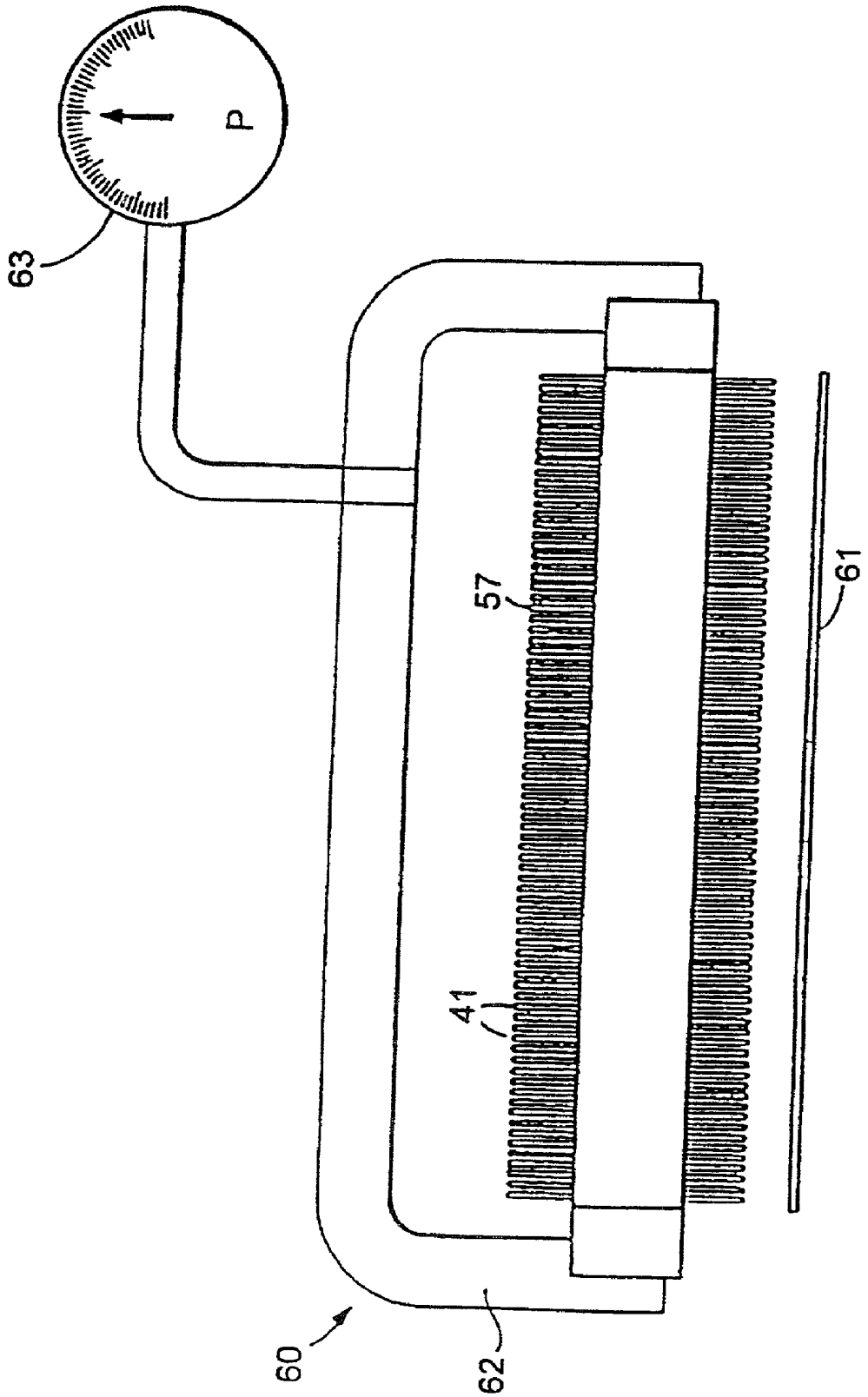


Fig. 20

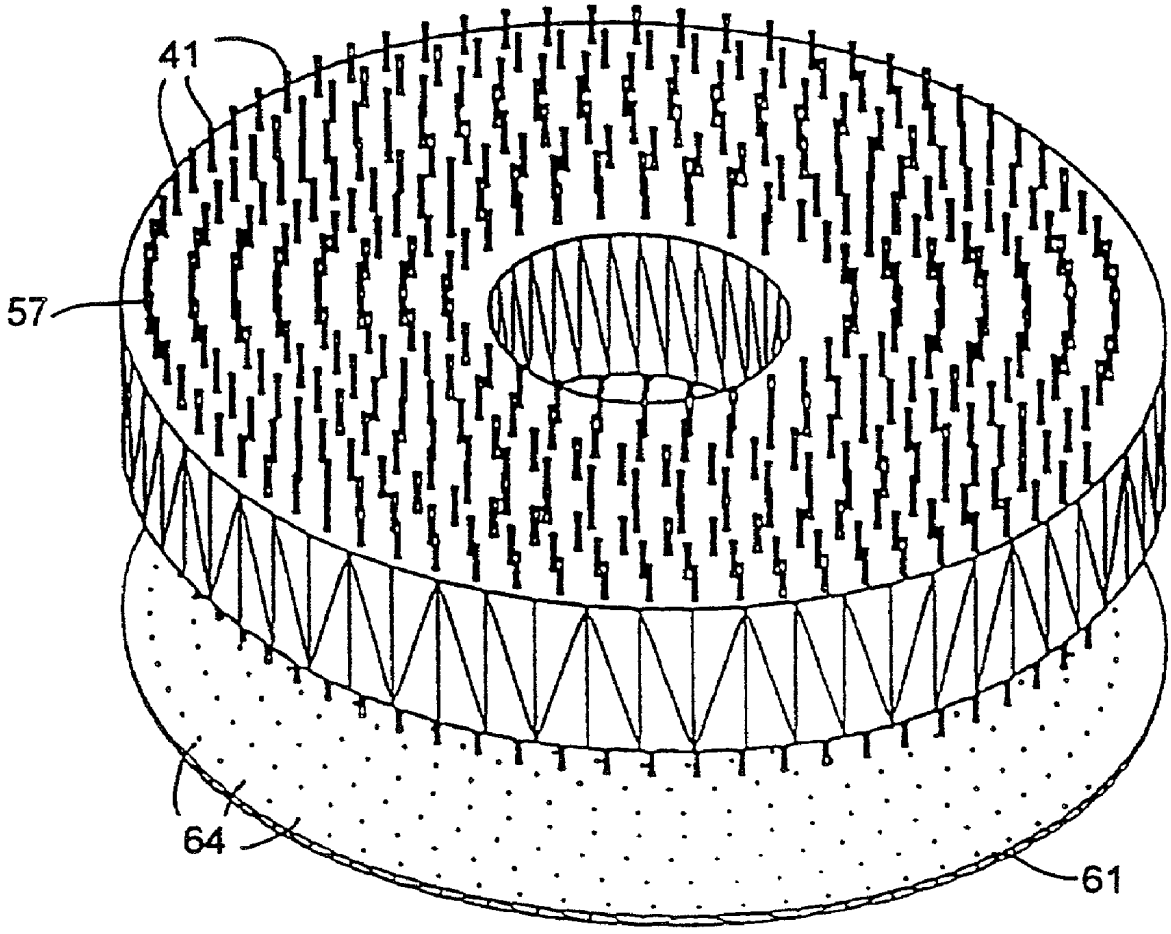


Fig. 21

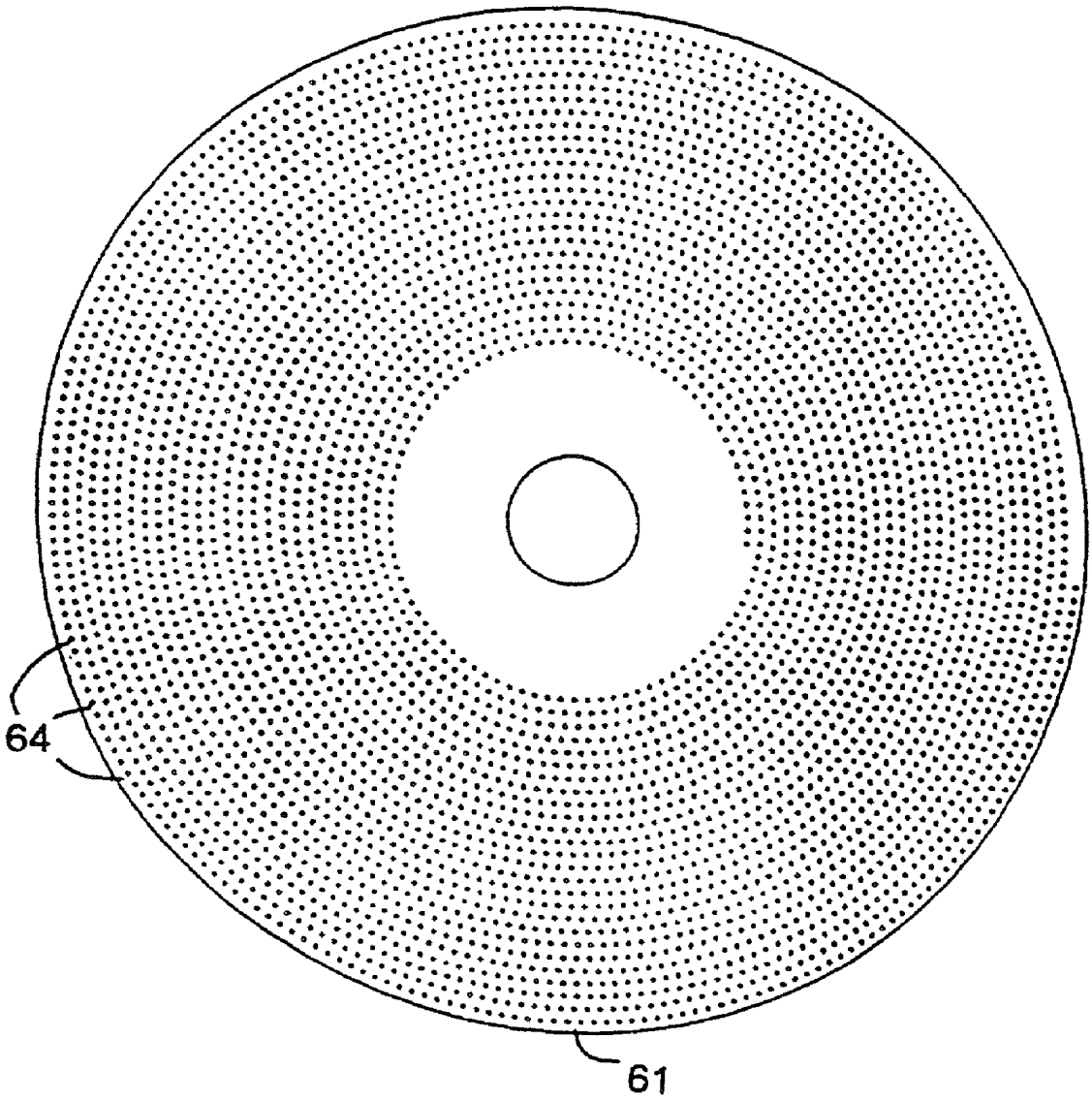


Fig. 22

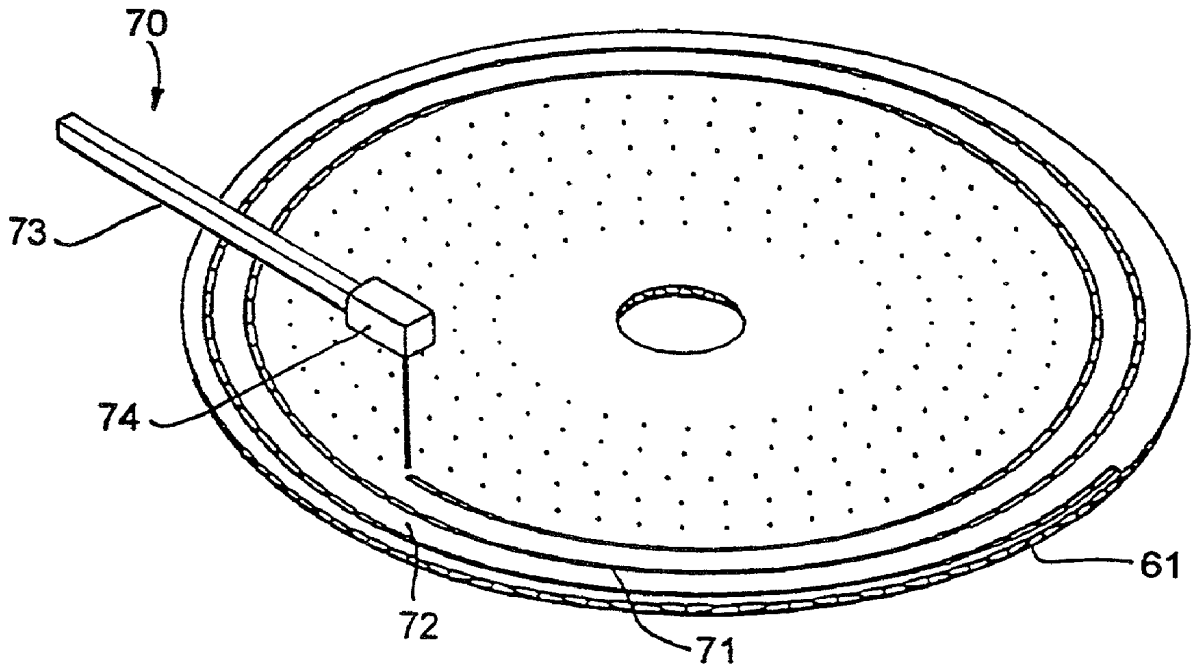


Fig. 23

APPARATUS FOR THE SIMULTANEOUS TRANSFER OF LIQUID ANALYTES

TECHNICAL FIELD

[0001] This invention relates to a method for the rapid screening of large numbers of analytes, including the rapid screening of chemical compounds in liquid form for use as potential drugs.

BACKGROUND ART

[0002] Currently the identification of potential compound candidates for use as drugs is achieved by screening programmes and/or rational drug design. Whereas the ab initio drug design concept originally offered wide expectations of success, this approach has not proved successful in practice, principally due to a lack of a sufficiently clear relationship between molecular structure and receptor sites.

[0003] Accordingly, compound screening is still the technology of choice for the rapid identification and selection of lead compounds as candidate drugs. Various methods of high throughput screening (HTS) are currently used in the screening of compounds as potential drug candidates. However, the speed and cost-effectiveness of HTS is limited by the unavailability of equipment which can simultaneously handle large numbers of compounds in liquid form.

[0004] In current HTS, the microtitre plate plays a central role being the standard device in which assays are performed. The microtitre plate has determined the design of liquid handling equipment such as programmable liquid handling work stations and has also led to the development of microtitre plate peripherals for filtration, washing, reading and other operations involved in HTS.

[0005] To accommodate ever increasing high throughput requirements. microtitre plate-based equipment has been integrated with robotic manipulators. The central role of the microtitre plate and the continuously increasing throughput requirements has led to ever larger and more complex HTS-modules. Such complex modules include robot rails to access wider surfaces, automatic and larger microtitre plate incubators and storage devices for disposables. Additionally, complex operations scheduling programmes and integration software are required for optimal management of all of the hardware components.

[0006] The central role of the microtitre plate also determines other aspects of a typical screening process such as the chemical library aspects. Traditionally, compounds are synthesised stored and catalogued at central chemical repositories from which new compounds are dispatched upon request from biological assay laboratories in or outside an organisation. Often, each new compound has to be weighed and dissolved prior to each new assay. Therefore, it is essential that the compounds can be individually accessed and routinely retrieved. Driven by the latest HTS-system layouts and capacities, efforts are being made to adapt the physical storage conditions of compounds to the microtitre plate format. Hence, the development of dimethylsulfoxide-(DMSO) dissolved stock compound stock solutions which are stored in microtitre plate format and whereby each compound is addressable through its location.

[0007] The basis of the HTS-infrastructure is still the 96-well standard microtitre plate and the majority of screening systems developed to date have been developed for use with this format.

[0008] However, higher density microtitre plates (e.g. 384-, 864-, 1536 and 9600 well plates) are incompatible with most equipment designed for the 96-well plate. As screening formats are likely to change considerably in the future, screening systems must have sufficient flexibility to meet this challenge.

[0009] The pace of HTS development to date has been determined by the development of fast liquid handling systems capable of handling small volumes of liquid to allow for the miniaturisation of assays in microtitre plate formats of which the number of wells has been scaled up dramatically in order to cope with ever increasing numbers of compounds to be screened. However, as the number of wells increases, the volume capacity of the wells decreases dramatically. Therefore it is obvious that this trend is becoming a self-limiting one because of the physical limitations imposed for example by biochemical equilibria relating to tissue culture in general where the control of pH, carbon dioxide exchange, humidity and temperature are of the utmost importance and which parameters are very difficult to control in small volumes of the type employed in high density microtitre plates.

[0010] One problem with the use of high density microtitre plate formats is that it is not possible to readily achieve serial dilution where a dose response curve is required. For example, it is not possible to carry out a serial dilution in the wells as such, so that the serial dilution must be carried out externally of the wells. However, even when the serial dilution is carried out in this way, one still has the problem of the liquid handling aspect of the screening process. Thus, for serial dilution one is currently effectively confined to the use of the 96-well microtitre plate. Also, even when the handling of compounds is under robotic control, the number of compounds that can be handled at any given time is typically 8 or 12 with a maximum of 96.

[0011] We are unaware of any system to date which can simultaneously process large numbers of analytes for screening 100 or more and even up to 1000 analytes simultaneously, hereinafter referred to as a larger number of analytes.

[0012] There is a need therefore for a method for the simultaneous screening of large numbers of analytes and which method obviates the difficulties and limitations of current HTS systems.

[0013] There is also a need for a method for the simultaneous manipulation of large numbers of analytes with means for simplifying identification and retrieval of the analytes for HTS.

DISCLOSURE OF INVENTION

[0014] The invention provides a method for the rapid screening of analytes, comprising the steps of:

[0015] a) simultaneously applying, a plurality of analytes to be screened onto one or more solid support(s) such that the analytes remain isolated from one another,

[0016] b) contacting said analyte-carrying solid support(s) with targets provided in a semi-solid or liquid medium, whereby said analytes are released from the solid support(s) to the targets; and

[0017] measuring analyte-target interactions.

[0018] The method according to the invention allows for the manipulation of thousands of different analytes simultaneously.

[0019] Preferably, step a) of the method according to the invention comprises (i) disposing the analytes within individually identifiable containers, and (ii) transferring the analytes from the containers to the solid support(s) in such a manner as to maintain the transferred contents of each container separate from those of each other container.

[0020] The individually identifiable containers are preferably selected from tubes, including capillary tubes, pens, including plotter pens, and print heads or any container allowing for the storage and direct application of an analyte from the container to a given solid support.

[0021] Further, preferably, the individually identifiable containers are an array of capillary tubes each of which is identifiable according to its position within the array, and wherein transfer of the analytes to the solid support(s) occurs by dispensing thereof through the open ends of the capillary tubes.

[0022] Especially preferred arrays are individual containers disposed in concentric or spiral arrays.

[0023] Transfer of the analytes to the solid support can be achieved by dispensing said analytes from the open ends of the capilliary tubes to the solid support with or without direct contact between the capilliary tubes and the solid support.

[0024] The analytes can be transferred to the solid support in variable amounts. Thus, by varying the drop size transferred one can achieve a serial dilution, if required.

[0025] Simultaneous transfer of analyte from the capilliaries can be achieved by applying a stimulus such as a change of pressure using a piezoelectric element. Alternatively, one can use high frequency conditions to break a liquid column into droplets which are dispensed to the solid support, as required. The droplet size will typically be nanolitre or picolitre in volume.

[0026] Thus, one can achieve an assay format in accordance with the invention where the analyte is not pipetted in as is currently the norm but is applied directly from its individual container to the assay medium.

[0027] It will be appreciated that the individually identifiable containers described herein provide a means of storing stock compounds which can be accessed and used as required. Thus, the analyte application units can consist of individual identifiable containers assembled in addressable compartments that can be retrieved automatically as a whole and from which the analytes can be applied directly to a solid support.

[0028] For example,. a preferred embodiment of the identifiable container is a capilliary tube and stock compounds in solution are taken up in a plurality of such capilliary tubes by capilliary action.

[0029] In this way unlimited numbers of capillaries can be filled without special energy requirements. Capillary tubes filled with stock solutions of compounds can be stored at desired temperatures and conditions.

[0030] The method according to the invention allows for the simultaneous mass application of analytes onto a solid support. For example, as described in greater detail herein one can achieve simultaneous mass application of equal volumes of 10,000 or more compounds from an array of individual containers such as capilliaries to a solid phase. The amount of compound delivered can be determined by the contact time of the capilliaries with the solid support. The analyte containers can also suitably be individual addressable plotter pen-like devices allowing simultaneous drawing of parallel lines of analyte on a solid support of choice.

[0031] After compound delivery the compounds, which can be disposed as a pattern of discrete spots or lines, are left to dry on the solid phase.

[0032] In one embodiment, the solid support is of a substantially flat, disc-, rectangular- or square-shape.

[0033] The solid support can comprise a material which allows for spontaneous release of the analyte(s) when applied thereto.

[0034] Alternatively, the solid support can comprise a material which allows for controlled release of the analyte(s) when applied thereto.

[0035] In each case, the material can be said semi-solid medium.

[0036] Preferably when each analyte is applied to the solid support it diffuses thereon so as to produce a concentration gradient.

[0037] In this way one can achieve a serial dilution of analyte if a dose response curve for a candidate drug is required, rather than a simple positive or negative (yes/no) result.

[0038] Utilisation of retarded analyte diffusion in a semi-solid medium or matrix further eliminates the need for physical separation as in the case of the wells of a microtitre plate and the necessity of serial dilution of analytes, when such is required as a concentration gradient will be established by passive diffusion.

[0039] Preferably the surface of the solid support onto which the analytes are applied is selected from polymers, ceramics, metals, cellulose and glass.

[0040] Further, preferably, said semi-solid medium is disposed on a carrier.

[0041] In another embodiment, the solid support is in the form of a flexible film or tape onto which the target-containing semi-solid medium is applied, whereby the method can be automated using a system of rollers to progress the flexible film or tape through the various steps of the method.

[0042] In this embodiment, the carrier can be covered by a further layer of film or tape and is thereby sandwiched between the solid support and the covering layer.

[0043] Furthermore, the solid support or covering layer (if present) can be provided with a track for the recordal of information regarding the applied analytes, whereby the information can be read and processed simultaneously with the measurement of analyte-target interactions in an automated process.

[0044] In a further embodiment, the solid support is itself a detector or forms part of a detector.

[0045] In this embodiment, the solid support is preferably selected from a SiO_2 wafer, a charge-coupled device and a photographic film.

[0046] The surface of the solid support can be coated with a membrane, a molecular monolayer, a cellular monolayer or a Langmuir-Blodgett film.

[0047] All of these coatings can be used to control the release of analytes when applied thereto.

[0048] In another embodiment, the solid support is itself an information carrier which carries information in electronic, magnetic or digitised form.

[0049] In an alternative embodiment, the surface of the solid support is reflective. For example, the surface can be the reflective surface of a compact disc.

[0050] The method according to the invention can further comprise the step of copying said compact disc to a writable compact disc.

[0051] In another embodiment, the semi-solid medium comprises a substance which provides a semi-solid or viscous liquid environment allowing controlled release of said analytes to said target.

[0052] Preferably, the substance which provides a semi-solid or viscous liquid environment is selected from gelatin, polysaccharides such as agar and agarose, and polymers such as methylcellulose and polyacrylamide or a so-called intelligent material. Such substances can also be used to control the release of the analytes when applied thereto.

[0053] So-called intelligent materials are natural and synthetic polymer gels that are undergo phase transitions and critical phenomena, for example phase transitions accompanied by a reversible, discontinuous volume change as large as several hundred times, in response to infinitesimal chances in environmental conditions.

[0054] Examples of so-called intelligent materials are polymeric gel-type materials, more particularly hydrogels that can take up a fluid and subsequently release that fluid in response to a chemical or physical stimulus or trigger. An example of a chemical stimulus is a change of pH or ionic or solvent composition and an example of a physical stimulus is light of a particular wave-length or a laser beam, a change of temperature or a small electric field.

[0055] For example, a gel containing N-isopropylacrylamide (main constituent) and the light-sensitive chromophore, the trisodium salt of copper undergoes phase transitions induced by visible light (Suzuki, A. and Tanaka, T (Nature (1990); 346, 345-347).

[0056] A range of suitable thermo-sensitive polymers is described by Snowden, M. J. et al. (Chemistry & Industry (July, 1996); p.p. 531-534.

[0057] Other suitable gels are those sold under the Trade Mark THERA GEL marketed by Gel Sciences Inc., Boston, Mass., U.S.A.

[0058] In a further embodiment, steps a) and b) are carried out simultaneously.

[0059] In a still further embodiment, each analyte is applied to a single solid support.

[0060] In this embodiment, the solid support is preferably of a rod shape or a spherical shape.

[0061] Further, preferably each analyte-bearing solid support is contacted in step b) with a target in a separate compartment of a multi-compartmented apparatus, more especially said compartments are an arrangement of mini-wells in said apparatus.

[0062] In another preferred embodiment, the analyte containers are small inert solid supports onto the surfaces of which analytes have been applied. Dipping of the solid supports into a liquid phase or semi-solid phase results in time-dependent release of analyte from the solid support into the liquid or semi-solid phase. In this way, dilutions of minute amounts of analytes in liquid or semi-solid phase can be obtained without the use of liquid handling devices. The final concentration of analyte released into the liquid or semi-solid phase is determined by the contact time between the analyte-carrying solid support and the liquid- or semi-solid phase.

[0063] The analytes for rapid screening in the method according to the invention are preferably selected from chemical compounds, antigens, antibodies, DNA-probes, cells and beads and liposomes carrying an analyte of interest.

[0064] Further, preferably, the analytes, when applied to the solid support, are dissolved in an organic or inorganic solvent.

[0065] Suitably, the solvent includes a so-called intelligent material responsive to a chemical or physical parameter such that each analyte, following application to the solid support and drying, liquifies in response to said chemical or physical parameter.

[0066] The analyte in a preferred embodiment is a chemical compound for screening as a potential drug candidate.

[0067] Preferably, the targets are selected from prokaryotic cells, eukaryotic cells, viruses, molecules, receptors, beads, and combinations thereof.

[0068] As used herein compound(s) means any synthetic, semi-synthetic or naturally-occuring compound or combination thereof.

[0069] In one embodiment the targets are cells equipped with reporter functions.

[0070] In a preferred embodiment the targets are mammalian cells equipped with single or multiple reporter construct(s). The activity and/or expression of the reporter genes is dependent on the impact of the specific experimental conditions, for example the in vitro effect of compounds released from the solid support to a semi-solid phase.

[0071] The incubation unit used in this embodiment will typically be an incubator of the type that is generally used for tissue culture.

[0072] The detection unit used to measure the compound-cell interaction will suitably consist of an inverted microscope (e.g. Zeiss Axiovert 100) coupled to a video camera (e.g. dage-MTI CCD72E) and computer system (e.g. PC 300 Mhz Pentium) with KS400 basic software and the graphic

KS400 option. The inverted microscope can be further equipped with a scanning stage modified to fit all culture plate configurations (from 6-well up to 9600 well formats) as well as other formats of the type described herein and equipped with a stepper motor with a resolution of 17,600 steps per revolution. The KS400 software package provides easy to use menus for entering configuration, calibration and assay parameters or data analysis specifications.

[0073] Other detection means include the use of computer-assisted image analysers.

[0074] In the event that adherent cells are used as a target, these cells are grown confluently either covering the surface of the solid support or the surface of small beads which are then homogeneously suspended in the semi-solid phase.

[0075] Still further, preferably, said analyte-target interactions are measured using one or more of the following methods: microscopic, calorimetric, fluorometric, luminometric, densitometric, isotopic, and physical measurements.

[0076] For example, one can use a combination of microscopy and fluorescence. Accordingly,. the microscope can be configured for epifluorescence fitted with a stepper motor and a camera system. Image acquisition and interpretation can be PC controlled.

[0077] It will be appreciated that the method according to the invention provides for the coupling of the principle of compound diffusion in semi-solid media containing experimental biological systems to image microscopy systems. The invention also provides for visualising and determining the effect of compounds on prokaryotic and eukaryotic cell function. This visualisation can be achieved with genetically engineered cells embedded in a semi-solid medium and expressing single or multiple reporter genes of which the expression product(s) is/are an intracellular fluorophore(s) of specific excitation and emission wavelengths matching wavelength characteristics of a high density fluorescence image microscopy system.

[0078] By way of example, the method according to the invention typically comprises the steps of:

[0079] a) direct application of compounds to be screened as potential drug candidates onto a solid support;

[0080] b) layering of a semi-solid medium containing targets of interest over the compound carrying support;

[0081] c) diffusion of compounds into the semi-solid medium;

[0082] d) incubation;

[0083] e) visualisation and registration of the compound-target interaction.

[0084] By way of example if one wishes to screen potential drugs or drug combinations for use in the treatment of individuals infected with the HIV virus, having full-blown AIDS or ARC, then suitable targets for use in accordance with the invention are genetically engineered green fluorescent protein (GFP)-expressing MT4 cells containing the long term no repeat (LTR) promotor (pLTR-EGFP-C1) and deposited at the BCCM on Aug. 20, 1998, under Accession No. LMBP3879. pLTR-EGFP-C1 (4777 bp) is based on pEGFP-C1 (Clontech). In pEGFP-C1 expression of the enchanced green fluorescent protein (EGFP) is controlled by the strong human cytomegalovirus immediate early promotor (589 bp). In pLTR-EGFP-C1 the CMV promotor is replaced by the HIV-1 long terminal repeat LTR (652bp) containing the highly inducible promotor in the U_3 region.

[0085] Such cells when infected with HIV become fluorescent due to activation of LTR and expression of GFP. If a candidate drug inhibits HIV, for example by inhibiting the viral reverse transcriptase, protease or integrase fluorescence is reduced or totally inhibited and one observes progressively darkened regions in the area of the compound-target interaction.

[0086] Step b) of the method according to the invention will typically involve maintaining the analytes and the targets under appropriate assay conditions and for a sufficient period of time for the release of the analytes from the solid support and the diffusion of said analytes into the liquid or semi-solid phase containing the targets of interest. Maintaining appropriate assay conditions will include maintaining correct temperature, osmolality, pH, tonicity and the like. These conditions are further determined by the nature of the targets.

[0087] Typically, with mammalian cells as a target, the temperature can range from about 4° C. to about 50° C. and pH can range from about 6.5 to about 7.5. Osmolality and tonicity will be chosen in a manner so that optimal cell-analyte interactions can be obtained. The only limitation on physical conditions is that the conditions used do not adversely affect cell viability nor interfere with analyte target interactions.

[0088] It will be appreciated that the method according to the invention can be carried out in a fully integrated system.

[0089] The method according to the invention facilitates the use of a data management unit. Such a unit suitable comprises chemical compound library data with compound identification, history tracking and software technology allowing accessing and identification of compounds within the warehouse installation at all times.

[0090] In another preferred embodiment, analyte application, analyte target interaction, incubation, detection and data interpretation are linked in such a way that the entire screening method according to the invention is a fully automated method from start to finish. One typical example of an automated method of rapid screening of compounds according to the invention comprises the steps of:

[0091] 1. Application of compounds by means of capillary delivery producing either discrete spots or discrete lines of compounds onto the surface of a carrier, such as a transparent film or information carrier used in the audio-casette industry;

[0092] 2. Drying the surface of the carrier;

[0093] 3. Layering of cells embedded in a semi-solid matrix of constant thickness over the surface of a transparent film;

[0094] 4. Contacting the surface of the carrier containing spotted compounds or discrete compound lines and the surface of the film serving as the semi-solid matrix carrier;

[0095] 5. Winding of the contacted film surfaces;

[0096] 6. Incubation of the wound film;

[0097] 7. Unwinding of the film and exposure to a detection and information reading unit;

[0098] 8. Continuous reading of exposed film by means, for example, of a fluorescence microscope image analysis system: and

[0099] 9. Data analysis.

[0100] Outer surfaces: (non-compound carrying or which are not in contact with the semi-solid matrix) can contain digital information regarding the identity of the applied compounds which is read simultaneously during sample analysis as described in step 8. In this way, biological information and compound information can be read and processed at the same time.

BRIEF DESCRIPTION OF THE DRAWINGS

[0101] **FIG. 1** illustrates the principle of compound diffusion in a semisolid matrix as described in Example 1 at a cell density of 10E5/ml;

[0102] **FIG. 2** illustrates the principle of compound diffusion in a semisolid matrix as described in Example 1 at a cell density of 10E6/ml;

[0103] **FIG. 3** illustrates the principle of compound diffusion in a semisolid matrix as described in Example 1 at a cell density of 10E7/ml;

[0104] **FIG. 4** is a capillary tube holder device (8×12) as described in Example 2;

[0105] **FIG. 5** shows the fluorescence observed with GFP (green fluorescent protein) expressing MT4-cells (LTR (long term no repeat) promotor) in RPMI (Rosemount Park Memorial Institute) medium without phenol red, 10% FCS (fetal calf serum) and 1% Pen-Strep (penicillin-streptomycin) in the absence (a) and in the presence of HIV-1 (b) as described in Example 3;

[0106] FIGS. **6**(*a*) and (*b*) show the fluorescence observed with GFP expressing MT4-cells (LTR promotor) in semi-solid phase (RPMI medium without phenol red. 10% FCS, 1% Pen-Strep. agar 0.34%) in the absence (a) and in the presence of HIV-1 (b) as described in Example 3;

[0107] FIGS. **7**(*a*)-(*e*) show the fluorescence observed with GFP-expressing MT4-cells (LTR promotor) in RPMI medium (without phenol red and supplemented with 10% FCS, 1% Pen-Strep) in the absence (a) and in the presence of HIV-1 (b) and in the presence of HIV-1 and the reverse transcriptase inhibitors: AZT (c), 3TC (d) and Loviride (e) at a final concentration of 2.5 μM in a total volume of **20 μl** as described in Example 3;

[0108] FIGS. **8**(*a*)-(*e*) show the fluorescence observed with GFP-expressing MT4-cells (LTR promotor) in semi-solid phase 0.34% agar in RPMI medium (RPMI medium without phenol red, supplemented with 10% FCS and 1% Pen-Strep) in the absence (a) and in the presence of HIV-1 (b) and in the presence of HIV-1 and the reverse transcriptase inhibitors: AZT (c), 3TC (d) and Loviride (e) spotted (1 μl of a stock solution) onto a surface of a solid support at a concentration producing a 2.5 μM end concentration, assuming complete diffusion of the compounds in 20 μl semi-solid phase as described in Example 3;

[0109] **FIG. 9** shows the fluorescence observed with HIV-1 infected MT4-cells in semi-solid phase (0.34% agar in RPMI medium without phenol red, supplemented with 10% FCS and 1% Pen-Strep) when HIV-1 infected cells were admixed with a solid support onto which 1 μl of 2.5 μM, 250 nM and 2.5 nM of reverse transcriptase inhibitors were applied, left to dry and kept for one week at 4° C. prior to use as described in Example 3:

[0110] **FIG. 10** shows the fluorescence observed with HIV-1 infected MT4-cells in medium (RPMI without phenol red, supplemented with 10% FCS and 1% Pen-Strep) when HIV-1 infected cells were added in medium to the wells of a 384-well tissue culture plate containing 1 μl of 2.5 μM, 250 nM and 2.5 nM of the reverse transcriptase inhibitors AZT, 3TC and Loviride, respectively as described in Example 3;

[0111] **FIG. 11** is a schematic representation of a solid support carrying spotted compounds in the method according to the invention;

[0112] **FIG. 12** illustrates schematically how the distance between capillaries in an array can be varied and adapted to the specific requirements of the method according to the invention;

[0113] **FIG. 13** is a diffusion pattern of calcein in a semi-solid phase on a solid support in accordance with the invention;

[0114] **FIG. 14** illustrates the principle of automated on-line mega-throughput screening using the method according to the invention;

[0115] **FIG. 15** is a schematic representation with exploded detail of capilliary tube sorting means;

[0116] FIGS. **16**(*a*)-(*c*) show the movement of the capilliary tubes of **FIG. 15** to a microtitre plate for filling;

[0117] **FIG. 17** illustrates the filling of the capilliary tubes of **FIG. 15**;

[0118] **FIG. 18** shows a screw device used to reduce the spacing between the capilliary tubes of **FIG. 15**;

[0119] **FIG. 19** shows the individual capilliary tubes of **FIG. 15** being formed into a spiral array:

[0120] **FIG. 20** illustrates means by which the liquid in the capilliary tubes of **FIG. 15** is released onto a solid support;

[0121] **FIG. 21** illustrates a sample deposition pattern;

[0122] **FIG. 22** is a plan view of a solid support with analytes spotted thereon; and

[0123] **FIG. 23** illustrates the application of a semi-solid phase to the solid support.

MODES FOR CARRYING OUT THE INVENTION

[0124] The invention will be further illustrated by the following Examples:

EXAMPLE 1

Principle of Compound Diffusion and Interaction with Cells Embedded in a Semi-solid Medium

[0125] Calcein a cell viability marker was dissolved at a concentration of 5 mM in dimethyl sulfoxide (DMSO). A

glass capillary tube with a total volume capacity of 0.5 μl was dipped into the calcein solution and filled by capillary action. The tip of the capillary tube was then contacted with a polystyrene surface in a such a way that a small drop of calcein solution was delivered from the capillary tube to the plastic surface. After drying of the drop, 20 μl of a cell suspension in semisolid medium (MT4 cells suspended in RPMI (Rosemount Park Memorial Institute) 1640 medium, without phenol red, supplemented with 10% FCS (fetal calf serum), 1% Pen-Strep (penicillin-streptomycin) and containing 0.34% agar) was layered over the dried calcein spot. After an incubation time of 2 hours at 37° C. (humidified atmosphere and 5% carbon dioxide) the diffusion of the calcein into the semi-solid phase was observed by means of fluorescence microscopy and visualisation of the fluorescence produced by the embedded MT4 cells. The method of drop delivery, drying and layering of semi-solid matrix containing increasing densities of embedded MT4 cells is illustrated in **FIGS. 1, 2** and **3**.

[**0126**] It follows from these results that the distance over which diffusion of calcein takes place in a semi-solid matrix of constant density is also determined by the number of embedded cells.

EXAMPLE 2

Principle of High Density Compound Application and Diffusion in a Semisolid Matrix

[**0127**] A bundle of capillaries filled with calcein and arranged (8×12) in a holder device as depicted in **FIG. 4** was contacted with a surface of polystyrene so that a drop of calcein was delivered simultaneously from each capillary to the polystyrene surface. The holder device is indicated generally at **10** and comprises capillary tubes **11** mounted in plates **12, 13** for maintaining the capillary tubes **11** in the desired relationship with respect to each other. Following drying, a homogeneous suspension of MT4 cells in RPMI 1640 medium supplemented with 10% FCS, 1% Pen-Strep and 0.34% agar was layered over the spots. After an incubation period of 2 hours (humidified atmosphere. 5% carbon dioxide) it was found that for each of the spots the distance of diffusion of the calcein in the semi-solid matrix was reflected by the fluorescence of the embedded MT4 cells.

EXAMPLE 3

Compound Target Interaction: Effect of Anti-HIV Compounds on the Fluorescence of GFP-expressing MT4 Cells (LTR Promotor) in the Presence of HIV-1 in a Semi-solid Phase

[**0128**] Three compounds of well known activity against HIV-1 (AZT, 3TC and Loviride) were spotted (+/−1 μl from stock in a capillary tube as hereinbefore described) onto the bottom surface of the wells of a transparent 384-well polystyrene tissue culture plate. The compounds in the wells were left to dry and stored at 4° C.

[**0129**] One week later, MT4 cells were collected from tissue culture flasks and suspended at 10E7/ml in RPMI medium. This cell suspension was further equally divided into four tubes. These tubes were then centrifuged at 450 g, for 10 min. To the cell pellets obtained after centrifugation of two of these tubes. 200 μl HIV suspension in RPMI medium were added for a period of 2 hours at 37° C. The other two tubes were treated in the same way, except that no virus was added. After an incubation time of 2 hours, agar solution (39° C.) was added to one tube containing cells and HIV and to one tube only containing cells at a final concentration of 0.34%. Then, 20 μl of the cell suspension in agar and 20 μl of the cell/virus-agar suspension were added to the different wells of a 384-well plate containing the spotted compounds as set forth above.

[**0130**] To the cell pellets of the remaining two tubes, 200 μl medium and 200 μl virus containing medium were added respectively. After an incubation period of 2 hours at 37° C., the final volume was corrected (made equal to the final volume of the agar composition) and 20 μl of the non-infected and infected cell suspensions were added to the wells of the 384-well culture plate with spotted compounds as described above.

[**0131**] 1 μl volumes of compounds were added to the wells immediately before the HIV-infected cells were added. Total assay volume was 20 μl.

[**0132**] After a 3-day incubation period, the fluorescence of the GFP-expressing MT4 cells was evaluated by fluorescence microscopy and plate reading. The results are summarized in FIGS. **5-10**. These data show that following the application of compounds onto a solid support (well of a 384-well microtitre plate) and after storage for 1 week at 4° C., the activity (protective effect of these compounds against HIV-1 infection) did not differ from the situation where compound diffusion takes place in a liquid phase.

[**0133**] Furthermore, the results obtained show that the concentration-dependent effects of RT inhibitors on HIV-1 infection as reflected by GFP-expressing MT4-cells are observed in semi-solid phase and that the nature of the compound and its effectiveness for HIV-1 RT inhibition is not affected by the use of a semi-solid medium.

[**0134**] The invention will be further illustrated by the following description of embodiments thereof given by way of example only with reference to the accompanying drawings.

[**0135**] Referring to **FIG. 11**, there is indicated a solid support **20** carrying compounds **21** which have been spotted thereon from an array of 196 bundles of capillary tubess (not shown) each carrying 110 capillary tubes so that 21,560 compounds are spotted onto the solid support **20**.

[**0136**] **FIG. 12** illustrates how the distance between the capillaries in an array can be varied (1.414 mm, 1 mm, 2.236 mm, 2 mm, 3.623 mm, 3 mm) to meet the specific requirements of a screening method in accordance with the invention.

[**0137**] **FIG. 13** is a diffusion pattern of calcein in a semi-solid phase on a solid support. Calcein was spotted on a polystyrene surface using a capillary compound holder device as depicted in **FIG. 4** with capillary tubes arranged at a centre to centre distance of 2 mm. The density of overlying cells, suspended in the semi-solid phase (RPMI 1640 medium, **10**% FCS, 1% Pen-Strep, 0.34% agar) was 10E7 cells (MT4)/ml. Detection was carried out by fluorescence microscopy after a two hour incubation period.

[**0138**] **FIG. 14** is a schematic representation of an automated method for the rapid screening of compounds in accordance with the invention. An information carrier **30** in

the form of a film or tape and with compounds to be screened applied as discrete spots or lines on its surface **31** is brought into contact with surface **32** of an information carrier **33**, which is also a film or tape, bearing targets of interest embedded in a semi-solid matrix. The respective carriers **30**, **33** are then wound with their surfaces **31** and **32** in contact and incubated in a temperature, humidity and carbon dioxide controlled environment, such that the compounds are released to the surface **32** of the carrier **33**. The carriers are then unwound and the carrier **33** is then passed to an analysis—and information reading unit indicated generally at **34**.

[**0139**] In the following FIGS. **15-23** like parts are denoted by the same reference numerals.

[**0140**] Referring to **FIG. 15**, there is indicated generally at **40**, apparatus for feeding capillary tubes **41** from a supply thereof to a conveyor belt **42** with regularly spaced-apart transverse grooves **43**. The capillary tubes **41** enter a channel **44** which can accommodate a single layer of capillary tubes **41** in response to the anti-clockwise movement of a belt **45** and are delivered one at a time to the grooves **43** as the conveyor belt **42** moves in a clockwise direction. The capillary tubes **41** travel along the channel **44** through the combined effects of gravity and the belt **45**. Each time a groove **43** is positioned at end **46** a capillary tube **41** is delivered thereto.

[**0141**] Referring to **FIGS. 16***a-c* and **FIG. 17**, the steps involved in transferring the capillary tubes **41** to a microtitre plate **47** for filling are illustrated. Capillary tubes **41** are lifted from the belt **42** by a clamping device indicated generally at **48**. The clamping device **48** comprises two elongate members **49** which clamp a plurality of tubes **41** therebetween by applying a sideways force to the tubes. In this way, the capillary tubes **41** are transferred from the conveyer belt **42** to the microtitre plate **47** for filling. The spacing of the grooves **43** on the belt **42** is the same as the spacing of the wells **50** in the microtitre plate **47**. The clamping device **48** lifts and transports the tubes **41** in groups of **12** corresponding to the number of wells **50** in one row of the microtitre plate **47**. During the transporting step, the clamping device **48** is rotated through 90° so that one end of the capillary tubes **41** is lowered into the wells **50** of the microtitre plate **47**.

[**0142**] The clamping device **48** and the capillary tubes **41** are free to move along a vertical axis to allow the tubes **41** to be lowered into the wells **50** of the microtitre plate **47**. The capillary tubes **41** are held in the wells **50** for a period of time sufficient to allow liquid in the respective wells **50** to be drawn into the tubes **41** by capillary action. Once the given filling time has elapsed, the tubes **41** are withdrawn and the clamping device **48** is rotated through 90° to again assume a horizontal orientation.

[**0143**] Following filling of the capillary tubes **41**, said tubes are transferred to a screw—or worm device **51** which is used to reduce the spacing between the capillary tubes **41** as shown in **FIG. 18**. The capillary tubes **41** are delivered to the screw device **51** at end **52**. Thread **53** of the screw device **51** has a varying pitch, being larger at end **52** than at end **54**. As the screw device **51** turns, the capillary tubes **41** advance along its length in the direction of the arrow and due to the changing pitch also come closer together. At the end **54** tubes **41** are only separated by wall **55** of the thread **53**.

[**0144**] Referring to **FIG. 19**, the capillary tubes **41** are discharged from the screw device **51** onto a tape **56** which has a layer of adhesive to which the tubes **41** are stuck with their sides abutting. The tape **56** advances at the same rate as the tubes **41** leave the screw device **51**. The tape **56** is then progressively rolled up into a tightly packed spiral array as shown at **57**.

[**0145**] Referring to **FIG. 20**, there is indicated Generally at **60**, apparatus for releasing the liquid in the capillary tubes **41** defining the array **57** to the surface of a solid support **61** and which is movable relative thereto. The apparatus **60** comprises a housing **62** which is adapted to receive said array **57**. The housing **62** is connected to an air pump **63** and by creating an area of positive pressure relative to the exterior of the housing **62** forces the liquid out of the tubes **41** onto the surface of the solid support **61**, when required.

[**0146**] **FIG. 21** shows the array **57** of capillary tubes **41** disposed above the solid support **61** following application of droplets of liquid analytes from the tubes **41** and the pattern of discrete spots **64** disposed on said solid support **61**.

[**0147**] **FIG. 22** is a plan view of the solid support **61** showing the spiral arrangement of the discrete spots **64** of the applied liquid analyte.

[**0148**] Referring to **FIG. 23**, there is illustrated a device indicated generally at **70**, for applying target cells in a semi-solid phase **71** to surface **72** of the solid support **61** following drying of the liquid analytes. Device **70** comprises an arm **73** and a printing head **74**. The solid support **61** is free to rotate and the device **70** is free to move in the z plane and x or y planes, so that the printing head **74** is located by a combination of motion of the arm **73** and the solid support **61**.

BELGIAN COORDINATED COLLECTIONS OF MICROORGANISMS – BCCM™/ LMBP-COLLECTION
Page 1 of Form BCCM™/LMBP/BP/4/98-16 Receipt in the case of an original deposit

---

Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure

Receipt in the case of an original deposit issued pursuant to Rule 7.1 by the International Depositary Authority BCCM™/LMBP identified at the bottom of next page

International **Form BCCM™/LMBP/BP/4/98-16**

To : **Name of the depositor : TIBOTEC N.V.**

Address : Institute for Antiviral Research
Generaal De Wittelaan L11 B3
B-2800 Mechelen
Belgium

I. **Identification of the microorganism:**

I.1 **Identification reference given by the depositor:**

pLTR-EGFP-C1

I.2 **Accession number given by the International Depositary Authority:**

LMBP3879

BELGIAN COORDINATED COLLECTIONS OF MICROORGANISMS – BCCM™/ LMBP-COLLECTION
Page 2 of Form BCCM™/LMBP/BP/4/98-16 Receipt in the case of an original deposit

II. **Scientific description and/or proposed taxonomic designation**

The microorganism identified under I above was accompanied by:

(mark with a cross the applicable box(es)):

☒ a scientific description

☐ a proposed taxonomic designation

III. Receipt and acceptance

This International Depositary Authority accepts the microorganism identified under I above, which was received by it on (date of original deposit) : August 20, 1998

IV. International Depositary Authority

Belgian Coordinated Collections of Microorganisms (BCCM™)
Laboratorium voor Moleculaire Biologie - Plasmidencollectie (LMBP)
Universiteit Gent
K.L. Ledeganckstraat 35
B-9000 Gent, Belgium

Signature(s) of person(s) having the power to represent the International Depositary Authority or of authorized official(s):

BELGIAN COORDINATED COLLECTIONS OF MICROORGANISMS – BCCM™/ LMBP-COLLECTION
Page 1 of Form BCCM™/LMBP/BP/9/98-16 Viability statement

Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure

Viability statement issued pursuant to Rule 10.2 by the International Depositary Authority BCCM™/LMBP identified on the following page

***International* Form BCCM™/LMBP/BP/9/98-16**

To : **Party to whom the viability statement is issued:**

Name : Chris Roelant

Address : TIBOTEC N.V.
Institute for Antiviral Research
Generaal De Wittelaan L11 B3
B-2800 Mechelen
Belgium

I. **Depositor:**

I.1 **Name** : TIBOTEC N.V.

I.2 **Address** : Institute for Antiviral Research
Generaal De Wittelaan L11 B3
B-2800 Mechelen
Belgium

II. **Identification of the microorganism:**

II.1 Accession number given by the International Depositary Authority:

LMBP3879

II.2 Date of the original deposit (or where a new deposit or a transfer has been made, the most recent relevant date) : August 20, 1998

III. **Viability statement.**

The viability of the microorganism identified under II above was tested on

: August 25, 1998

(Give date. In the cases referred to in Rule 10.2(a)(ii) and (iii), refer to the most recent viability test).

On that date, the said microorganism was: (mark the applicable box with a cross)

- ● viable
- o no longer viable

BELGIAN COORDINATED COLLECTIONS OF MICROORGANISMS – BCCM™/ LMBP-COLLECTION
Page 2 of Form BCCM™/LMBP/BP/9/98-16 Viability statement

IV. Conditions under which the viability test has been performed:

(Fill in if the information has been requested and if the results of the test were negative).

V. International Depositary Authority

Belgian Coordinated Collections of Microorganisms (BCCM™)
Laboratorium voor Moleculaire Biologie - Plasmidencollectie (LMBP)
Universiteit Gent
K.L. Ledeganckstraat 35
B-9000 Gent, Belgium

Signature(s) of person(s) having the power to represent the International Depositary Authority or of authorized official(s):

1. A method for the rapid screening of analytes, comprising the steps of:

a) simultaneously applying a plurality of analytes to be screened onto one or more solid support(s) such that the analytes remain isolated from one another;

b) contacting said analyte-carrying solid support(s) with targets provided in a semi-solid or liquid medium, whereby said analytes are released from the solid support(s) to the targets; and

c) measuring analyte-target interactions.

2. A method according to claim 1, wherein step (a) comprises (i) disposing the analytes within individually identifiable containers, and (ii) transferring the analytes from the containers to the solid support(s) in such a manner as to maintain the transferred contents of each container separate from those of each other container.

3. A method according to claim 2, wherein the individually identifiable containers are selected from tubes, including capillary tubes, pens, including plotter pens, and print heads.

4. A method according to claim 3, wherein the individually identifiable containers are an array of capillary tubes each of which is identifiable according to its position within the array, and wherein transfer of the analytes to the solid support(s) occurs by dispensing thereof through the open ends of the capillary tubes.

5. A method according to any one of claims **1-4**, wherein the solid support is of a substantially flat, disc-, rectangular- or square-shape.

6. A method according to claim 5, wherein the solid support comprises a material which allows for spontaneous release of the analyte(s) when applied thereto.

7. A method according to claim 5, wherein the solid support comprises a material which allows for controlled release of the analyte(s) when applied thereto.

8. A method according to claim 6 or **7**, wherein said material is said semi-solid medium.

9. A method according to any preceding claim, wherein when each analyte is applied to the solid support it diffuses thereon so as to produce a concentration gradient.

10. A method according to any preceding claim, wherein the surface of the solid support onto which the analytes are applied is selected from polymers, ceramics, metals, cellulose and glass.

11. A method according to any preceding claim, wherein said semi-solid medium is disposed on a carrier.

12. A method according to claim 11, wherein the solid support is in the form of a flexible film or tape onto which the target-containing semi-solid medium is applied, whereby the method can be automated using a system of rollers to progress the flexible film or tape through the various steps of the method.

13. A method according to claim 12, wherein the carrier is covered by a further layer of film or tape and is thereby sandwiched between the solid support and the covering layer.

14. A method according to claim 12 or **13**, wherein the solid support or covering layer (if present) is provided with a track for the recordal of information regarding the applied analytes, whereby the information can be read and processed simultaneously with the measurement of analyte-target interactions in an automated process.

15. A method according to any one of claims **1-10**, wherein the solid support is itself a detector or forms part of a detector.

16. A method according to claim 15, wherein the solid support is selected from a SiO_2 wafer, a charge-coupled device, and a photographic film.

17. A method according to any preceding claim, wherein the surface of the solid support is coated with a membrane, a molecular monolayer, a cellular monolayer or a Langmuir-Blodgett film.

18. A method according to any preceding claim, wherein the solid support is itself an information carrier which carries information in electronic, magnetic or digitised form.

19. A method according to any preceding claim, wherein said surface of the solid support is reflective.

20. A method according to claim 19, when dependent on claim 17, wherein said surface is the reflective surface of a compact disc.

21. A method according to claim 20, further comprising the step of copying said compact disc to a writable compact disc.

22. A method according to any preceding claim, wherein the semi-solid medium comprises a substance which provides a semi-solid or viscous liquid environment allowing controlled release of said anlaytes to said target.

23. A method according to claim 22, wherein said substance is selected from gelatin, polysaccharides such as agar and agarose, and polymers such as methylcellulose and polyacrylamide or a so-called intelligent material.

24. A method according to any preceding claim, wherein steps a) and b) are carried out simultaneously.

25. A method according to claim 1, wherein each analyte is applied to a single solid support.

26. A method according to claim 25, wherein the solid support is of a rod shape or a spherical shape.

27. A method according to claim 25 or **26**, wherein each analyte-bearing solid support is contacted in step b) with a target in a separate compartment of a multi-compartmented apparatus.

28. A method according to claim 27, wherein said compartments are an arrangement of mini-wells in said apparatus.

29. A method according to any preceding claim, wherein the analytes are selected from chemical compounds, antigens, antibodies, DNA-probes, cells and beads and liposomes carrying an analyte of interest.

30. A method according to claim 29, wherein the analytes, when applied to the solid support, are dissolved in an organic or inorganic solvent.

31. A method according to claim 30, wherein the solvent includes a so-called intelligent material responsive to a chemical or physical parameter such that each analyte following application to the solid support and drying liquifies in response to said chemical or physical parameter.

32. A method according to any one of claims **29-31**, wherein the analyte is a chemical compound.

33. A method according to any preceding claim, wherein said targets are selected from prokaryotic cells, eukaryotic cells, viruses, molecules, receptors, beads, and combinations thereof.

34. A method according to claim 33, wherein the targets are cells equipped with reporter functions.

35. A method according to claim 34, wherein said analyte-target interactions are measurable by the effects of the analytes on the reporter functions of the cells.

36. A method according to any preceding claim, wherein said analyte-target interactions are measured using one or more of the following methods: microscopic, colorimetric, fluorometric, luminometric, densitometric, isotopic, and physical measurements.

37. A method according to claim 1, substantially as hereinbefore described with reference to and as illustrated in the accompanying drawings.

38. A method according to claim 1, substantially as hereinbefore described with reference to the accompanying Examples.

31. A method according to claim 30, wherein the solvent includes a so-called intelligent material responsive to a chemical or physical parameter such that each analyte following application to the solid support and drying liquifies in response to said chemical or physical parameter.

32. A method according to any one of claims **29-31**, wherein the analyte is a chemical compound.

33. A method according to any preceding claim, wherein said targets are selected from prokaryotic cells, eukaryotic cells, viruses, molecules, receptors, beads, and combinations thereof.

34. A method according to claim 33, wherein the targets are cells equipped with reporter functions.

35. A method according to claim 34, wherein said analyte-target interactions are measurable by the effects of the analytes on the reporter functions of the cells.

36. A method according to any preceding claim, wherein said analyte-target interactions are measured using one or more of the following methods: microscopic, calorimetric, fluorometric, luminometric, densitometric, isotopic, and physical measurements.

37. A method according to any one of claims **12-36**, wherein:

a) a first information carrier, in the form of a film or tape, having analytes to be screened applied to a surface thereof as discrete spots or lines, is brought into contact with a second information carrier, which carrier is also in the form of a film or tape, having targets of interest embedded in a semi-solid matrix on a surface thereof;

b) the respective carriers are wound with their respective analyte- and target-bearing surfaces in contact;

c) the wound carriers are incubated under conditions at which the analytes are released from the first carrier to the target-bearing surface;

d) the first and second carriers are unwound; and

e) the second information carrier is passed to an analysis and information reading unit.

38. A method for the rapid screening of analytes which comprises:

a) bringing a first information carrier, in the form of a film or tape, having analytes to be screened applied to a surface thereof as discrete spots or lines, into contact with a second information carrier, which carrier is also in the form of a film or tape, having targets of interest embedded in a semisolid matrix on a surface thereof;

b) winding the respective carriers with their respective analyte- and target-bearing surfaces in contact;

c) incubating the wound carriers under conditions at which the analytes are released from the first carrier to the target-bearing surface;

d) unwinding the first and second carriers; and

e) passing the second information carrier to an analysis and information reading unit.

39. An apparatus comprising:

a) an array of capillary tubes from which be simultaneously released to a surface of b said support being movable relative to the an

b) a housing adapted to receive the array, said h connected to an air pump capable of expelling from their respective capillary tubes onto the by means of a pressure change.

40. An apparatus according to claim 39, wherein said in liquid form.

41. A method according to claim 1, substantially as he described with reference to and as illustrated in the accompanyi drawings.

42. A method according to claim 1, substantially as her described with reference to the accompanying Examples.

43. A method according to claim 38, substantially as he described and exemplified.

44. An apparatus according to claim 39, substantially as hereinbefore described with particular reference to and as illustrat of the accompanying drawings.

* * * * *

专利名称(译)	用于同时转移液体分析物的装置		
公开(公告)号	US20020081629A1	公开(公告)日	2002-06-27
申请号	US10/025391	申请日	2001-12-19
[标]申请(专利权)人(译)	悌柏泰克公司		
申请(专利权)人(译)	Tibotec公司N.V.		
当前申请(专利权)人(译)	Tibotec公司N.V.		
[标]发明人	PAUWELS RUDI WILFRIED JAN ROELANT CHRISTIAAN HUBERT SIMON VAN ACKER KOENRAAD LODEWIJK AUGUST		
发明人	PAUWELS, RUDI WILFRIED JAN ROELANT, CHRISTIAAN HUBERT SIMON VAN ACKER, KOENRAAD LODEWIJK AUGUST		
IPC分类号	C40B60/14 G01N33/543 G01N33/53 C12M1/34		
CPC分类号	B01J2219/00274 G01N33/54366 C40B60/14 B01J2219/00369		
优先权	PCT/IB1998/001399 1998-09-08 WO		
外部链接	Espacenet USPTO		

摘要(译)

用于快速筛选分析物（例如潜在候选药物）的方法包括以下步骤：将待筛选的多种分析物施加到一个或多个固体支持物（61）上，使得分析物保持彼此隔离;使所述携带分析物的固体支持物（61）与在半固体或液体介质中提供的靶接触，由此所述分析物从固体支持物（61）释放到靶上;并测量分析物 - 目标相互作用。该方法允许同时操纵数千种不同的分析物。当分析物被施加到固体支持物（61）上时，如果需要候选药物的剂量响应曲线，它可以在其上扩散以产生浓度梯度和分析物的连续稀释。所描述的方法可以容易地自动化。