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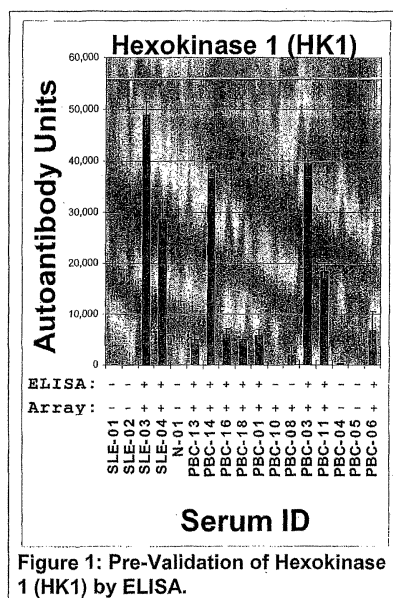
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(54) **A METHOD FOR DIAGNOSING PRIMARY BILIARY CIRRHOSIS (PBC) USING NOVEL AUTOANTIGENS**

(57) Methods and compositions are described for the diagnosis of primary biliary cirrhosis. Novel autoantigens are described for use in assays which employ test samples from individuals.



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Description**FIELD OF THE INVENTION**

5 **[0001]** This invention relates to molecular biology, biochemistry, cell biology, medicine and medical diagnostics. Specifically, the invention relates to novel nucleic acid molecules, proteins and polypeptide fragments encoded thereby, polyclonal and monoclonal antibodies thereto, and methods of using the nucleic acid molecules, proteins/polypeptides and antibodies in diagnostic, prognostic, staging and therapeutic regimens for the control of autoimmune disorders, viral diseases and cancers.

BACKGROUND OF THE INVENTION

10 **[0002]** More than 80 illnesses have been described that are associated with activation of auto-reactive lymphocytes and the production of autoantibodies directed against normal tissue or cellular components (autoantigens) [von Muhlen and Tan (1995) *Semin Arthritis Rheum* 24: 323-58; Mellors (2002) 2005]. Collectively referred to as autoimmune diseases, they are estimated to afflict 14.7-23.5 million people, up to 8% of the total U.S. population and constitute a major economic and health burden [Jacobson, Gange, Rose and Graham (1997) *Clin Immunol Immunopathol* 84: 223-43]. For unknown reasons, the number of people afflicted by autoimmune diseases is on the rise. An autoimmune diagnosis means a lifetime of illness and treatment, possible organ damage, debilitation and an increased chance of mortality. The chronic and often debilitating nature of autoimmune diseases results in poor patient health, increased medical costs, and decreased productivity. The root causes of the immune dysfunction underpinning autoimmune disease are still not well understood. Consequently, autoimmune diseases generally remain difficult to diagnose, due to the wide variability of clinical presentation, which typically involves a constellation of symptoms.

20 **[0003]** Autoimmune diseases are disorders in which an individual's immune system targets and destroys apparently normal tissue. Examples of autoimmune diseases include rheumatoid arthritis (RA), systemic lupus erythematosus (SLE), scleroderma (SCL), Sjogren's syndrome (SjS), polymyositis (PM), dermatomyositis (DM), mixed connective tissue disease (MCTD), pemphigus vulgaris (PV) and primary biliary cirrhosis (PBC). Autoantibodies are commonly directed against cellular proteins and nucleic acids. In certain diseases, such as PV, the target of autoantibodies is known and the autoantibody is thought to play a role in the pathogenesis of the disease. In other diseases, such as SLE, the targets of many different autoantibodies have been identified but the role of autoantibodies in the pathogenesis of SLE is as yet uncertain.

25 **[0004]** Detection of autoantibodies in the serum of patients assists in the diagnosis of autoimmune diseases. Rheumatoid factor (IgM antibodies directed against human IgG) is detected in the majority of patients with RA and supports that diagnosis in a given individual [Kelly, W.N., et al. 1985. *Textbook of Rheumatology*. 2nd ed. Saunders. pp. 667]. Antinuclear antibodies (ANA) are present in approximately 98% of individuals with active SLE. Although ANA are not specific for the diagnosis of SLE, the absence of these antibodies argues against the diagnosis of SLE in a given patient [Kelly et al., 1985 supra pp. 691].

30 **[0005]** Liver and biliary diseases collectively rank in the top ten causes of mortality in the U.S. Chronic liver diseases affect between 5 and 10 percent of Americans and cause 1 to 2 percent of deaths in the United States. Chronic liver disease and cirrhosis cost an estimated \$1.6 billion per year [(2004)]. General causes of liver and biliary diseases include infectious agents, inherited defects, metabolic disturbances, alcohol, toxins and environmental toxicants. The most common liver diseases are chronic hepatitis C, alcohol liver disease, nonalcoholic fatty liver disease, chronic hepatitis B, autoimmune liver diseases and drug-induced liver diseases. Many of these conditions can be prevented or treated, but if not, they can lead to progressive liver injury, liver fibrosis and ultimately cirrhosis, portal hypertension, end-stage liver disease and, in some instances, liver cancer. Currently, the only therapy for end-stage liver disease is liver transplantation. More than 5,000 liver transplants are done in the U.S. each year. At least 17,000 persons are on a waiting list for liver transplantation and as many as 1,500 die yearly while waiting [(2004)]. Liver disease research presents many challenging needs. Autoimmune liver diseases include primary biliary cirrhosis (PBC), autoimmune hepatitis and primary sclerosing cholangitis. These chronic liver diseases can all lead to end-stage liver disease. Collectively, autoimmune liver diseases are responsible for 13% of adult liver transplants per year in the U.S. [(2004)].

40 **[0006]** PBC is a progressive cholestatic liver disease, with an estimated prevalence in the U.S. of approximately 40 adults per 100,000 population (incidence 2.7 per 100,000 U.S. population) [Kim, Lindor et al. (2000) *Gastroenterology* 119: 1631-6; Feld and Heathcote (2003) *J Gastroenterol Hepatol* 18: 1118-28; 2004]. Women between the ages of 40 and 65 are predominantly affected by PBC, with a female to male ratio of 9:1 [Kaplan and Gershwin (2005) *N Engl J Med* 353: 1261-73], as is typical for autoimmune disease. PBC is characterized by the gradual progressive destruction of intrahepatic biliary ductules leading to hepatic fibrosis and liver failure (reviewed in [Kaplan (1996) *N Engl J Med* 335: 1570-80; Heathcote (2000) *Hepatology* 31: 1005-13; Kaplan (2002) *Gastroenterology* 123: 1392-4; Talwalkar and Lindor (2003) *Lancet* 362: 53-61]). PBC is a significant indication for liver transplantation, and PBC patients constitute 11% of

all patients undergoing liver transplantation for cirrhosis [Milkiewicz (2008) *Clin Liver Dis* 12: 461-72; xi].

[0007] Treatment of PBC is accomplished with ursodeoxycholic acid (ursodiol), a natural bile acid that is not toxic to the liver, to replace the bile acids which are reduced by PBC. While the mechanisms are not fully understood, this treatment ultimately reduces intracellular build up of other liver-toxic bile acids (which was caused by bile duct destruction).

Although ursodiol slows progression to cirrhosis, ursodiol treatment functions best when implemented early in the course of PBC, highlighting the importance of a rapid, reliable PBC diagnostic test. In fact, a study showed that ursodiol treatment at stages III and IV did not result in significant slowing of liver progression while patients treated early at histological stages I and II did show significant slowing of liver destruction with ursodiol treatment. This highlights the need for an early PBC diagnostic, to allow prompt medical treatment [Heathcote (2000) *Hepatology* 31: 1 005-13; Poupon, Lindor, Pares, Chazouilleres, Poupon and Heathcote (2003) *J Hepatol* 39: 12-6].

[0008] Roughly half of PBC patients first present with an abnormal blood test which triggers the eventual PBC diagnosis. Generally, diagnostic testing is initially activated by abnormal liver function tests and signs of bile disease, followed by testing for serum anti-mitochondrial autoantibodies (AMA), for which an estimated 87-95% of PBC patients test positive [Heathcote (2000) *Hepatology* 31: 1 005-13; Yang, Yu, Nakajima, Neuberger, Lindor and Bloch (2004) *Clin Gastroenterol Hepatol* 2: 1116-22; Kaplan and Gershwin (2005) *N Engl J Med* 353: 1261-73; Liu, Shi, Zhang, Zhang and Gao (2008) *Liver Int* 28:233-9]. Bile duct imaging tests are used to rule out other causes of biliary tract disease, and liver biopsies confirm diagnosis and provide a gauge of disease stage (based upon the degree of fibrosis).

[0009] However, the other roughly half of PBC patients will present only with a variety of relatively non-specific physical symptoms, highlighting the difficulties facing the general practitioner or specialist responsible for diagnosis. The most common of such symptoms are pruritis, fatigue and musculoskeletal pain [Prince, Chetwynd, Newman, Metcalf and James (2002) *Gastroenterology* 123: 1044-51]. Furthermore, numerous autoimmune disorders may be found in association with PBC, including autoimmune hepatitis(AIH) [Czaja (2006) *J Hepatol* 44: 251-2], thyroid dysfunction, sicca symptoms, Raynaud's syndrome, systemic lupus erythematosus (SLE) and rheumatoid arthritis [Heathcote (2000) *Hepatology* 31: 1005-13; Gershwin, Selmi, Worman, Gold, Watnik, Utts, Lindor, Kaplan and Vierling (2005) *Hepatology* 42: 1194-202]. In one study, 19% of PBC patients were found to have features of another disease [Czaja (1998) *Hepatology* 28: 360-5], thereby clouding diagnosis. Of concern, the proper testing may not be ordered in many patients due to unrecognized etiology, especially when patients present with vague symptoms of pruritis or joint discomfort.

[0010] Autoantibodies have the potential to serve not only as diagnostic tools, but also as harbingers of the future development of PBC. In fact, anti-mitochondrial autoantibodies (AMA) have been shown to pre-date clinical manifestations and diagnosis of PBC [Metcalf, Mitchison, Palmer, Jones, Bassendine and James (1996) *Lancet* 348: 1399-402]. This demonstrates that it may be possible to diagnose PBC at an earlier stage using autoantibody biomarkers. The serological hallmark of PBC are AMA, which can be detected in 87-95% of patients [Kaplan (1996) *N Engl J Med* 335: 1570-80; Nishio, Keeffe and Gershwin (2002) *Semin Liver Dis* 22: 291-302]. The major autoantigens targeted by these AMA include the E2 subunits of the pyruvate dehydrogenase complex (PDC-E2); the branched/chain 2-oxo-acid dehydrogenase complex (BCOADC-E2) and the the 2-oxo-glutarate dehydrogenase complex (OGDC-E2) [Fussey, Guest, James, Bassendine and Yeaman (1988) *Proc Natl Acad Sci U S A* 85: 8654-8; Nishio, Keeffe et al. (2002) *Semin Liver Dis* 22: 291-302].

[0011] Anti-nuclear autoantibodies (ANA) are present in ~50% of PBC patients. Autoantibodies recognizing proteins of the nuclear core complex and multiple nuclear dots (MND) are useful PBC markers in AMA-negative patients, with a prevalence of 13-44% [Manuel Lucena, Montes Cano, Luis Caro, Respaldiza, Alvarez, Sanchez-Roman, Nunez-Roldan and Wichmann (2007) *Ann N Y Acad Sci* 1109: 203-11]. Additionally, ANA can serve as prognostic indicators, with anti-centromere and/or anti-nuclear pore glycoprotein 210 (gp210) autoantibodies being associated with liver failure in PBC [Yang, Yu et al. (2004) *Clin Gastroenterol Hepatol* 2: 1116-22; Nakamura, Kondo et al. (2007) *Hepatology* 45: 118-27].

[0012] The nuclear body(NB, also known as nuclear domain 10, PML oncogenic domain, and Kr body) is a nuclear organelle whose function is unknown [Ascoli, C. A., and Maul, G. G., *J. Cell. Biol.* 112:785-795 (1991); Brasch, K., and Ochs, R. L., *Exp. Cell Res.* 202:211-223 (1992); Dyck, J. A. et al., *Cell* 76:333-343 (1994)]. Using immunohistochemical staining, NBs appear as 5 to 30 discrete, punctate, dot-like regions within the nucleus. The NB is distinct from other nuclear domains including those involved in DNA replication and mRNA processing. In addition, components of the NB do not co-localize with kinetochores or centromeres [Brasch, K., and Ochs, R. L., *Exp. Cell Res.* 202:211-223 (1992)]. The number of NBs in the cell, and the intensity of antibody staining of these structures, increase in response to stimuli including interferons (IFNs), heat shock and viral infection [Ascoli, C. A., and Maul, G. G., *J. Cell. Biol.* 112:785-795 (1991)].

[0013] The NB is a target of autoantibodies in the serum of patients with the autoimmune disease primary biliary cirrhosis (PBC). Approximately 40% of patients with PBC have antibodies directed against this structure [Evans, J., et al., *Arthr. Rheum.* 347:31-736 (1991); Szosteki, C. et al., *Scand. J. Immunol.* 36:555-564 (1992)]. Serum from patients with PBC was used to identify and characterize a 100-kDa component of the NB which was designated Sp100 (Speckled, 100 kDa) [Szosteki, C. et al., *J. Immunol.* 145:4338-4347 (1990)]. The fusion of Sp100 to the LexA DNA binding domain has been shown to activate gene transcription in *Saccharomyces cerevisiae*, and it has been suggested that Sp100 may participate in activation of transcription of specific regions in the genome [Xie, K. et al., *Mol. Cell. Biol.* 13:6170-6179

(1993)].

[0014] A second component of the NB, designated NDP52, was characterized using a murine monoclonal antibody that reacted with the NB [Korioth, F., et al., *J. Cell Biol.* 130:1-13 (1995)]. A cDNA encoding NDP52 was identified and the predicted amino acid sequence contained coiled coil, leucine zipper and zinc finger motifs. One or more of these domains may be involved in interactions between NDP52 and other components of the NB [Korioth, F., et al., *J. Cell Biol.* 130:1-13 (1995)].

[0015] A third component of the NB, PML, was identified by several investigators studying the t(15;17) translocation associated with human acute promyelocytic leukemia (APL) [de The, H. et al., *Nature (London)* 347:558-561 (1990); Borrow, J. et al., *Science* 249:1577-1580 (1990); Longo, L. et al., *J. Exp. Med.* 172:1571-1575 (1990); Kakizuka, A. et al., *Cell* 66:663-674 (1991)]. In this translocation, the amino terminal portion of PML is fused to retinoic acid receptor alpha. PML was found to co-localize with Sp100 in the NB [Weis, K. et al., *Cell* 76:345-356 (1994); Koken, M. H. M. et al., *EMBO* 13:1073-1083 (1994)]. Expression of the PML-alpha fusion protein in APL cells appears to disrupt the NB; in these cells, the NB antigens are detected in numerous smaller regions in the nucleus described as "microspeckles." Treatment of APL cells with retinoic acid (RA) results in differentiation of myeloid precursor cells and reformation of NBs [Dyck, J. A. et al., *Cell* 76:333-343 (1994); Weis, K. et al., *Cell* 76:345-356 (1994); Koken, M. H. M. et al., *EMBO* 13:1073-1083 (1994)]. In patients with APL, treatment with RA results in differentiation of leukemic cells and temporary disease remission [Warrell, R. P. et al., *N. Eng. J. Med.* 329:177-189 (1993)].

[0016] It is important to note however, that ANA are also found in a variety of other prevalent autoimmune disorders and a wide range of cancers [Bei, Masuelli, Palumbo, Modesti and Modesti (2008) *Cancer Lett.*]

[0017] Indirect immunofluorescence (IIF) and solid-phase immunoassay are the two formats used to establish the presence or absence of autoantibodies in patients. Both methods have their pros and cons as discussed below:

[0018] For the past several decades, indirect immunofluorescence (IIF) has been the method of choice by physicians for the detection of autoantibodies present in the serum of autoimmune patients. Importantly, it remains the gold standard for AMA and ANA testing, including for PBC. Typically, patient serum is serially diluted in two-fold increments and allowed to bind to a cell substrate on a microscope slide (e.g. HEp-2 liver cells), which is then fluorescently stained to detect bound autoantibodies and examined under the microscope by a trained technician to identify the cellular/tissue staining patterns. IIF does have the advantage that as a cell/tissue based substrate, it can in theory "universally" cover all cellular autoantigens (pending their expression and preservation in the substrate). This, in part, is evidenced by the high diagnostic sensitivity of the IIF test, e.g. 93% (ANA) for systemic lupus erythematosus (SLE) [Solomon, Kavanaugh and Schur (2002) *Arthritis Rheum* 47: 434-44] and 90% (AMA) for PBC [Tanaka, Miyakawa, Luketic, Kaplan, Storch and Gershwin (2002) *Cell Mol Biol (Noisy-le-grand)* 48: 295-9].

[0019] Although IIF based AMA is a sensitive marker for PBC, the tradeoff may be specificity. Asymptomatic patients have been deemed AMA positive, and while a large portion only develop symptoms years later, some never develop symptoms at all [Metcalf, Mitchison et al. (1996) *Lancet* 348: 1399-402]. Moreover, one study found that 34% of AIH patients tested positive for AMA [Nezu, Tanaka, Yasui, Imamura, Nakajima, Ishida and Takahashi (2006) *J Gastroenterol Hepatol* 21: 1448-54].

[0020] Furthermore, the IIF assay is problematic overall when used as a routine diagnostic screening tool, as it is difficult to standardize owing to variations in the substrate and fixation process, variations in the microscopy apparatus, and due to the highly subjective interpretation of results [Jaskowski, Schroder, Martins, Mouritsen, Litwin and Hill (1996) *Am J Clin Pathol* 105: 468-73]. The consensus statement in 2004 from the Committee for Autoimmune Serology of the International Autoimmune Hepatitis Group (IAIHG) recommended that IIF be performed on three different organs from rodents [Vergani, Alvarez, Bianchi, Cancado, Mackay, Manns, Nishioka and Penner (2004) *J Hepatol* 41: 677-83]. Both AMA and anti-liver kidney microsomal-1 (LKM1) antibodies stain the renal tubules of the kidney, with differences only apparent to the trained eye, and this confusion can lead to a diagnosis of autoimmune hepatitis (AIH) instead of PBC [Bogdanos, Invernizzi, Mackay and Vergani (2008) *World J Gastroenterol* 14: 3374-87]. Moreover, some autoantigens are lost (unrecognizable) by diffusion or denaturation during the fixation process of IIF. Another confounding factor is that multiple autoimmune diseases can often occur together in the same patient, and the overlapping IIF patterns can lead to confusion in the correct diagnosis of each [Assassi, Fritzler et al. (2009) *J Rheumatol*; Norman, Bialek, Encabo, Butkiewicz, Wiechowska-Kozłowska, Brzoska, Shums and Milkiewicz (2009) *Dig Liver Dis* 41: 762-4]. Finally, IIF is slow, laborious and not amenable to high-throughput automation [Ulvestad, Kanestrom, Madland, Thomassen, Raga and Vollset (2000) *Scand J Immunol* 52: 309-15].

[0021] Although IIF remains the gold standard in AMA testing, solid-phase immunoassays, such as ELISA (Enzyme Linked Immunosorbent Assay), are gaining popularity, especially in high-throughput laboratories [Fritzler and Fritzler (2006) *Curr Med Chem* 13: 2503-12]. These methods have the advantage of high throughput automation, high analytical sensitivity, purely objective scoring, reliability, and the ability to test for specific autoantigen species, including in a multiplexed fashion [Fritzler and Fritzler (2006) *Curr Med Chem* 13: 2503-12]. With a resolution at the individual antigen level, these methods have the potential for greater disease specificity, if the correct marker panel is chosen. The drawback, however, is that a sufficient number of autoantigens needs to be both discovered and clinically validated to match the

diagnostic sensitivity of the cellular substrate based IIF assay.

[0022] In one example of a commercial solid-phase immunoassay for PBC, INOVA Diagnostics Inc. (San Diego, CA) markets the MIT3 assay, an FDA-approved ELISA-based immunoassay for PBC based on the detection of AMAs. The MIT3 utilizes a recombinant protein containing the immunodominant epitopes of all three E2 subunits of the pyruvate dehydrogenase complex [Moteki, Leung, Cappel, Dickson, Kaplan, Munoz and Gershwin (1996) *Hepatology* 24: 97-103]. The overall goal of these tests is to mimic the cellular IIF-based AMA test for PBC, but with all the aforementioned benefits of solid-phase immunoassays of individual antigens. Still, this test is only meant to be diagnostic aid, together with clinicopathological findings for PBC. In one study, the AMA-based MIT3 ELISA assay had a reported a diagnostic sensitivity of 81.6%, however, it is important to note that serum samples with AMA-negative PBC disease were excluded [Gabeta, Norman, Liaskos, Papamichalis, Zografos, Garagounis, Rigopoulou and Dalekos (2007) *J Clin Immunol* 27: 378-87]. In another study, it was shown that the MIT3 assay, for instance, lacks all the necessary mitochondrial autoantigens for maximum diagnostic sensitivity of PBC [Dahnrich, Pares et al. (2009) *Clin Chem* 55: 978-85].

[0023] This highlights the need for the discovery and validation of additional autoantigen biomarkers to be used in solid-phase immunoassays for the optimal diagnosis of autoimmune diseases such as PBC. The most effective methods for the discovery of autoantigens are proteomics based. Proteomics can be defined as the global (e.g. parallel or simultaneous) analysis of the entire expressed protein complement of the genome [Wasinger, Cordwell et al. (1995) *Electrophoresis* 16: 1090-4]. Proteomics methods allow for the discovery of novel autoantigens in an unbiased fashion. Common proteomics methods for discovery of novel autoantigens include SEREX (serological identification of antigens by recombinant expression cloning) [Krebs, Kurrer, Sahin, Tureci and Ludewig (2003) *Autoimmun Rev* 2: 339-45] and human proteome microarrays ("chips", commonly the dimensions of standard microscope slides, containing thousands of purified recombinant human proteins printed to their surface in an ordered array of microscopic spots, e.g. spots of 100 micron in diameter) [Robinson, DiGennaro et al. (2002) *Nat Med* 8: 295-301; Robinson, Steinman and Utz (2002) *Arthritis Rheum* 46: 885-93].

SUMMARY OF THE INVENTION

[0024] The present invention relates to methods of using the novel autoantigens (Tables I and V) human hexokinase 1 (HK1) and/or kelch-like 12 (KLHL12), or fragments thereof comprising an epitope, in the diagnostic, prognostic, staging and therapeutic regimens of the autoimmune liver disease Primary Biliary Cirrhosis (PBC). The present invention also relates to methods of using homologs, family members, transcript variants and isoforms (e.g. Table VI), preferably at least 70% identical, more preferably at least 90% identical and most preferably at least 95% identical, of human hexokinase 1 (HK1) and/or kelch-like 12 (KLHL12), or fragments thereof comprising an epitope, in the diagnostic, prognostic, staging and therapeutic regimens of the autoimmune liver disease Primary Biliary Cirrhosis (PBC).

[0025] The present invention further provides isolated antibodies that bind specifically to the above-described polypeptides, or fragments thereof comprising an epitope. Antibodies provided herein may be polyclonal or monoclonal, may be affinity purified, may be immobilized onto a solid support, and may be detectably labeled. The invention also provides methods for detecting the presence of an autoimmune disease in an animal, preferably a human, comprising the steps of isolating a body fluid sample, preferably blood, serum or plasma, from the animal, incubating the serum with an isolated HK1 and/or KLHL12 polypeptide described above, and detecting the binding of autoantibodies in the serum sample to the isolated polypeptide. The invention also provides alternative methods for detecting the presence of an autoimmune disease in an animal comprising the steps of isolating a body fluid sample from the animal, preferably blood, serum or plasma, and immobilizing components of the serum on a solid support, contacting the immobilized serum components with an isolated polypeptide described above under conditions favoring the formation of a complex between the serum components and isolated polypeptide, contacting the formed complex with an antibody that binds specifically to HK1 and/or KLHL12, and detecting the binding of the antibody to the complex. Autoimmune diseases that may be diagnosed by the methods of the present invention include primary biliary cirrhosis (PBC) and systemic lupus erythematosus (SLE). Cancers that may be diagnosed by the methods of the present invention include colorectal cancer (CRC). The present invention also provides methods of determining prognosis, disease stage and treatment regimens using the aforementioned methods of detecting autoantibodies against HK1 and/or KLHL12.

[0026] In a preferred embodiment, heterogeneous or homogenous immunoassays, singleplex or multiplex, are used to detect autoantibodies present in body fluids directed against said autoantigens. Other preferred embodiments of the present invention will be apparent to one of ordinary skill in light of the following drawings (Figures) and description of the invention, and of the claims.

Experimental

Example 1: Proteome Microarray Based Discovery of Novel Primary Biliary Cirrhosis (PBC) Autoantigens

5 *Serum Screening on Microarrays*

[0027] Patient sera were screened against commercial human proteome microarrays comprised of ~8,000 unique human recombinant (eukaryotically expressed) proteins printed in duplicate at high density to a "chip" the size of a standard microscope slide (Human ProtoArray® v4.0, Invitrogen, Carlsbad, CA) [Sheridan (2005) Nat Biotechnol 23: 3-4]. Microarrays were performed according to the manufacturer's instructions. Microarrays were imaged on an Array-WoRx^e BioChip fluorescence reader (Applied Precision, LLC, Issaquah, Washington) using the appropriate standard built-in filter sets. Image analysis and data acquisition was performed using the GenePix Pro v6.1 software package (Molecular Devices, Sunnyvale, CA) according to the instructions of the microarray manufacturer (Human ProtoArray® v4.0, Invitrogen, Carlsbad; CA).

15 [0028] 92 different serum samples from normal individuals and patients with various diseases were individually screened against the proteome microarrays in order to detect the presence of autoantibodies against the arrayed proteins (potential autoantigens). For this, 2 different lots of microarrays were used in 2 sequential studies. The composition of the entire patient population was as follows: Microarray Lot #1 (80 unique samples) - 18 Primary Biliary Cirrhosis (PBC) patients versus 62 non-PBC control samples [13 normal, 25 colorectal cancer (CRC), 22 systemic lupus erythematosus (SLE), 2 Sjögrens syndrome (SjS)]. Microarray Lot# 2 (12 unique samples) - 3 more PBC and 9 non-PBC controls
20 [4 normal and 5 autoimmune hepatitis (AIH)]. The normal sera were approximately age and gender matched to the PBC cohort. The AIH sera were used because it is an autoimmune liver disease different from PBC yet known to be associated with autoantibodies. The CRC sera were used because cancer patients are also known to have various autoantibodies against so-called tumor associated autoantigens (TAA), including a common repertoire of nuclear autoantibodies observed in both cancers and autoimmune disease [Bei, Masuelli, Palumbo, Modesti and Modesti (2008) Cancer Lett].
25 Archived sera were obtained from the repositories of the following sources: Our collaborator, Dr. Donald Bloch, M.D., Center for Immunology and Inflammatory Diseases, Massachusetts General Hospital, Assistant Professor of Medicine, Harvard Medical School provided 12 of the SLE sera as well as the SjS and PBC sera. Remaining SLE sera and all the AIH sera were from Bioreclamation Inc. (Hicksville, NY), normal sera were from ProMedDx, LLC (Norton, MA) and CRC sera were from Asterand Inc. (Detroit, MI).
30

Biostatistical Analyses of Microarrays Data

35 [0029] In order to identify the autoantigen biomarkers from the microarray data, the biostatistical methods used were the standard approaches provided by the microarray manufacturer in the form of the ProtoArray® Prospector v4.0 software package (Invitrogen, Carlsbad, CA) using the Immune Response Profiling (IRP) add-on [Hudson, Pozdnyakova, Haines, Mor and Snyder (2007) Proc Natl Acad Sci USA 104: 17494-9]. Two of the biostatistical methods from this software package were used to create two corresponding PBC autoantigen lists as follows:

40 [0030] "Hit Calling" Autoantigen List: To convert the data to binary format, proteins (i.e. potential autoantigens) on each microarray (1 serum/microarray) were scored as a "hit" (i.e. positive) or not a hit (i.e. negative). Autoantigen hits were called on a per microarray basis using the Z-score with a cutoff threshold of 3 standard deviations above the microarray mean. The number of hits in the PBC and control groups for each autoantigen were used to determine the percent prevalence of each autoantigen. Autoantigens ultimately placed on this list had to have greater percent prevalence in the PBC cohort than the control cohort (i.e. all non-PBC samples).

45 [0031] M-Statistics Autoantigen List: This approach uses quantile normalized microarray data and performs a pairwise t-test for each protein between the two patient groups (i.e. PBC group and the control group corresponding to all non-PBC patients). This algorithm also estimates the autoantigen prevalence based on cutoffs set by the quantile normalized data. Autoantigens ultimately placed on this list had to have greater percent prevalence in the PBC cohort than the control cohort (i.e. all non-PBC samples) and had to have M-Statistics p-values of <0.1.

50 [0032] Microarray Lots # 1 and 2 were analyzed separately. To comprise a single final list of microarray-derived PBC autoantigens, those observed as overlapping on both aforementioned biostatistical lists for Microarray Lot #1 (only) were taken. Next, any markers on this compiled list that were positive in any of the AIH patients (Microarray Lot # 2), as determined by the "Hit Calling" method, were eliminated. Finally, the list was then prioritized based on the M-Statistics p-value as well as diagnostic sensitivity and specificity.

55 *Results:*

[0033] Two of the PBC autoantigen markers, human Hexokinase 1 (HK1) and human Kelch-Like 12 (KLHL12), identified

from the proteome microarrays and claimed in this patent, are listed in **Table I**, along with their M-Statistics p-values as well as their diagnostic sensitivities and specificities (calculated from Microarray Lot #1). Quantile normalized microarray data (normalized autoantibody signal intensity) for all 92 samples (i.e. all 92 microarrays) are shown in **Figure 16** and **Figure 17** for HK1 and KLHL12 respectively. In summary (**Table I**), the presence of serum autoantibodies against either autoantigen is strongly correlated with the PBC cohort, showing highly significant p-values (1×10^{-10} and 8×10^{-5} for HK1 and KLHL12 respectively) as well as sensitivities of 85-89% and 33-40% for HK1 and KLHL12 respectively, and, specificities 84-90% and 97-98% for HK1 and KLHL12 respectively (see **Table I** for details). By definition (see "Biostatistical Analysis of Microarray Data" above in this Example), none of the 5 Autoimmune Hepatitis (AIH) sera were positive for HK1 or KLHL12 (see also **Figure 16** and **Figure 17**; Microarray Lot #2). The HK1 and KLHL12 autoantigen biomarkers were also the subject of further validation as detailed in other experimental Examples.

[0034] It should also be noted that HK1 autoantibodies are also observed with low prevalence in systemic lupus erythematosus (SLE) and colorectal cancer (CRC) (**Figure 16**). N-03 is the only "normal" serum sample to be positive for HK1 (**Figure 16**; red bar). N-03 is also the only "normal" serum sample to be positive for KLHL12 (**Figure 17**; red bar). Thus, in fact, it is believed that N-03 may in fact have yet undiagnosed or unreported/undocumented PBC (note that autoantibodies have been shown to pre-date clinical symptoms/manifestations of autoimmune disease, including in PBC).

Example 2: Pre-Validation of Novel Primary Biliary Cirrhosis (PBC) Autoantigens HK1 and KLHL12 Using an ELISA

[0035] It should be noted that the ELISA assay described here in this Example and used in many subsequent Examples is termed T²-ELISA, and is based on the use of dual-epitope tagged cell-free expressed protein antigens. In this Example, those antigens are HK1 and KLHL12 and the T²-ELISA used as a tool for clinical pre-validation (and eventually validation in later Examples) of these microarray-derived novel autoantigens.

Autoantigens Expression

[0036] The entire Open Reading Frames (ORFs) of human HK1 and KLHL12 were cloned, using standard and accepted molecular biology practices, into a plasmid vector compatible with cell-free protein expression, containing the T7 RNA polymerase promoter, a Kozak (ribosome binding) sequence, a start codon, an N-terminal VSV-G epitope tag (YTDIEM-NRLGK(SEQ ID NO: 19)), and a C-terminal HSV epitope tag (QPELAPEDPED(SEQ ID NO: 20)) in addition to the ORF insert. As source DNA for cloning into the expression vector, full-length sequence-verified clones were purchased from OpenBiosystems (Huntsville, AL) [catalog OHS1770-9381021 (UniGene Hs.370365) for HK1 and MHS1011-61211 (UniGene Hs.706793) for KLHL12]. Expression vectors were verified for the correct ORF insert using standard EcoRI digestion methods and/or DNA sequencing.

[0037] Autoantigens were produced from the aforementioned plasmid clones by cell-free protein expression. Cell-free protein expression reactions were performed using a transcription/translation coupled rabbit reticulocyte lysate system (TNT[®] T7 Quick for PCR DNA; Promega, Madison, WI) according to the manufacturer's instructions. Autoantigen expression reactions contained the cognate plasmid DNA while blank expression reactions lacked only the plasmid DNA. Expression reactions were stopped by diluting 1/20 in TDB [1% BSA (w/v) and 0.1% (v/v) Triton X-100 in TBS-T (50 mM Tris, pH 7.5, 200 mM NaCl, 0.05% (v/v) Tween-20)].

Dual-Tag Enzyme-Linked Immunosorbent Assay (T²-ELISA) of Autoantigens

[0038] Nunc Brand 96-well Polysorp™ Microwell™ white opaque, flat bottom, untreated polystyrene microtiter plates (Nunc Brand from Thermo-Fisher Scientific, Rochester, NY) were used for a sandwich type Enzyme-Linked Immunosorbent Assay (ELISA). Plates were coated with 0.5 μg/mL of a mouse monoclonal anti-HSV[®] tag capture antibody (EMD Biosciences, Inc., San Diego, CA) in sodium carbonate/bicarbonate pH 9.3 for 30 min with shaking (50 μL/well). Plates were then washed 6x in TBS-T (wells filled to maximum) on an ELx405 Select Robotic Plate Washer (BioTek, Winoski, VT). All plate washes were performed in this manner unless noted otherwise. Plates were then blocked for 30 min at 300 μL/well in 1% BSA (w/v) in TBS-T. The solution was removed from the plates and the aforementioned stopped (i.e. diluted) cell-free expression reactions (autoantigen and blank reactions) were then added at 100 μL/well and shaken for 30 min. Plates were washed and serum samples (diluted at 1/1,000 in 1% BSA (w/v) in TBS-T) were added at 100 μL/well and shaken for 30 min. Each serum sample was run against triplicate wells of autoantigen and triplicate wells of the cell-free expression blank. Additionally, one set of triplicate wells of autoantigen and one set of triplicate wells of the cell-free expression blank were designated for VSV-G epitope tag detection, and therefore received plain 1% BSA (w/v) in TBS-T instead of diluted serum. To avoid contamination of the robotic plate washer with human serum, plates were subsequently washed 4x by manual addition of TBS-T (wells filled to maximum) followed by vacuum aspiration and then washed 6x in the robotic plate washer as described earlier in this Example. Wells designated for

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detection of the VSV-G epitope tag then received an anti-VSV-G horseradish peroxidase (HRP) labeled monoclonal antibody (Clone P5D4, Roche Applied Science, Indianapolis, IN) diluted 1/20,000 in 1% BSA/TBS-T. Wells designated for detection of serum autoantibody received a mouse anti-[human IgG] HRP labeled monoclonal secondary antibody (minimum cross-reactivity with mouse immunoglobulin; Jackson ImmunoResearch Laboratories, Inc, West Grove, PA) diluted 1/20,000 in 1% BSA/TBS-T. Plates were shaken for 30 min. The solutions were then manually dumped from the plates by inversion followed by vigorous patting of the plates inverted on a dry paper towel to remove residual fluid. Plates were then washed in the robotic plate washer as described earlier in this Example. Chemiluminescence signal was generated by the addition of 50 μ L/well of SuperSignal ELISA Pico Chemiluminescence Substrate (Pierce Brand from Thermo Fisher Scientific, Rockford, IL). Plates were developed by shaking for 15 min and then read on a LumiCount luminescence plate reader (1s exposure, PMT of 650V, gain 1) (Packard/PerkinElmer Life and Analytical Sciences, Inc., Boston, MA).

Results:

[0039] For this pre-validation of the new PBC autoantigen markers listed in **Table I**, randomly selected sera that were detected as positive or negative for a given autoantigen in the microarray analyses (see Example 1) were also analyzed here by T²-ELISA.

[0040] Calculation of Autoantibody Units from the T²-ELISA, in short, was achieved by background subtracting the data and normalizing to the detection of the common VSV-G epitope tag for each antigen on each assay (i.e. each plate). More specifically, for each serum-autoantigen pair, for each of the triplicate wells from the T²-ELISA data, Autoantibody Units were calculated as follows: **[autoantibody signal from one well (i.e. serum versus autoantigen)]** minus **[average background from triplicates (i.e. same serum versus average of all three blank expression wells)]** to yield triplicate Background Subtracted Values (BSV) for each serum-autoantigen pair. Note that one assay is defined as one 96-well microtiter ELISA plate. To normalize for inter-assay variances (day-to-day and assay-to-assay) for each autoantigen, wells on each assay, for each autoantigen on that assay, were dedicated solely for detection of the common VSV-G epitope tag. The VSV-G Normalization Factor (VNF) was calculated as follows: **[average VSV-G signal for triplicate wells (i.e. autoantigen wells probed with VSV-G antibody)]** minus **[average VSV-G background for triplicate wells (i.e. blank expression wells probed with VSV-G antibody)]**. On a per assay basis, the triplicate BSV for all serum-autoantigen pairs were then divided by the VNF for that assay and multiplied by 100, yielding triplicate Autoantibody Unit values for each serum-autoantigen pair (i.e. expressed as a percent of the VNF). Note that a floor of zero was set for the Autoantibody Units. The average and standard deviation (errors bars) were calculated and plotted in **Figures 1 and 2** for the new PBC autoantigens HK1 and KLHL12 respectively.

[0041] Sera were scored "analytically", as positive or negative in the T²-ELISA in order to check concordance with the microarrays. For this, both of the following criteria must have been met for each serum-autoantigen pair to have been scored as analytically positive in the T²-ELISA: i) a p-value ≤ 0.05 in a 1-tailed homoscedastic unpaired t-test on the raw T²-ELISA values from the triplicate wells of the autoantibody signal (i.e. serum versus autoantigen) compared to background (i.e. same serum versus blank expression wells); ii) autoantibody signal-to-background ratio ≥ 2 . In **Figures 1 and 2**, T²-ELISA scores and microarray ("Array") scores are denoted as positive (+) or negative (-). For HK1 (**Figure 1**), of 12 randomly selected sera that were positive by the microarray analyses, 10 were positive by ELISA for 83% concordance. Additionally for HK1 (**Figure 1**), 5 sera were randomly selected that were negative on the microarrays, all of which were also negative by T²-ELISA for a 100% concordance. For KLHL12, of the 7 negative and 4 positive sera randomly chosen from the microarray analyses (see Example 1), there was full 100% concordance with the T²-ELISA results as shown in **Figure 2**.

Example 3: Validation of Novel Primary Biliary Cirrhosis (PBC) Autoantigens HK1 and KLHL12 Using an ELISA on a New AMA-Positive PBC Patient Cohort Not-Previously Screened by Microarrays

Autoantigens Expression

[0042] As in Example 2.

Dual-Tag Enzyme-Linked Immunosorbent Assay (T²-ELISA) of Autoantigens

[0043] As in Example 2.

Results:

[0044] A critical validation of the newly discovered markers is to perform studies on a new patient cohort (22 PBC

samples), never before screened on the proteome microarrays. In this Example, this has been done with both of the new PBC autoantigens, HK1 and KLHL12 (previously listed in **Table I**).

[0045] The new PBC sera were obtained from our collaborator, Dr. Donald Bloch, M.D., Center for Immunology and Inflammatory Diseases, Massachusetts General Hospital, Assistant Professor of Medicine, Harvard Medical School and the normal sera were from ProMedDx, LLC (Norton, MA).

[0046] Calculation of Autoantibody Units from the T²-ELISA, in short, was achieved by background subtracting the data and normalizing to the positive control on each assay (i.e. each plate), whereby the positive control is set to 1,000 Autoantibody Units. More specifically, for each serum-autoantigen pair, for each of the triplicate wells from the T²-ELISA data, Autoantibody Units were calculated as follows: **[autoantibody signal from one well (i.e. serum versus autoantigen)] minus [average background from triplicates (i.e. same serum versus average of all three blank expression wells)]**. This yields triplicate Background Subtracted Values (BSV) for each serum-autoantigen pair. Note that one assay is defined as one 96-well microtiter ELISA plate. To normalize for inter-assay variances (day-to-day and assay-to-assay) for each autoantigen, a common positive control PBC serum for HK1 and KLHL12 was run on every assay (selected from the microarray PBC cohort in Example 1). The positive control T²-ELISA data were processed in the aforementioned manner on a per assay basis and the triplicate BSV averaged to yield the Positive Control Normalization Factor (PCNF) for each assay. On a per assay basis, the triplicate BSV for all serum-autoantigen pairs were then divided by the PCNF for that assay and multiplied by 1,000, yielding triplicate Autoantibody Unit values for each serum-autoantigen pair. Importantly, the VSV-G common epitope tag detection (Example 2) was still used to verify successful and consistent autoantigen expression, but was not used here in the calculation of Autoantibody Units.

[0047] In order to set diagnostic scoring thresholds for a given autoantigen, the T²-ELISA assay was run on a group of 22 normal patient sera and the cutoffs then set at 2 standard deviations above the mean for this normal cohort, for -95% statistical confidence. The use of this method at 2-3 standard deviations is common practice (e.g. [Liu, Wang, Li, Xu, Dai, Wang and Zhang (2009) Scand J Immunol 69: 57-63]). However, a critical requirement of this standard deviation based cutoff calculation method is that the data follows a Gaussian distribution, yet a Shapiro-Wilk test for normality determined this was not the case. As a solution, we log₂ transformed the Autoantibody Units and set the floor to 0 (i.e. non-transformed values of ≤0 were left as 0 without transformation) yielding a Gaussian distribution (of the >0 values) and allowing cutoffs to be set based on the aforementioned standard deviation methodology. Autoantibody Unit values of ≤0 were excluded from the cutoff calculations because background subtraction is used in the calculation of Autoantibody Units, meaning patient samples yielding ≤0 values would by definition have to be scored as autoantibody negative regardless (i.e. a cutoff is not needed nor relevant to ≤0 values).

[0048] As seen by the data in **Figure 3** for HK1, using a cutoff of 2.0, an 82% diagnostic sensitivity (100% specificity) on this new sample cohort is in good agreement with the microarray analyses performed on the original sample cohort (see Table I). As seen by the data in **Figure 4** for KLHL12, using a cutoff of 2.5, a 36% diagnostic sensitivity. (100% specificity) on this new sample cohort is in good agreement with the microarray analyses performed on the original sample cohort (see **Table I**).

Example 4: Validation of Novel Primary Biliary Cirrhosis (PBC) Autoantigens HK1 and KLHL12 Using an ELISA on a New Anti-Mitochondrial Antibody (AMA)-Negative PBC Patient Cohort Not Previously Screened by Microarrays

[0049] Patients with suspected PBC but an antimitochondrial antibody (AMA)-negative status make up approximately 5-20% of all PBC patients [Oertelt, Rieger et al. Hepatology 2007; 45:659-665], and AMA-negative PBC patients are particularly difficult to confirm diagnostically based on serotesting. Employing the known and validated autoantigens Sp 100 and gp210 only results in the detection of a fraction of the AMA-negative PBC patients (e.g. 17-33% in one recent study [Liu, Shi, Zhang, Zhang and Gao (2008) Liver Int 28: 233-9]), showing a need for specific autoantigens which can detect AMA-negative PBC patients.

[0050] To test the ability of our novel autoantigens, HK1 and KLHL12, to detect AMA-negative PBC patients, we utilized 17 patient sera which were AMA-negative by indirect immunofluorescence (IIF) but with confirmed PBC by conventional methods [Heathcote (2000) Hepatology 31: 1005-13], and by liver biopsy. The new AMA-negative PBC sera were obtained from our collaborator, Dr. Donald Bloch, M.D., Center for Immunology and Inflammatory Diseases, Massachusetts General Hospital, Assistant Professor of Medicine, Harvard Medical School. We compared the ability of our novel autoantigens, HK1 and KLHL12, with the available commercial tests to detect these patients with confirmed PBC but a known AMA-negative status.

Autoantigens Expression

[0051] As in Example 2.

Dual-Tag Enzyme-Linked Immunosorbent Assay (T²-ELISA) of Autoantigens

[0052] As in Example 2.

5 *FDA-Approved Commercial PBC ELISAs*

[0053] FDA-approved commercial ELISAs for PBC diagnostics were also run and were the Quanta Lite™ M2 EP (MIT3), Quanta Lite™ sp100, Quanta Lite™ gp210 and Quanta Lite™ PBC Screen IgG/IgA assays from INOVA Diagnostics (San Diego, CA); and were performed according to the manufacturer's instructions.

10

Results:

[0054] For scoring purposes, Autoantibody Unit calculations and diagnostic thresholds established in Example 3 were once again employed here for each autoantigen (HK1 and KLHL12).

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[0055] As illustrated by the data in **Figure 5** for HK1, 4 out of 17 AMA-negative PBC sera tested positive for this autoantigen (24% sensitivity). As seen by the data in **Figure 6** for KLHL12, 6 of the 17 AMA-negative PBC sera tested diagnostically positive (35% sensitivity).

20

[0056] We also tested the aforementioned 17 AMA negative PBC sera on all four of INOVA Diagnostics' commercially available FDA-approved PBC tests, namely, Quanta. Lite™ M2 EP (MIT3), Quanta Lite™ sp100, Quanta Lite™ gp210 and Quanta Lite™ PBC Screen IgG/IgA ELISA. The results of these tests, as well as our T²-ELISA results with HK1 and KLHL12, are summarized in **Table II**. INOVA's tests were unable to detect 3 of the 17 patients (18%). Strikingly however, HK1 and KLHL12 were each able to detect one of the previously undetectable AMA-negative PBC sera (PB-AMN-044 and PB-AMN-263 respectively). The third patient (PB-AMN-084) remained undetected by the aforementioned autoantigens but was detected by Sp140 (see Example 6 for details). These results are summarized in **Figure 7** as a Venn Diagram, illustrating overlap (or lack thereof) between the various biomarkers. Note that the results of the Quanta Lite™ PBC Screen IgG/IgA ELISA are not shown in the Venn Diagram (**Figure 7**), however, as seen in **Table II**, this assay did not increase detection as compared to the other INOVA assays. Together, these findings indicate that our two novel autoantigens, HK1 and KLHL12, are diagnostically very significant. It suggests that adding our novel biomarkers to the existing panel of PBC biomarkers could result in. the improved detection, and therefore earlier treatment and improved outcome of PBC patients, in particular for AMA-negative PBC patients.

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Example 5: Assessing HK1 and KLHL12 in Patients with Atypical Indirect Immunofluorescent (IIF) Staining

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[0057] We propose that the number of PBC patients may be higher than previously suspected, due to the extreme difficulty in drawing a conclusive diagnosis of PBC in the. absence of definitive AMA staining or the proper anti-nuclear autoantibody (ANA) staining pattern as determined by indirect immunofluorescence (IIF). To test this theory, we examined sera from undiagnosed patients with diffuse cytoplasmic or nuclear membrane IIF staining patterns. These new patient sera were obtained from our collaborator, Dr. Donald Bloch, M.D., Center for Immunology and Inflammatory Diseases, Massachusetts General Hospital, Assistant Professor of Medicine, Harvard Medical School.

40

Autoantigens Expression

[0058] As in Example 2.

45

Dual-Tag Enzyme-Linked Immunosorbent Assay (T²-ELISA) of Autoantigens

[0059] As in Example 2.

50

Quanta Lite™ M2 EP (MIT3) ELISA

[0060] Assay was performed according to manufacturer's instructions (INOVA Diagnostics, San Diego, CA).

Results:

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[0061] We ran HK1, KLHL12 and the M2 EP (MIT3) Quanta Lite™ Assay (INOVA Diagnostics, San Diego, CA) on 20 patients, the results of which are shown in **Figure 8**. Serum samples prefixed with "Cyto" or "NM" are from patients with diffuse cytoplasmic or nuclear membrane IIF staining, respectively. Calculation of Autoantibody Units for the T²-ELISA as run on HK1 and KLHL12 was done as in Example 2. Scoring for the T²-ELISA assay was done according to the

"analytical" method described in Example 2 (note that any serum sample with a graphed bar in **Figure 8** is positive). To avoid scale effects, graphed data for each antigen in **Figure 8** is normalized as a percent of the patient having the maximum autoantibody units for that antigen (that patient is marked with a blue arrow for each antigen). We set the Y-axis to INOVA's MIT3 cut-off of 25 units (based on the low positive control; cutoff determined per manufacturer's instructions), which corresponded to 17%, so all bars shown represent positive results.

[0062] One patient is detected by all three markers. Novel autoantigen KLHL12 detects two nuclear membrane patients that no other markers detect. Finally, MIT3 detects one nuclear membrane and several cytoplasmic patients that no other marker detects. These results strongly suggest that detection of the HK1, KLHL12 and MIT3 antigens may be useful in revealing a large number of previously undiagnosed patients suffering from PBC, but with atypical IIF staining.

Example 6: Improved Diagnostic Sensitivity by ELISA for Primary Biliary Cirrhosis (PBC) by Detection of Sp140

[0063] Antinuclear antibodies reacting with 5-20 nuclear dots are detected in 20-30% of patients with primary biliary cirrhosis (PBC). The "multiple nuclear dot" (MND) staining pattern produced by these antibodies is directed against promyelocytic leukemia protein nuclear body (PML NB) components, one of which was recently identified as Sp140. Sp140 has been reported to be present in 13% of PBC patients, with a larger proportion of AMA-negative compared with AMA positive PBC patients (53% versus 8%) [Granito, A. Yang, W. et. al, 2009, Am J Gastroenterol, In Press]. We therefore tested Sp140 in our T²-ELISA.

[0064] The PBC patient sera were obtained from our collaborator, Dr. Donald Bloch, M.D., Center for Immunology and Inflammatory Diseases, Massachusetts General Hospital, Assistant Professor of Medicine, Harvard Medical School. Sp140 status was initially determined by IIF on Sp140 expressing cells versus negative cells.

Autoantigens Expression

[0065] As in Example 2.

Dual-Tag Enzyme-Linked Immunosorbent Assay (T²-ELISA) of Autoantigens

[0066] As in Example 2.

QUANTA Lite™ Sp100 ELISA

[0067] Assay was performed according to manufacturer's instructions (INOVA Diagnostics, San Diego, CA).

Results:

[0068] T²-ELISA Autoantibody Unit calculations and "analytical" scoring were performed as in Example 2. Scoring for the INOVA Diagnostics Sp100 ELISA were performed according to the manufacturer's instructions. Results are in **Table III**. Notably, although Sp100 was unable to be detected in PBC patients PB-AMP-020 or PB-AMN-084 (orange shading) by either our T²-ELISA or INOVA's assay, the T²-ELISA platform was able to detect these PBC patients using the Sp140 autoantigen. The detection of PB-AMN-084 is most notable, since this patient was not detected by any of the following: the Sp140 indirect immunofluorescence (IIF) methods (not shown) any of INOVA's available PBC ELISA tests, or either of the novel autoantigens HK1 and KLHL12 as determined by T²-ELISA (see earlier in Example 4 and **Table II** for these ELISA results).

[0069] Together then, HK1, KLHL12 and Sp140 may serve as a powerful diagnostic panel of autoantigens which enable the rapid and accurate diagnosis of previously missed PBC patients.

[0070] This Example also demonstrates another important result, that is, with respect to Sp100, our T²-ELISA platform is essentially 100% concordant with INOVA's FDA approved Sp100 ELISA. The only discordant results were 2 cases where the T²-ELISA gave a negative result and the INOVA assay an equivocal result, that is, too close to INOVA's designated cutoff to be conclusive (per the manufacturer's scoring methods).

Example 7: Colorimetric Versus Chemiluminescent ELISA Detection of Autoantibodies Against the Novel Primary Biliary Cirrhosis (PBC) Autoantigens HK1 and KLHL12 Using PBC Patient Serum

[0071] ELISA experiments exploring the binding between autoantigens and autoantibodies usually employ one of two detection strategies. Chemiluminescence is generally accepted to be more sensitive and has a broader dynamic range, while colorimetric is generally accepted to be more stable and consistent. The purpose of these experiments was to perform the exact same experiment twice and then to develop it in parallel, once by colorimetric detection, and once by

chemiluminescent detection.

Autoantigens Expression and T2-ELISA

5 **[0072]** Performed as in Example 2 except that for the colorimetric ELISA detection, the following reagents from the INOVA Diagnostics QUANTA Lite™ ELISA platform (San Diego, CA) were utilized: HRP Sample Diluent, HRP Wash Concentrate, HRP IgG Conjugate, TMB Chromogen, HRP Stop Solution. Instructions were followed per the manufacturer. The diagnostic scoring for the chemiluminescent ELISA were those as already determined in Example 4 for the same sera.

10 *Results:*

[0073] ELISA results of HK1 on sera from PBC patients are shown in **Figure 9A** and KLHL12 in **Figure 9B**, demonstrating both colorimetric and chemiluminescent detection. Colorimetric assay results are plotted as signal minus background, with the background being the same serum run against an expression blank (no autoantigen expressed). The chemiluminescence ELISA score is listed under the X-Axis as "+" (positive) or "-" (negative). Note that the scores for the chemiluminescent ELISA were those as already determined in Example 4 for the same sera (with sera PB-AMN-044 and PB-AMN-263, green outline in **Figures 9A and B**, being the ones that scored previously negative for all available PBC ELISA assays from INOVA Diagnostics but positive for HK1 and KLHL12 respectively). These results clearly demonstrate concordance between the chemiluminescent and colorimetric ELISA readout methods.

20 Example 8: Feasibility of Point-of-Care Diagnostics Colorimetric Dot Blot Detection of Autoantibodies Against the Novel Primary Biliary Cirrhosis (PBC) Autoantigen HK1 Using PBC Patient Serum

25 **[0074]** The purpose of this example is to show proof-of-principle for use of autoantigens in a point-of-care (POC) autoantibody based diagnostic assay for autoimmune disease (i.e. an assay that is rapidly and readily performed in the doctor's office, e.g. by an internist, general practitioner or rheumatologist).

30 **[0075]** One common format of a solid-phase immunoassay for point-of-care (POC) diagnostics is the lateral flow based immuno-chromatographic method, performed on a porous solid membrane matrix, such as nitrocellulose. For example, a blood sample as well as a colorimetrically labeled detector reagent (commonly a colloidal gold label) are allowed to flow by capillary action across the length of a nitrocellulose strip, subsequently contacting the test area where, for example, an antigen, capture antibody or other capture agent had been previously immobilized (i.e. striped). A positive result is visualized as a colored stripe in the test area.

35 **[0076]** The most ubiquitously recognized form of such an assay is the "home" pregnancy test, however, various formats for rapidly detecting antibodies in human blood, e.g. for detection of pathogen infection, are possible [Biagini, Sammons, Smith, MacKenzie, Striley, Snawder, Robertson and Quinn (2006) Clin Vaccine Immunol 13: 541-6; Laderman, Whitworth, Dumaul, Jones, Hudak; Hogrefe, Carney and Groen (2008) Clin Vaccine Immunol 15: 159-63].

40 **[0077]** To mimic this type of device and show feasibility with the new PBC autoantigen HK1 reported in this patent, a dot blot assay was performed. In this assay, autoantigen is immobilized on a nitrocellulose membrane which is then probed with patient serum. Detection of bound autoantibody is achieved with a colloidal-gold labeled anti-human IgG detector antibody. Details of the procedure and results are as follows:

Colorimetric Dot Blot of Autoantigen

45 **[0078]** Recombinant purified human Hexokinase 1 protein (HK-1, Alpha Diagnostic, International, San Antonio, TX) was diluted to 200 ng/μL in TBS (50 mM Tris, pH 7.5, 200 mM NaCl). Human IgG was diluted to 250 ng/μL in PBS (50 mM sodium phosphate, pH 7.5, 100 mM NaCl).

50 **[0079]** Nitrocellulose (HiFlow Plus, Millipore Corporation, Bedford, MA) was cut to form 0.5 cm x 3 cm strips. 1 μL each of TBS, HK1 and human IgG were individually spotted onto the nitrocellulose and allowed to dry thoroughly by incubation for 1 h at 37°C. Strips were then treated in Block buffer [1% BSA (w/v) in TBS-T (TBS with 0.05% v/v Tween-20)] for 30 min at room temperature (RT). Block was vacuum aspirated. Patient serum was diluted 1:100 in Block and then incubated with nitrocellulose strips for 30 min at RT. Serum was aspirated and the strips were washed with 1.5 mL TBS-T: 4 x 5 min each. Strips were probed with colloidal gold conjugated secondary antibody [Anti-Human IgG (H+L) antibody, Gold labeled (40nm), KPL, Gaithersburg, MD] diluted 1:10 in Block shaking at RT for 3 hours.

55 *Results:*

[0080] Lateral flow immunoassays offer a simple, accurate, fast result-reporting and ease-of-use format and thus are a popular point-of-care (POC) diagnostic platform. Lateral flow-based devices use immunochromatographic principles

to assay bio-fluids such as blood for various analytes in a matter of minutes, under "field" conditions with no special instrumentation or expertise. To test the feasibility of a colorimetric lateral flow POC assay of PBC autoantigens, we performed a model dot blot experiment.

[0081] Recombinant purified human HK1 was spotted onto nitrocellulose, as well as carrier buffer (negative control) and human IgG (positive control). Diluted sera (1:100) from a PBC patient and normal patient was allowed to bind and washed before adding colloidal gold labeled anti-human IgG. Results are shown in **Figure 10**. After 1 h 20 min, all IgG spots (positive controls) had turned pink. The HK1 spot turned pink with 1:100 dilution of PBC patient serum but was negative (no color) with normal serum. Negative control spots (carrier buffer only) remained colorless.

Example 9: A Dual-Epitope Tag Based Solid-Phase Heterogeneous Assay (T²-ELISA) as a Tool for Detecting Protein Interactions

[0082] We have developed a novel, high throughput and internally normalized solid-phase heterogeneous assay which is based on dual-epitope tagged cell-free (*in vitro*) expressed target proteins captured on a surface. The assay can detect the binding of "probes" (e.g. drugs, oligonucleotides or antibodies) to the surface-immobilized cell-free expressed target proteins while being able to normalize for the amount of target protein on the same surface. Although the Example shown here relates to detection of autoantibody binding from human serum to cell-free expressed autoantigens as the target proteins, the methodology is broadly applicable. Furthermore, although the assay format used in this Example is a micro-well (microtiter) plate based ELISA format, various assay formats are possible.

[0083] One embodiment of our novel assay, which we shall call the T²-ELISA method, comprises the capture of an autoantigen (target protein) onto the microtiter plate well with one epitope tag (capture tag) followed by reading the autoantibody (probe) signal in the same well, while using the other tag (detection tag) to normalize for the amount of protein expressed in separate wells. In order to compare our T²-ELISA assay with an FDA-approved, commercially available, semi-quantitative ELISA assay for the detection of anti-sp100 IgG antibodies in human serum (QUANTA Lite™ sp100; INOVA Diagnostics, San Diego, CA) we set up the following experiment: Briefly, autoantigens are cell-free expressed, purified in-line with the microtiter plate based assay (i.e. captured on well surface) and screened against patient sera for autoantibody binding using a traditional sandwich ELISA format. Enzyme-tagged detector antibodies (each having a different chemiluminescent substrate) are added in series, after which two different chemiluminescent substrates are added to the appropriate wells one at a time in order to read both autoantibody binding as well as the detection tag (normalization signal).

Autoantigens Expression

[0084] The entire Open Reading Frame (ORF) of the putative autoantigen (in this case human Sp100) was cloned, using standard and accepted molecular biology practices, into a plasmid vector compatible with cell-free protein expression, containing the T7 RNA polymerase promoter, a Kozak (ribosome binding) sequence, a start codon, an N-terminal VSV-G epitope tag (YTDIEMNRLGK(SEQ ID NO: 19)), and a C-terminal HSV epitope tag (QPELAPEDPED(SEQ ID NO: 20)) in addition to the ORF insert. As source DNA for cloning into the expression vector, full-length sequence-verified clones were purchased from OpenBiosystems (Huntsville, AL). Expression vectors were verified for the correct ORF insert using standard EcoRI digestion methods.

[0085] Autoantigens were produced from the aforementioned plasmid clones by cell-free protein expression. Cell-free protein expression reactions were performed using a transcription/translation coupled rabbit reticulocyte lysate system (TNT® T7 Quick for PCR DNA; Promega, Madison, WI) according to the manufacturer's instructions: Autoantigen expression reactions contained the cognate plasmid DNA while blank expression reactions lacked only the plasmid DNA. Expression reactions were stopped by diluting 1/20 in TDB [1% BSA (w/v) and 0.1% (v/v) Triton X-100 in TBS-T (50 mM Tris, pH 7.5, 200 mM NaCl, 0.05% (v/v) Tween-20)].

Enzyme-Linked Immunosorbent Assay (ELISA) of Autoantigens

[0086] Nunc Brand 96-well Polysorp™ Microwell™ white opaque, flat bottom, untreated polystyrene microtiter plates (Nunc Brand from Thermo-Fisher Scientific; Rochester, NY) were used for a sandwich type Enzyme-Linked Immunosorbent Assay (ELISA). Plates were coated with 0.5 μg/mL of a mouse monoclonal anti-HSV® tag capture antibody (EMD Biosciences, Inc., San Diego, CA) in sodium carbonate/bicarbonate pH 9.3 for 30 min with shaking (50 μL/well). All plate washing consisted of manual addition of TBS-T (wells filled to maximum, i.e. 300 μL) followed by vacuum aspiration, repeated 4x. All plate washes were performed in this manner unless noted otherwise. Plates were then blocked for 30 min at 300 μL/well in 1% BSA (w/v) in TBS-T. The solution was removed from the plates and the aforementioned stopped (i.e. diluted) cell-free expression reactions (autoantigen and blank reactions) were then added at 100 μL/well and shaken for 30 min. Plates were washed and serum samples (diluted at 1/1,000 in 1% BSA (w/v) in

TBS-T) were added at 100 μ L/well and shaken for 30 min. Plates were washed and serum samples (diluted at 1/1,000 in 1% BSA (w/v) in TBS-T) were added at 100 μ L/well and shaken for 30 min. Each serum sample was run against duplicate wells of autoantigen and duplicate wells of the cell-free expression blank with an additional set of duplicate wells of the cell-free expression blank designated for VSV-G epitope tag detection [thus received plain 1% BSA (w/v) in TBS-T instead of diluted serum]. Wells designated for detection of the VSV-G epitope tag then received an anti-VSV-G horseradish peroxidase (HRP) labeled monoclonal antibody, while wells designated for detection of serum autoantibody received a mouse anti-[human IgG] HRP labeled monoclonal secondary antibody. Plates were subsequently washed 4x by manual addition of TBS-T (wells filled to maximum) followed by vacuum aspiration as described earlier in this Example. Wells designated for detection of the VSV-G epitope tag then received an anti-VSV-G horseradish peroxidase (HRP) labeled monoclonal antibody (Clone P5D4, Roche Applied Science, Indianapolis, IN) diluted 1/20,000 in 1% BSA/TBS-T. Wells designated for detection of serum autoantibody received a mouse anti-[human IgG] HRP labeled monoclonal secondary antibody (minimum cross-reactivity with mouse immunoglobulin; Jackson ImmunoResearch Laboratories, Inc, West Grove, PA) diluted 1/20,000 in 1% BSA/TBS-T. Plates were shaken for 30 min. The solutions were then manually dumped from the plates by inversion followed by vigorous patting of the plates inverted on a dry paper towel to remove residual fluid. Plates were then washed as described earlier in this Example. Chemiluminescence signal was generated by the addition of 50 μ L/well of SuperSignal ELISA Pico Chemiluminescence Substrate (Pierce Brand from Thermo Fisher Scientific, Rockford, IL). Plates were developed by shaking for 15 min and then read on a LumiCount luminescence plate reader (1s exposure, PMT of 650V, gain 1) (Packard/PerkinElmer Life and Analytical Sciences, Inc., Boston, MA).

QUANTA Lite™ sp100 ELISA

[0087] Assay was performed according to manufacturer's instructions (INOVA Diagnostics, San Diego, CA).

Results:

[0088] We compared our T²-ELISA to a commercial ELISA to test concordance (**Figure 11**). This was done by testing 35 primary biliary cirrhosis (PBC) sera for autoantibodies against the known autoantigen Sp100. The commercial ELISA (INOVA Diagnostics, San Diego, CA) is an FDA-approved colorimetric ELISA comprised of autoantigen immobilized on the plate surface and was performed according to the manufacturer's instructions. Data are shown in **Figure 11** using a subset of the PBC cohort. The INOVA standard positive control serum used to calculate "Units" was run on both assays to convert the signals of each assay to the same scale (Units/ μ L of Neat Serum). Both assays were scored using the INOVA methodology, i.e. positive when units >25; which is what the "Low Positive" standard positive control serum is set to. As **Figure 11** indicates, in terms of scoring sera positive or negative, there is perfect concordance. However, the INOVA assay saturates very quickly, while the T²-ELISA displays at least a 5-fold wider dynamic range.

Example 10: Comparison of T²-ELISA with a Conventional Commercial ELISA for p53 Tumor Associated Autoantibody Detection From Cancer Sera in Order to Assess Concordance

Autoantigens Expression for T²-ELISA

[0089] The entire Open Reading Frame (ORF) of human p53 was cloned, using standard and accepted molecular biology practices, into a plasmid vector compatible with cell-free protein expression, containing the T7 RNA polymerase promoter, a Kozak (ribosome binding) sequence, and C-terminal HSV (QPELAPEDPED (SEQ ID NO: 20)) and 6X His epitope tags, in addition to the ORF insert. Expression vectors were verified for the correct ORF insert using DNA sequencing.

[0090] The p53 autoantigen was produced from the aforementioned plasmid clone by cell-free protein expression. Cell-free protein expression reactions were performed using a transcription/translation coupled rabbit reticulocyte lysate system (TNT[®] T7 Quick for PCR DNA; Promega, Madison, WI) according to the manufacturer's instructions. Autoantigen expression reactions contained the cognate plasmid DNA while blank expression reactions lacked only the plasmid DNA. Expression reactions were stopped by diluting 1/20 in TDB [1% BSA (w/v) and 0.1% (v/v) Triton X-100 in TBS-T (50 mM Tris, pH 7.5, 200 mM NaCl, 0.05% (v/v) Tween-20)].

Enzyme-Linked Immunosorbent Assay (T²-ELISA) of Autoantigens

[0091] Sera (ProMedDx, Norton, MA) from 34 patients diagnosed with colorectal cancer (CRC) of varying stages (ranging from AJCC/UICC Stage I to Stage IV) and from 7 disease-free individuals were screened in duplicate for autoantibodies against the p53 tumor autoantigen using a commercial ELISA (EMD Biosciences, Inc., San Diego, CA)

comprised of recombinant human cellular expressed p53 and the T²-ELISA. For the commercial ELISA, sera, pre-cleared with a 5 minute spin at 16,000 x g in a microcentrifuge at 4°C, were diluted 1:100 and run in duplicate following instructions provided by the manufacturer and described in the literature [Oshikawa and Sugiyama (2000) *Respir Med* 94: 1085-91]. A validated negative control sera (provided by the manufacturer) was also run in duplicate and used to determine assay background. Absorbance readings at 450 nm for each well were collected on a SpectraMax Plus384 microplate spectrophotometer (Molecular Devices, Sunnyvale, CA).

[0092] For screening sera with the T²-ELISA, the following protocol was used. Nunc Brand 96-well Polysorp™ Microwell™ white opaque, flat bottom, untreated polystyrene microtiter plates (Nunc Brand from Thermo-Fisher Scientific, Rochester, NY) were used for a sandwich type Enzyme-Linked Immunosorbent Assay (ELISA). Plates were coated with 0.5 μg/mL of a mouse monoclonal anti-HSV® tag capture antibody (EMD Biosciences, Inc., San Diego, CA) in sodium carbonate/bicarbonate pH 9.3 for 30 min with shaking (50 μL/well). Plates were then manually washed 4x in 300 μL TBS-T using a multichannel pipette to add the wash buffer and inversion of the plates followed by vigorous patting of the inverted plates on a dry paper towel to remove the wash buffer and residual fluid. Blocking was performed for 30 min with 300 μL/well in 1% BSA (w/v) in TBS-T. The solution was removed from the plates as just described and the aforementioned stopped (i.e. diluted) cell-free expression reactions (autoantigen and blank reactions) were then added at 100 μL/well and shaken for 30 min. Plates were washed as above and serum samples (pre-cleared with a 5 minute spin at 16,000 x g in a microcentrifuge at 4°C) were diluted at 1/2,000 in 1% BSA (w/v) in TBS-T. A volume of 100 μL serum/well was added and plates were shaken for 30 minutes at room temperature. Each serum sample was run against duplicate wells on each of two separate plates, one containing cell-free expressed autoantigen and the other containing cell-free expression blank (expression reaction minus DNA template). Following serum incubation, serum was removed by vacuum aspiration and plates were washed 4x with TBS-T. For serum autoantibody detection, 100 μL of a mouse anti-[human IgG] HRP labeled monoclonal secondary antibody (minimum cross-reactivity with mouse immunoglobulin; Jackson ImmunoResearch Laboratories, Inc, West Grove, PA) diluted 1/20,000 in 1% BSA/TBS-T was added to each well. Plates were shaken for 30 min at room temperature followed by washing 4x in 300 μL TBS-T as described above. Chemiluminescence signal was generated by the addition of 50 μL/well of SuperSignal ELISA FEMTO Chemiluminescence Substrate (Pierce Brand from Thermo Fisher Scientific, Rockford, IL). Plates were developed by shaking for 15 seconds at room temperature and then read on a LumiCount luminescence plate reader (1s exposure, PMT of 693V, gain 1) (Packard/PerkinElmer Life and Analytical Sciences, Inc., Boston, "MA").

Results:

[0093] To test concordance of our T²-ELISA with the commercial ELISA in detecting autoantibodies against p53, a known tumor autoantigen, 34 sera from CRC patients (**Figure 12**, 1-34) and 7 sera from disease-free, "normal" individuals [**Figure 12**, N1-N7 (outlined by green box)] were tested in duplicate on each of the two assays. After running each ELISA, signal minus background values were first calculated for each sera. For the commercial ELISA, background was calculated as the average of the raw values from each of the two wells probed with a validated negative sera provided by the manufacturer. This background value was then subtracted from the raw values of each of the test wells probed with either CRC or "normal" sera, yielding duplicate signal-minus-background values for each sera. Note that a floor of zero was set for these signal-minus-background values (i.e. any negative values were set to zero). The duplicate signal-minus background values for each sera were then averaged yielding a single, average, signal-minus-background value. For the T²-ELISA, background was determined as the average of the duplicate wells for each serum run against the cell-free expression blank (minus DNA template reaction). This background value was then independently subtracted from each of the duplicate raw values for the same serum run against cell-free expressed autoantigen (p53) yielding two signal-minus-background values for each sera. As with the analysis of the commercial ELISA data, a floor of zero was once again set for the signal-minus-background values. The duplicate signal-minus-background values for each sera were then averaged yielding a single, average, signal-minus-background value for each sera. Next, for both the commercial ELISA and T²-ELISA, sera were simply scored as analytically positive or negative (**Figure 12** shows only those sera scored as analytically positive) in order to check concordance between the two assays. For this, both of the following criteria must have been met for each serum-autoantigen pair in order for that pair to have been scored as analytically positive in the ELISA: i) a p-value ≤0.05 in a 1-tailed homoscedastic unpaired t-test on the raw ELISA values from the duplicate wells of the autoantibody signal (serum versus autoantigen) compared to values from the duplicate wells of the background signal (same serum versus blank expression wells); ii) autoantibody signal-to-background ratio ≥2. Serum-autoantigen pairs not passing these criteria are set to 0. Finally, for each assay independently, the average signal-minus-background values of those sera scored as analytically positive were normalized to the serum with the highest value in that same assay (CRC 12 for the commercial ELISA and CRC19 for the T²-ELISA), which was set to 100%. These normalized values were then plotted with error bars representing standard deviations (**Figure 12**). As can be noted in **Figure 12**, all sera that scored positive for p53 autoantibodies in the commercial ELISA also scored positive (with an approximately equal relative strength of signal, also) in the T²-ELISA. Additionally, one additional CRC serum

(serum 18), but no additional normal serum, was scored slightly positive by the T²-ELISA and negative by the commercial ELISA. Together, the data suggest that the T²-ELISA is at least as sensitive as the commercial ELISA, and perhaps may even be slightly more sensitive as indicated by the ability to identify one additional CRC sample. Neither assay detected an autoantibody signal in any of the normal sera, suggesting a very good concordance with respect to specificity, also.

Example 11: A Dual-Epitope Tag and Dual-Reporter Based Solid-Phase Heterogeneous Assay as a Tool for Detecting Interactions with Proteins

[0094] The dual-tagged T²-ELISA described in Example 2 utilizes a single-reporter system for autoantibody detection and target protein normalization. Whereas Example 2 demonstrates using separate wells for probe readout (autoantibody in that case) and epitope tag readout, this Example illustrates the ability of the assay to detect the binding of "probes" (e.g. drugs, oligonucleotides or antibodies) to the surface-immobilized cell-free expressed target proteins while being able to normalize for the amount of target protein on the same surface (i.e. same well), using a dual-reporter system. Although the Example shown here relates to detection of autoantibody binding from human serum to cell-free expressed autoantigens as the target proteins, the methodology is broadly applicable. Furthermore, although the assay format used in this Example is a micro-well (microtiter) plate-based ELISA format, various assay formats are possible.

[0095] In order to show that it is possible to capture an autoantigen (target protein) onto the microtiter plate well with one epitope tag (capture tag) and normalize with the other (detection tag), while still reading the autoantibody (probe) signal in the same well, we performed the T²-ELISA assay as described in Example 2, with the following exceptions: following cell-free expression and antigen capture, and the sequential addition of enzyme-tagged antibodies, two different chemiluminescent substrates were also added sequentially, thereby enabling both autoantibody binding signals and detection tag (normalization) signals to be read sequentially within the same well.

[0096] In addition to showing that dual detection within the same well is possible, we directly compare dual-well detection to single-well detection on a variety of autoantigens with various patient sera, in order to demonstrate the potential advantages of per-well-normalization, namely, by normalizing for possible protein expression or capture variations.

Autoantigens Expression

[0097] Performed as in Example 2, with the exception of Rap55, which was expressed from column-purified PCR product. Rap55 was PCR-amplified from cDNA using standard and accepted molecular biology practices. Primers were designed to yield a PCR product compatible with cell-free protein expression, containing the T7 RNA polymerase promoter, a Kozak (ribosome binding) sequence, a start codon, an N-terminal VSV-G epitope tag (YTDIEMNRLGK(SEQ ID NO: 19)), and a C-terminal HSV epitope tag (QPELAPEDPED(SEQ ID NO: 20)) in addition to the Rap55 insert.

Enzyme-Linked Immunosorbent Assay (T2-ELISA) of Autoantigens

[0098] Performed as in Example 2, with the following exceptions. For the dual-reporter assay (different from the single-reporter assay as described in Example 2) there were no additional wells set aside for VSV-G epitope tag detection, since the tag and the probe (autoantibody) were detected sequentially in the same well. The enzyme-tagged antibodies were added sequentially to all the wells, followed each time by washing, as described here: First a mouse anti-[human IgG] alkaline phosphatase (AP) labeled monoclonal secondary antibody (minimum cross-reactivity with mouse immunoglobulin; Jackson ImmunoResearch Laboratories, Inc, West Grove, PA) diluted 1/20,000 in 1% BSA/TBS-T was added. Plates were then shaken for 30 min. The solutions were then manually dumped from the plates by inversion followed by vigorous patting of the plates inverted on a dry paper towel to remove residual fluid. Plates were then washed manually as described earlier in Example 8. This process was repeated for an anti-VSV-G horseradish peroxidase (HRP) labeled monoclonal antibody (Clone P5D4, Roche Applied Science, Indianapolis, IN) diluted 1/20,000 in 1% BSA/TBS-T. An AP chemiluminescence signal was generated by the addition of 50 μ L/well of BM Chemiluminescence ELISA Substrate (Alkaline Phosphatase Detection; Roche Diagnostics, GmbH, Mannheim, Germany) following the manufacturer's instructions. After allowing the signal to develop, plates were read as described in Example 8, followed by a second reading where PMT was set relative to the highest signal on the plate. After reading the plate, the plate was washed manually followed by the addition of 50 μ L/well of SuperSignal ELISA Pico Chemiluminescence Substrate (Pierce Brand from Thermo Fisher Scientific, Rockford, IL). Plates were developed by shaking for 15 min and then read as described in Example 1, followed by a second reading where PMT was set relative to the highest signal on the plate.

[0099] Different from the data in **Table IV**, the dual-reporter and single-reporter ELISAs performed for **Figure 13** were washed with the aid of a robotic plate washer. Specifically, Plates were washed 6x in TBS-T (wells filled to maximum) on an ELx405 Select Robotic Plate Washer (BioTek, Winooski, VT). Following the addition of serum, in order to avoid

contamination of the robotic plate washer with human serum, plates were subsequently washed 4x by manual addition of TBS-T (wells filled to maximum) followed by vacuum aspiration and then washed 6x in the robotic plate washer as described earlier in this Example.

5 *Results:*

[0100] First, in order to establish that the dual-detection process of the T²-ELISA is as efficient as single detection, we directly compared this using Rap55, a known PBC autoantigen, and a PBC patient serum sample. As seen in **Table IV-A**, the autoantibody (AP) signal [calculated as AP signal- noise (i.e. same serum versus blank expression wells)] from the dual-reporter assay was calculated as a percent of the corresponding autoantibody signal from the single-reporter (AP) assay. Both methods yielded almost identical results (dual reporter AP signal was 97% of corresponding single reporter, dual reporter HRP signal was 96% of corresponding single reporter), clearly demonstrating that detection of the VSV-G epitope tag (HRP) does not inhibit the subsequent detection of the autoantibody signal (AP) in the same well. Likewise, autoantibody (AP) detection does not significantly interfere with VSV-G epitope tag (HRP) detection in the same well. We also calculated signal-to-noise ratios for the autoantibody (AP) signal: [calculated as AP signal/noise (i.e. same serum versus blank expression wells)] from the dual-reporter assay as compared to the single-reporter assay (**Table IV-B**) and demonstrated that dual detection within the same well does not decrease the signal-to-noise ratios in the slightest.

[0101] Second, dual-reporter and single-reporter T²-ELISA assays were compared for several serum-antigen pairs. **Figure 13** shows example data from T²-ELISA for systemic lupus erythematosus (SLE), PBC and normal patient sera versus a variety of known autoantigens (CENPB, Ro-60, Smith B, and Sp140). As a reference, samples were already known to be positive for the various autoantigens as reported by clinical annotation of samples. Autoantibody Unit ELISA values were determined for each serum-autoantigen pair, for which the average and standard deviation (errors bars) was calculated and plotted in **Figure 13** individually for the aforementioned autoantigens. Note that a floor of zero was set for the Autoantibody Units. Normal sera tested with CENPB are indeed negative as expected. Signal-to-noise ratios of positive results ranged from 3:1 (Smith B vs. SLE-H) to 300:1 (SP140 vs. PBC-I-21). This experiment also compares the dual-reporter assay to a single-reporter assay whereby separate wells were used solely for the detection of the VSV-G normalization epitope tag. The potential advantage of dual-reporter detection is that each autoantibody signal is normalized per well for possible protein expression (e.g. day-to-day) or capture variations (intra- or inter-assay). The data shows no significant detriment to using the dual-reporter assay. Furthermore, as expected, standard deviations of the dual-reporter assay, which is a per-well normalization, are significantly less than the single-reporter assay, which normalizes only on a per assay (per plate) basis.

35 Example 12: Detection of Autoantibodies Against the Novel Primary Biliary Cirrhosis (PBC) Autoantigens HK1 and KLHL12 Recombinantly Expressed in a Wheat Germ Based System and Assayed Using a Direct Autoantigen Coating to the Surface of the ELISA Plate

Autoantigen and ELISA Assay

[0102] In this Example, a key feature is that the ELISA assay was performed on polystyrene microtiter plates directly coated with pre-purified recombinantly expressed autoantigens (instead of antibody mediated in situ capture/purification to ELISA plate surface as in T²-ELISA). Another notable feature is that HK1 and KLHL12 were expressed in a different system as compared to previous Examples. Human HK1 and KLHL12 full-length recombinant proteins expressed in a cell-free wheat germ based system and purified by their N-terminal GST fusion tag were purchased from Abnova (Taiwan). The plates were coated overnight with 100 μ L per well of 0.5 μ g/mL recombinant protein diluted in PBS. As detailed in Example 2, plates were then washed 6x in TBS-T (wells filled to maximum) and then were blocked for 30 min at 300 μ L/well in 1% BSA (w/v) in TBS-T. The block solution was removed from the plates and serum samples (diluted at 1/100) (diluent from INOVA Diagnostics' QUANTA Lite™ ELISA system; San Diego, CA) were added at 50 μ L/well and shaken for 30 min at room temperature. Plate washing and addition of the secondary antibody is described in Example 2. The ELISA was developed using the colorimetric substrate and stop solution from INOVA Diagnostics' QUANTA Lite™ ELISA system (San Diego, CA) according to the manufacturer's instructions.

Results:

[0103] **Figure 14** shows that the colorimetric assay works well for HK1 versus several PBC and normal sera and results are 100% concordant with the expected results (based on the microarray and T²-ELISA results; see Examples 1 and 2). Note these expected scores are indicated by "+" and "-" in the graph. Note that the red line is the cutoff for this assay (set at 2 standard deviations above the mean for the 4 expected negative samples). Also note that this is direct

plate coating with a recombinant antigen and there is no background subtraction here (it is not needed with no capture antibody present). Finally, note that N-03 is in fact supposed to be positive (and PBC-04 and PBC-05 negative) based on previous results from Examples 1 and 2.

[0104] Similarly, **Figure 15** for KLHL12 shows colorimetric assay results that are 100% concordant with the expected results (based on the microarray and T²-ELISA, results; see Examples 1 and 2). Note these expected scores are indicated by "+" and "-" in the graph. The cutoff is indicated as the red line and was set 2 standard deviations above the mean for the 4 expected negative samples. N-03 is expected to be positive and PBC-02 and PBC-07 negative based on previous results from Examples 1 and 2.

Example 13: Detection of Autoantibodies in Primary Biliary Cirrhosis (PBC) Using Homologs of HK1 and KLHL12

[0105] Information in the following paragraphs was obtained from the publically available UniProt database [The-UniProt-Consortium (2009) Nucleic Acids Res 37: D169-74] as well as the various publically available NCBI databases [National (United States) Center for Biotechnology Information].

[0106] Hexokinase 1 (HK1) is a protein which localizes to the outer membrane of mitochondria. Alternative splicing the gene encoding HK1 results in five transcript variants which encode different isoforms. Each isoform has a distinct N terminus but the remainder of the protein is identical among all isoforms [NCBI RefSeq]. Therefore, it is reasonable to assume that any of the aforementioned isoforms would be sufficient for detection of autoantibodies to hexokinase 1 in Primary Biliary Cirrhosis (PBC).

[0107] Furthermore, Hexokinase 1 is one member of a family of proteins, which includes Hexokinase 2, Hexokinase 3, Glucokinase (Hexokinase 4), and Hexokinase Domain Containing 1. The aforementioned proteins demonstrate significant sequence homology, (e.g. using the NCBI BLAST engine, human HK1 and HK2 have 73% identities and 86% positives; NCBI Accessions BC008730.2 coding sequence and NP_000180.2, respectively) as well as share common conserved domains, including hexokinase domains_1 and_2 (pfam00349 and pfam03727, respectively), as well as the conserved multi-domain COG5026 Hexokinase [carbohydrate transport and metabolism].

[0108] Kelch-like 12 (KLHL12) is a protein involved in the ubiquitin ligase conjugation and wnt cell-signaling pathway. It contains 6 kelch repeat domains and a BTB (POZ) domain. Several Kelch-like and other proteins exist containing the aforementioned domains (e.g. see Table VI).

[0109] Due to both protein sequence similarity and the phenomena of intra- and inter-molecular epitope spreading [Vanderlugt and Miller (2002) Nat Rev Immunol2: 85-95], we fully expect that the aforementioned HK1 and KLHL12 homologs (see also Examples in Table VI) would show a similar performance with respect to the detection of disease-specific autoantibodies in Primary Biliary Cirrhosis (PBC). Furthermore, the use of homologs may increase diagnostic sensitivity and/or specificity. In this Example, this will be evaluated.

Autoantigens Expression

[0110] Will be performed as in Example 3 except that homologs of HK1 and KLHL12 will be expressed and used as autoantigens for detection of autoantibodies, such as those mentioned above in this Example and the examples of homologs listed in Table VI.

Dual-Tag Enzyme-Linked Immunosorbent Assay (T2-ELISA) of Autoantigens

[0111] Will be performed as in Example 3.

Results:

[0112] As in Example 3, in order to set diagnostic scoring thresholds for a given autoantigen species, the T²-ELISA assay will be run on a group of 22 normal patient sera and the cutoffs will then be set at 2 standard deviations above the mean for this normal cohort, for ~95% statistical confidence. The use of this method at 2-3 standard deviations is common practice (e.g. [Liu, Wang, Li, Xu, Dai, Wang and Zhang (2009) Scand J Immunol 69: 57-63]). The T²-ELISA will then be run on 22 PBC patient sera (e.g. 22 AMA-negative and/or 22 AMA-positive). The autoantigen-specific cutoffs will then be used to score both the normal and PBC patients as autoantibody negative or positive. Autoantibody Unit calculations and data processing will be performed as in Example 3. Calculations of diagnostic sensitivity and specificity for each autoantigen species will then be performed as in Example 3.

[0113] Due to both protein sequence similarity and the phenomena of intra- and inter-molecular epitope spreading [Vanderlugt and Miller (2002) Nat Rev Immunol 2: 85-95], the expectation is that at least some of the HK1 and KLHL12 homologs will show similar diagnostic performance as in Example 3 for AMA-positive and Example 4 for AMA-negative PBC where human HK1 and KLHL12 themselves were used. It is also expected that some may perform better, either

in diagnostic sensitivity or specificity, or both.

Description of the Figures

5 [0114]

[00114] **Figure 1: ELISA Based Pre-Validation of the PBC Autoantigen Hexokinase 1 (HK1) on Positive and Negative Serum Samples Randomly Selected from the Microarray Analyses.** The graphed data are from the ELISA. The "+" and "-" denote if a given serum was positive or negative for HK1 autoantibodies based on either the "ELISA" assay or microarray ("Array") analyses. Serum samples prefixed with "N" are from healthy individuals, "PBC" from primary biliary cirrhosis patients, and "SLE" from systemic lupus erythematosus patients. Calculation of Autoantibody Units from the ELISA assay is detailed in Example 2.

15 [00115] **Figure 2: ELISA Based Pre-Validation of the PBC Autoantigen Kelch-Like 12 (KLHL12) on Positive and Negative Serum Samples Randomly Selected from the Microarray Analyses.** The graphed data are from the ELISA. The "+" and "-" denote if a given serum was positive or negative for KLHL12 autoantibodies based on either the "ELISA" assay or microarray ("Array") analyses. Serum samples prefixed with "N" are from healthy individuals and "PBC" from primary biliary cirrhosis patients. Calculation of Autoantibody Units from the ELISA assay is detailed in Example 2.

20 [00116] **Figure 3: ELISA Based Validation of the PBC Autoantigen Hexokinase 1 (HK1) on a new PBC Patient Cohort Never Before Tested on the Proteome Microarrays.** The graphed data are the Log₂ transformed Autoantibody Units from the ELISA analysis. Calculation of Autoantibody Units from the ELISA assay is detailed in Example 2. Patient samples were scored as HK1 negative or positive based on the cutoff values (dotted red line) which were calculated as detailed in Example 3. The red boxed region indicates the PBC cohort and the unboxed region the normal cohort.

25 [00117] **Figure 4: ELISA Based Validation of the PBC Autoantigen Kelch-Like 12 (KLHL12) on a New PBC Patient Cohort Never Before Tested on the Proteome Microarrays.** The graphed data are the Log₂ transformed Autoantibody Units from the ELISA analysis. Calculation of Autoantibody Units from the ELISA assay is detailed in Example 2. Patient samples were scored as KLHL12 negative or positive based on the cutoff values (dotted red line) which were calculated as detailed in Example 3. The red boxed region indicates the PBC cohort and the unboxed region the normal cohort.

30 [00118] **Figure 5: Detection of the PBC Autoantigen Hexokinase 1 (HK1) on a New PBC Antimitochondrial Antibody (AMA)-Negative Cohort.** The graphed data are the Log₂ transformed Autoantibody Units from the ELISA assay, as calculated in Example 2. Dotted red line indicates the diagnostic scoring threshold, as previously determined in Example 3. HK1 detected 4 of 17 AMA-negative PBC patients (24% sensitivity). Of note, one AMA-negative PBC patient (green bar) was detected by HK1 but undetected by any of the commercially available FDA-approved ELISA assays from INOVA Diagnostics for PBC.

35 [00119] **Figure 6: Detection of the PBC Autoantigen Kelch-like 12 (KLHL12) on a New PBC Antimitochondrial Antibody (AMA)-Negative Cohort.** The graphed data are the Log₂ transformed Autoantibody Units from the ELISA assay, as calculated in Example 2. Dotted red line indicates the diagnostic scoring threshold, as previously determined in Example 3. KLHL12 detected 6 of 17 AMA-negative PBC patients (35% sensitivity). Of note, one AMA-negative PBC patient (green bar) was detected by KLHL12 but undetected by any of the commercially available FDA-approved ELISA assays from INOVA Diagnostics for PBC.

40 [00120] **Figure 7: Venn Diagram - Novel PBC-Specific Autoantigens, HK1 and KLHL12, Capture Previously Undetectable AMA-Negative PBC Patients.** Each number represents a patient.

45 [00121] **Figure 8: Detection of Hexokinase 1 (HK1) and Kelch-like 12 (KLHL12), in Addition to INOVA Diagnostic's MIT3 Assay, May Reveal a Large Number of Previously Undiagnosed PBC Patients With Atypical Indirect Immunofluorescence Staining (IIF).** Serum samples prefixed with "Cyto" or "NM" are from patients with diffuse cytoplasmic or nuclear membrane IIF staining, respectively. To avoid scale effects, graphed data for each antigen is normalized as a percent of the patient having the maximum autoantibody units for that antigen (that patient is marked with a blue arrow for each antigen). We set the Y-axis to INOVA's MIT3 cut-off of 25 units, which corresponded to 17%. All bars shown in the graph represent positive results and the lack of a bar a negative result. The

"High Positive" is a selected positive control serum for each of the autoantigens.

[00122] Figure 9A: HK1 Detection By Colorimetric ELISA in Selected PBC Patients - Concordance with Chemiluminescence ELISA Readout. Colorimetric ELISA results are plotted as the signal minus background, with the background being the same serum run against an expression blank (no expressed autoantigen). The chemiluminescence ELISA score is indicated below the X-Axis by a "+" (positive) or "-" (negative). The scores for the chemiluminescent ELISA were those as already determined in Example 4 for the same sera. The bar with the green outline corresponds to the same sample from Example 4 (PB-AMN-044) to score negative on all available PBC ELISA assays from INOVA Diagnostics but positive for HK1.

[00123] Figure 9B: KLHL12 Detection By Colorimetric ELISA in Selected PBC Patients - Concordance with Chemiluminescence ELISA Readout. Colorimetric ELISA results are plotted as the signal minus background, with the background being the same serum run against an expression blank (no expressed autoantigen). The chemiluminescence ELISA score is indicated below the X-Axis by a "+" (positive) or "-" (negative). The scores for the chemiluminescent ELISA were those as already determined in Example 4 for the same sera. The bar with the green outline corresponds to the same sample from Example 4 (PB-AMN-263) to score negative on all available PBC ELISA assays from INOVA Diagnostics but positive for KLHL12.

[00124] Figure 10: Colorimetric Dot Blot of PBC Autoantigen HK1 Probed with PBC and Normal Patient Sera. Newly discovered PBC Autoantigen HK1 was spotted onto nitrocellulose, as well as buffer (negative control) and human IgG (positive control). Diluted sera from a PBC patient and normal patient was allowed to bind and washed before adding colloidal gold labeled anti-human IgG. "hIgG" is human IgG positive control; "AAg" is new PBC autoantigen HK1; "Ctrl" is negative control (carrier buffer).

[00125] Figure 11: Comparison of T²-ELISA to a Commercial (INOVA Diagnostics) ELISA Using the Sp100 Autoantigen and PBC Sera. Serum samples prefixed with "PBC" are from primary biliary cirrhosis patients. Red boxed region represents INOVA ELISA results; yellow boxed region represents T²-ELISA results. *Units above the "Low Positive" control (red line) are scored as diagnostically positive.

[00126] Figure 12: T²-ELISA Versus Conventional ELISA for p53 Autoantibody Detection Cancer Sera. Normal sera are prefixed with an "N" (green box) and all others are CRC sera. Data are normalized as a percent of the maximum sera for that assay.

[00127] Figure 13: Dual-Reporter and Single-Reporter T²-ELISA Assays Against Various Serum-Antigen Pairs. The graphed data are the Autoantibody Units from the ELISA analysis. Calculation of Autoantibody Units from the ELISA assay is detailed in Example 12. Blue text denotes the antigen. Serum samples prefixed with "N" are normal (from healthy individuals), "SLE" systemic lupus erythematosus and "PBC" primary biliary cirrhosis.

[00128] Figure 14: Autoantibody Detection in ELISA with Pre-Purified Human Hexokinase 1 Autoantigen (HK1) Coated Directly to Polystyrene Microtiter Plate Surface. Pre-purified expressed recombinant protein autoantigen was bound directly to the polystyrene microtiter ELISA plate surface and used to assay patient serum for the presence of autoantibodies. The expected result, based on previous microarray and T²-ELISA data (Examples 1 and 2), is listed below the X-Axis as "+" (autoantibody positive) or "-" (autoantibody negative). The actual result of the assay in this Example, is shown based on the scoring cutoff in the bar graph (red dotted line), which was calculated as 2 standard deviations above the mean for the 4 expected negative samples.

[00129] Figure 15: Autoantibody Detection in ELISA with Pre-Purified Human Kelch-Like 12 Autoantigen (KLHL12) Coated Directly to Polystyrene Microtiter Plate Surface. Pre-purified expressed recombinant protein autoantigen was bound directly to the polystyrene microtiter ELISA plate surface and used to assay patient serum for the presence of autoantibodies. The expected result, based on previous microarray and T²-ELISA data (Examples 1 and 2), is listed below the X-Axis as "+" (autoantibody positive) or "-" (autoantibody negative). The actual result of the assay in this Example, is shown based on the scoring cutoff in the bar graph (red dotted line), which was calculated as 2 standard deviations above the mean for the 4 expected negative samples.

[00130] Figure 16: Quantile Normalized Proteome Microarray (ProtoArray) Autoantibody Data for Human Hexokinase 1 (HK1) for 92 Distinct Serum Samples. Autoantibody fluorescence signal intensity, "Array Signal" (quantile normalized across the entire 92-member microarray set on a per lots basis), for each of the patient serum samples is shown for the novel autoantigen human HK1. PBC = Primary Biliary Cirrhosis; Normal or Norm = Healthy

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Individuals; SLE = Systemic Lupus Erythematosus; SjS = Sjogren's Syndrome; CRC = Colorectal Cancer; AIH = Autoimmune Hepatitis.

[00131] Figure 17: Quantile Normalized Proteome Microarray (ProtoArray) Autoantibody Data for Human Kelch-Like 12 (KLHL12) for 92 Distinct Serum Samples. Autoantibody fluorescence signal intensity, "Array Signal" (quantile normalized across the entire 92-member microarray set on a per lots basis), for each of the patient serum samples is shown for the novel autoantigen human KLHL12. PBC = Primary Biliary Cirrhosis; Normal or Norm = Healthy Individuals; SLE = Systemic Lupus Erythematosus; SjS =Sjogren's Syndrome; CRC = Colorectal Cancer; AIH Autoimmune Hepatitis.

NCBI GenBank Accession (ID of Nucleic Acid Coding for Protein)	Gene	Fasta Header/Description	M-Statistics P-Value	Diagnostic Sensitivity (S) & Specificity (P) in PBC vs. all Non-PBC
BC008730.2	HK1	>gi 33869444 gb BC008730.2 Homo sapiens hexokinase 1, transcript variant 1, mRNA (cDNA clone MGC:1724 IMAGE:3163058), complete cds	0.0000000012	By Hit Calling Method: (S) 89% (P) 84% By M-Statistics Method: (S) 85% (P) 90%
N_021633.2	KLHL12	>gi 21361889 ref NM_021633.2 Homo sapiens kelch-like 12 (Drosophila) (KLHL12), mRNA	0.000076	By Hit Calling Method: (S) 33% (P) 98% By M-Statistics Method: (S) 40% (P) 97%

PBC-Positive AMA-Negative Sera						
Sera ID	INOVA				AmberGen	
	MIT3	Gp210	Sp100	PBC Screen IgG/IgA	HK1	KLHL12
PB-AMN-005	-	-	+	+	+	-
PB-AMN-031	+	-	+	+	-	-
PB-AMN-033	+	-	E	+	-	-
*PB-AMN-044	-	-	-	-	+	-
PB-AMN-077	-	-	+	E	-	+
ΔPB-AMN-084	-	-	-	-	-	-

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Table II: Compiled ELISA Results for PBC-Specific Autoantigens on Antimitochondrial Antibody (AMA)-Negative PBC Sera

PBC-Positive AMA-Negative Sera						
INOVA					AmberGen	
Sera ID	MIT3	Gp210	Sp100	PBC Screen IgG/IgA	HK1	KLHL12
PB-AMN-095	-	+	-	+	E	-
PB-AMN-105	-	+	-	-	-	-
PB-AMN-109	-	-	+	+	-	+
PB-AMN-110	+	-	+	+	-	+
PB-AMN-120	+	+	-	+	+	+
PB-AMN-217	-	-	+	+	-	-
PB-AMN-223	-	-	+	+	-	-
PB-AMN-224	-	-	+	+	-	+
PB-AMN-225	-	-	+	+	-	-
PB-AMN-262	-	-	+	+	+	-
*PB-AMN-263	-	-	-	-	-	+
*	Negative by all 4 INOVA tests but detected by AmberGen					
Δ	Negative by INOVA and AmberGen tests					
E	Equivocal-presence or absence of autoantibodies unable to be determined					

Table III: ELISA Scores for PBC Patient Sera. E = equivocal, i.e. inconclusive (too close to cutoff; only used in INOVA assay). Asterisks indicate samples negative for Sp100 but positive for Sp140.

Serum ID	Sp100		Sp140
	INOVA Score	T ² -ELISA Score	T ² -ELISA Score
PB-AMP-002	+	+	+
PB-AMN-005	+	+	-
PB-AMP-006	-	-	-
PB-AM P-011	-	-	-
PB-AMP-018	+	+	-
*PB-AMP-020	-	-	+
PB-AMP-021	+	+	+
PB-AMP-024	+	+	-
PB-AMP-029	+	+	-
PB-AMN-031	+	+	-
PB-AMN-033	E	-	-
PB-AMP-035	+	+	-
PB-AMP-036	+	+	-

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Table III: ELISA Scores for PBC Patient Sera. E = equivocal, i.e. inconclusive (too close to cutoff; only used in INOVA assay). Asterisks indicate samples negative for Sp100 but positive for Sp140.

Serum ID	Sp100		Sp140
	INOVA Score	T ² -ELISA Score	T ² -ELISA Score
PB-AMP-039	+	+	+
PB-AMP-046	-	-	-
PB-AMP-047	+	+	+
PB-AMP-048	+	+	-
PB-AMP-059	+	+	-
PB-AMP-063	+	+	-
PB-AMP-066	+	+	-
PB-AMP-068	E	-	-
PB-AMN-077	+	+	+
PB-AMP-080	+	+	-
*PB-AM N-084	-	-	+
PB-AMP-102	+	+	-
PB-AMN-109	+	+	+
PB-AMN-110	+	+	-
PB-AMP-113	+	+	-
PB-AMP-122	+	+	+
PB-AMN-217	+	+	+
PB-AMP-218	+	+	-
PB-AMN-223	+	+	+
PB-AMN-224	+	+	-
PB-AMN-225	+	+	-
PB-AMN-262	+	+	+

Table IV: Dual detection ELISA is as efficient as single detection ELISA

A. Single Reporter (Control) Versus Dual Reporter (Percent of Control) T²-ELISA Against Rap55 Autoantigen and PBC Patient Serum

Percent of Control		
Reporter Labeled Probes Added	AP Detection (Autoantibody)	HRP Detection (Rap55 Autoantigen Expression)
anti-VSV-HRP	0.02	100.00 (control)
anti-human-AP	100.00 (control)	0.23
anti-VSV-AP, then anti-human AP	97.38	96.48

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B. Single Reporter Versus Dual Reporter T²-ELISA Against Rap55 Autoantigen and PBC Patient Serum (Signal to Noise)

Signal to Noise		
Reporter Labeled Probes Added	AP Detection (Autoantibody)	HRP Detection (Rap55 Autoantigen Expression)
anti-VSV-HRP	3.98	697.86 (control)
anti-human-AP	20.15 (control)	4.51
anti-VSV-AP, then anti-human AP	20.22	760.00

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NCBI GenBank or Protein Accession	Fasta Header/Description	Sequence
<p>Table V: Human Hexokinase 1 (HK1) and Human Kelch-Like 12 (KLHL12) Sequences on ProtoArray v.4.0 (Invitrogen, Carlsbad, CA) (Example 1), on the T2-ELISA (several Examples) and on the Conventional ELISA of Example 12.</p>		
<p><u>BC008730.2</u> SEQ NO: 1</p>	<p>>gj 33869444 gb BC008730.2 Homo sapiens hexokinase 1, transcript variant 1, mRNA (cdna clone MGC:1724 IMAGE:3163058), complete cds</p>	<pre> MIAAQLAYVTELEKDDQVKIKDKLYAMRLSDTELLDITMTRFRKEMKNGLSRDNFNTA TVKMLPFTVRSIPDGSSEKGFIALDLGGSFRLLRVQVNHKKNQNVHMESEVYDTPENI VHGSQSQLEDFHVAFLQDFMEKRIKDKKLPVGFTEFSPCCQSKIDEAELLITWTRRKA SGEGADVVKLLNKAIKRKYDANIIVAVNDIVGTMTCGDDQCEVGLIIGTGINA CYMEFLRHIDLVEGDEGRMCINTEWGFDDGSLERTEFDREIDRGSNLPKQJLFEK MVSQMYLGEELVRLILVKMAKEGLLEFEGRIPELLETRKGFNTSDVSAIEKNKEGLHNAKE ILTRLGVPEPDDDCVSVQHVCTIVSFRSANLVAATLGAIALNRLDNKGTPRLRTIVGVD GSLYKTHPQYRRRHKTLRRLVDPDVRFLLESESGSGKGAAMVIAVAYRLAEQHRQIEE TLAHPHLTKDMLLEVKRMAEWELEGLRQKTHNNAVVKMLPFFVRRTPDGTENGDFLAL DLGGNFRVLLVIRSGKRRVEMHNTIYAIPIEIMQGTGEELEFDHIVSCLSDFLDYMG IKPRMPLGFTFSFPCQOTS LDAGILITWTKGFKATDCVGHVWVTLRLDATRREFFDL DVAVVNDIVGTMTCAYEPTCEVGLIVGIGSNACYMEEMKNVEMVEGQGMCMINME WGAFDNGCLDDIRTHYDRLVDEYSLNAGKQRYEKMISGMVILGEIVRNILIDFTKKGFL FRGQISETLTKTRGIFETKFLSQIESDRLLQVRAILLQQLGLNSTCDDSLIVKIVCGVV SRAAQLCGAGMAAVVDKIRENRGLDRLNVTIVGVDGTLTKLHPHFSRIMHQTVKELSPK CNVSEFLLEDGSGKGAALLIIVAVGVRLRTEASS </pre>
<p><u>NM_021633.2</u> SEQ NO: 2</p>	<p>>gj 21361889 ref NM_021633.2 Homo sapiens kelch-like 12 (Drosophila) (KLHL12), mRNA</p>	<pre> MGGIMAPKDIMTNTHAKSLLNSMNSLRKSNITLDCDVTILRVEQKDFPAHRIVLAACSDYFC AMFTSELSEKGPVVDIQGLTASTIMEILLDFVYTEIVHTVENVQELLPAACLLQIKGV KQACCFFLESQLEDFSNCLGIRDFAEATHNCVDLMOAAEVFSQKHFPEVWQHEEFILLISQ EVEKLIKDELIQVDSEEVFEAVINWVHAKKEREESLPNLLQYVEMPLLITRYITDVI DAEPPIRCSLQCRDLVDEAKKFLHRLPRLSQMGPRTRARLGAENEVILLVVGFGSQSP IDVVEKYDFKTQWSEFLPSITRKRYYASVSLHDRIYVIGGYDGRSELSSVECLDYTAD EDGVWYSVAFPMNRRRGLAGATTLGDMIVYSGGFDSRRHTSMERYDPNIDQWSMLGDMQ TAREGAGLVASGVTYCLGGYDGLNLSVEKYDPHTGHWTNVTPTMAKRSAGVALLN DHIYVVGDFDGTALHSVVEAYNIRIDSWITVTSMTPTFCYVVGATVLRGLYAIAGYDGN SLLSSIECYDPIIDSWVEVVTSMGTQRCDAVCVIREK </pre>

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<p>T²-ELISA - Recombinant human HK1 and KLHL12 cell-free expressed in a rabbit reticulocyte lysate. Note that the underlined sequences are exogenously added N-terminal and C-terminal epitope tags as well as vector-derived sequences.</p>	<p>CV026580 (EST) SEQ NO: 3</p> <p>>gi 51484591 gb CV026580.1 ICV026580 4566 Full Length cDNA from the Mammalian Gene Collection Homo sapiens cDNA 5' similar to BC008730 (HK1), mRNA sequence</p>	<p>MAIYTDIEMNRLGKMIAGLLAYFTELEKDDQVKKIDKYLAMRLSDETLIDIMTRFRK EMKNGLSRDFNFATVVKMLP TFVRS IPDGSEKGF TALDLGSSFRILRVQVNHKKNQ VHMESEYDTPENIVHSGSOLFEDHVAECLGDFMEKRRKIKDKLVPVGF TF SFCQQS DEAILITWTRFKASGVEADVVKLLNKAIKRGGDYANIIVAVVNDIVGTMMITCGYDDQ HCEVGLIIGTGTNACIMEELRHLDLVEGDEGRMCINTEWGAFGDDGSLERTEFDREI DRGSLNPKQGFLEKRWSGMYLGEVLVRLILVKAKEGLLEGRITPELLTRGKFNISDVS AIEKNKEGLHNAKEILTRLGVEFSDDDCVSQHVCTIVSFRANLVAATLGAALNRLRD NKGTFRLTITVGDGSLYKTHPOYSRFRPHKTLRRLVDSDFRFLSES GSGKAAMVTA VAYRLAEQHRQIEETLAHFHLTKDMLLEVKRMRAMELGLRQKHNNNAVVKMLP SFVR RTPDGTENGDFLALDLGGTFRVLLVTRSGKRTVEMHNKIYAIPIEIMQGTGEELFD HIVSISDFLDYMGIKGPRMPLGFTTF SFCQQTSLDAGILLITWIKGFKAIDCVGHDDVT LLRDAIKRREFFDLVAVVNDIVGTMMITCAVEEPTCEVGLIVGTGSNACYMEEMKNVE MVEGDQGM CINMEWGAFGDNGCLDDIRHYDRLVDEYSLNAGKQRYEKMI SGMVIGEI VRNILDFTKKGFLFRGQISEITLKTGRFFETKFLSQFESDRLALLQVRAILQQLGLNST CDDSLIVKIVCGVVSRRAAQLCGAGMAAVVDKIRENRGLRLNVTVGVDGTLYKLLHFE SRIMHQTVKELSPKCNVSEFLLEDGSGKGAALITAVGVRLRTEASSLSRELVDPNVQA RLQVDVGTIDTRSKLAAALYTRASQPELAPEDPEDELEHHHHHH</p> <p>MYTDEMNRLLGKMGIMAPKDIMTINTHAKSII LNSMNSLRKSNITL CDVTLRVEQKDFPAH RIVIAACSDYFCAMFTSELSEKGPYVDIQGLTASTMEILLDFVYTEIVHVTVENVQEL LPAACILQLKGVKQACCFLESQLDPSNCLGTRDFAE THNCVLDLQAEEVFSQKHPEV VQHEEFILLSQGEVEKLIKDEIQVDSEEPVEFAVINWVKAHAKEREESLPNLLQYVRM PLLTFRYITDVIDAEPFTRCSLQCRDLVDEAKKFLHRLPELRSQMCGPTRARLIGANEVL LVVGGFDSQOSP IDVVEKYDKPTQEWSEFLPSITIRKRRYAVASVSLHDRIYVILGGYDGRSR LSSVECLDYTADEDGVWYSVAPMNVRRGLAGATTLGDMIVYSGGFDGSRRHITSMERYDP NIDQWSMLGDMQTAREGAGLVASGVICYLGGYDGLNILNSVEKYDPHTGHWINVTPMA TKRSGAGVALLNDHIYVVGDFGTAHLSSVEAYNIRTDSTWITVTSMTTPRCYVGAIVLR GRLYAAGYDGNLSLSTIECYDPIIDSWEVVTSMGTQRCDAQVCLREKQPELAPEDPE D</p>
<p>3C003183.1 SEQ NO: 4</p>	<p>>gi 13112018 gb BC003183.1 Homo sapiens kelch-like 12 (Drosophila), mRNA (cDNA clone MGC:4435 IMAGE:2958852), complete cds</p>	<p>MYTDEMNRLLGKMGIMAPKDIMTINTHAKSII LNSMNSLRKSNITL CDVTLRVEQKDFPAH RIVIAACSDYFCAMFTSELSEKGPYVDIQGLTASTMEILLDFVYTEIVHVTVENVQEL LPAACILQLKGVKQACCFLESQLDPSNCLGTRDFAE THNCVLDLQAEEVFSQKHPEV VQHEEFILLSQGEVEKLIKDEIQVDSEEPVEFAVINWVKAHAKEREESLPNLLQYVRM PLLTFRYITDVIDAEPFTRCSLQCRDLVDEAKKFLHRLPELRSQMCGPTRARLIGANEVL LVVGGFDSQOSP IDVVEKYDKPTQEWSEFLPSITIRKRRYAVASVSLHDRIYVILGGYDGRSR LSSVECLDYTADEDGVWYSVAPMNVRRGLAGATTLGDMIVYSGGFDGSRRHITSMERYDP NIDQWSMLGDMQTAREGAGLVASGVICYLGGYDGLNILNSVEKYDPHTGHWINVTPMA TKRSGAGVALLNDHIYVVGDFGTAHLSSVEAYNIRTDSTWITVTSMTTPRCYVGAIVLR GRLYAAGYDGNLSLSTIECYDPIIDSWEVVTSMGTQRCDAQVCLREKQPELAPEDPE D</p>

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<p>Conventional ELISA - Recombinant human HK1 and KLHL12 (Abnova, Taipei City, 114, Taiwan) cell-free expressed in a wheat germ based system. Note that HK1 and KLHL12 contained an N-terminal GST fusion tag (sequence not shown) commonly known to those skilled in the art.</p>	<p>AAH08730 SEQ NO: 5</p> <p>>gij14250554 gb AAH08730.1 Hexokinase 1 [Homo sapiens]</p>	<pre> : MIAQLLAYFTTELKDDQVKDKYLWRLSDETLIDIMTRFRKEMKNGLSRDNPETA : TVKMLPFFVRSIDPGEKGFIAIDLGGSFILRVQVNHKQNVHMESEVYDFPNI : VHGSGQLFDHVAECLGDFMEKRKIKDKKLPVGFTEFPCCQSKIDEAIIITWKRKA : SGEGADVVKLLNKAIKRKYDANIYAVVNDIVGTMTCGGYDDQCEVGLIIGTINA : CYMEELRHIDLVEGDEGRMCINTEWGAFGDDGLEDIRTEFDREIDRGSINPKQJFEK : MVSVMYLGEIVRLIIVKMAKEGLIFEGRIPELITRKGKNTSDVSAIEKNKEGLHNAKE : ILTRLGVPESSDDDCVSVQHVCTIVSFRSANLVAATLGAIIINRLDNKGTPLRTTVGVD : GSLYKTHPQYSRRFHKLRLRPDSDVRFLLSESGSGKAAMVAVAYRAEQHRQIEE : TLAHFHLTKDMLLEVKRMAEMELGLRKQTHNNVAVKMLP SFVRRTPDGTENGDFLAL : DLGGTNRVILVLRSGRKRIVEMHNKIYAIPIEIMQGTGEELEFDHIVSCLSDFLDYMG : IKGPRMPLGFTFSPCCQTSLDAGILITWTKGFATDCVGHVWVILLRDAIKRREFFDL : DWAVVNDIVGTMTCAYEPTCEVGLIVGTSNACYMEEMKVENVEGQGMCMNME : WGAFGDNGCLDDIRTHYDRLVDEYSLNAGKQRYEKMI SGMYLGEIVRNILIDFTKKGFL : FRGQISETLITRIGFETKFLSQIESDRIALLOVRAILQQGLNSICDSDSIIIVKIVCGVV : SRRAAQLCGAGMAAVVDKIRENRGLDRLNVTYGVDTLYKLPHPFSRIMHQTVKELSPK : CNVSEFLLEDGSGKGAALITAVGVRLRTEASS : MGGIMAPKDIMTNTTHAKSLLNSMNSLRKSNITLDCDVTILRVEQKDFPAHRIVLAACSDYFC : AMFTSELSEKGPYVDIQGLTASIMEILLDFYETEIVHVTIVENVQELLPACLQLKGV : KQACCEFLFSQLDFSNCLGRDFAETHNCVDLMQAAEVFSOKHFPEVQVQHEEFILLISQ : EVEKLIKDEIQVDSEEPVEAVINWVKHAKKEREESLPNLLQYVRMPLLPTRYITDVI : DAEPIRCSLQCRDLDVEAKKFLHRLPELRSQMGPRTRARLGANEVLLVWGGFGSQSP : IDVVEKYDEKIQEWSFLPSITRKRRYVASVSLHDIRIYVIGGYDGRSRLSVECLDPTAD : EDGVWYSVAPMNVRRGLAGATTLGDMIVYVSGGFDGRRHTSMERYDENIDQWSMLGDMQ : TAREGAGLVVASGVYICLGGYDGLNINLSVEKYDPHTGHWTNVTIPMAIKRSAGVALLN : DHIYVYVGGFDGTAHLSVFEAYNIRITDSWTVTSMITPCRYVGATVLRGRLYAIAGYDGN : SLLSSIECYDPIIDSWEEVVTSMGTQRCDAGVCVLRREK </pre>
<p>NP_067646.1 SEQ NO: 6</p>	<p>>gij11056006 ref NP_067646.1 ketch-like 12 [Homo sapiens]</p>	<pre> : MGGIMAPKDIMTNTTHAKSLLNSMNSLRKSNITLDCDVTILRVEQKDFPAHRIVLAACSDYFC : AMFTSELSEKGPYVDIQGLTASIMEILLDFYETEIVHVTIVENVQELLPACLQLKGV : KQACCEFLFSQLDFSNCLGRDFAETHNCVDLMQAAEVFSOKHFPEVQVQHEEFILLISQ : EVEKLIKDEIQVDSEEPVEAVINWVKHAKKEREESLPNLLQYVRMPLLPTRYITDVI : DAEPIRCSLQCRDLDVEAKKFLHRLPELRSQMGPRTRARLGANEVLLVWGGFGSQSP : IDVVEKYDEKIQEWSFLPSITRKRRYVASVSLHDIRIYVIGGYDGRSRLSVECLDPTAD : EDGVWYSVAPMNVRRGLAGATTLGDMIVYVSGGFDGRRHTSMERYDENIDQWSMLGDMQ : TAREGAGLVVASGVYICLGGYDGLNINLSVEKYDPHTGHWTNVTIPMAIKRSAGVALLN : DHIYVYVGGFDGTAHLSVFEAYNIRITDSWTVTSMITPCRYVGATVLRGRLYAIAGYDGN : SLLSSIECYDPIIDSWEEVVTSMGTQRCDAGVCVLRREK </pre>

Table VI: Examples of Homologous Sequences for HK1 and KLHL12

NCBI Protein Accession (Gene Name)	Fasta Header/Description	Sequence
HEXOKINASE 1 and Homologs		
NP_277031.1 (HK1) SEQ NO: 7	>gi 15991827 ref NP_277031.1 hexokinase 1 isoform HK1-R (transcript variant 2) [Homo sapiens]	<pre> MDCHELSLPCRGAEAEWIGIDKYLAMRLSDTLIDIMTRFRKMKNGLSRDFNPATVTK MLPFPVRSIPDGSSEKGFIALDLGGSSFRILRVQVNHKKNQVHMESEVYDTPENIVHGSG SQLFDHVAECLGDFMEKRKIKDKKLPVGFTFSPFCQSQSKIDEAIIITWTKRFFKASGVEGAD VVKLLNKAIKKRGDYDANIVAVVNDIVGTMTCGYDDQHCVEVGLIIGTGNACYMEEELRHI DLVEGDEGRMCINTEWGAFFDGSLEDIRTEFDREIDRGSINPGKQLEFKMVSVMYLGELIV RLIILVKMAKEGLLFEGRITPELLTRGFENISDVSAIEKNKEGLHNAKEIILRLGVPEP SDDD CVSVQHVCTIVFRSANLVAATLGAAILNRLRDNKGTPLRITTVGVDGSLYKTHPQYRRRFH KTLRRLVPSDVRFLLSSESGSGKAAMVAVAYRLAEQHRQIEETLAHFHLTKDMLLELVYKK RMRAEMELGLRQTHNNAVVMPLSFVRRTPDGTENGDFLALDLGGTFRVLLVVKIRSGKK RTVEMHNTIYAIPIEIMQGTGEEELFDHIVSCLISDFLDYMGTKGPRMPLGFTFFSFCQQTSL DAGILLITWTKGKATDCVGHVVTLRDAIKRREFFLDVVAVVNDIVGTMTCAYEPEP TC EVGLIVGTSNACYMEEKMVEVGEQGMQINMEWGAFGDNGCLDDIRTHYDRLLVDEYS LNAGQRVEKMI SGMYLGEIVRNILIDFTKKGFLFRGQISETLKTRGIFETKFLSQIESDR LALLQVRAILQQLGLNSTCDDSLIVKIVCGVVSRRAAQLCGAGMAAVDKIRENRGLDRIN VTVGVDGTLTKLHPHFSTRIMHQTVKELSPKCNVSFLLSEDSGSGKGAALIITAVGVRLRTEAS S </pre>
NP_000180.2 (HK2) (SEQ NO:8)	>gi 15553127 ref NP_000180.2 hexokinase 2 [Homo sapiens]	<pre> MIASHLLAYFFTELNHDQVKVDQYLXHMRLSDETLLEISKRFRKEMEKGIGATTHPTAAV KMLPFPVRSPTPDGTEHGEFLALDLGGINFRVLWVKVTDNGLQKVMENQIYAIPEDIMRGS GTQLFDHIAECLANFMDKLIQDKKLPVGFTFSPFCQTKLDESLVSWTKGFKSGVEGR DVALIRKALQRRGDFDIDIVAVVNDIVGTMTCGYDDHNCIEIGLIVGTSNACYMEEEMRH IDMVEGDEGRMCINMEWGAFGDDGSLNDIRTEFDQEIDMGSLNPGKQLEFKMISGMYMGEI VRLIILVKMAKEELLFGKLSPELLNTGRFETKDISDIEGEKDGIRKAREVLMRLGLDPTQE DCVATHRICOIVSTRSASLCAATAAVALQRIKENKGERLRSIIGVDGVSVMYKHFHFAKRL HKTVRLVFGCDVRFRLSEDSGSGKAAMVAVAYRLADQHRARQKLEHLQLSHDQLLEVK RRMKVEMERGLSKETHASAPVKMLPTVYCAITPDGTEKGFALDLGGTFRVLLVVRNGK WGGVEMHNTIYAIPIQEVMHGTGDELFDHIVQCIADFLIYMGMKVSLPLGFTFFSFCQQNS LDESILLKWTGFKFASGCEGEDVVLLKEAIIHRRREFDLVVAVVNDIVGTMTCGFEDPH CEVGLIVGTSNACYMEEKMVEVGEGRMCMWGAFGDNGCLDDIRTEFDVADEL SLNPGKQRFKMSGMYLGEIVRNILIDFTKRGILLFRGRIERLKRTRGIFETKFLSQIESD CLALLQVRAILQHLGLESICDDSLIVKEVCTIVARRAAQLCGAGMAAVDRIRENRGLDAL KVTVGVDGTLTKLHPHFARVMHETVKDLAPKCDVSLQSEDSGSGKGAALIITAVAGRIREAG QR </pre>

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NCBI Protein Accession (Gene Name)	Fasta Header/Description	Sequence
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NP_002106.2 (HK3) SEQ NO: 9	>gj 194097330 ref NP_002106.2 hexokinase 3 [Homo sapiens]	<pre> MDSIGSSGLRQGEETLSCSEEGLPGPSDSSELYQECLQQFKVTRAQLQQIQASLLGSMEOA LRQASPAVAVRMLPTTYGFS TPHGTEQGDVFLVLELGTAGTASLRVLWVTLTIGIEGHRVPEPRS QEFVTPQEVMLGAGQQDFDAAHCLSEFLDAQFVNKQGLQGFSEFPFCHQTGLDRSLLIS WTKFRCSGVEGQDVQLLRDAIRROGAYNIDVAVVNDIVGTMGCEPVGVPCEVGLVVD TGTNACYMEEARHVAVLDEDRGRVCSVVEWGSFSDDGALGPVLTTFDHLDEHSLNP GAQR FEKMI GGLXILGELVRLVLAHLCRCVLFPGCTSPALLSQGSI LLEHVAEMEDP SICAARVH AILQLGLSP GASDVEIVQHVCAAVCTRAAQLCAAALAAVLSCLQHSREQQTLQVAVATGG RVCERHPFC SVLQGTVMLLAF ECDVSLIP SVDDGGRGVAVTAVAAARLAAHRRLLLEETLA PFRINHDLAAVQAQMRKAMAKGLRGEASSLRMLPTFVRA TEDGSEKDFLALDLGGTNR VLLVRVTIGVQITSEIYSIPETVAQSGGQQLFDHIVDCIVDFQQKGLSGQSLPLGF TFSF PCRQLDQGLLLNWKGFKASDCEGQDVVSLLEAI TRRQAVELNVVAIYNDIVGTMMS GYEDPERCEI GLIVGTGNACYMEELRNAGVPGDSGRMCINMEWGAFGDDGSLAMLS TRFD ASVDQASINP GKQREFKMI SGMYLGEI VRHILLHLTSLGLFRGQQIQRLOTRDIEFKFL SEIESDSLALRQVRAI LEDLGLPLTSDDALMVEVCOAVSQRAAQLCGAGVAAVVEKIREN RGLLELAVSVGVDGTLTKLHPFSSLVAA TVRELIAPRCVWTF LQSEDDSGGKAALVTAVAC RLAQLTRV </pre>
NP_277042.1 (HK4) SEQ NO: 10	>gj 15967159 ref NP_277042.1 glucokinase isoform 2 [Homo sapiens]	<pre> MAMDVTRSAQTALTLVEQLAEFQLQEDDLKVMRRMQEMDRGLRLETHEEASVKMLPTI YVRSTPEGSEVGF LSLDLGGINFRVMLVKGEGEGOWSVKTKHQMYSIFEDAMITGTAEM LFDYI SECTSDFLDKHQMKKKLPLGF TFSFVRHED IDKGLLLNWKGFKASGAEGNVW GLLRDAI KRRGDFEMDVAVMNDVATMI SCYYEDHQCEVGMIVGTGCNACYMEEFMQVYEL VEGDGRMCVNT EWGAF GDSGELDEF LLEYDRLVDESSANFGQQLYEKLLGGKYMGEIVRL VLLRLVDENLLFHGEASEQLRTRGAFETRFVSVQVE SDTGDRKQIYNILSTLGLRP STIDCD IVRRACEVSTRAAHMCSAGLAGVINRMRESSEDMRIIVGVDGVSVKLHPSPKEREHAS VRRLTFSCELTTF IESEGSGRGAALVSAVACKKACMLGQ </pre>

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Table Vi: Examples of Homologous Sequences for HK1 and KLHL12		
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HEXOKINASE 1 and Homologs		
NP_079406.3 (HKDC1) SEQ NO: 11	>gi 156151420 ref NP_079406.3 hexokinase domain containing 1 [Homo sapiens]	MFVHLMAFYFSKLEKEDQIKKVDRLYHMRLSDDTLIDIMRRFRAEMEKGLAKDTNP TAAV KMLPTFVRAIPDGSSEGEFLSLDLGGSKFRVLKQVAEEGKRHVQMESQFYFPNLEIRGN GTELFYVADCLADFMKTKDKLKHKKLPGLTFSPFCROTKEEGVLLSWTKRFKARGVQDI DVSRLTKAMRRHKMDVDLALVNDITVGMTICAYDDPYCEVGVIIIGTGINACYMEDMSN IDLVEDEGRMCINTEWGAFGDDGALEDIETDFDLGSLNPKQLFKMI SGLYLGEI VRLILKMAKAGLLFGEEKS SAHITKGIETRHVAMKEYKEGLANTREILVDLGLPESEA DCIAVQHVCTIVSFRSANLCAALAAAILTRLENKVERLRTVGMGTLYKIHPQYPKRL HKVVKLVFSCDVRFLLESSEGSTGAAMVAVASRVQQRQIDRVLALFQLITREQLVDVQ AKMRAELEYGLKKSHGLATVRMLPTTYVCGLPDGTGKGLADLDLGGTFRVLLVIRISGR RSVRMINKIFAIPLIEMQGTGEELFHHVQCIADFLDYMLKGLASLFLGFTFSFCRQMSI DKGTLIGWTKGFKATDCGEGDVDMLEAKRRNEFDLIVAVNDITVGMTTCGYEDPNC EIGLTAGTGNMCMYMEDMRNIEMVGEGGKMCINTEWGGFGDNGCIDDIRIRYDIEVDEGS LNPKQRYEKMTSGMYLGEIVRQLIDLTKQGLFRGQISERLRTGRGFEIKFSLQIESDR LALLQVRRILQQLGLDSTCEDSIIVKVEYCGAVSRAAQLCGAGLAAIVKRRREDQGLEHLR ITVGVDTLYKLEHFSRILQETVKEIAPRCDVTFMLEDGSGKGAALITAVAKRLQQAQK EN
KELCH-LIKE 12 and Homologs		
NP_067646.1 (KLHL12) SEQ NO: 12	>gi 11056006 ref NP_067646.1 kelch-like 12 [Homo sapiens]	:MGGIMAPKDIMTNTHAKSLLNSMNSLRKSNITLDCDVLRLVEQKDFFAHRIVLAACSDYFCAM FTSELSEKGPYVDIQGLTASTMEILLDFVYETVHVTVENVQELLPAAACLLQLKGVKQAC CEFLSQDLDFSNCLGIRDFAETHNCVDIMQAAEFLSQKHFPEVWQHEEFLILSQGEVEKLI KCDDEIQVDSSEEPVEAVINWVKHAKKEREESLPLNLIQYVRMELLPYIITDVIDAEFFTRC SLQCRDLVDEAKKFLHRELPRLSQMQGFRTRARLIGAMEVLLVVGFGSQSQSFIDVVEKYDPK :TQWSEFLPITIRRRRYVASVLIHDIRIYVIGGYDGRSRLSSVECLDYTADEDDGVMYVVAEPM VRRGLAGATTLGDMIVYSGGFDGSRRTSMERYDPNIDQWSMLGDMQIAREGAGLWVASGV IYCLGGYDGLNIIINSVEKYDPTHGHWINVTMAITKRSAGYVALLNDHIYVVGDFDGTALHS SVEAYNIRIDSWITVTSMITPRCYVGTALRGRLYATAGYDGNLSLSSIECYDPIIDSWEV VTSMTQRCGDAGCVLREK
NP_055273.2 (KLHL20) SEQ NO: 13	>gi 40807500 ref NP_055273.2 kelch-like 20 [Homo sapiens]	MEGPMRRCNTNIGETGMVDVTSCTLGDPNKILPEGVFPQPARMPYISDKKHPQRTLEVINLL RKHRELCDVVIVGAKKIYAHRVILSACSPYFRAMFTGELAESRQTEVWIRIDERAMELL IDFAYTSQITVEEGNVQIILLPAACLLQAEIQEACCFELKROLDPSNCLGRFAFADTHSCR ELLRIADKFTQHNEQEVMESEEFMLIPANQLIDIISSDELNVRSSEEQVFNAVMWVKYSIQ ERRPQLFQVLQHVRLPLLSFKLVGTVGSDPLIKSDEECRDLVDEAKNLLLPQERLMQG PRTREKPIRCGEVILFVGGWCSGDAISSVERYPQINWRMVA SMSKRCRCGVGVSVLDDL LYAVGGHDSYLLNSVERYPKINQWSBDVAPTSCRTSVGAVLGGFLYAVGGQDGVSCIL NIVERYPKENKWTIRVMSMSTRRLGVAVLGGFLYAVGGSDGTSPLNITVERYNPQENRWH TIAPMGTTRKHLGCAVYQDMIVAVGGRDDITELSSAERYNPRINQWSPFWAMTSRSVGVGL AVVNGQLMAVGGFDGITYLKTIEVDFDPDANTWRLYGGMNYRRLGGVGVYKMTHCESHIV

(continued)

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NP_059111.2 (KLHL3) SEQ NO: 14	>gj166235129 ref NP_059111.2 kelch-like 3 [Homo sapiens]	MEGESVKLSQTLLIQAQDDEKQKRIITVNFAMHGKAFKVMNELRSKQLLCLDVMIVAEVDVEI EHRVVLACSPFCAMPTGDMSESKAKKIEIKVDGQTLTKSLIDYITAEIEVTEENQV LLPAAASLLQMLMDVRQNCDFLOQLHPINCLGIRAFADVHTCDLLQQANAYAQHFPEVM LGEFFLSLDQVCSLISDDKLVSEKVFVAIWINYKETEIRLEHMAKLEHVRIPLL PRDYLQIVVEEALIKNNNTCKDFLEAMKYHLLPDLQRLIKNPKTKPTPVS LPKVMIV VGGQPKAIRSVECYDFEEDRWQIAELP SRRCRAGVVFVAGHVAVGGNGSLRVRIVDV YDGVKQWTSIASMQERRSTLGAAVLNDLLYAVGGDGTGLASVEAYS YKTNWFFVAFM NTRRSVGVGVEGKLYAVGGYDASRQCLSTVEQNPATNEWIYVADMS TRRSAGVGVL SGQLYATGGHDGFLVRKSVVEYDFGTINTKQVADMMCRNAGVCAVNGLLIYVGGDDGSC NLASVEYINPVTDEWTLLEP TMSIGRSYAGVAIVHKS L
NP_938073.1 (KLHL17) SEQ NO: 15	>gj38194229 ref NP_938073.1 kelch-like 17 [Homo sapiens]	MQP RSERPAGRTQSPFEGHSPFGPEAPPPFPFPAEERTPRQARPAPMEGAVQLLS REGHSVAHNSKRHYHDAEFVAMSRMQRLGCLLIVLHVAAKEIRAHKVVILASCSPYFHAMFT NEMESRQTHVTLHDIDFQALDQIVQFAYTAETIVWGEQVQTLLEPAASLLQLNGVRDACC FLLSOLDP SNCLGIRGFADAHSCSLLKAAHRYVLQHFVDYAKTEEFMLLP LKQVLELYSS DSLNVFSEEEVYRAVLSWKYHVDYDARRQHPRLMKCVRLP LLSRDFLLGHVDAESLVRHHP DCKDLLIEALKFHLLEPQRGVIGTSRTFPRCEGAGEVLFVAVGGGSLFAHGDCEAYDTRI DRWHVAMS TRRARVGVAAVGNRLYAVGGYDGTSDIATVES YDPVINTWQPEVSMGTRRS CLGVAALHGLLYSAGYDASCLNSAERYDPLTGTWTSVAAMSTRRRYVAVATLDGNLYAV GGYDSSHLATVEKYEPQVNVSPFVASM LSRRSAGVAVLEFALYVAGGNDGTSLNSVER YSPKAGAWESVAPMNI RRS THDLVMDGWL YAVGGNDGSSLSINS IEKYNPTNKWVAASC M FTRRSVGVAVLELLINFPFPPSSPTLSVSS TSL
NP_001154993.1 (KLHL2 isoform 2) SEQ NO: 16	>gj239835722 ref NP-001154993.1 kelch-like 2, Mayven isoform 2 [Homo sapiens]	MVWLEARPOLFVCTKQGHOKPLDSKDDNTEKHCPVTVNPWHMKKAFKVMNELRSQNLLCD VTI VAEEDMEISAHRVVLAACSPYFHAMFTGEMSESRARVRIKEVDGWTLRMLIDYVYTAE IQVTEENVQVLLPAAAGLLQLODVKTKCCEFLFESQLHPVNCILGIRAFADHACHTDILNKNANT YAEQHFADVVLSEEFNLGIEQVCSLLISSDKLTI SSEEKVFVAI AWVNHDKDVRQEFMAR LMEHVRLP LLLPREYIVQRVEEALVKNS SACKDYLITAMKYHLLPTEQRLIMKSVTRLRIT PMNLP KLMVVVGGQAPKALRSVECYDFKEERWHQVLELPSRRCRAGVMYAGLVFVAVGGFN GSLRVRTVDS YDPVKDQWTSVANMRDRRS TLGAAVNLGLLYAVGGDGTGLSSVEAYNIK SNEWFHVAPMNTRRRSVGVVGGGLLYAVGGYDASRQCLSTVECYNATNEWIYIAEMST RRSGAGVGVNLLIYAVGGHDGFLVRKSVVEYDFGTINAWRQVADMMCRNAGVCAVNGLL YVVGDDGSCNLASVEYINPITIDKWTIVVSSCMSITGRSYAGTVIDKPL

(continued)

NCBI Protein Accession (Gene Name)	Fasta Header/Description	Sequence
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<p>NP_009177.3 (KLHL2 soform 1) SEQ NO: 17</p>	<p>>gj 239835720 ref NP_009177.3 kelch-like 2, Mayven isoform 1 [Homo sapiens]</p>	<pre> METPLPFACTKQGHQKPLD SKDDNTEKHCPTVNPWHMKKAFKVMNELRSQNLLCDVTIV AEDMEISAHRVWLAACSPYFHAMFTGEMSESRKRVRKEVDGWTLRMLIDYVYTAELQVTI EENVQVLLFAAGLLQLQDVKTCCEFLFSQLHFVNCIGIRAFADMHACTDLINKANTYAEQ HFADVLSSEFLNLGIEQVCSLISDDKLLISSEKVF EAVIAWVNHDKDVRQEFMARLMEH VRLPLIPREYLVQRVEEALVKNSACKDYLIEAMKYHLLPTEQRIIMKSVTRLRTPMNL PKLMVVGGQAPKALRSECYDFEERWHQVLELPSRRCRAGMVMYAGLVFAVGGFNGSLR VRTVDSYDPVKDQWTSVANMRDRRSILGAAVINGLLYAVGGDGTGLSSVEAYNIKSNW FHVAFMNTRRSSVGVVGGLLYAVGGYDGASRQCLSTVECYNATTHWEVYIAEMSTRRS AGVGVINLLYAVGGHDGFLVRKSVVEYDFITINAWRQVADMMNCRRNAGVCVAVNGLLLYVVG GDDGSCNLA SVEYYNPTTDKWTIVVSSCMSITGRSYAGVIVIDKPL </pre>
<p>NP_079286.2 (KLHL18) SEQ NO: 18</p>	<p>>gj 55925604 ref NP_79286.2 kelch-like 18 [Homo sapiens]</p>	<pre> MVEDGAEELDLVHFSVSELP SRGYGVWEEIRRQKLCVTLKI GDHKFSAHRIVLAASIP YFHAMFTNDMMECKQDEITVMQGMDFSALEALINFAYNGNLAI DQQNVQSLIMGASFLLQQS IKDACCTFLRERLHPKNCILGVRQFAETMMCAVLYDANSEF THQHFVEVSMSEEF LALPLED VLELVSRDELNVKSEEQVFEALAWVRDYDREQRP YLPELLSNIRLPLCRPQFLS DRVQOD DLVRCCHKCRDLVDEAKDYHLMFERREHLPFAFRTRPCCTSIAGLLYAVGGLNSAGDSLNV VEVDFPIANCWERCPRMTARSRVGVA VVNGLLYAI GGYDQRLRS TVEAYNPETDTWTRV GSMNSKRSMGTVVLDGQIYVCGGYDGNSSLSSVEIYSPETDKWIVVT SMSNRSRAAGTV FEGRIYVSGGHDLGLQIFSSVEHYNHHTATWHEPAAGMLNKRGRHGAASLGSKMFVCGGYDGS GFLSTAEMYSVADQWCLIVPMHTRRSVSLVASCGRLYAVGGYDQSNLSSVEMYPDPEID CWTFMAPMACHEGGVGVGCIPLLLTI </pre>

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5 <110> AmberGen, Inc.
 Lim, Mark J.
 Ostendorff, Heather P.
 Rothschild, Kenneth J.
 Bloch, Donald B.

10 <120> A Method for Diagnosing Primary Biliary Cirrhosis (PBC) Using
 Novel Autoantigens

<130> INOVA-1076610

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15 <150> PCT/US10/51475
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 Ile Val Leu Ala Ala Cys Ser Asp Tyr Phe Cys Ala Met Phe Thr Ser
 30 50 55 60
 Glu Leu Ser Glu Lys Gly Lys Pro Tyr Val Asp Ile Gln Gly Leu Thr
 35 65 70 75 80
 Ala Ser Thr Met Glu Ile Leu Leu Asp Phe Val Tyr Thr Glu Thr Val
 40 85 90 95
 His Val Thr Val Glu Asn Val Gln Glu Leu Leu Pro Ala Ala Cys Leu
 45 100 105 110
 Leu Gln Leu Lys Gly Val Lys Gln Ala Cys Cys Glu Phe Leu Glu Ser
 115 120 125
 Gln Leu Asp Pro Ser Asn Cys Leu Gly Ile Arg Asp Phe Ala Glu Thr
 50 130 135 140
 His Asn Cys Val Asp Leu Met Gln Ala Ala Glu Val Phe Ser Gln Lys
 145 150 155 160
 55 His Phe Pro Glu Val Val Gln His Glu Glu Phe Ile Leu Leu Ser Gln
 165 170 175

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Gly Glu Val Glu Lys Leu Ile Lys Cys Asp Glu Ile Gln Val Asp Ser
 180 185 190
 5 Glu Glu Pro Val Phe Glu Ala Val Ile Asn Trp Val Lys His Ala Lys
 195 200 205
 10 Lys Glu Arg Glu Glu Ser Leu Pro Asn Leu Leu Gln Tyr Val Arg Met
 210 215 220
 Pro Leu Leu Thr Pro Arg Tyr Ile Thr Asp Val Ile Asp Ala Glu Pro
 225 230 235 240
 15 Phe Ile Arg Cys Ser Leu Gln Cys Arg Asp Leu Val Asp Glu Ala Lys
 245 250 255
 20 Lys Phe His Leu Arg Pro Glu Leu Arg Ser Gln Met Gln Gly Pro Arg
 260 265 270
 Thr Arg Ala Arg Leu Gly Ala Asn Glu Val Leu Leu Val Val Gly Gly
 275 280 285
 25 Phe Gly Ser Gln Gln Ser Pro Ile Asp Val Val Glu Lys Tyr Asp Pro
 290 295 300
 30 Lys Thr Gln Glu Trp Ser Phe Leu Pro Ser Ile Thr Arg Lys Arg Arg
 305 310 315 320
 Tyr Val Ala Ser Val Ser Leu His Asp Arg Ile Tyr Val Ile Gly Gly
 325 330 335
 35 Tyr Asp Gly Arg Ser Arg Leu Ser Ser Val Glu Cys Leu Asp Tyr Thr
 340 345 350
 40 Ala Asp Glu Asp Gly Val Trp Tyr Ser Val Ala Pro Met Asn Val Arg
 355 360 365
 45 Arg Gly Leu Ala Gly Ala Thr Thr Leu Gly Asp Met Ile Tyr Val Ser
 370 375 380
 Gly Gly Phe Asp Gly Ser Arg Arg His Thr Ser Met Glu Arg Tyr Asp
 385 390 395 400
 50 Pro Asn Ile Asp Gln Trp Ser Met Leu Gly Asp Met Gln Thr Ala Arg
 405 410 415
 55 Glu Gly Ala Gly Leu Val Val Ala Ser Gly Val Ile Tyr Cys Leu Gly
 420 425 430

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Gly Tyr Asp Gly Leu Asn Ile Leu Asn Ser Val Glu Lys Tyr Asp Pro
435 440 445

5 His Thr Gly His Trp Thr Asn Val Thr Pro Met Ala Thr Lys Arg Ser
450 455 460

10 Gly Ala Gly Val Ala Leu Leu Asn Asp His Ile Tyr Val Val Gly Gly
465 470 475 480

15 Phe Asp Gly Thr Ala His Leu Ser Ser Val Glu Ala Tyr Asn Ile Arg
485 490 495

20 Thr Asp Ser Trp Thr Thr Val Thr Ser Met Thr Thr Pro Arg Cys Tyr
500 505 510

25 Val Gly Ala Thr Val Leu Arg Gly Arg Leu Tyr Ala Ile Ala Gly Tyr
515 520 525

30 Asp Gly Asn Ser Leu Leu Ser Ser Ile Glu Cys Tyr Asp Pro Ile Ile
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35 Asp Ser Trp Glu Val Val Thr Ser Met Gly Thr Gln Arg Cys Asp Ala
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40 Gly Val Cys Val Leu Arg Glu Lys
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<223> The residues in these positions are exogenously added N-terminal
and C-terminal epitope tags as well as vector-derived sequences.

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<223> The residues in these positions are exogenously added N-terminal
and C-terminal epitope tags as well as vector-derived sequences.

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Met Ala Ile Tyr Thr Asp Ile Glu Met Asn Arg Leu Gly Lys Met Ile
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65 Ala Ala Gln Leu Leu Ala Tyr Tyr Phe Thr Glu Leu Lys Asp Asp Gln

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5	Val	Lys	Lys	Ile	Asp	Lys	Tyr	Leu	Tyr	Ala	Met	Arg	Leu	Ser	Asp	Glu			
			35					40					45						
10	Thr	Leu	Ile	Asp	Ile	Met	Thr	Arg	Phe	Arg	Lys	Glu	Met	Lys	Asn	Gly			
		50					55					60							
15	Leu	Ser	Arg	Asp	Phe	Asn	Pro	Thr	Ala	Thr	Val	Lys	Met	Leu	Pro	Thr			
	65					70					75					80			
20	Phe	Val	Arg	Ser	Ile	Pro	Asp	Gly	Ser	Glu	Lys	Gly	Asp	Phe	Ile	Ala			
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25	Leu	Asp	Leu	Gly	Gly	Ser	Ser	Phe	Arg	Ile	Leu	Arg	Val	Gln	Val	Asn			
				100					105					110					
30	His	Glu	Lys	Asn	Gln	Asn	Val	His	Met	Glu	Ser	Glu	Val	Tyr	Asp	Thr			
			115					120					125						
35	Pro	Glu	Asn	Ile	Val	His	Gly	Ser	Gly	Ser	Gln	Leu	Phe	Asp	His	Val			
		130					135					140							
40	Ala	Glu	Cys	Leu	Gly	Asp	Phe	Met	Glu	Lys	Arg	Lys	Ile	Lys	Asp	Lys			
	145					150					155					160			
45	Lys	Leu	Pro	Val	Gly	Phe	Thr	Phe	Ser	Phe	Pro	Cys	Gln	Gln	Ser	Lys			
					165					170					175				
50	Ile	Asp	Glu	Ala	Ile	Leu	Ile	Thr	Trp	Thr	Lys	Arg	Phe	Lys	Ala	Ser			
				180					185					190					
55	Gly	Val	Glu	Gly	Ala	Asp	Val	Val	Lys	Leu	Leu	Asn	Lys	Ala	Ile	Lys			
			195					200					205						
60	Lys	Arg	Gly	Asp	Tyr	Asp	Ala	Asn	Ile	Val	Ala	Val	Val	Asn	Asp	Thr			
		210					215					220							
65	Val	Gly	Thr	Met	Met	Thr	Cys	Gly	Tyr	Asp	Asp	Gln	His	Cys	Glu	Val			
	225					230					235					240			
70	Gly	Leu	Ile	Ile	Gly	Thr	Gly	Thr	Asn	Ala	Cys	Tyr	Met	Glu	Glu	Leu			
					245					250					255				
75	Arg	His	Ile	Asp	Leu	Val	Glu	Gly	Asp	Glu	Gly	Arg	Met	Cys	Ile	Asn			
				260					265					270					

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Thr Glu Trp Gly Ala Phe Gly Asp Asp Gly Ser Leu Glu Asp Ile Arg
 275 280 285

5 Thr Glu Phe Asp Arg Glu Ile Asp Arg Gly Ser Leu Asn Pro Gly Lys
 290 300

10 Gln Leu Phe Glu Lys Met Val Ser Gly Met Tyr Leu Gly Glu Leu Val
 305 310 315 320

Arg Leu Ile Leu Val Lys Met Ala Lys Glu Gly Leu Leu Phe Glu Gly
 325 330 335

15 Arg Ile Thr Pro Glu Leu Leu Thr Arg Gly Lys Phe Asn Thr Ser Asp
 340 345 350

20 Val Ser Ala Ile Glu Lys Asn Lys Glu Gly Leu His Asn Ala Lys Glu
 355 360 365

Ile Leu Thr Arg Leu Gly Val Glu Pro Ser Asp Asp Asp Cys Val Ser
 370 375 380

25 Val Gln His Val Cys Thr Ile Val Ser Phe Arg Ser Ala Asn Leu Val
 385 390 395 400

30 Ala Ala Thr Leu Gly Ala Ile Leu Asn Arg Leu Arg Asp Asn Lys Gly
 405 410 415

35 Thr Pro Arg Leu Arg Thr Thr Val Gly Val Asp Gly Ser Leu Tyr Lys
 420 425 430

Thr His Pro Gln Tyr Ser Arg Arg Phe His Lys Thr Leu Arg Arg Leu
 435 440 445

40 Val Pro Asp Ser Asp Val Arg Phe Leu Leu Ser Glu Ser Gly Ser Gly
 450 455 460

45 Lys Gly Ala Ala Met Val Thr Ala Val Ala Tyr Arg Leu Ala Glu Gln
 465 470 475 480

His Arg Gln Ile Glu Glu Thr Leu Ala His Phe His Leu Thr Lys Asp
 485 490 495

50 Met Leu Leu Glu Val Lys Lys Arg Met Arg Ala Glu Met Glu Leu Gly
 500 505 510

55 Leu Arg Lys Gln Thr His Asn Asn Ala Val Val Lys Met Leu Pro Ser
 515 520 525

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Phe Val Arg Arg Thr Pro Asp Gly Thr Glu Asn Gly Asp Phe Leu Ala
 530 535 540

5
 Leu Asp Leu Gly Gly Thr Asn Phe Arg Val Leu Leu Val Lys Ile Arg
 545 550 555 560

10
 Ser Gly Lys Lys Arg Thr Val Glu Met His Asn Lys Ile Tyr Ala Ile
 565 570 575

15
 Pro Ile Glu Ile Met Gln Gly Thr Gly Glu Glu Leu Phe Asp His Ile
 580 585 590

20
 Val Ser Cys Ile Ser Asp Phe Leu Asp Tyr Met Gly Ile Lys Gly Pro
 595 600 605

25
 Arg Met Pro Leu Gly Phe Thr Phe Ser Phe Pro Cys Gln Gln Thr Ser
 610 615 620

30
 Leu Asp Ala Gly Ile Leu Ile Thr Trp Thr Lys Gly Phe Lys Ala Thr
 625 630 635 640

35
 Asp Cys Val Gly His Asp Val Val Thr Leu Leu Arg Asp Ala Ile Lys
 645 650 655

40
 Arg Arg Glu Glu Phe Asp Leu Asp Val Val Ala Val Val Asn Asp Thr
 660 665 670

45
 Val Gly Thr Met Met Thr Cys Ala Tyr Glu Glu Pro Thr Cys Glu Val
 675 680 685

50
 Gly Leu Ile Val Gly Thr Gly Ser Asn Ala Cys Tyr Met Glu Glu Met
 690 695 700

55
 Lys Asn Val Glu Met Val Glu Gly Asp Gln Gly Gln Met Cys Ile Asn
 705 710 715 720

60
 Met Glu Trp Gly Ala Phe Gly Asp Asn Gly Cys Leu Asp Asp Ile Arg
 725 730 735

65
 Thr His Tyr Asp Arg Leu Val Asp Glu Tyr Ser Leu Asn Ala Gly Lys
 740 745 750

70
 Gln Arg Tyr Glu Lys Met Ile Ser Gly Met Tyr Leu Gly Glu Ile Val
 755 760 765

75
 Arg Asn Ile Leu Ile Asp Phe Thr Lys Lys Gly Phe Leu Phe Arg Gly
 770 775 780

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Gln Ile Ser Glu Thr Leu Lys Thr Arg Gly Ile Phe Glu Thr Lys Phe
 785 790 795 800
 5 Leu Ser Gln Ile Glu Ser Asp Arg Leu Ala Leu Leu Gln Val Arg Ala
 805 810 815
 10 Ile Leu Gln Gln Leu Gly Leu Asn Ser Thr Cys Asp Asp Ser Ile Leu
 820 825 830
 15 Val Lys Thr Val Cys Gly Val Val Ser Arg Arg Ala Ala Gln Leu Cys
 835 840 845
 20 Gly Ala Gly Met Ala Ala Val Val Asp Lys Ile Arg Glu Asn Arg Gly
 850 855 860
 25 Leu Asp Arg Leu Asn Val Thr Val Gly Val Asp Gly Thr Leu Tyr Lys
 865 870 875 880
 30 Leu His Pro His Phe Ser Arg Ile Met His Gln Thr Val Lys Glu Leu
 885 890 895
 35 Ser Pro Lys Cys Asn Val Ser Phe Leu Leu Ser Glu Asp Gly Ser Gly
 900 905 910
 40 Lys Gly Ala Ala Leu Ile Thr Ala Val Gly Val Arg Leu Arg Thr Glu
 915 920 925
 45 Ala Ser Ser Leu Ser Arg Glu Leu Val Asp Pro Asn Ser Val Gln Ala
 930 935 940
 50 Arg Leu Gln Asp Val Asp Gly Thr Ile Asp Thr Arg Ser Lys Leu Ala
 945 950 955 960
 55 Ala Ala Gln Leu Tyr Thr Arg Ala Ser Gln Pro Glu Leu Ala Pro Glu
 965 970 975
 60 Asp Pro Glu Asp Leu Glu His His His His His His
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 75 Met Tyr Thr Asp Ile Glu Met Asn Arg Leu Gly Lys Met Gly Gly Ile
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Met Ala Pro Lys Asp Ile Met Thr Asn Thr His Ala Lys Ser Ile Leu
20 25 30

5 Asn Ser Met Asn Ser Leu Arg Lys Ser Asn Thr Leu Cys Asp Val Thr
35 40 45

10 Leu Arg Val Glu Gln Lys Asp Phe Pro Ala His Arg Ile Val Leu Ala
50 55 60

15 Ala Cys Ser Asp Tyr Phe Cys Ala Met Phe Thr Ser Glu Leu Ser Glu
65 70 75 80

Lys Gly Lys Pro Tyr Val Asp Ile Gln Gly Leu Thr Ala Ser Thr Met
85 90 95

20 Glu Ile Leu Leu Asp Phe Val Tyr Thr Glu Thr Val His Val Thr Val
100 105 110

Glu Asn Val Gln Glu Leu Leu Pro Ala Ala Cys Leu Leu Gln Leu Lys
115 120 125

25 Gly Val Lys Gln Ala Cys Cys Glu Phe Leu Glu Ser Gln Leu Asp Pro
130 135 140

30 Ser Asn Cys Leu Gly Ile Arg Asp Phe Ala Glu Thr His Asn Cys Val
145 150 155 160

35 Asp Leu Met Gln Ala Ala Glu Val Phe Ser Gln Lys His Phe Pro Glu
165 170 175

40 Val Val Gln His Glu Glu Phe Ile Leu Leu Ser Gln Gly Glu Val Glu
180 185 190

Lys Leu Ile Lys Cys Asp Glu Ile Gln Val Asp Ser Glu Glu Pro Val
195 200 205

45 Phe Glu Ala Val Ile Asn Trp Val Lys His Ala Lys Lys Glu Arg Glu
210 215 220

50 Glu Ser Leu Pro Asn Leu Leu Gln Tyr Val Arg Met Pro Leu Leu Thr
225 230 235 240

Pro Arg Tyr Ile Thr Asp Val Ile Asp Ala Glu Pro Phe Ile Arg Cys
245 250 255

55 Ser Leu Gln Cys Arg Asp Leu Val Asp Glu Ala Lys Lys Phe His Leu

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	260					265					270					
5	Arg	Pro	Glu	Leu	Arg	Ser	Gln	Met	Gln	Gly	Pro	Arg	Thr	Arg	Ala	Arg
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10	Leu	Gly	Ala	Asn	Glu	Val	Leu	Leu	Val	Val	Gly	Gly	Phe	Gly	Ser	Gln
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15	Gln	Ser	Pro	Ile	Asp	Val	Val	Glu	Lys	Tyr	Asp	Pro	Lys	Thr	Gln	Glu
	305				310						315				320	
20	Trp	Ser	Phe	Leu	Pro	Ser	Ile	Thr	Arg	Lys	Arg	Arg	Tyr	Val	Ala	Ser
					325					330					335	
25	Val	Ser	Leu	His	Asp	Arg	Ile	Tyr	Val	Ile	Gly	Gly	Tyr	Asp	Gly	Arg
				340					345					350		
30	Ser	Arg	Leu	Ser	Ser	Val	Glu	Cys	Leu	Asp	Tyr	Thr	Ala	Asp	Glu	Asp
			355					360					365			
35	Gly	Val	Trp	Tyr	Ser	Val	Ala	Pro	Met	Asn	Val	Arg	Arg	Gly	Leu	Ala
		370					375					380				
40	Gly	Ala	Thr	Thr	Leu	Gly	Asp	Met	Ile	Tyr	Val	Ser	Gly	Gly	Phe	Asp
	385					390					395					400
45	Gly	Ser	Arg	Arg	His	Thr	Ser	Met	Glu	Arg	Tyr	Asp	Pro	Asn	Ile	Asp
					405					410					415	
50	Gln	Trp	Ser	Met	Leu	Gly	Asp	Met	Gln	Thr	Ala	Arg	Glu	Gly	Ala	Gly
				420					425					430		
55	Leu	Val	Val	Ala	Ser	Gly	Val	Ile	Tyr	Cys	Leu	Gly	Gly	Tyr	Asp	Gly
			435					440					445			
60	Leu	Asn	Ile	Leu	Asn	Ser	Val	Glu	Lys	Tyr	Asp	Pro	His	Thr	Gly	His
	450						455				460					
65	Trp	Thr	Asn	Val	Thr	Pro	Met	Ala	Thr	Lys	Arg	Ser	Gly	Ala	Gly	Val
	465					470					475					480
70	Ala	Leu	Leu	Asn	Asp	His	Ile	Tyr	Val	Val	Gly	Gly	Phe	Asp	Gly	Thr
				485						490					495	
75	Ala	His	Leu	Ser	Ser	Val	Glu	Ala	Tyr	Asn	Ile	Arg	Thr	Asp	Ser	Trp
				500					505					510		

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Thr Thr Val Thr Ser Met Thr Thr Pro Arg Cys Tyr Val Gly Ala Thr
 515 520 525
 5 Val Leu Arg Gly Arg Leu Tyr Ala Ile Ala Gly Tyr Asp Gly Asn Ser
 530 535 540
 10 Leu Leu Ser Ser Ile Glu Cys Tyr Asp Pro Ile Ile Asp Ser Trp Glu
 545 550 555 560
 15 Val Val Thr Ser Met Gly Thr Gln Arg Cys Asp Ala Gly Val Cys Val
 565 570 575
 20 Leu Arg Glu Lys Gln Pro Glu Leu Ala Pro Glu Asp Pro Glu Asp
 580 585 590
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 30 Asp Gln Val Lys Lys Ile Asp Lys Tyr Leu Tyr Ala Met Arg Leu Ser
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 35 Asp Glu Thr Leu Ile Asp Ile Met Thr Arg Phe Arg Lys Glu Met Lys
 35 40 45
 40 Asn Gly Leu Ser Arg Asp Phe Asn Pro Thr Ala Thr Val Lys Met Leu
 50 55 60
 45 Pro Thr Phe Val Arg Ser Ile Pro Asp Gly Ser Glu Lys Gly Asp Phe
 65 70 75 80
 50 Ile Ala Leu Asp Leu Gly Gly Ser Ser Phe Arg Ile Leu Arg Val Gln
 85 90 95
 55 Val Asn His Glu Lys Asn Gln Asn Val His Met Glu Ser Glu Val Tyr
 100 105 110
 Asp Thr Pro Glu Asn Ile Val His Gly Ser Gly Ser Gln Leu Phe Asp
 115 120 125
 60 His Val Ala Glu Cys Leu Gly Asp Phe Met Glu Lys Arg Lys Ile Lys
 130 135 140

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Asp Lys Lys Leu Pro Val Gly Phe Thr Phe Ser Phe Pro Cys Gln Gln
 145 150 155 160
 5 Ser Lys Ile Asp Glu Ala Ile Leu Ile Thr Trp Thr Lys Arg Phe Lys
 165 170 175
 Ala Ser Gly Val Glu Gly Ala Asp Val Val Lys Leu Leu Asn Lys Ala
 10 180 185 190
 Ile Lys Lys Arg Gly Asp Tyr Asp Ala Asn Ile Val Ala Val Val Asn
 15 195 200 205
 Asp Thr Val Gly Thr Met Met Thr Cys Gly Tyr Asp Asp Gln His Cys
 20 210 215 220
 Glu Val Gly Leu Ile Ile Gly Thr Gly Thr Asn Ala Cys Tyr Met Glu
 25 225 230 235 240
 Glu Leu Arg His Ile Asp Leu Val Glu Gly Asp Glu Gly Arg Met Cys
 30 245 250 255
 Ile Asn Thr Glu Trp Gly Ala Phe Gly Asp Asp Gly Ser Leu Glu Asp
 35 260 265 270
 Ile Arg Thr Glu Phe Asp Arg Glu Ile Asp Arg Gly Ser Leu Asn Pro
 40 275 280 285
 Gly Lys Gln Leu Phe Glu Lys Met Val Ser Gly Met Tyr Leu Gly Glu
 45 290 295 300
 Leu Val Arg Leu Ile Leu Val Lys Met Ala Lys Glu Gly Leu Leu Phe
 50 305 310 315 320
 Glu Gly Arg Ile Thr Pro Glu Leu Leu Thr Arg Gly Lys Phe Asn Thr
 55 325 330 335
 Ser Asp Val Ser Ala Ile Glu Lys Asn Lys Glu Gly Leu His Asn Ala
 340 345 350
 Lys Glu Ile Leu Thr Arg Leu Gly Val Glu Pro Ser Asp Asp Asp Cys
 355 360 365
 Val Ser Val Gln His Val Cys Thr Ile Val Ser Phe Arg Ser Ala Asn
 370 375 380
 Leu Val Ala Ala Thr Leu Gly Ala Ile Leu Asn Arg Leu Arg Asp Asn
 385 390 395 400

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Lys Gly Thr Pro Arg Leu Arg Thr Thr Val Gly Val Asp Gly Ser Leu
 405 410 415
 5 Tyr Lys Thr His Pro Gln Tyr Ser Arg Arg Phe His Lys Thr Leu Arg
 420 425 430
 10 Arg Leu Val Pro Asp Ser Asp Val Arg Phe Leu Leu Ser Glu Ser Gly
 435 440 445
 Ser Gly Lys Gly Ala Ala Met Val Thr Ala Val Ala Tyr Arg Leu Ala
 450 455 460
 15 Glu Gln His Arg Gln Ile Glu Glu Thr Leu Ala His Phe His Leu Thr
 465 470 475 480
 20 Lys Asp Met Leu Leu Glu Val Lys Lys Arg Met Arg Ala Glu Met Glu
 485 490 495
 Leu Gly Leu Arg Lys Gln Thr His Asn Asn Ala Val Val Lys Met Leu
 500 505 510
 25 Pro Ser Phe Val Arg Arg Thr Pro Asp Gly Thr Glu Asn Gly Asp Phe
 515 520 525
 30 Leu Ala Leu Asp Leu Gly Gly Thr Asn Phe Arg Val Leu Leu Val Lys
 530 535 540
 35 Ile Arg Ser Gly Lys Lys Arg Thr Val Glu Met His Asn Lys Ile Tyr
 545 550 555 560
 Ala Ile Pro Ile Glu Ile Met Gln Gly Thr Gly Glu Glu Leu Phe Asp
 565 570 575
 40 His Ile Val Ser Cys Ile Ser Asp Phe Leu Asp Tyr Met Gly Ile Lys
 580 585 590
 45 Gly Pro Arg Met Pro Leu Gly Phe Thr Phe Ser Phe Pro Cys Gln Gln
 595 600 605
 Thr Ser Leu Asp Ala Gly Ile Leu Ile Thr Trp Thr Lys Gly Phe Lys
 610 615 620
 50 Ala Thr Asp Cys Val Gly His Asp Val Val Thr Leu Leu Arg Asp Ala
 625 630 635 640
 55 Ile Lys Arg Arg Glu Glu Phe Asp Leu Asp Val Val Ala Val Val Asn
 645 650 655

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Asp Thr Val Gly Thr Met Met Thr Cys Ala Tyr Glu Glu Pro Thr Cys
660 665 670

5 Glu Val Gly Leu Ile Val Gly Thr Gly Ser Asn Ala Cys Tyr Met Glu
675 680 685

10 Glu Met Lys Asn Val Glu Met Val Glu Gly Asp Gln Gly Gln Met Cys
690 695 700

15 Ile Asn Met Glu Trp Gly Ala Phe Gly Asp Asn Gly Cys Leu Asp Asp
705 710 715 720

Ile Arg Thr His Tyr Asp Arg Leu Val Asp Glu Tyr Ser Leu Asn Ala
725 730 735

20 Gly Lys Gln Arg Tyr Glu Lys Met Ile Ser Gly Met Tyr Leu Gly Glu
740 745 750

25 Ile Val Arg Asn Ile Leu Ile Asp Phe Thr Lys Lys Gly Phe Leu Phe
755 760 765

Arg Gly Gln Ile Ser Glu Thr Leu Lys Thr Arg Gly Ile Phe Glu Thr
770 775 780

30 Lys Phe Leu Ser Gln Ile Glu Ser Asp Arg Leu Ala Leu Leu Gln Val
785 790 795 800

35 Arg Ala Ile Leu Gln Gln Leu Gly Leu Asn Ser Thr Cys Asp Asp Ser
805 810 815

Ile Leu Val Lys Thr Val Cys Gly Val Val Ser Arg Arg Ala Ala Gln
820 825 830

40 Leu Cys Gly Ala Gly Met Ala Ala Val Val Asp Lys Ile Arg Glu Asn
835 840 845

45 Arg Gly Leu Asp Arg Leu Asn Val Thr Val Gly Val Asp Gly Thr Leu
850 855 860

50 Tyr Lys Leu His Pro His Phe Ser Arg Ile Met His Gln Thr Val Lys
865 870 875 880

Glu Leu Ser Pro Lys Cys Asn Val Ser Phe Leu Leu Ser Glu Asp Gly
885 890 895

55 Ser Gly Lys Gly Ala Ala Leu Ile Thr Ala Val Gly Val Arg Leu Arg

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5	Lys	Glu	Arg	Glu	Glu	Ser	Leu	Pro	Asn	Leu	Leu	Gln	Tyr	Val	Arg	Met
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10	Pro	Leu	Leu	Thr	Pro	Arg	Tyr	Ile	Thr	Asp	Val	Ile	Asp	Ala	Glu	Pro
	225					230					235					240
15	Phe	Ile	Arg	Cys	Ser	Leu	Gln	Cys	Arg	Asp	Leu	Val	Asp	Glu	Ala	Lys
					245					250					255	
20	Lys	Phe	His	Leu	Arg	Pro	Glu	Leu	Arg	Ser	Gln	Met	Gln	Gly	Pro	Arg
				260					265					270		
25	Thr	Arg	Ala	Arg	Leu	Gly	Ala	Asn	Glu	Val	Leu	Leu	Val	Val	Gly	Gly
			275					280					285			
30	Phe	Gly	Ser	Gln	Gln	Ser	Pro	Ile	Asp	Val	Val	Glu	Lys	Tyr	Asp	Pro
	290						295					300				
35	Lys	Thr	Gln	Glu	Trp	Ser	Phe	Leu	Pro	Ser	Ile	Thr	Arg	Lys	Arg	Arg
	305					310					315					320
40	Tyr	Val	Ala	Ser	Val	Ser	Leu	His	Asp	Arg	Ile	Tyr	Val	Ile	Gly	Gly
					325					330					335	
45	Tyr	Asp	Gly	Arg	Ser	Arg	Leu	Ser	Ser	Val	Glu	Cys	Leu	Asp	Tyr	Thr
				340					345					350		
50	Ala	Asp	Glu	Asp	Gly	Val	Trp	Tyr	Ser	Val	Ala	Pro	Met	Asn	Val	Arg
			355					360					365			
55	Arg	Gly	Leu	Ala	Gly	Ala	Thr	Thr	Leu	Gly	Asp	Met	Ile	Tyr	Val	Ser
		370					375					380				
60	Gly	Gly	Phe	Asp	Gly	Ser	Arg	Arg	His	Thr	Ser	Met	Glu	Arg	Tyr	Asp
	385					390					395					400
65	Pro	Asn	Ile	Asp	Gln	Trp	Ser	Met	Leu	Gly	Asp	Met	Gln	Thr	Ala	Arg
					405					410					415	
70	Glu	Gly	Ala	Gly	Leu	Val	Val	Ala	Ser	Gly	Val	Ile	Tyr	Cys	Leu	Gly
				420					425					430		
75	Gly	Tyr	Asp	Gly	Leu	Asn	Ile	Leu	Asn	Ser	Val	Glu	Lys	Tyr	Asp	Pro
			435					440					445			

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His Thr Gly His Trp Thr Asn Val Thr Pro Met Ala Thr Lys Arg Ser
 450 455 460

5
 Gly Ala Gly Val Ala Leu Leu Asn Asp His Ile Tyr Val Val Gly Gly
 465 470 475 480

10
 Phe Asp Gly Thr Ala His Leu Ser Ser Val Glu Ala Tyr Asn Ile Arg
 485 490 495

15
 Thr Asp Ser Trp Thr Thr Val Thr Ser Met Thr Thr Pro Arg Cys Tyr
 500 505 510

20
 Val Gly Ala Thr Val Leu Arg Gly Arg Leu Tyr Ala Ile Ala Gly Tyr
 515 520 525

25
 Asp Gly Asn Ser Leu Leu Ser Ser Ile Glu Cys Tyr Asp Pro Ile Ile
 530 535 540

30
 Asp Ser Trp Glu Val Val Thr Ser Met Gly Thr Gln Arg Cys Asp Ala
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 Gly Val Cys Val Leu Arg Glu Lys
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 Trp Glu Ile Gly Ile Asp Lys Tyr Leu Tyr Ala Met Arg Leu Ser Asp
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50
 Glu Thr Leu Ile Asp Ile Met Thr Arg Phe Arg Lys Glu Met Lys Asn
 35 40 45

55
 Gly Leu Ser Arg Asp Phe Asn Pro Thr Ala Thr Val Lys Met Leu Pro
 50 55 60

65
 Thr Phe Val Arg Ser Ile Pro Asp Gly Ser Glu Lys Gly Asp Phe Ile
 65 70 75 80

85
 Ala Leu Asp Leu Gly Gly Ser Ser Phe Arg Ile Leu Arg Val Gln Val
 85 90 95

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	Asn	His	Glu	Lys	Asn	Gln	Asn	Val	His	Met	Glu	Ser	Glu	Val	Tyr	Asp
				100					105					110		
5	Thr	Pro	Glu	Asn	Ile	Val	His	Gly	Ser	Gly	Ser	Gln	Leu	Phe	Asp	His
			115					120					125			
10	Val	Ala	Glu	Cys	Leu	Gly	Asp	Phe	Met	Glu	Lys	Arg	Lys	Ile	Lys	Asp
		130					135					140				
15	Lys	Lys	Leu	Pro	Val	Gly	Phe	Thr	Phe	Ser	Phe	Pro	Cys	Gln	Gln	Ser
	145					150					155					160
20	Lys	Ile	Asp	Glu	Ala	Ile	Leu	Ile	Thr	Trp	Thr	Lys	Arg	Phe	Lys	Ala
					165					170					175	
25	Ser	Gly	Val	Glu	Gly	Ala	Asp	Val	Val	Lys	Leu	Leu	Asn	Lys	Ala	Ile
				180					185					190		
30	Lys	Lys	Arg	Gly	Asp	Tyr	Asp	Ala	Asn	Ile	Val	Ala	Val	Val	Asn	Asp
			195					200					205			
35	Thr	Val	Gly	Thr	Met	Met	Thr	Cys	Gly	Tyr	Asp	Asp	Gln	His	Cys	Glu
		210					215					220				
40	Val	Gly	Leu	Ile	Ile	Gly	Thr	Gly	Thr	Asn	Ala	Cys	Tyr	Met	Glu	Glu
	225					230					235					240
45	Leu	Arg	His	Ile	Asp	Leu	Val	Glu	Gly	Asp	Glu	Gly	Arg	Met	Cys	Ile
				245						250					255	
50	Asn	Thr	Glu	Trp	Gly	Ala	Phe	Gly	Asp	Asp	Gly	Ser	Leu	Glu	Asp	Ile
				260					265						270	
55	Arg	Thr	Glu	Phe	Asp	Arg	Glu	Ile	Asp	Arg	Gly	Ser	Leu	Asn	Pro	Gly
			275					280					285			
60	Lys	Gln	Leu	Phe	Glu	Lys	Met	Val	Ser	Gly	Met	Tyr	Leu	Gly	Glu	Leu
		290					295					300				
65	Val	Arg	Leu	Ile	Leu	Val	Lys	Met	Ala	Lys	Glu	Gly	Leu	Leu	Phe	Glu
	305					310					315					320
70	Gly	Arg	Ile	Thr	Pro	Glu	Leu	Leu	Thr	Arg	Gly	Lys	Phe	Asn	Thr	Ser
					325					330					335	
75	Asp	Val	Ser	Ala	Ile	Glu	Lys	Asn	Lys	Glu	Gly	Leu	His	Asn	Ala	Lys
				340					345					350		

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Glu Ile Leu Thr Arg Leu Gly Val Glu Pro Ser Asp Asp Asp Cys Val
 355 360 365
 5 Ser Val Gln His Val Cys Thr Ile Val Ser Phe Arg Ser Ala Asn Leu
 370 375 380
 10 Val Ala Ala Thr Leu Gly Ala Ile Leu Asn Arg Leu Arg Asp Asn Lys
 385 390 395 400
 15 Gly Thr Pro Arg Leu Arg Thr Thr Val Gly Val Asp Gly Ser Leu Tyr
 405 410 415
 20 Lys Thr His Pro Gln Tyr Ser Arg Arg Phe His Lys Thr Leu Arg Arg
 420 425 430
 25 Leu Val Pro Asp Ser Asp Val Arg Phe Leu Leu Ser Glu Ser Gly Ser
 435 440 445
 30 Gly Lys Gly Ala Ala Met Val Thr Ala Val Ala Tyr Arg Leu Ala Glu
 450 455 460
 35 Gln His Arg Gln Ile Glu Glu Thr Leu Ala His Phe His Leu Thr Lys
 465 470 475 480
 40 Asp Met Leu Leu Glu Val Lys Lys Arg Met Arg Ala Glu Met Glu Leu
 485 490 495
 45 Gly Leu Arg Lys Gln Thr His Asn Asn Ala Val Val Lys Met Leu Pro
 500 505 510
 50 Ser Phe Val Arg Arg Thr Pro Asp Gly Thr Glu Asn Gly Asp Phe Leu
 515 520 525
 55 Ala Leu Asp Leu Gly Gly Thr Asn Phe Arg Val Leu Leu Val Lys Ile
 530 535 540
 60 Arg Ser Gly Lys Lys Arg Thr Val Glu Met His Asn Lys Ile Tyr Ala
 545 550 555 560
 65 Ile Pro Ile Glu Ile Met Gln Gly Thr Gly Glu Glu Leu Phe Asp His
 565 570 575
 70 Ile Val Ser Cys Ile Ser Asp Phe Leu Asp Tyr Met Gly Ile Lys Gly
 580 585 590
 75 Pro Arg Met Pro Leu Gly Phe Thr Phe Ser Phe Pro Cys Gln Gln Thr
 595 600 605

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Ser Leu Asp Ala Gly Ile Leu Ile Thr Trp Thr Lys Gly Phe Lys Ala
 610 615 620

5
 Thr Asp Cys Val Gly His Asp Val Val Thr Leu Leu Arg Asp Ala Ile
 625 630 635 640

10
 Lys Arg Arg Glu Glu Phe Asp Leu Asp Val Val Ala Val Val Asn Asp
 645 650 655

15
 Thr Val Gly Thr Met Met Thr Cys Ala Tyr Glu Glu Pro Thr Cys Glu
 660 665 670

20
 Val Gly Leu Ile Val Gly Thr Gly Ser Asn Ala Cys Tyr Met Glu Glu
 675 680 685

25
 Met Lys Asn Val Glu Met Val Glu Gly Asp Gln Gly Gln Met Cys Ile
 690 695 700

30
 Asn Met Glu Trp Gly Ala Phe Gly Asp Asn Gly Cys Leu Asp Asp Ile
 705 710 715 720

35
 Arg Thr His Tyr Asp Arg Leu Val Asp Glu Tyr Ser Leu Asn Ala Gly
 725 730 735

40
 Lys Gln Arg Tyr Glu Lys Met Ile Ser Gly Met Tyr Leu Gly Glu Ile
 740 745 750

45
 Val Arg Asn Ile Leu Ile Asp Phe Thr Lys Lys Gly Phe Leu Phe Arg
 755 760 765

50
 Gly Gln Ile Ser Glu Thr Leu Lys Thr Arg Gly Ile Phe Glu Thr Lys
 770 775 780

55
 Phe Leu Ser Gln Ile Glu Ser Asp Arg Leu Ala Leu Leu Gln Val Arg
 785 790 795 800

60
 Ala Ile Leu Gln Gln Leu Gly Leu Asn Ser Thr Cys Asp Asp Ser Ile
 805 810 815

65
 Leu Val Lys Thr Val Cys Gly Val Val Ser Arg Arg Ala Ala Gln Leu
 820 825 830

70
 Cys Gly Ala Gly Met Ala Ala Val Val Asp Lys Ile Arg Glu Asn Arg
 835 840 845

75
 Gly Leu Asp Arg Leu Asn Val Thr Val Gly Val Asp Gly Thr Leu Tyr

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	850		855		860												
5	Lys	Leu	His	Pro	His	Phe	Ser	Arg	Ile	Met	His	Gln	Thr	Val	Lys	Glu	
	865					870					875					880	
	Leu	Ser	Pro	Lys	Cys	Asn	Val	Ser	Phe	Leu	Leu	Ser	Glu	Asp	Gly	Ser	
10					885					890					895		
	Gly	Lys	Gly	Ala	Ala	Leu	Ile	Thr	Ala	Val	Gly	Val	Arg	Leu	Arg	Thr	
				900					905					910			
15	Glu	Ala	Ser	Ser													
			915														
	<210>	8															
	<211>	917															
20	<212>	PRT															
	<213>	Homo sapiens															
	<400>	8															
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	1				5					10					15		
	Asp	Gln	Val	Gln	Lys	Val	Asp	Gln	Tyr	Leu	Tyr	His	Met	Arg	Leu	Ser	
30				20					25					30			
	Asp	Glu	Thr	Leu	Leu	Glu	Ile	Ser	Lys	Arg	Phe	Arg	Lys	Glu	Met	Glu	
			35					40					45				
35	Lys	Gly	Leu	Gly	Ala	Thr	Thr	His	Pro	Thr	Ala	Ala	Val	Lys	Met	Leu	
	50						55					60					
	Pro	Thr	Phe	Val	Arg	Ser	Thr	Pro	Asp	Gly	Thr	Glu	His	Gly	Glu	Phe	
40	65					70					75					80	
	Leu	Ala	Leu	Asp	Leu	Gly	Gly	Thr	Asn	Phe	Arg	Val	Leu	Trp	Val	Lys	
				85						90					95		
45	Val	Thr	Asp	Asn	Gly	Leu	Gln	Lys	Val	Glu	Met	Glu	Asn	Gln	Ile	Tyr	
				100					105					110			
	Ala	Ile	Pro	Glu	Asp	Ile	Met	Arg	Gly	Ser	Gly	Thr	Gln	Leu	Phe	Asp	
50			115					120					125				
	His	Ile	Ala	Glu	Cys	Leu	Ala	Asn	Phe	Met	Asp	Lys	Leu	Gln	Ile	Lys	
	130						135					140					
55	Asp	Lys	Lys	Leu	Pro	Leu	Gly	Phe	Thr	Phe	Ser	Phe	Pro	Cys	His	Gln	

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Lys Gly Glu Glu Arg Leu Arg Ser Thr Ile Gly Val Asp Gly Ser Val
 405 410 415
 5 Tyr Lys Lys His Pro His Phe Ala Lys Arg Leu His Lys Thr Val Arg
 420 425 430
 10 Arg Leu Val Pro Gly Cys Asp Val Arg Phe Leu Arg Ser Glu Asp Gly
 435 440 445
 Ser Gly Lys Gly Ala Ala Met Val Thr Ala Val Ala Tyr Arg Leu Ala
 450 455 460
 15 Asp Gln His Arg Ala Arg Gln Lys Thr Leu Glu His Leu Gln Leu Ser
 465 470 475 480
 20 His Asp Gln Leu Leu Glu Val Lys Arg Arg Met Lys Val Glu Met Glu
 485 490 495
 Arg Gly Leu Ser Lys Glu Thr His Ala Ser Ala Pro Val Lys Met Leu
 500 505 510
 25 Pro Thr Tyr Val Cys Ala Thr Pro Asp Gly Thr Glu Lys Gly Asp Phe
 515 520 525
 30 Leu Ala Leu Asp Leu Gly Gly Thr Asn Phe Arg Val Leu Leu Val Arg
 530 535 540
 35 Val Arg Asn Gly Lys Trp Gly Gly Val Glu Met His Asn Lys Ile Tyr
 545 550 555 560
 Ala Ile Pro Gln Glu Val Met His Gly Thr Gly Asp Glu Leu Phe Asp
 565 570 575
 40 His Ile Val Gln Cys Ile Ala Asp Phe Leu Glu Tyr Met Gly Met Lys
 580 585 590
 45 Gly Val Ser Leu Pro Leu Gly Phe Thr Phe Ser Phe Pro Cys Gln Gln
 595 600 605
 50 Asn Ser Leu Asp Glu Ser Ile Leu Leu Lys Trp Thr Lys Gly Phe Lys
 610 615 620
 Ala Ser Gly Cys Glu Gly Glu Asp Val Val Thr Leu Leu Lys Glu Ala
 625 630 635 640
 55 Ile His Arg Arg Glu Glu Phe Asp Leu Asp Val Val Ala Val Val Asn
 645 650 655

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Asp Thr Val Gly Thr Met Met Thr Cys Gly Phe Glu Asp Pro His Cys
 660 665 670
 5
 Glu Val Gly Leu Ile Val Gly Thr Gly Ser Asn Ala Cys Tyr Met Glu
 675 680 685
 10
 Glu Met Arg Asn Val Glu Leu Val Glu Gly Glu Glu Gly Arg Met Cys
 690 695 700
 Val Asn Met Glu Trp Gly Ala Phe Gly Asp Asn Gly Cys Leu Asp Asp
 705 710 715 720
 15
 Phe Arg Thr Glu Phe Asp Val Ala Val Asp Glu Leu Ser Leu Asn Pro
 725 730 735
 20
 Gly Lys Gln Arg Phe Glu Lys Met Ile Ser Gly Met Tyr Leu Gly Glu
 740 745 750
 Ile Val Arg Asn Ile Leu Ile Asp Phe Thr Lys Arg Gly Leu Leu Phe
 755 760 765
 25
 Arg Gly Arg Ile Ser Glu Arg Leu Lys Thr Arg Gly Ile Phe Glu Thr
 770 775 780
 30
 Lys Phe Leu Ser Gln Ile Glu Ser Asp Cys Leu Ala Leu Leu Gln Val
 785 790 795 800
 35
 Arg Ala Ile Leu Gln His Leu Gly Leu Glu Ser Thr Cys Asp Asp Ser
 805 810 815
 Ile Ile Val Lys Glu Val Cys Thr Val Val Ala Arg Arg Ala Ala Gln
 820 825 830
 40
 Leu Cys Gly Ala Gly Met Ala Ala Val Val Asp Arg Ile Arg Glu Asn
 835 840 845
 45
 Arg Gly Leu Asp Ala Leu Lys Val Thr Val Gly Val Asp Gly Thr Leu
 850 855 860
 Tyr Lys Leu His Pro His Phe Ala Lys Val Met His Glu Thr Val Lys
 865 870 875 880
 50
 Asp Leu Ala Pro Lys Cys Asp Val Ser Phe Leu Gln Ser Glu Asp Gly
 885 890 895
 55
 Ser Gly Lys Gly Ala Ala Leu Ile Thr Ala Val Ala Cys Arg Ile Arg
 900 905 910

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Glu Ala Gly Gln Arg
915

5 <210> 9
<211> 923
<212> PRT
<213> Homo sapiens

10 <400> 9

Met Asp Ser Ile Gly Ser Ser Gly Leu Arg Gln Gly Glu Glu Thr Leu
1 5 10 15

15 Ser Cys Ser Glu Glu Gly Leu Pro Gly Pro Ser Asp Ser Ser Glu Leu
20 25 30

20 Val Gln Glu Cys Leu Gln Gln Phe Lys Val Thr Arg Ala Gln Leu Gln
35 40 45

Gln Ile Gln Ala Ser Leu Leu Gly Ser Met Glu Gln Ala Leu Arg Gly
50 55 60

25 Gln Ala Ser Pro Ala Pro Ala Val Arg Met Leu Pro Thr Tyr Val Gly
65 70 75 80

30 Ser Thr Pro His Gly Thr Glu Gln Gly Asp Phe Val Val Leu Glu Leu
85 90 95

35 Gly Ala Thr Gly Ala Ser Leu Arg Val Leu Trp Val Thr Leu Thr Gly
100 105 110

Ile Glu Gly His Arg Val Glu Pro Arg Ser Gln Glu Phe Val Ile Pro
115 120 125

40 Gln Glu Val Met Leu Gly Ala Gly Gln Gln Leu Phe Asp Phe Ala Ala
130 135 140

45 His Cys Leu Ser Glu Phe Leu Asp Ala Gln Pro Val Asn Lys Gln Gly
145 150 155 160

50 Leu Gln Leu Gly Phe Ser Phe Ser Phe Pro Cys His Gln Thr Gly Leu
165 170 175

Asp Arg Ser Thr Leu Ile Ser Trp Thr Lys Gly Phe Arg Cys Ser Gly
180 185 190

55 Val Glu Gly Gln Asp Val Val Gln Leu Leu Arg Asp Ala Ile Arg Arg
195 200 205

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Gln Gly Ala Tyr Asn Ile Asp Val Val Ala Val Val Asn Asp Thr Val
 210 215 220

5
 Gly Thr Met Met Gly Cys Glu Pro Gly Val Arg Pro Cys Glu Val Gly
 225 230 235 240

10
 Leu Val Val Asp Thr Gly Thr Asn Ala Cys Tyr Met Glu Glu Ala Arg
 245 250 255

15
 His Val Ala Val Leu Asp Glu Asp Arg Gly Arg Val Cys Val Ser Val
 260 265 270

20
 Glu Trp Gly Ser Phe Ser Asp Asp Gly Ala Leu Gly Pro Val Leu Thr
 275 280 285

25
 Thr Phe Asp His Thr Leu Asp His Glu Ser Leu Asn Pro Gly Ala Gln
 290 295 300

30
 Arg Phe Glu Lys Met Ile Gly Gly Leu Tyr Leu Gly Glu Leu Val Arg
 305 310 315 320

35
 Leu Val Leu Ala His Leu Ala Arg Cys Gly Val Leu Phe Gly Gly Cys
 325 330 335

40
 Thr Ser Pro Ala Leu Leu Ser Gln Gly Ser Ile Leu Leu Glu His Val
 340 345 350

45
 Ala Glu Met Glu Asp Pro Ser Thr Gly Ala Ala Arg Val His Ala Ile
 355 360 365

50
 Leu Gln Asp Leu Gly Leu Ser Pro Gly Ala Ser Asp Val Glu Leu Val
 370 375 380

55
 Gln His Val Cys Ala Ala Val Cys Thr Arg Ala Ala Gln Leu Cys Ala
 385 390 395 400

60
 Ala Ala Leu Ala Ala Val Leu Ser Cys Leu Gln His Ser Arg Glu Gln
 405 410 415

65
 Gln Thr Leu Gln Val Ala Val Ala Thr Gly Gly Arg Val Cys Glu Arg
 420 425 430

70
 His Pro Arg Phe Cys Ser Val Leu Gln Gly Thr Val Met Leu Leu Ala
 435 440 445

75
 Pro Glu Cys Asp Val Ser Leu Ile Pro Ser Val Asp Gly Gly Gly Arg

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	450		455		460												
5	Gly 465	Val	Ala	Met	Val	Thr 470	Ala	Val	Ala	Ala	Arg 475	Leu	Ala	Ala	His	Arg 480	
10	Arg	Leu	Leu	Glu	Glu 485	Thr	Leu	Ala	Pro	Phe 490	Arg	Leu	Asn	His	Asp 495	Gln	
15	Leu	Ala	Ala	Val	Gln 500	Ala	Gln	Met	Arg 505	Lys	Ala	Met	Ala	Lys 510	Gly	Leu	
20	Arg	Gly	Glu	Ala	Ser	Ser	Leu	Arg 520	Met	Leu	Pro	Thr	Phe 525	Val	Arg	Ala	
25	Thr	Pro 530	Asp	Gly	Ser	Glu	Arg 535	Gly	Asp	Phe	Leu	Ala 540	Leu	Asp	Leu	Gly	
30	Gly 545	Thr	Asn	Phe	Arg 550	Val	Leu	Leu	Val	Arg 555	Val	Thr	Thr	Gly	Val	Gln 560	
35	Ile	Thr	Ser	Glu	Ile 565	Tyr	Ser	Ile	Pro	Glu 570	Thr	Val	Ala	Gln	Gly 575	Ser	
40	Gly	Gln	Gln	Leu	Phe 580	Asp	His	Ile	Val 585	Asp	Cys	Ile	Val	Asp 590	Phe	Gln	
45	Gln	Lys	Gln	Gly	Leu	Ser	Gly	Gln 600	Ser	Leu	Pro	Leu	Gly 605	Phe	Thr	Phe	
50	Ser	Phe 610	Pro	Cys	Arg	Gln	Leu 615	Gly	Leu	Asp	Gln	Gly 620	Ile	Leu	Leu	Asn	
55	Trp 625	Thr	Lys	Gly	Phe 630	Lys	Ala	Ser	Asp	Cys	Glu 635	Gly	Gln	Asp	Val	Val 640	
60	Ser	Leu	Leu	Arg	Glu 645	Ala	Ile	Thr	Arg	Arg 650	Gln	Ala	Val	Glu	Leu 655	Asn	
65	Val	Val	Ala	Ile	Val 660	Asn	Asp	Thr	Val 665	Gly	Thr	Met	Met	Ser	Cys 670	Gly	
70	Tyr	Glu	Asp 675	Pro	Arg	Cys	Glu	Ile 680	Gly	Leu	Ile	Val	Gly 685	Thr	Gly	Thr	
75	Asn 690	Ala	Cys	Tyr	Met	Glu	Glu 695	Leu	Arg	Asn	Val	Ala 700	Gly	Val	Pro	Gly	

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Asp Ser Gly Arg Met Cys Ile Asn Met Glu Trp Gly Ala Phe Gly Asp
 705 710 715 720
 5 Asp Gly Ser Leu Ala Met Leu Ser Thr Arg Phe Asp Ala Ser Val Asp
 725 730 735
 10 Gln Ala Ser Ile Asn Pro Gly Lys Gln Arg Phe Glu Lys Met Ile Ser
 740 745 750
 Gly Met Tyr Leu Gly Glu Ile Val Arg His Ile Leu Leu His Leu Thr
 755 760 765
 15 Ser Leu Gly Val Leu Phe Arg Gly Gln Gln Ile Gln Arg Leu Gln Thr
 770 775 780
 20 Arg Asp Ile Phe Lys Thr Lys Phe Leu Ser Glu Ile Glu Ser Asp Ser
 785 790 795 800
 25 Leu Ala Leu Arg Gln Val Arg Ala Ile Leu Glu Asp Leu Gly Leu Pro
 805 810 815
 Leu Thr Ser Asp Asp Ala Leu Met Val Leu Glu Val Cys Gln Ala Val
 820 825 830
 30 Ser Gln Arg Ala Ala Gln Leu Cys Gly Ala Gly Val Ala Ala Val Val
 835 840 845
 35 Glu Lys Ile Arg Glu Asn Arg Gly Leu Glu Glu Leu Ala Val Ser Val
 850 855 860
 Gly Val Asp Gly Thr Leu Tyr Lys Leu His Pro Arg Phe Ser Ser Leu
 865 870 875 880
 40 Val Ala Ala Thr Val Arg Glu Leu Ala Pro Arg Cys Val Val Thr Phe
 885 890 895
 45 Leu Gln Ser Glu Asp Gly Ser Gly Lys Gly Ala Ala Leu Val Thr Ala
 900 905 910
 50 Val Ala Cys Arg Leu Ala Gln Leu Thr Arg Val
 915 920
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 <211> 466
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 55 <400> 10

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1 Met Ala Met Asp Val Thr Arg Ser Gln Ala Gln Thr Ala Leu Thr Leu
 5 Val Glu Gln Ile Leu Ala Glu Phe Gln Leu Gln Glu Glu Asp Leu Lys
 10 Lys Val Met Arg Arg Met Gln Lys Glu Met Asp Arg Gly Leu Arg Leu
 15 Glu Thr His Glu Glu Ala Ser Val Lys Met Leu Pro Thr Tyr Val Arg
 20 Ser Thr Pro Glu Gly Ser Glu Val Gly Asp Phe Leu Ser Leu Asp Leu
 25 Gly Gly Thr Asn Phe Arg Val Met Leu Val Lys Val Gly Glu Gly Glu
 30 Glu Gly Gln Trp Ser Val Lys Thr Lys His Gln Met Tyr Ser Ile Pro
 35 Glu Asp Ala Met Thr Gly Thr Ala Glu Met Leu Phe Asp Tyr Ile Ser
 40 Glu Cys Ile Ser Asp Phe Leu Asp Lys His Gln Met Lys His Lys Lys
 45 Leu Pro Leu Gly Phe Thr Phe Ser Phe Pro Val Arg His Glu Asp Ile
 50 Asp Lys Gly Ile Leu Leu Asn Trp Thr Lys Gly Phe Lys Ala Ser Gly
 55 Ala Glu Gly Asn Asn Val Val Gly Leu Leu Arg Asp Ala Ile Lys Arg
 60 Arg Gly Asp Phe Glu Met Asp Val Val Ala Met Val Asn Asp Thr Val
 65 Ala Thr Met Ile Ser Cys Tyr Tyr Glu Asp His Gln Cys Glu Val Gly
 70 Met Ile Val Gly Thr Gly Cys Asn Ala Cys Tyr Met Glu Glu Met Gln
 75 Asn Val Glu Leu Val Glu Gly Asp Glu Gly Arg Met Cys Val Asn Thr
 80

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Glu Trp Gly Ala Phe Gly Asp Ser Gly Glu Leu Asp Glu Phe Leu Leu
 260 265 270
 5 Glu Tyr Asp Arg Leu Val Asp Glu Ser Ser Ala Asn Pro Gly Gln Gln
 275 280 285
 10 Leu Tyr Glu Lys Leu Ile Gly Gly Lys Tyr Met Gly Glu Leu Val Arg
 290 295 300
 Leu Val Leu Leu Arg Leu Val Asp Glu Asn Leu Leu Phe His Gly Glu
 305 310 315 320
 15 Ala Ser Glu Gln Leu Arg Thr Arg Gly Ala Phe Glu Thr Arg Phe Val
 325 330 335
 20 Ser Gln Val Glu Ser Asp Thr Gly Asp Arg Lys Gln Ile Tyr Asn Ile
 340 345 350
 Leu Ser Thr Leu Gly Leu Arg Pro Ser Thr Thr Asp Cys Asp Ile Val
 355 360 365
 25 Arg Arg Ala Cys Glu Ser Val Ser Thr Arg Ala Ala His Met Cys Ser
 370 375 380
 30 Ala Gly Leu Ala Gly Val Ile Asn Arg Met Arg Glu Ser Arg Ser Glu
 385 390 395 400
 35 Asp Val Met Arg Ile Thr Val Gly Val Asp Gly Ser Val Tyr Lys Leu
 405 410 415
 His Pro Ser Phe Lys Glu Arg Phe His Ala Ser Val Arg Arg Leu Thr
 420 425 430
 40 Pro Ser Cys Glu Ile Thr Phe Ile Glu Ser Glu Glu Gly Ser Gly Arg
 435 440 445
 45 Gly Ala Ala Leu Val Ser Ala Val Ala Cys Lys Lys Ala Cys Met Leu
 450 455 460
 Gly Gln
 50 465
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 55 <213> Homo sapiens
 <400> 11

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1 Met Phe Ala Val His Leu Met Ala Phe Tyr Phe Ser Lys Leu Lys Glu
 5
 5 Asp Gln Ile Lys Lys Val Asp Arg Phe Leu Tyr His Met Arg Leu Ser
 20
 10 Asp Asp Thr Leu Leu Asp Ile Met Arg Arg Phe Arg Ala Glu Met Glu
 35
 15 Lys Gly Leu Ala Lys Asp Thr Asn Pro Thr Ala Ala Val Lys Met Leu
 50
 20 Pro Thr Phe Val Arg Ala Ile Pro Asp Gly Ser Glu Asn Gly Glu Phe
 65
 25 Leu Ser Leu Asp Leu Gly Gly Ser Lys Phe Arg Val Leu Lys Val Gln
 85
 30 Val Ala Glu Glu Gly Lys Arg His Val Gln Met Glu Ser Gln Phe Tyr
 100
 35 Pro Thr Pro Asn Glu Ile Ile Arg Gly Asn Gly Thr Glu Leu Phe Glu
 115
 40 Tyr Val Ala Asp Cys Leu Ala Asp Phe Met Lys Thr Lys Asp Leu Lys
 130
 45 His Lys Lys Leu Pro Leu Gly Leu Thr Phe Ser Phe Pro Cys Arg Gln
 145
 50 Thr Lys Leu Glu Glu Gly Val Leu Leu Ser Trp Thr Lys Lys Phe Lys
 165
 55 Ala Arg Gly Val Gln Asp Thr Asp Val Val Ser Arg Leu Thr Lys Ala
 180
 60 Met Arg Arg His Lys Asp Met Asp Val Asp Ile Leu Ala Leu Val Asn
 195
 65 Asp Thr Val Gly Thr Met Met Thr Cys Ala Tyr Asp Asp Pro Tyr Cys
 210
 70 Glu Val Gly Val Ile Ile Gly Thr Gly Thr Asn Ala Cys Tyr Met Glu
 225
 75 Asp Met Ser Asn Ile Asp Leu Val Glu Gly Asp Glu Gly Arg Met Cys
 245

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Ile Asn Thr Glu Trp Gly Ala Phe Gly Asp Asp Gly Ala Leu Glu Asp
 260 265 270

5 Ile Arg Thr Glu Phe Asp Arg Glu Leu Asp Leu Gly Ser Leu Asn Pro
 275 280 285

10 Gly Lys Gln Leu Phe Glu Lys Met Ile Ser Gly Leu Tyr Leu Gly Glu
 290 295 300

15 Leu Val Arg Leu Ile Leu Leu Lys Met Ala Lys Ala Gly Leu Leu Phe
 305 310 315 320

Gly Gly Glu Lys Ser Ser Ala Leu His Thr Lys Gly Lys Ile Glu Thr
 325 330 335

20 Arg His Val Ala Ala Met Glu Lys Tyr Lys Glu Gly Leu Ala Asn Thr
 340 345 350

25 Arg Glu Ile Leu Val Asp Leu Gly Leu Glu Pro Ser Glu Ala Asp Cys
 355 360 365

Ile Ala Val Gln His Val Cys Thr Ile Val Ser Phe Arg Ser Ala Asn
 370 375 380

30 Leu Cys Ala Ala Ala Leu Ala Ala Ile Leu Thr Arg Leu Arg Glu Asn
 385 390 395 400

35 Lys Lys Val Glu Arg Leu Arg Thr Thr Val Gly Met Asp Gly Thr Leu
 405 410 415

Tyr Lys Ile His Pro Gln Tyr Pro Lys Arg Leu His Lys Val Val Arg
 420 425 430

40 Lys Leu Val Pro Ser Cys Asp Val Arg Phe Leu Leu Ser Glu Ser Gly
 435 440 445

45 Ser Thr Lys Gly Ala Ala Met Val Thr Ala Val Ala Ser Arg Val Gln
 450 455 460

50 Ala Gln Arg Lys Gln Ile Asp Arg Val Leu Ala Leu Phe Gln Leu Thr
 465 470 475 480

Arg Glu Gln Leu Val Asp Val Gln Ala Lys Met Arg Ala Glu Leu Glu
 485 490 495

55 Tyr Gly Leu Lys Lys Lys Ser His Gly Leu Ala Thr Val Arg Met Leu

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Val Arg Gln Ile Leu Ile Asp Leu Thr Lys Gln Gly Leu Leu Phe Arg
755 760 765

5 Gly Gln Ile Ser Glu Arg Leu Arg Thr Arg Gly Ile Phe Glu Thr Lys
770 775 780

10 Phe Leu Ser Gln Ile Glu Ser Asp Arg Leu Ala Leu Leu Gln Val Arg
785 790 795 800

15 Arg Ile Leu Gln Gln Leu Gly Leu Asp Ser Thr Cys Glu Asp Ser Ile
805 810 815

20 Val Val Lys Glu Val Cys Gly Ala Val Ser Arg Arg Ala Ala Gln Leu
820 825 830

25 Cys Gly Ala Gly Leu Ala Ala Ile Val Glu Lys Arg Arg Glu Asp Gln
835 840 845

30 Gly Leu Glu His Leu Arg Ile Thr Val Gly Val Asp Gly Thr Leu Tyr
850 855 860

35 Lys Leu His Pro His Phe Ser Arg Ile Leu Gln Glu Thr Val Lys Glu
865 870 875 880

40 Leu Ala Pro Arg Cys Asp Val Thr Phe Met Leu Ser Glu Asp Gly Ser
885 890 895

45 Gly Lys Gly Ala Ala Leu Ile Thr Ala Val Ala Lys Arg Leu Gln Gln
900 905 910

Ala Gln Lys Glu Asn
915

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<213> Homo sapiens

<400> 12

50 Met Gly Gly Ile Met Ala Pro Lys Asp Ile Met Thr Asn Thr His Ala
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Lys Ser Ile Leu Asn Ser Met Asn Ser Leu Arg Lys Ser Asn Thr Leu
20 25 30

55 Cys Asp Val Thr Leu Arg Val Glu Gln Lys Asp Phe Pro Ala His Arg
35 40 45

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Ile Val Leu Ala Ala Cys Ser Asp Tyr Phe Cys Ala Met Phe Thr Ser
50 55 60

5 Glu Leu Ser Glu Lys Gly Lys Pro Tyr Val Asp Ile Gln Gly Leu Thr
65 70 75 80

Ala Ser Thr Met Glu Ile Leu Leu Asp Phe Val Tyr Thr Glu Thr Val
10 85 90 95

His Val Thr Val Glu Asn Val Gln Glu Leu Leu Pro Ala Ala Cys Leu
100 105 110

15 Leu Gln Leu Lys Gly Val Lys Gln Ala Cys Cys Glu Phe Leu Glu Ser
115 120 125

20 Gln Leu Asp Pro Ser Asn Cys Leu Gly Ile Arg Asp Phe Ala Glu Thr
130 135 140

His Asn Cys Val Asp Leu Met Gln Ala Ala Glu Val Phe Ser Gln Lys
145 150 155 160

25 His Phe Pro Glu Val Val Gln His Glu Glu Phe Ile Leu Leu Ser Gln
165 170 175

30 Gly Glu Val Glu Lys Leu Ile Lys Cys Asp Glu Ile Gln Val Asp Ser
180 185 190

Glu Glu Pro Val Phe Glu Ala Val Ile Asn Trp Val Lys His Ala Lys
35 195 200 205

Lys Glu Arg Glu Glu Ser Leu Pro Asn Leu Leu Gln Tyr Val Arg Met
210 215 220

40 Pro Leu Leu Thr Pro Arg Tyr Ile Thr Asp Val Ile Asp Ala Glu Pro
225 230 235 240

Phe Ile Arg Cys Ser Leu Gln Cys Arg Asp Leu Val Asp Glu Ala Lys
45 245 250 255

Lys Phe His Leu Arg Pro Glu Leu Arg Ser Gln Met Gln Gly Pro Arg
50 260 265 270

Thr Arg Ala Arg Leu Gly Ala Asn Glu Val Leu Leu Val Val Gly Gly
275 280 285

55 Phe Gly Ser Gln Gln Ser Pro Ile Asp Val Val Glu Lys Tyr Asp Pro
290 295 300

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Lys Thr Gln Glu Trp Ser Phe Leu Pro Ser Ile Thr Arg Lys Arg Arg
 305 310 315 320
 5 Tyr Val Ala Ser Val Ser Leu His Asp Arg Ile Tyr Val Ile Gly Gly
 325 330 335
 10 Tyr Asp Gly Arg Ser Arg Leu Ser Ser Val Glu Cys Leu Asp Tyr Thr
 340 345 350
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 370 375 380
 25 Gly Gly Phe Asp Gly Ser Arg Arg His Thr Ser Met Glu Arg Tyr Asp
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 405 410 415
 35 Glu Gly Ala Gly Leu Val Val Ala Ser Gly Val Ile Tyr Cys Leu Gly
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 40 Gly Tyr Asp Gly Leu Asn Ile Leu Asn Ser Val Glu Lys Tyr Asp Pro
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 45 His Thr Gly His Trp Thr Asn Val Thr Pro Met Ala Thr Lys Arg Ser
 450 455 460
 50 Gly Ala Gly Val Ala Leu Leu Asn Asp His Ile Tyr Val Val Gly Gly
 465 470 475 480
 55 Phe Asp Gly Thr Ala His Leu Ser Ser Val Glu Ala Tyr Asn Ile Arg
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 60 Thr Asp Ser Trp Thr Thr Val Thr Ser Met Thr Thr Pro Arg Cys Tyr
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 65 Val Gly Ala Thr Val Leu Arg Gly Arg Leu Tyr Ala Ile Ala Gly Tyr
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 70 Asp Gly Asn Ser Leu Leu Ser Ser Ile Glu Cys Tyr Asp Pro Ile Ile
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 75 Asp Ser Trp Glu Val Val Thr Ser Met Gly Thr Gln Arg Cys Asp Ala
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Leu Pro Glu Gly Val Pro Gln Pro Ala Arg Met Pro Tyr Ile Ser Asp
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Lys His Pro Arg Gln Thr Leu Glu Val Ile Asn Leu Leu Arg Lys His
50 55 60

25 Arg Glu Leu Cys Asp Val Val Leu Val Val Gly Ala Lys Lys Ile Tyr
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30 Ala His Arg Val Ile Leu Ser Ala Cys Ser Pro Tyr Phe Arg Ala Met
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Phe Thr Gly Glu Leu Ala Glu Ser Arg Gln Thr Glu Val Val Ile Arg
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Asp Ile Asp Glu Arg Ala Met Glu Leu Leu Ile Asp Phe Ala Tyr Thr
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40 Ser Gln Ile Thr Val Glu Glu Gly Asn Val Gln Thr Leu Leu Pro Ala
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45 Ala Cys Leu Leu Gln Leu Ala Glu Ile Gln Glu Ala Cys Cys Glu Phe
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Leu Lys Arg Gln Leu Asp Pro Ser Asn Cys Leu Gly Ile Arg Ala Phe
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Ala Asp Thr His Ser Cys Arg Glu Leu Leu Arg Ile Ala Asp Lys Phe
180 185 190

55 Thr Gln His Asn Phe Gln Glu Val Met Glu Ser Glu Glu Phe Met Leu
195 200 205

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Leu Pro Ala Asn Gln Leu Ile Asp Ile Ile Ser Ser Asp Glu Leu Asn
 210 215 220

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 Val Arg Ser Glu Glu Gln Val Phe Asn Ala Val Met Ala Trp Val Lys
 225 230 235 240

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 Tyr Ser Ile Gln Glu Arg Arg Pro Gln Leu Pro Gln Val Leu Gln His
 245 250 255

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 Val Arg Leu Pro Leu Leu Ser Pro Lys Phe Leu Val Gly Thr Val Gly
 260 265 270

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 Ser Asp Pro Leu Ile Lys Ser Asp Glu Glu Cys Arg Asp Leu Val Asp
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 Glu Ala Lys Asn Tyr Leu Leu Leu Pro Gln Glu Arg Pro Leu Met Gln
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 Gly Pro Arg Thr Arg Pro Arg Lys Pro Ile Arg Cys Gly Glu Val Leu
 305 310 315 320

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 Phe Ala Val Gly Gly Trp Cys Ser Gly Asp Ala Ile Ser Ser Val Glu
 325 330 335

40
 Arg Tyr Asp Pro Gln Thr Asn Glu Trp Arg Met Val Ala Ser Met Ser
 340 345 350

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 Lys Arg Arg Cys Gly Val Gly Val Ser Val Leu Asp Asp Leu Leu Tyr
 355 360 365

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 Ala Val Gly Gly His Asp Gly Ser Ser Tyr Leu Asn Ser Val Glu Arg
 370 375 380

55
 Tyr Asp Pro Lys Thr Asn Gln Trp Ser Ser Asp Val Ala Pro Thr Ser
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Thr Cys Arg Thr Ser Val Gly Val Ala Val Leu Gly Gly Phe Leu Tyr
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Ala Val Gly Gly Gln Asp Gly Val Ser Cys Leu Asn Ile Val Glu Arg
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Tyr Asp Pro Lys Glu Asn Lys Trp Thr Arg Val Ala Ser Met Ser Thr
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Arg Arg Leu Gly Val Ala Val Ala Val Leu Gly Gly Phe Leu Tyr Ala

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40	Gln	His	Phe	Pro 180	Glu	Val	Met	Leu	Gly 185	Glu	Glu	Phe	Leu	Ser 190	Leu	Ser
45	Leu	Asp	Gln 195	Val	Cys	Ser	Leu	Ile 200	Ser	Ser	Asp	Lys	Leu 205	Thr	Val	Ser
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Val Val Gly Gly Gln Ala Pro Lys Ala Ile Arg Ser Val Glu Cys Tyr
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5 Asp Phe Glu Glu Asp Arg Trp Asp Gln Ile Ala Glu Leu Pro Ser Arg
325 330 335

10 Arg Cys Arg Ala Gly Val Val Phe Met Ala Gly His Val Tyr Ala Val
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15 Gly Gly Phe Asn Gly Ser Leu Arg Val Arg Thr Val Asp Val Tyr Asp
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20 Gly Val Lys Asp Gln Trp Thr Ser Ile Ala Ser Met Gln Glu Arg Arg
370 375 380

25 Ser Thr Leu Gly Ala Ala Val Leu Asn Asp Leu Leu Tyr Ala Val Gly
385 390 395 400

30 Gly Phe Asp Gly Ser Thr Gly Leu Ala Ser Val Glu Ala Tyr Ser Tyr
405 410 415

35 Lys Thr Asn Glu Trp Phe Phe Val Ala Pro Met Asn Thr Arg Arg Ser
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40 Ser Val Gly Val Gly Val Val Glu Gly Lys Leu Tyr Ala Val Gly Gly
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45 Tyr Asp Gly Ala Ser Arg Gln Cys Leu Ser Thr Val Glu Gln Tyr Asn
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50 Pro Ala Thr Asn Glu Trp Ile Tyr Val Ala Asp Met Ser Thr Arg Arg
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55 Ser Gly Ala Gly Val Gly Val Leu Ser Gly Gln Leu Tyr Ala Thr Gly
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60 Gly His Asp Gly Pro Leu Val Arg Lys Ser Val Glu Val Tyr Asp Pro
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65 Gly Thr Asn Thr Trp Lys Gln Val Ala Asp Met Asn Met Cys Arg Arg
515 520 525

70 Asn Ala Gly Val Cys Ala Val Asn Gly Leu Leu Tyr Val Val Gly Gly
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75 Asp Asp Gly Ser Cys Asn Leu Ala Ser Val Glu Tyr Tyr Asn Pro Val
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 30 Pro Ala Ala Pro Met Glu Gly Ala Val Gln Leu Leu Ser Arg Glu Gly
 50 55 60
 35 His Ser Val Ala His Asn Ser Lys Arg His Tyr His Asp Ala Phe Val
 65 70 75 80
 40 Ala Met Ser Arg Met Arg Gln Arg Gly Leu Leu Cys Asp Ile Val Leu
 85 90 95
 45 His Val Ala Ala Lys Glu Ile Arg Ala His Lys Val Val Leu Ala Ser
 100 105 110
 50 Cys Ser Pro Tyr Phe His Ala Met Phe Thr Asn Glu Met Ser Glu Ser
 115 120 125
 55 Arg Gln Thr His Val Thr Leu His Asp Ile Asp Pro Gln Ala Leu Asp
 130 135 140
 60 Gln Leu Val Gln Phe Ala Tyr Thr Ala Glu Ile Val Val Gly Glu Gly
 145 150 155 160
 65 Asn Val Gln Thr Leu Leu Pro Ala Ala Ser Leu Leu Gln Leu Asn Gly
 165 170 175
 70 Val Arg Asp Ala Cys Cys Lys Phe Leu Leu Ser Gln Leu Asp Pro Ser
 180 185 190

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Asn Cys Leu Gly Ile Arg Gly Phe Ala Asp Ala His Ser Cys Ser Asp
 195 200 205

5 Leu Leu Lys Ala Ala His Arg Tyr Val Leu Gln His Phe Val Asp Val
 210 215 220

Ala Lys Thr Glu Glu Phe Met Leu Leu Pro Leu Lys Gln Val Leu Glu
 225 230 235 240

10 Leu Val Ser Ser Asp Ser Leu Asn Val Pro Ser Glu Glu Glu Val Tyr
 245 250 255

15 Arg Ala Val Leu Ser Trp Val Lys His Asp Val Asp Ala Arg Arg Gln
 260 265 270

His Val Pro Arg Leu Met Lys Cys Val Arg Leu Pro Leu Leu Ser Arg
 275 280 285

20 Asp Phe Leu Leu Gly His Val Asp Ala Glu Ser Leu Val Arg His His
 290 295 300

25 Pro Asp Cys Lys Asp Leu Leu Ile Glu Ala Leu Lys Phe His Leu Leu
 305 310 315 320

30 Pro Glu Gln Arg Gly Val Leu Gly Thr Ser Arg Thr Arg Pro Arg Arg
 325 330 335

Cys Glu Gly Ala Gly Pro Val Leu Phe Ala Val Gly Gly Gly Ser Leu
 340 345 350

35 Phe Ala Ile His Gly Asp Cys Glu Ala Tyr Asp Thr Arg Thr Asp Arg
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40 Trp His Val Val Ala Ser Met Ser Thr Arg Arg Ala Arg Val Gly Val
 370 375 380

Ala Ala Val Gly Asn Arg Leu Tyr Ala Val Gly Gly Tyr Asp Gly Thr
 385 390 395 400

45 Ser Asp Leu Ala Thr Val Glu Ser Tyr Asp Pro Val Thr Asn Thr Trp
 405 410 415

50 Gln Pro Glu Val Ser Met Gly Thr Arg Arg Ser Cys Leu Gly Val Ala
 420 425 430

Ala Leu His Gly Leu Leu Tyr Ser Ala Gly Gly Tyr Asp Gly Ala Ser
 435 440 445

55

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Cys Leu Asn Ser Ala Glu Arg Tyr Asp Pro Leu Thr Gly Thr Trp Thr
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 5 Ser Val Ala Ala Met Ser Thr Arg Arg Arg Tyr Val Arg Val Ala Thr
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 10 Leu Asp Gly Asn Leu Tyr Ala Val Gly Gly Tyr Asp Ser Ser Ser His
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 15 Leu Ala Thr Val Glu Lys Tyr Glu Pro Gln Val Asn Val Trp Ser Pro
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 20 Val Ala Ser Met Leu Ser Arg Arg Ser Ser Ala Gly Val Ala Val Leu
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5 His Cys Pro Val Thr Val Asn Pro Trp His Met Lys Lys Ala Phe Lys
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10 Val Met Asn Glu Leu Arg Ser Gln Asn Leu Leu Cys Asp Val Thr Ile
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15 Val Ala Glu Asp Met Glu Ile Ser Ala His Arg Val Val Leu Ala Ala
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20 Cys Ser Pro Tyr Phe His Ala Met Phe Thr Gly Glu Met Ser Glu Ser
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25 Arg Ala Lys Arg Val Arg Ile Lys Glu Val Asp Gly Trp Thr Leu Arg
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30 Met Leu Ile Asp Tyr Val Tyr Thr Ala Glu Ile Gln Val Thr Glu Glu
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35 Asn Val Gln Val Leu Leu Pro Ala Ala Gly Leu Leu Gln Leu Gln Asp
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40 Val Lys Lys Thr Cys Cys Glu Phe Leu Glu Ser Gln Leu His Pro Val
 145 150 155 160

45 Asn Cys Leu Gly Ile Arg Ala Phe Ala Asp Met His Ala Cys Thr Asp
 165 170 175

50 Leu Leu Asn Lys Ala Asn Thr Tyr Ala Glu Gln His Phe Ala Asp Val
 180 185 190

55 Val Leu Ser Glu Glu Phe Leu Asn Leu Gly Ile Glu Gln Val Cys Ser
 195 200 205

60 Leu Ile Ser Ser Asp Lys Leu Thr Ile Ser Ser Glu Glu Lys Val Phe
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65 Glu Ala Val Ile Ala Trp Val Asn His Asp Lys Asp Val Arg Gln Glu
 225 230 235 240

70 Phe Met Ala Arg Leu Met Glu His Val Arg Leu Pro Leu Leu Pro Arg
 245 250 255

75 Glu Tyr Leu Val Gln Arg Val Glu Glu Glu Ala Leu Val Lys Asn Ser

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	260					265					270					
5	Ser	Ala	Cys	Lys	Asp	Tyr	Leu	Ile	Glu	Ala	Met	Lys	Tyr	His	Leu	Leu
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15	Thr	Pro	Met	Asn	Leu	Pro	Lys	Leu	Met	Val	Val	Val	Gly	Gly	Gln	Ala
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20	Pro	Lys	Ala	Ile	Arg	Ser	Val	Glu	Cys	Tyr	Asp	Phe	Lys	Glu	Glu	Arg
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25	Trp	His	Gln	Val	Ala	Glu	Leu	Pro	Ser	Arg	Arg	Cys	Arg	Ala	Gly	Met
				340					345					350		
30	Val	Tyr	Met	Ala	Gly	Leu	Val	Phe	Ala	Val	Gly	Gly	Phe	Asn	Gly	Ser
			355					360					365			
35	Leu	Arg	Val	Arg	Thr	Val	Asp	Ser	Tyr	Asp	Pro	Val	Lys	Asp	Gln	Trp
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40	Thr	Ser	Val	Ala	Asn	Met	Arg	Asp	Arg	Arg	Ser	Thr	Leu	Gly	Ala	Ala
	385					390					395					400
45	Val	Leu	Asn	Gly	Leu	Leu	Tyr	Ala	Val	Gly	Gly	Phe	Asp	Gly	Ser	Thr
					405					410					415	
50	Gly	Leu	Ser	Ser	Val	Glu	Ala	Tyr	Asn	Ile	Lys	Ser	Asn	Glu	Trp	Phe
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55	His	Val	Ala	Pro	Met	Asn	Thr	Arg	Arg	Ser	Ser	Val	Gly	Val	Gly	Val
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65	Gln	Cys	Leu	Ser	Thr	Val	Glu	Cys	Tyr	Asn	Ala	Thr	Thr	Asn	Glu	Trp
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70	Thr	Tyr	Ile	Ala	Glu	Met	Ser	Thr	Arg	Arg	Ser	Gly	Ala	Gly	Val	Gly
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75	Val	Leu	Asn	Asn	Leu	Leu	Tyr	Ala	Val	Gly	Gly	His	Asp	Gly	Pro	Leu
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Val Arg Lys Ser Val Glu Val Tyr Asp Pro Thr Thr Asn Ala Trp Arg
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5 Gln Val Ala Asp Met Asn Met Cys Arg Arg Asn Ala Gly Val Cys Ala
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10 Val Asn Gly Leu Leu Tyr Val Val Gly Gly Asp Asp Gly Ser Cys Asn
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15 Leu Ala Ser Val Glu Tyr Tyr Asn Pro Thr Thr Asp Lys Trp Thr Val
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Ile Asp Lys Pro Leu
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40 Leu Arg Ser Gln Asn Leu Leu Cys Asp Val Thr Ile Val Ala Glu Asp
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45 Met Glu Ile Ser Ala His Arg Val Val Leu Ala Ala Cys Ser Pro Tyr
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Phe His Ala Met Phe Thr Gly Glu Met Ser Glu Ser Arg Ala Lys Arg
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50 Val Arg Ile Lys Glu Val Asp Gly Trp Thr Leu Arg Met Leu Ile Asp
100 105 110

55 Tyr Val Tyr Thr Ala Glu Ile Gln Val Thr Glu Glu Asn Val Gln Val
115 120 125

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Leu Leu Pro Ala Ala Gly Leu Leu Gln Leu Gln Asp Val Lys Lys Thr
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 5 Cys Cys Glu Phe Leu Glu Ser Gln Leu His Pro Val Asn Cys Leu Gly
 145 150 155 160
 10 Ile Arg Ala Phe Ala Asp Met His Ala Cys Thr Asp Leu Leu Asn Lys
 165 170 175
 Ala Asn Thr Tyr Ala Glu Gln His Phe Ala Asp Val Val Leu Ser Glu
 180 185 190
 15 Glu Phe Leu Asn Leu Gly Ile Glu Gln Val Cys Ser Leu Ile Ser Ser
 195 200 205
 20 Asp Lys Leu Thr Ile Ser Ser Glu Glu Lys Val Phe Glu Ala Val Ile
 210 215 220
 25 Ala Trp Val Asn His Asp Lys Asp Val Arg Gln Glu Phe Met Ala Arg
 225 230 235 240
 Leu Met Glu His Val Arg Leu Pro Leu Leu Pro Arg Glu Tyr Leu Val
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 30 Gln Arg Val Glu Glu Glu Ala Leu Val Lys Asn Ser Ser Ala Cys Lys
 260 265 270
 35 Asp Tyr Leu Ile Glu Ala Met Lys Tyr His Leu Leu Pro Thr Glu Gln
 275 280 285
 Arg Ile Leu Met Lys Ser Val Arg Thr Arg Leu Arg Thr Pro Met Asn
 290 295 300
 40 Leu Pro Lys Leu Met Val Val Val Gly Gly Gln Ala Pro Lys Ala Ile
 305 310 315 320
 45 Arg Ser Val Glu Cys Tyr Asp Phe Lys Glu Glu Arg Trp His Gln Val
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 50 Ala Glu Leu Pro Ser Arg Arg Cys Arg Ala Gly Met Val Tyr Met Ala
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 Gly Leu Val Phe Ala Val Gly Gly Phe Asn Gly Ser Leu Arg Val Arg
 355 360 365
 55 Thr Val Asp Ser Tyr Asp Pro Val Lys Asp Gln Trp Thr Ser Val Ala
 370 375 380

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Asn Met Arg Asp Arg Arg Ser Thr Leu Gly Ala Ala Val Leu Asn Gly
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 Leu Leu Tyr Ala Val Gly Gly Phe Asp Gly Ser Thr Gly Leu Ser Ser
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 Val Glu Ala Tyr Asn Ile Lys Ser Asn Glu Trp Phe His Val Ala Pro
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 Met Asn Thr Arg Arg Ser Ser Val Gly Val Gly Val Val Gly Gly Leu
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 Leu Tyr Ala Val Gly Gly Tyr Asp Gly Ala Ser Arg Gln Cys Leu Ser
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 Thr Val Glu Cys Tyr Asn Ala Thr Thr Asn Glu Trp Thr Tyr Ile Ala
 465 470 475 480

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 Glu Met Ser Thr Arg Arg Ser Gly Ala Gly Val Gly Val Leu Asn Asn
 485 490 495

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 Leu Leu Tyr Ala Val Gly Gly His Asp Gly Pro Leu Val Arg Lys Ser
 500 505 510

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 Val Glu Val Tyr Asp Pro Thr Thr Asn Ala Trp Arg Gln Val Ala Asp
 515 520 525

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 Met Asn Met Cys Arg Arg Asn Ala Gly Val Cys Ala Val Asn Gly Leu
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50

 Leu Tyr Val Val Gly Gly Asp Asp Gly Ser Cys Asn Leu Ala Ser Val
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Met Ser Thr Gly Arg Ser Tyr Ala Gly Val Thr Val Ile Asp Lys Pro
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Leu

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<400> 18

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 Arg Gln Gly Lys Leu Cys Asp Val Thr Leu Lys Ile Gly Asp His Lys
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 Phe Ser Ala His Arg Ile Val Leu Ala Ala Ser Ile Pro Tyr Phe His
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 Ala Met Phe Thr Asn Asp Met Met Glu Cys Lys Gln Asp Glu Ile Val
 65 70 75 80
 Met Gln Gly Met Asp Pro Ser Ala Leu Glu Ala Leu Ile Asn Phe Ala
 20 85 90 95
 Tyr Asn Gly Asn Leu Ala Ile Asp Gln Gln Asn Val Gln Ser Leu Leu
 100 105 110
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 Met Gly Ala Ser Phe Leu Gln Leu Gln Ser Ile Lys Asp Ala Cys Cys
 115 120 125
 Thr Phe Leu Arg Glu Arg Leu His Pro Lys Asn Cys Leu Gly Val Arg
 130 135 140
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 Gln Phe Ala Glu Thr Met Met Cys Ala Val Leu Tyr Asp Ala Ala Asn
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 Ser Phe Ile His Gln His Phe Val Glu Val Ser Met Ser Glu Glu Phe
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 180 185 190
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 Leu Asn Val Lys Ser Glu Glu Gln Val Phe Glu Ala Ala Leu Ala Trp
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 Val Arg Tyr Asp Arg Glu Gln Arg Gly Pro Tyr Leu Pro Glu Leu Leu
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 Ser Asn Ile Arg Leu Pro Leu Cys Arg Pro Gln Phe Leu Ser Asp Arg
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 Val Gln Gln Asp Asp Leu Val Arg Cys Cys His Lys Cys Arg Asp Leu
 245 250 255

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Val Asp Glu Ala Lys Asp Tyr His Leu Met Pro Glu Arg Arg Pro His
 260 265 270

5 Leu Pro Ala Phe Arg Thr Arg Pro Arg Cys Cys Thr Ser Ile Ala Gly
 275 280 285

10 Leu Ile Tyr Ala Val Gly Gly Leu Asn Ser Ala Gly Asp Ser Leu Asn
 290 295 300

15 Val Val Glu Val Phe Asp Pro Ile Ala Asn Cys Trp Glu Arg Cys Arg
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20 Pro Met Thr Thr Ala Arg Ser Arg Val Gly Val Ala Val Val Asn Gly
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25 Leu Leu Tyr Ala Ile Gly Gly Tyr Asp Gly Gln Leu Arg Leu Ser Thr
 340 345 350

30 Val Glu Ala Tyr Asn Pro Glu Thr Asp Thr Trp Thr Arg Val Gly Ser
 355 360 365

35 Met Asn Ser Lys Arg Ser Ala Met Gly Thr Val Val Leu Asp Gly Gln
 370 375 380

40 Ile Tyr Val Cys Gly Gly Tyr Asp Gly Asn Ser Ser Leu Ser Ser Val
 385 390 395 400

45 Glu Thr Tyr Ser Pro Glu Thr Asp Lys Trp Thr Val Val Thr Ser Met
 405 410 415

50 Ser Ser Asn Arg Ser Ala Ala Gly Val Thr Val Phe Glu Gly Arg Ile
 420 425 430

55 Tyr Val Ser Gly Gly His Asp Gly Leu Gln Ile Phe Ser Ser Val Glu
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60 His Tyr Asn His His Thr Ala Thr Trp His Pro Ala Ala Gly Met Leu
 450 455 460

65 Asn Lys Arg Cys Arg His Gly Ala Ala Ser Leu Gly Ser Lys Met Phe
 465 470 475 480

70 Val Cys Gly Gly Tyr Asp Gly Ser Gly Phe Leu Ser Ile Ala Glu Met
 485 490 495

75 Tyr Ser Ser Val Ala Asp Gln Trp Cys Leu Ile Val Pro Met His Thr

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500 505 510

5 Arg Arg Ser Arg Val Ser Leu Val Ala Ser Cys Gly Arg Leu Tyr Ala
515 520 525

10 Val Gly Gly Tyr Asp Gly Gln Ser Asn Leu Ser Ser Val Glu Met Tyr
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25 <220>
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<400> 19

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35 <213> Artificial Sequence

<220>
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40 <400> 20

Gln Pro Glu Leu Ala Pro Glu Asp Pro Glu Asp
1 5 10

45 **Claims**

1. A method of diagnosing primary biliary cirrhosis (PBC) in an individual comprising:
 - 50 a. contacting a test sample from the individual with one or more target antigens, each comprising an epitope of kelch-like 12 or of a homolog of kelch-like 12; and
 - b. detecting binding of the one or more target antigens to one or more antibodies in the test sample, wherein the presence of the one or more antibodies bound to the one or more target antigens is indicative of primary biliary cirrhosis (PBC).
- 55 2. The method of claim 1, wherein the one or more target antigens are immobilized on a solid support.
3. The method of claim 1, wherein the test sample is cells, tissues or body fluids.

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4. The method of claim 1, wherein the test sample is blood, plasma or serum.
5. The method of claim 1, wherein the test sample is contacted with two or more of the target antigens.
- 5 6. The method of claim 1, wherein the test sample is contacted with three or more of the target antigens.
7. The method of claim 1, wherein the test sample is contacted with four or more of the target antigens.
8. The method of claim 1, wherein the test sample is contacted with five or more of the target antigens.
- 10 9. The method of any one of claims 1 to 8, wherein said kelch-like 12 or homolog of kelch-like 12 comprises a sequence selected from the group consisting of SEQ ID NOS. 12, 13, 14, 15, 16, 17 and 18.
- 15 10. The method of any one of claims 1 to 8, wherein said target antigen comprises a sequence selected from the group consisting of SEQ ID NOS. 12, 13, 14, 15, 16, 17 and 18.
11. The method of claim 10, wherein said target antigen comprises the sequence of SEQ ID NO. 12.
12. The method of any one of claims 1 to 11, wherein said target antigen comprises a recombinant antigen.
- 20 13. The method of any one of claims 1 to 12, wherein said target antigen further comprises a tag sequence.
14. The method of claim 13, wherein said tag sequence is located at the C-terminal or N-terminal, or at both the C-terminal and N-terminal of said target antigen.
- 25 15. The method of any one of claims 1 to 14, wherein said detection step further comprises a labeled anti-immunoglobulin (IG) antibody and wherein the target antigen, antibody, and anti-IG antibody form a complex.

30

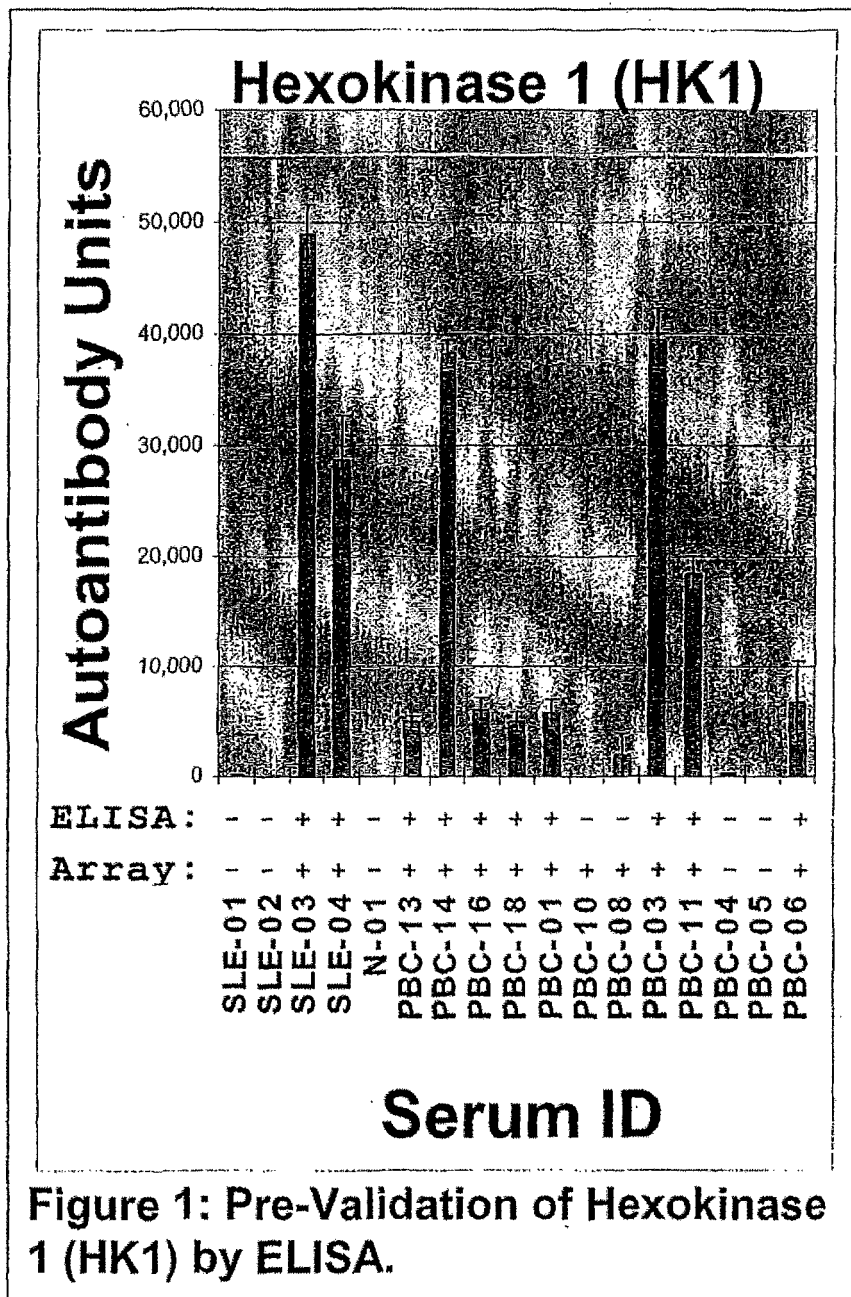
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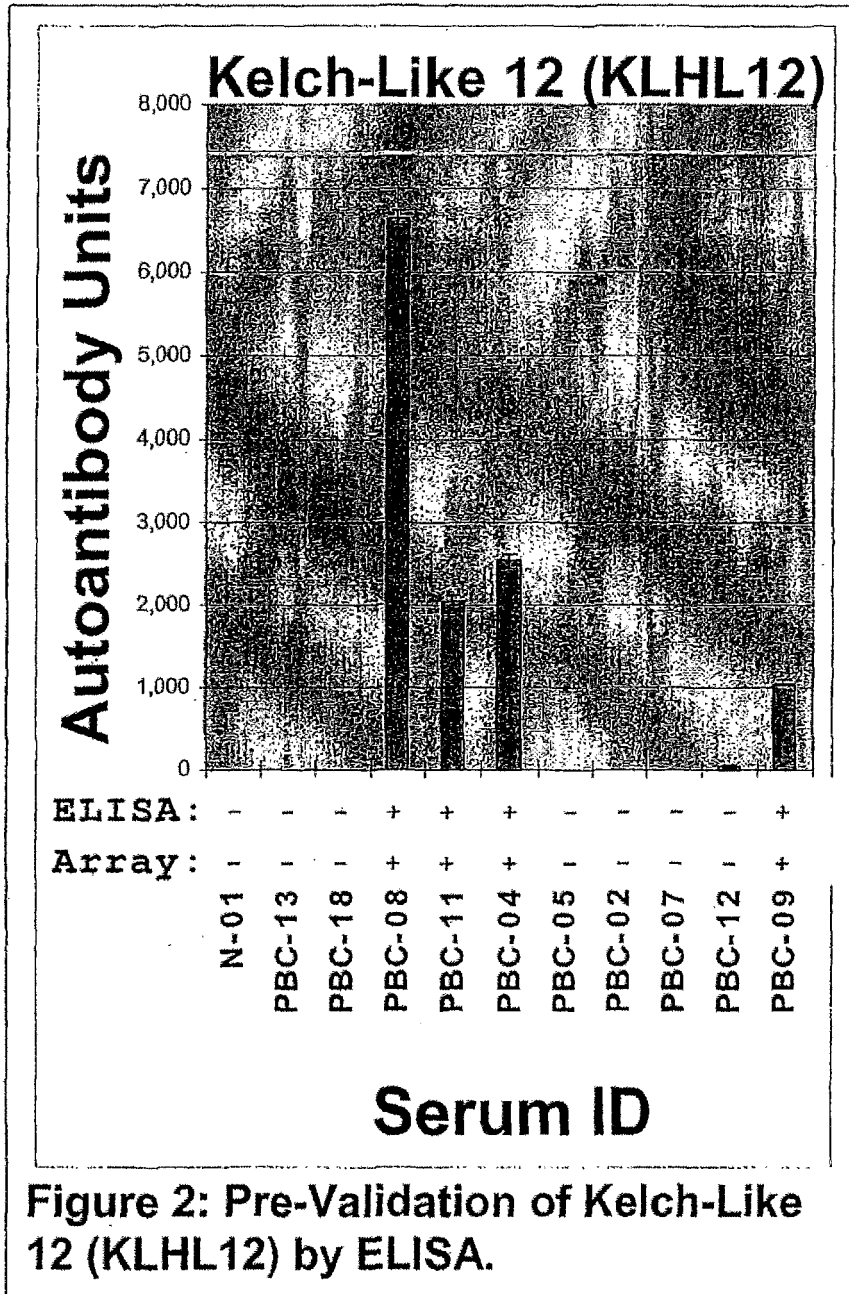
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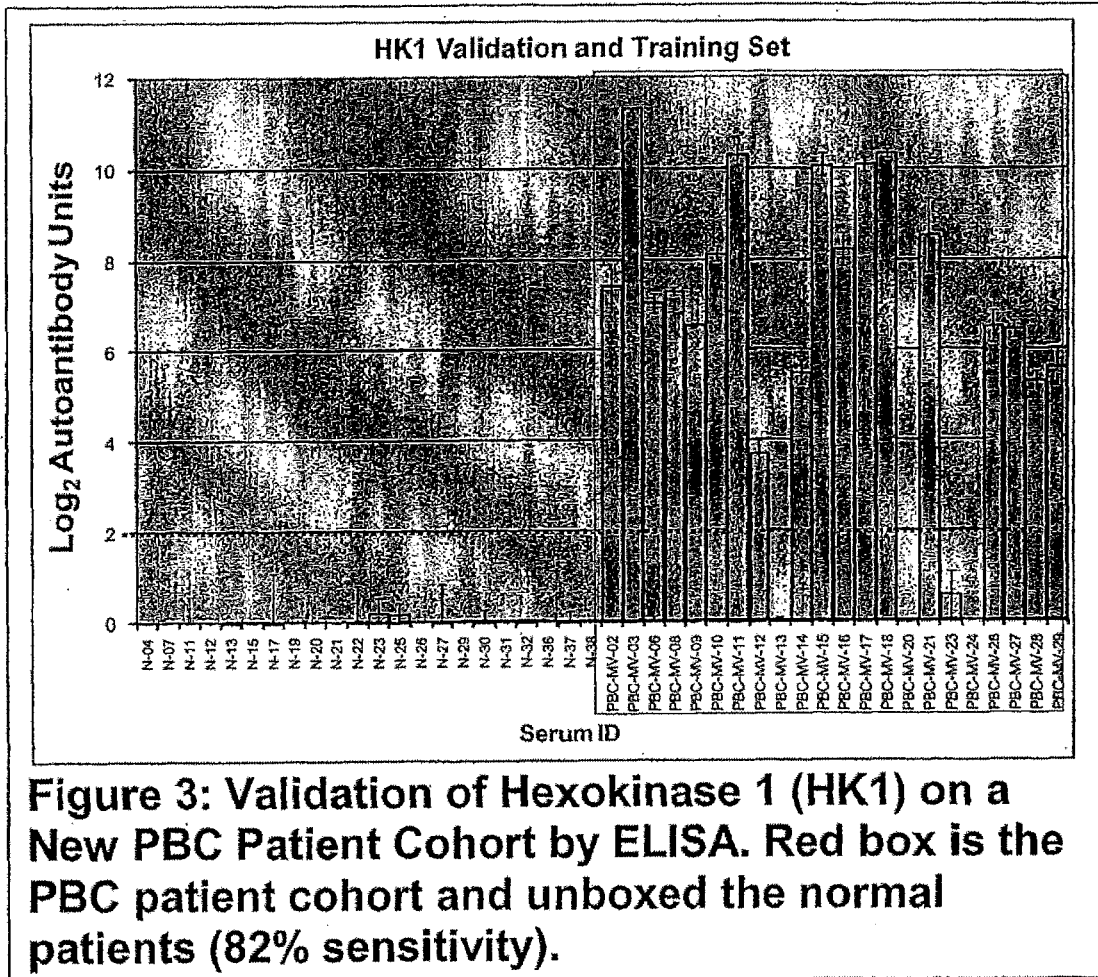
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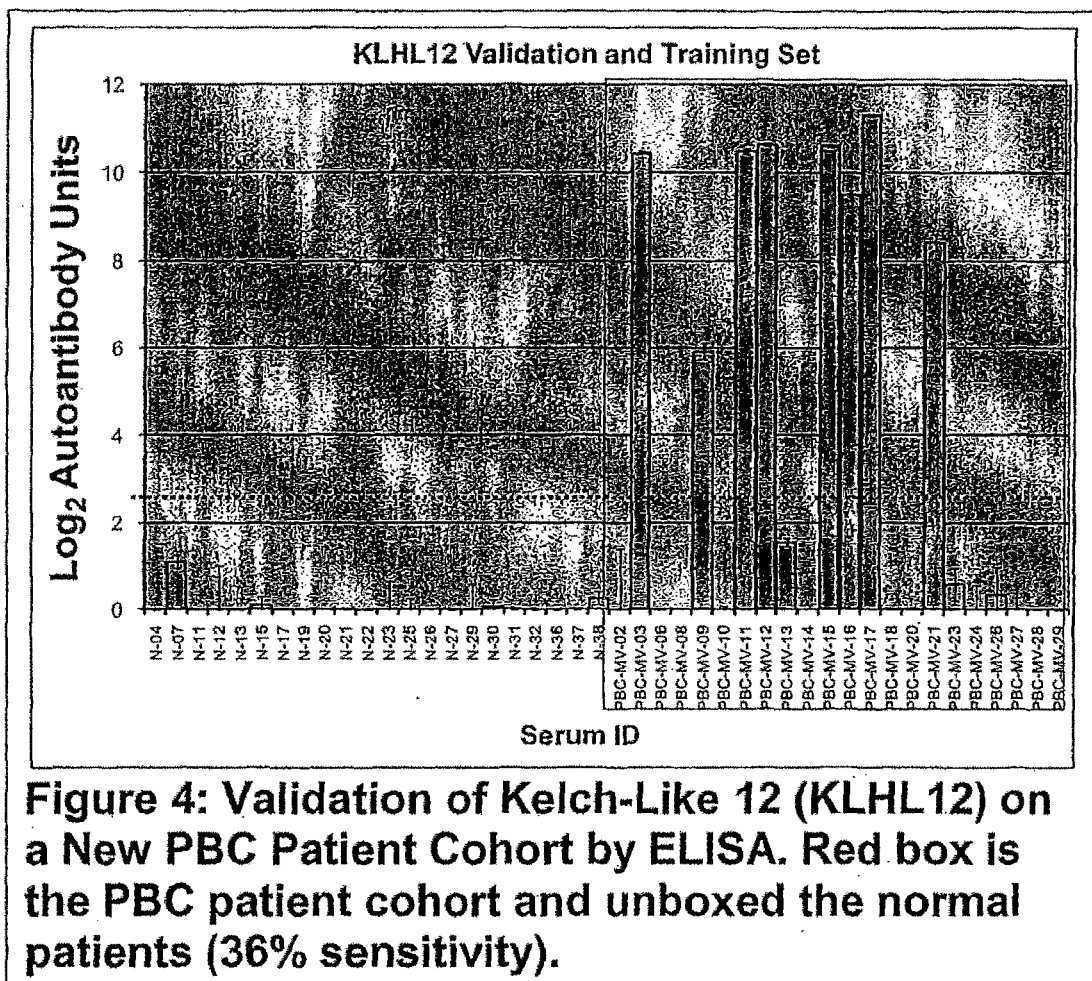
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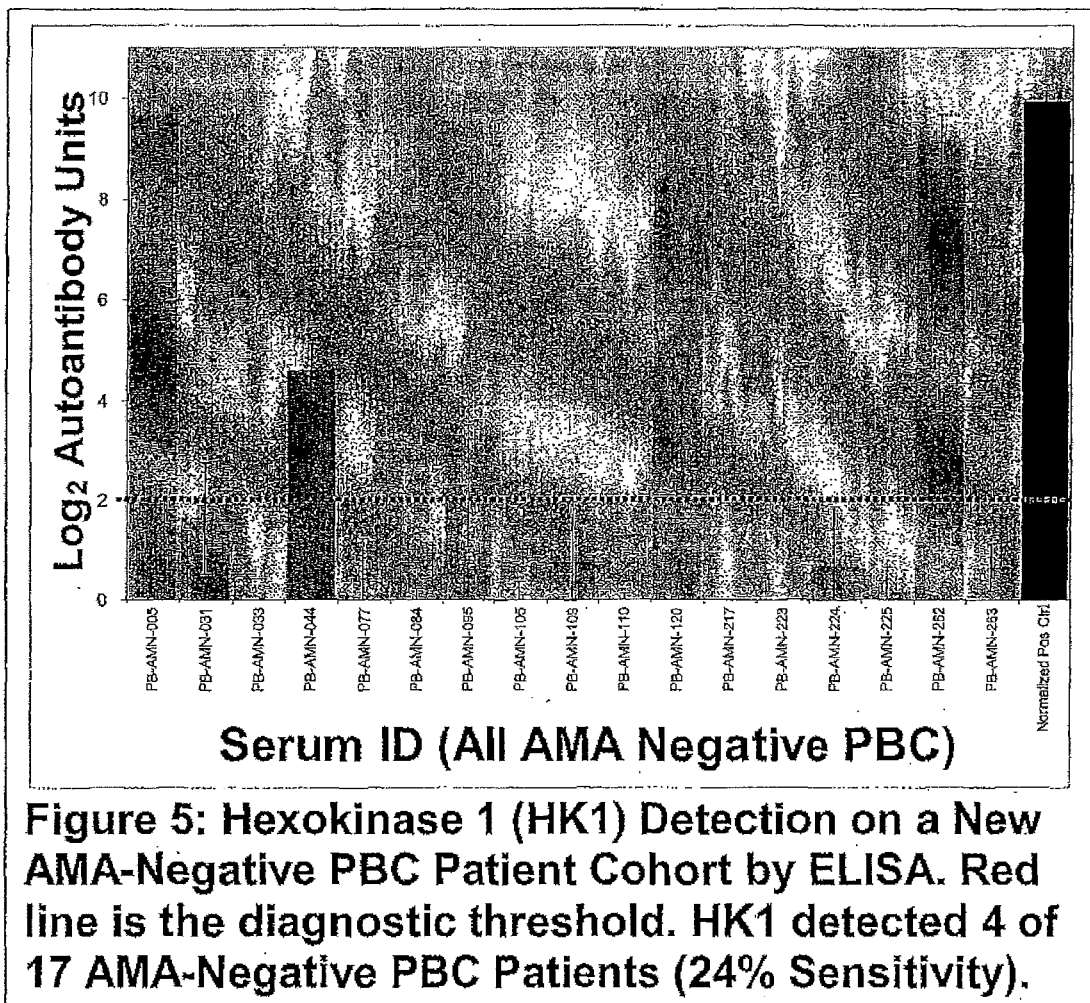
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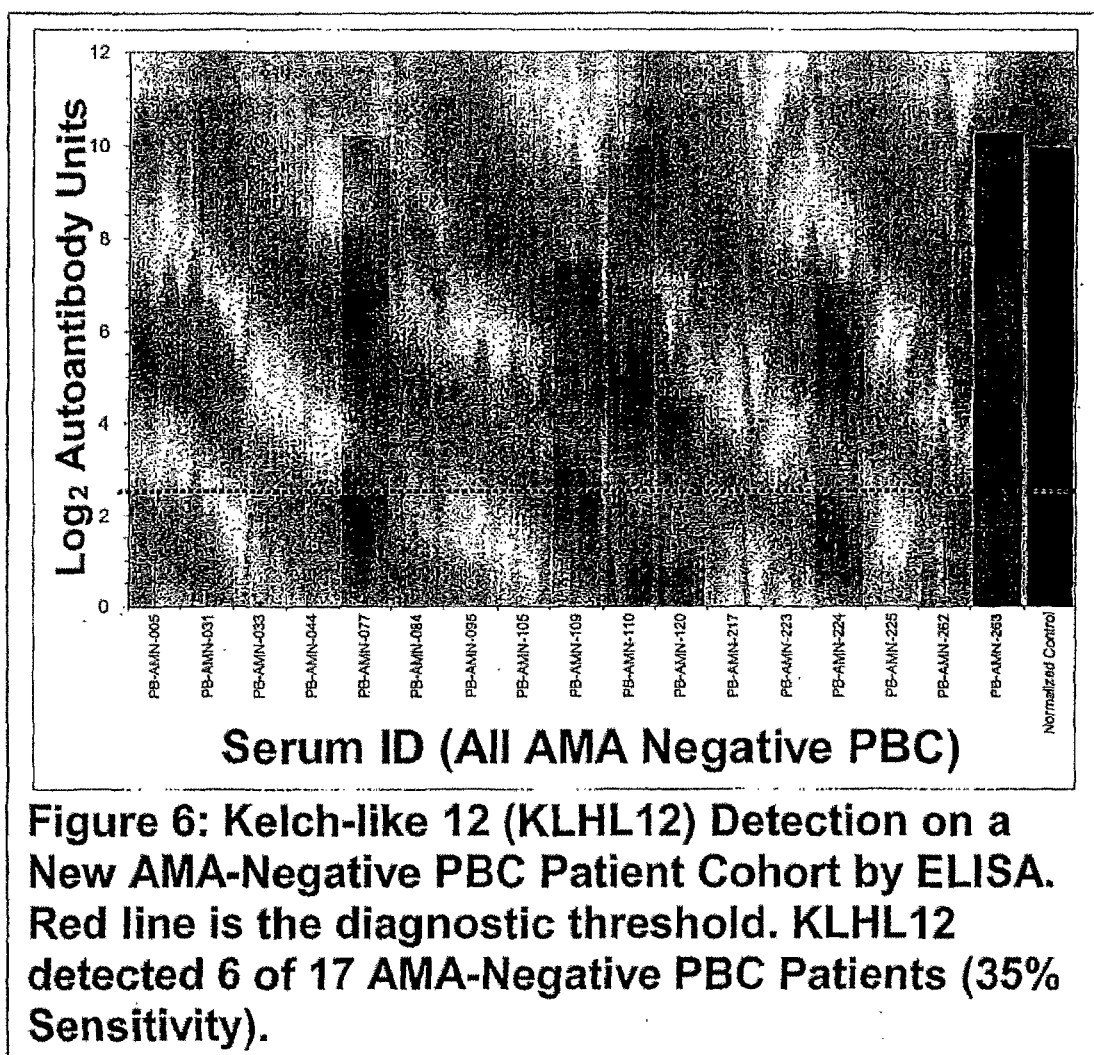


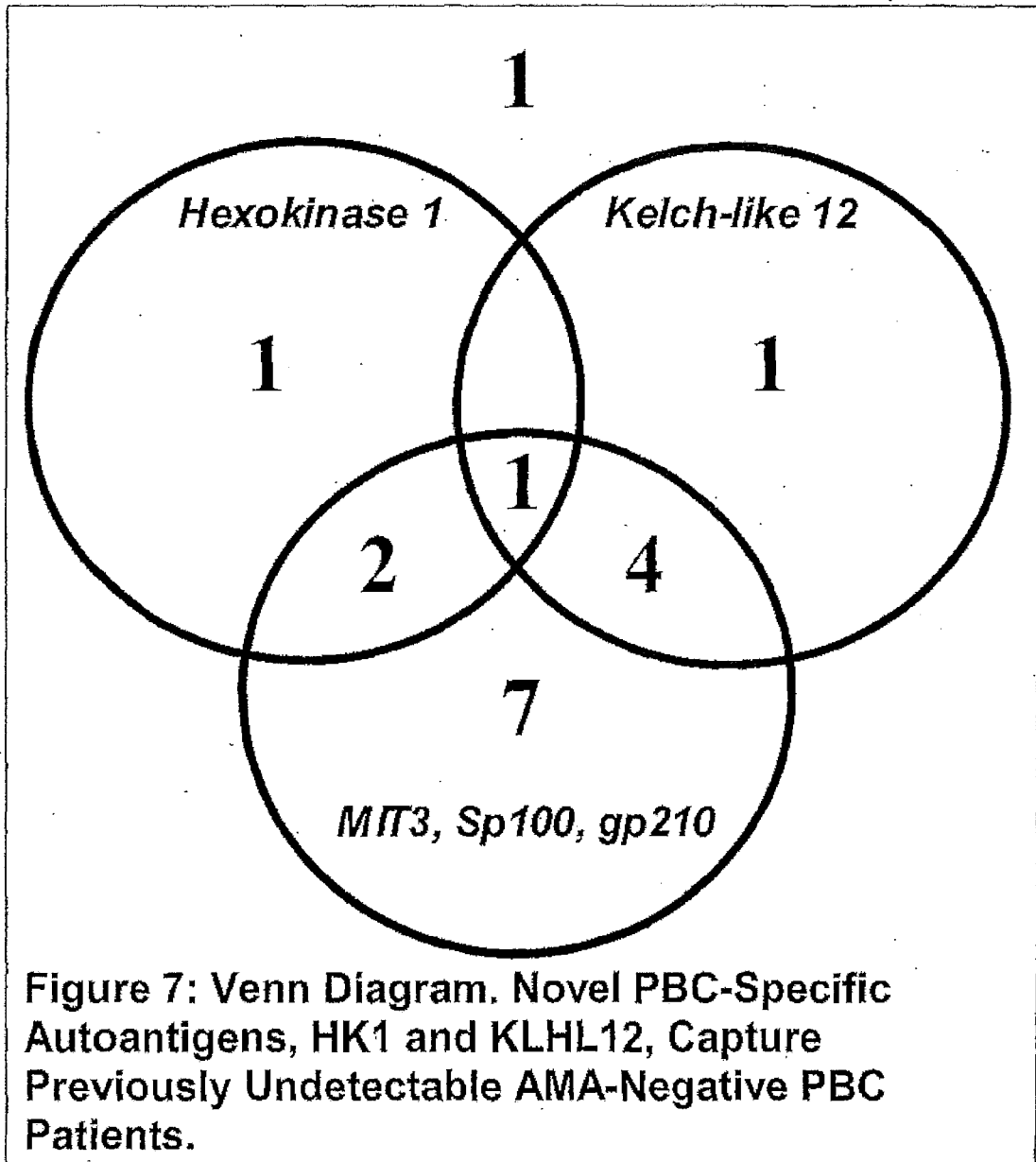


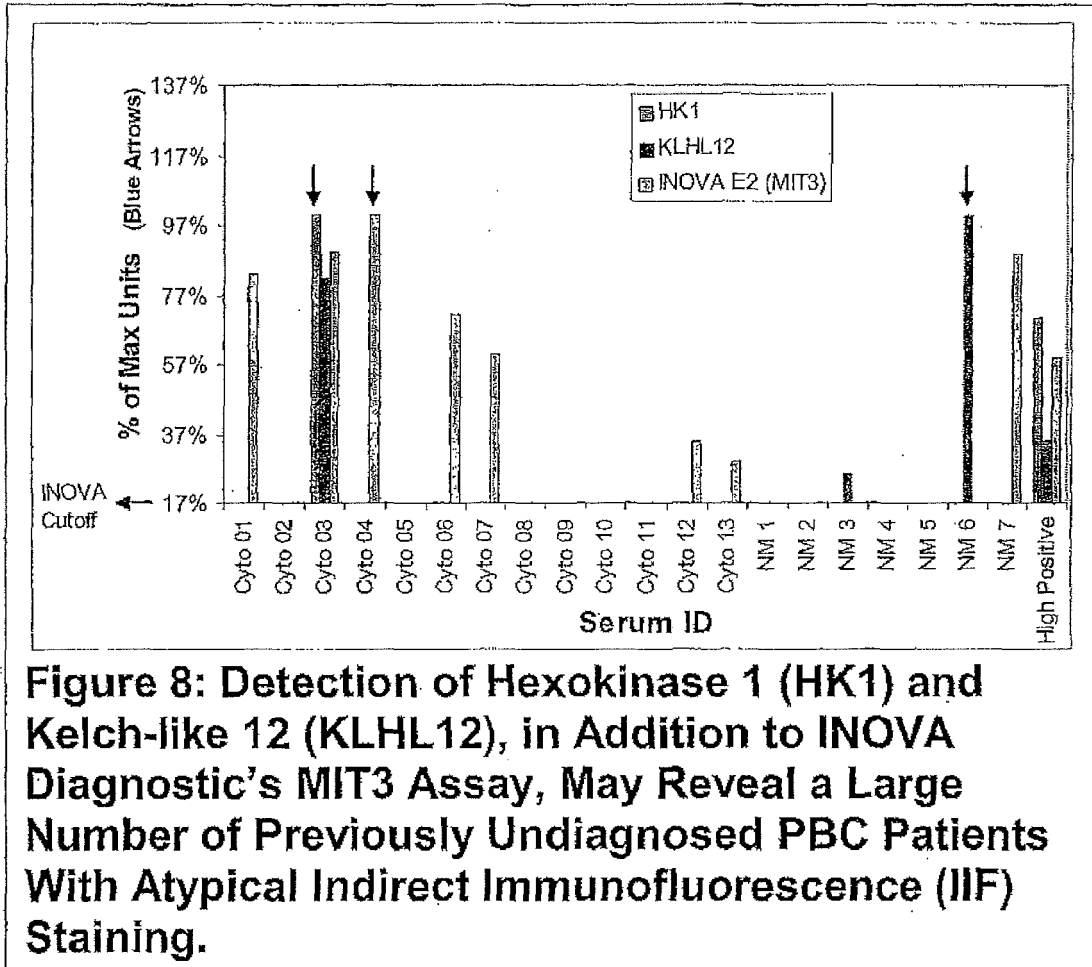


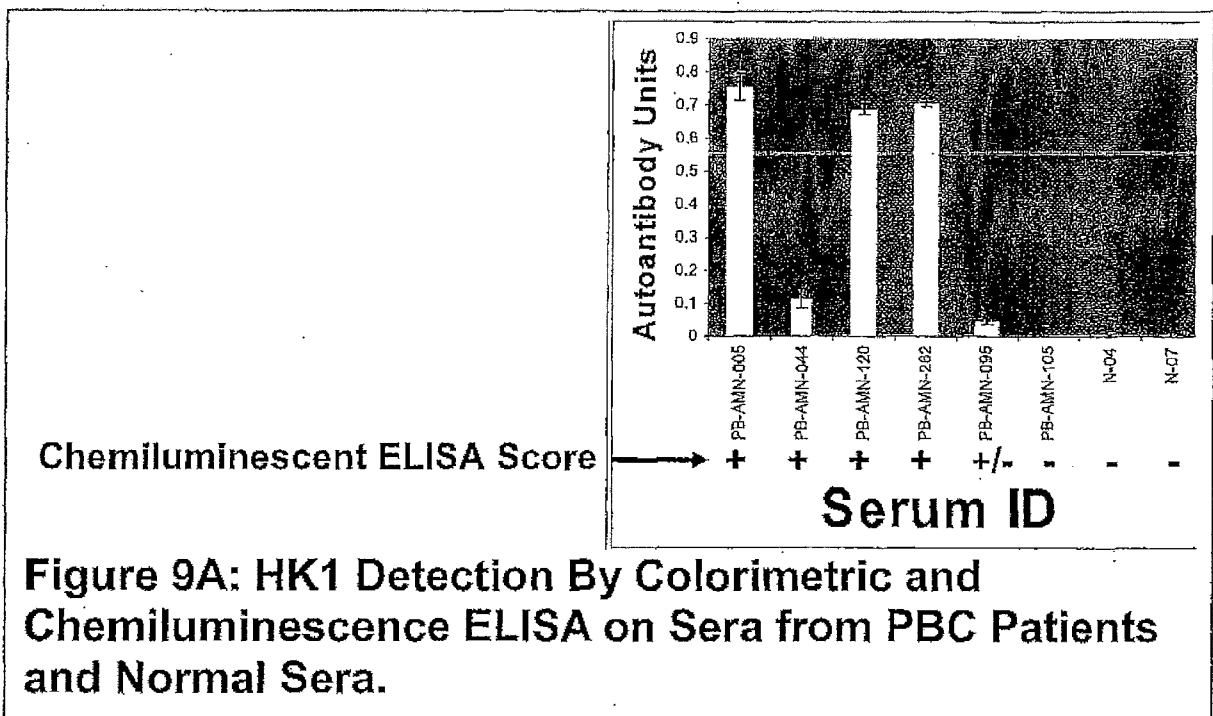


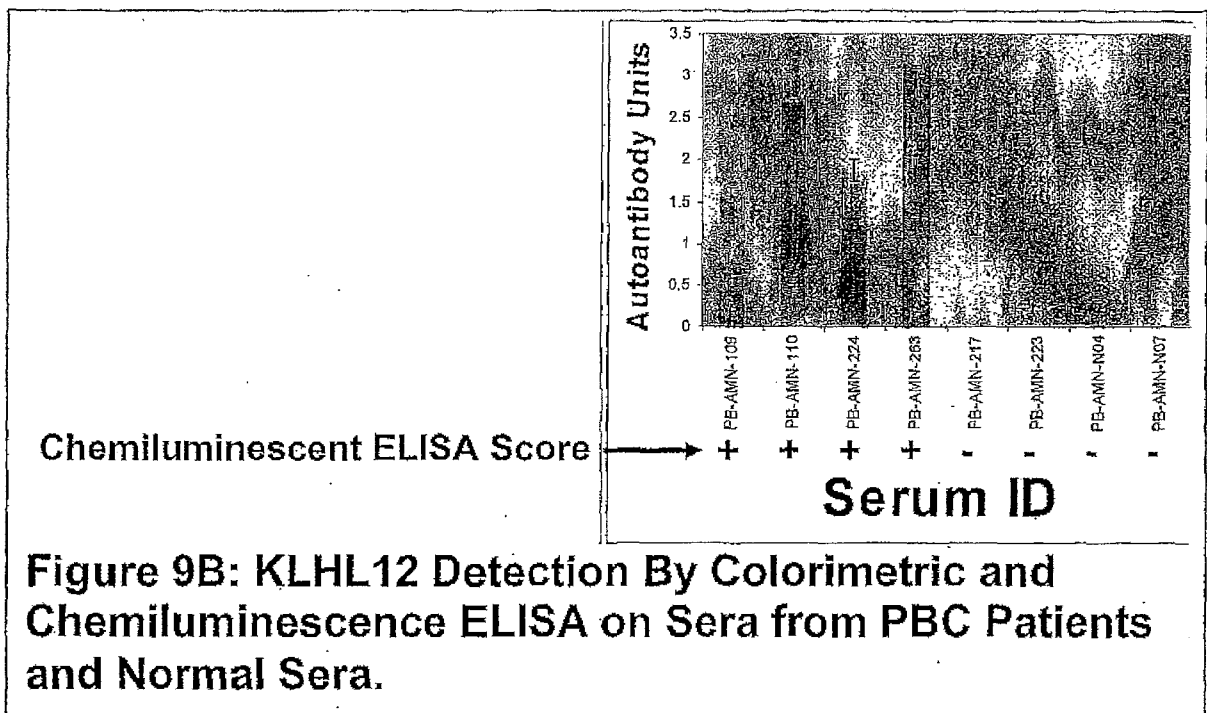


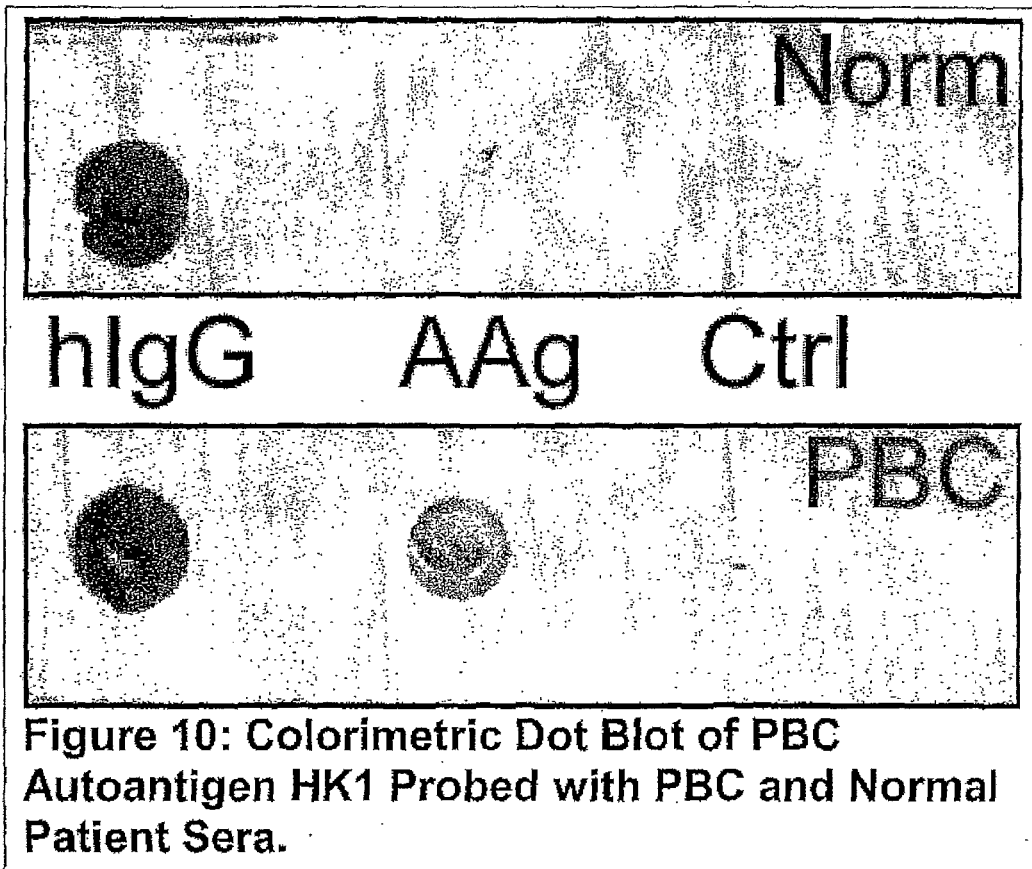


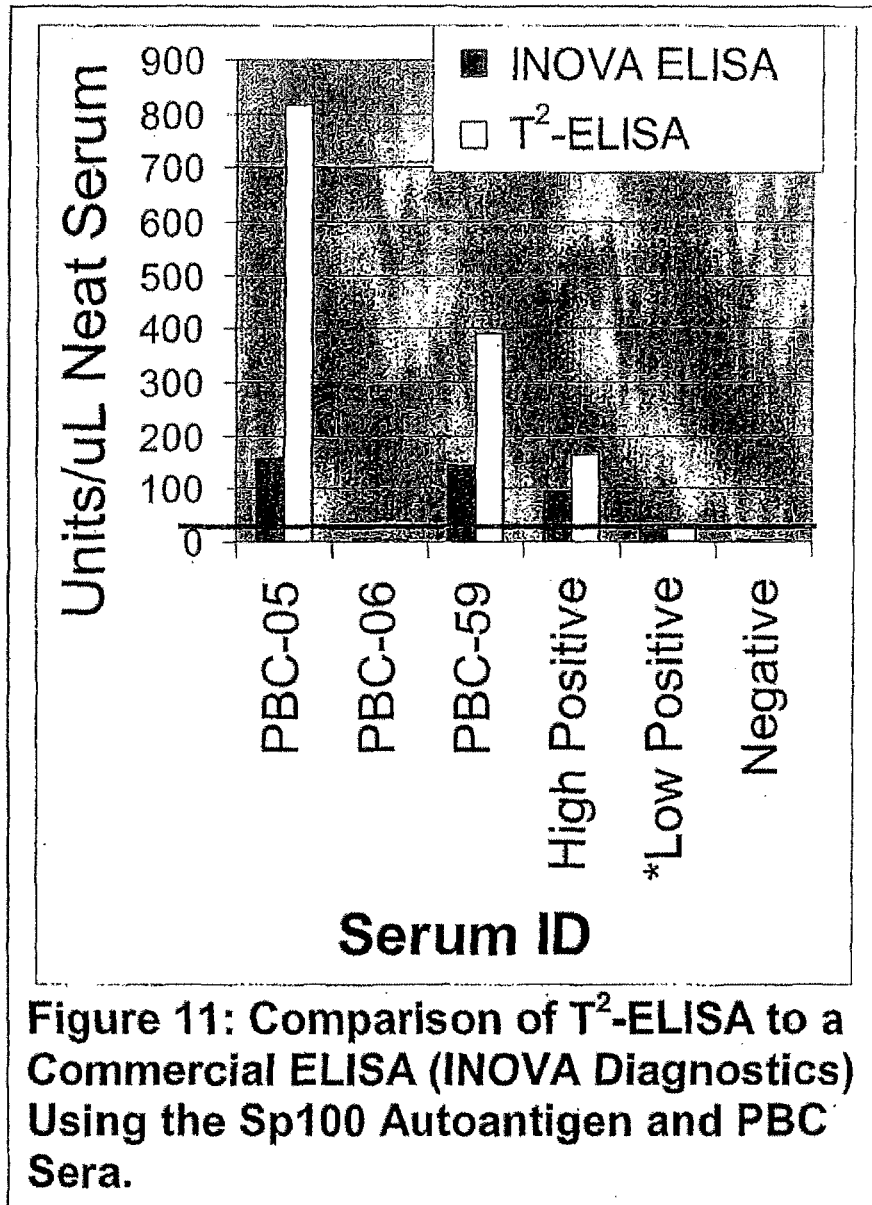












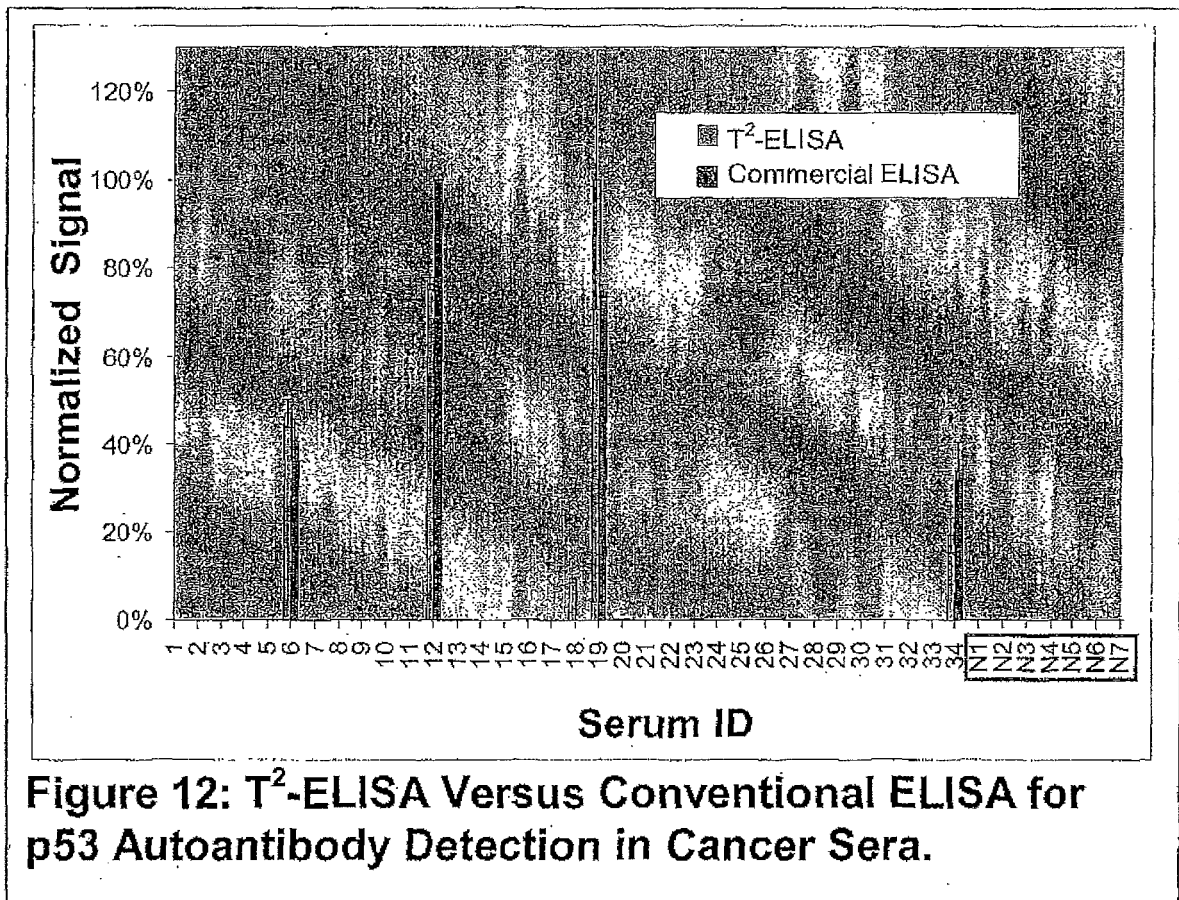
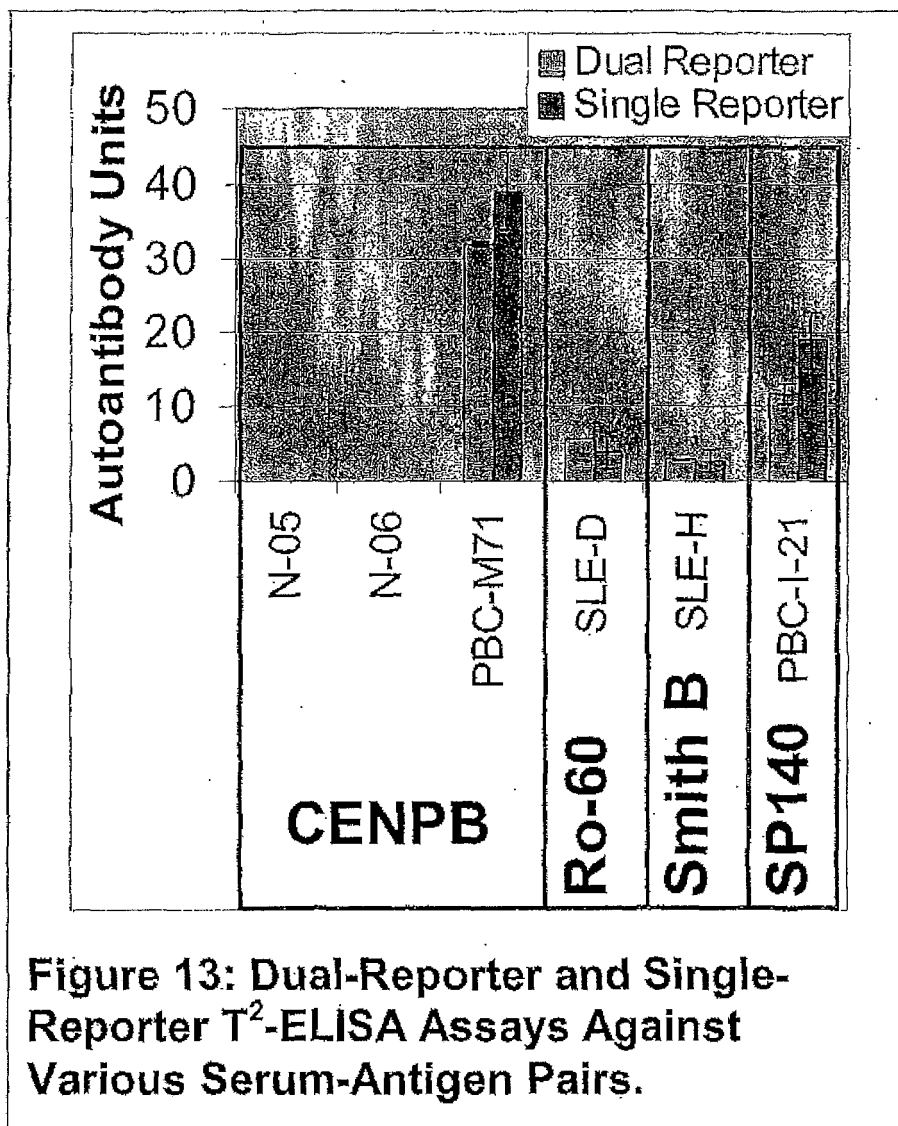
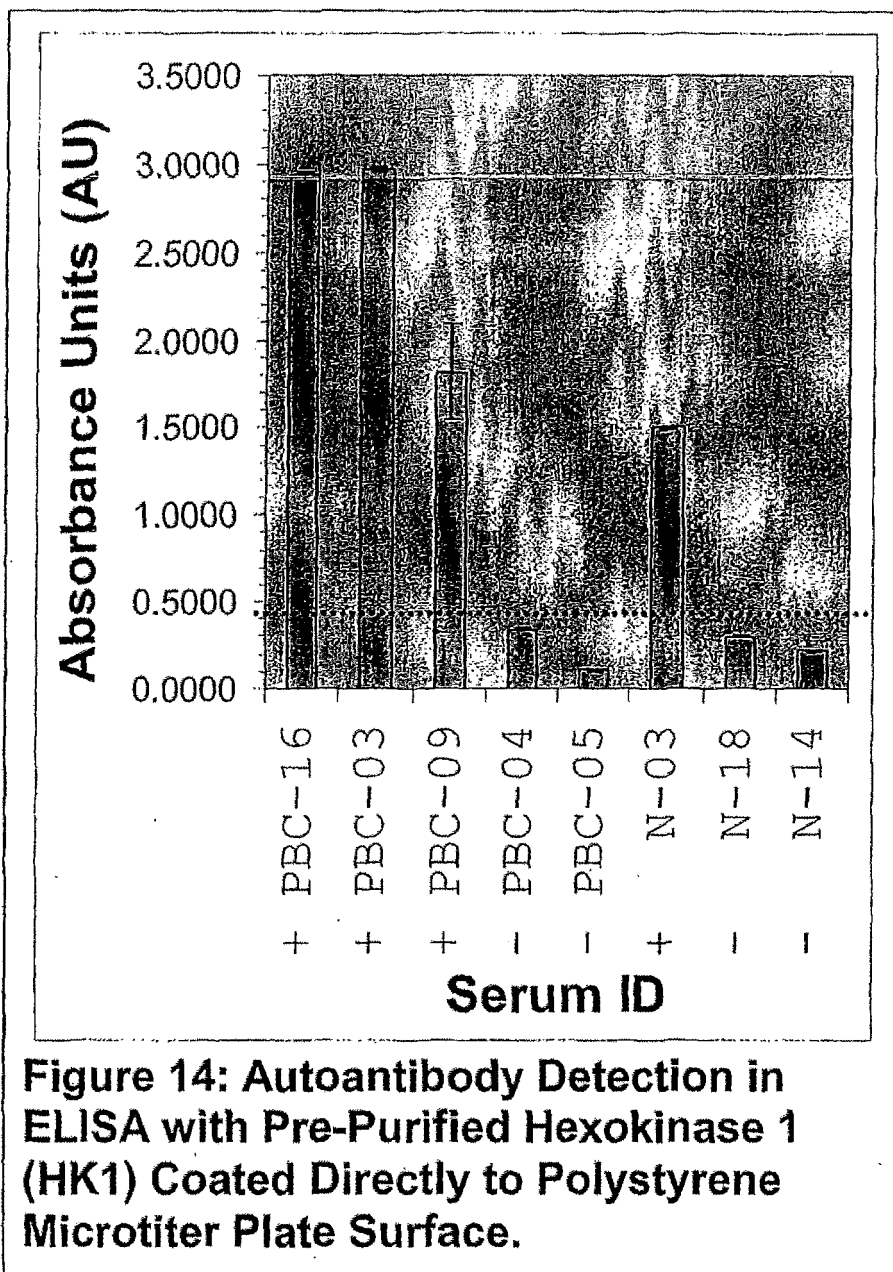


Figure 12: T²-ELISA Versus Conventional ELISA for p53 Autoantibody Detection in Cancer Sera.





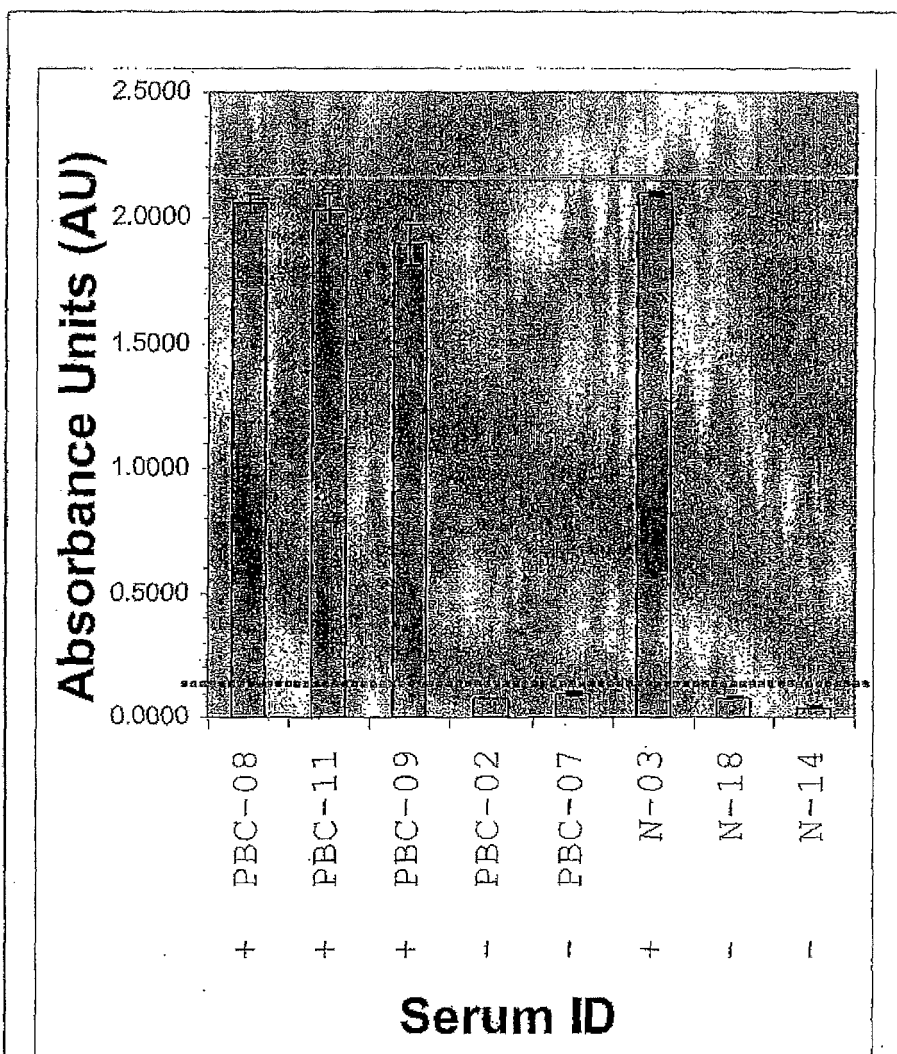
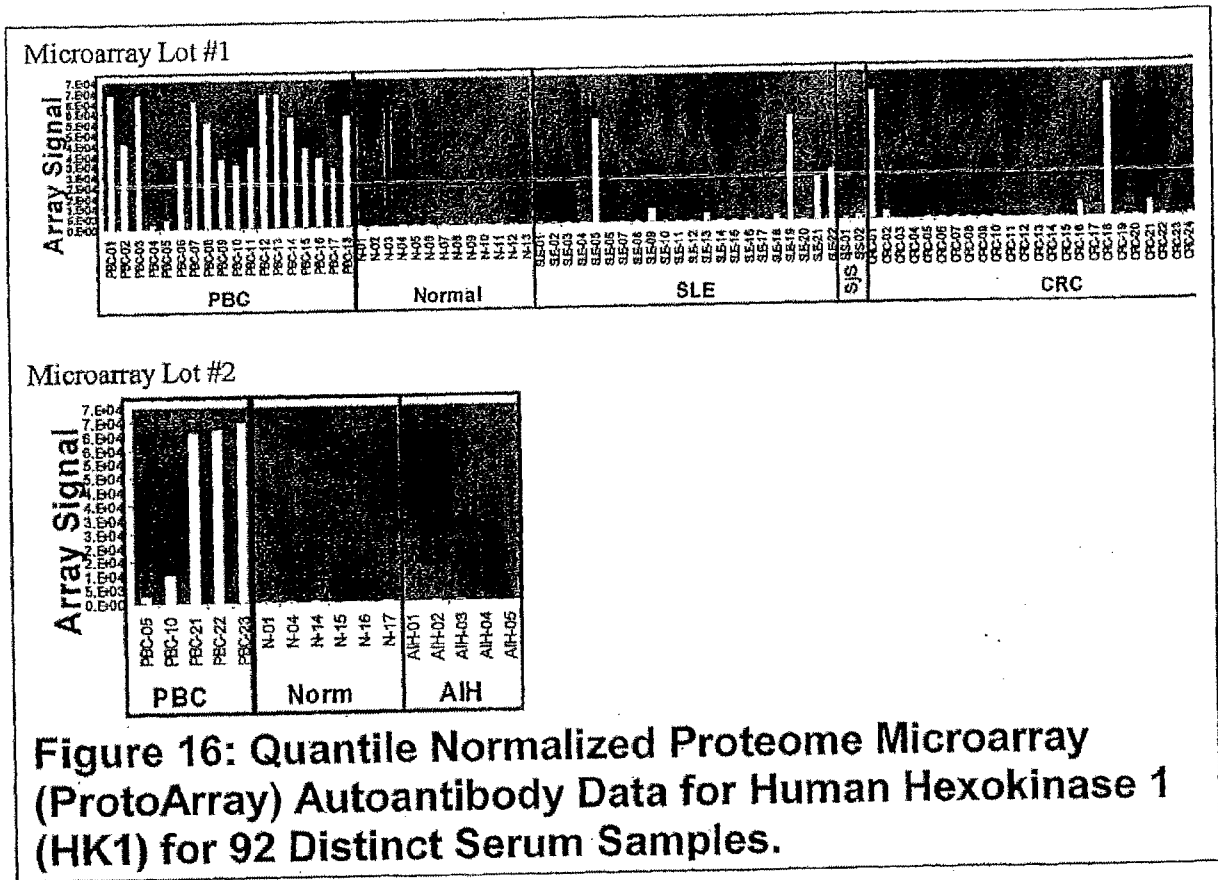
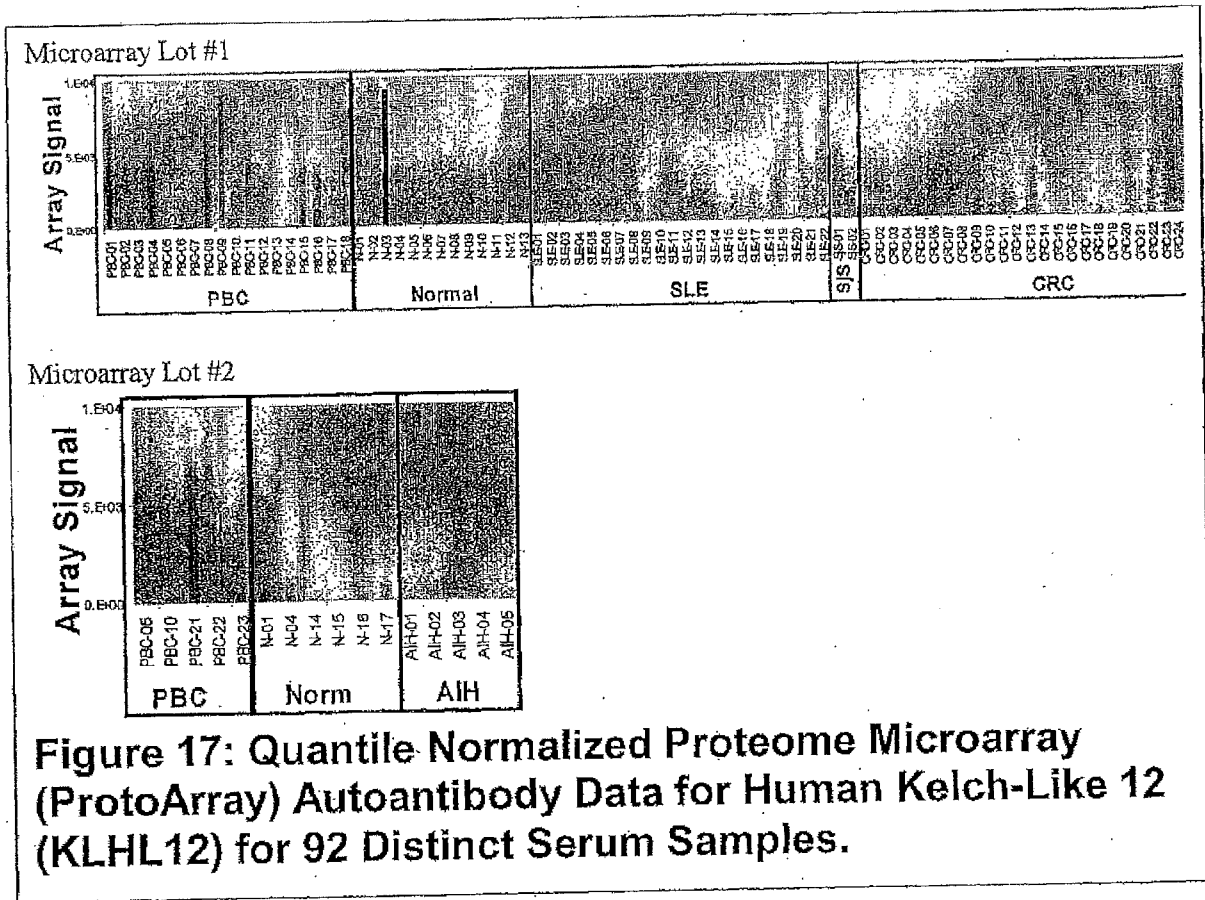


Figure 15: Autoantibody Detection in ELISA with Pre-Purified Kelch-Like 12 (KLHL12) Coated Directly to Polystyrene Microtiter Plate Surface.







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EUROPEAN SEARCH REPORT

Application Number
EP 15 19 4007

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DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (IPC)
A	<p>DATABASE MEDLINE [Online] US NATIONAL LIBRARY OF MEDICINE (NLM), BETHESDA, MD, US; April 2005 (2005-04), SELM I CARLO ET AL: "The enigma of primary biliary cirrhosis.", XP009166968, Database accession no. NLM15879614 * the whole document * & SELMI CARLO ET AL: "The enigma of primary biliary cirrhosis.", CLINICAL REVIEWS IN ALLERGY & IMMUNOLOGY APR 2005, vol. 28, no. 2, April 2005 (2005-04), pages 73-81, ISSN: 1080-0549</p>	1-14	INV. G01N33/53
A	<p>----- Eirini I Rigopoulou ET AL: "Molecular diagnostics of primary biliary cirrhosis", Expert Opin. Med. Diagn, 1 January 2008 (2008-01-01), pages 621-634, XP055243573, DOI: 10.1517/17530050802133669 Retrieved from the Internet: URL:https://www.researchgate.net/profile/E irini_Rigopoulou/publication/236050185_Mol ecular_diagnostics_of_primary_biliary_cirr hosis/links/02bfe50f57dfe25931000000.pdf * the whole document * ----- -/--</p>	1-15	TECHNICAL FIELDS SEARCHED (IPC) G01N
The present search report has been drawn up for all claims			
Place of search Munich		Date of completion of the search 21 January 2016	Examiner Moreno de Vega, C
CATEGORY OF CITED DOCUMENTS X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document		T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document	

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EUROPEAN SEARCH REPORT

Application Number
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DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (IPC)
A	<p>KAZUO UCHIDA ET AL: "Identification of specific autoantigens in Sjogren's syndrome by SEREX", IMMUNOLOGY, WILEY-BLACKWELL PUBLISHING LTD, GB, vol. 116, no. 1, 1 January 2005 (2005-01-01), pages 53-63, XP009166940, ISSN: 0019-2805, DOI: 10.1111/J.1365-2567.2005.02197.X [retrieved on 2005-06-27] * abstract *</p> <p style="text-align: center;">-----</p>	1-15	
			TECHNICAL FIELDS SEARCHED (IPC)
The present search report has been drawn up for all claims			
Place of search Munich		Date of completion of the search 21 January 2016	Examiner Moreno de Vega, C
<p>CATEGORY OF CITED DOCUMENTS</p> <p>X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document</p>		<p>T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document</p>	

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EPO FORM 1503 03.82 (P04C01)

REFERENCES CITED IN THE DESCRIPTION

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专利名称(译)	一种使用新型自身抗原诊断原发性胆汁性肝硬化 (PBC) 的方法		
公开(公告)号	EP3012631A1	公开(公告)日	2016-04-27
申请号	EP2015194007	申请日	2010-10-05
[标]申请(专利权)人(译)	AMBERGEN		
申请(专利权)人(译)	AMBERGEN INC. 马萨诸塞州总医院		
当前申请(专利权)人(译)	AMBERGEN INC. 马萨诸塞州总医院		
[标]发明人	LIM MARK J OSTENDORFF HEATHER P ROTHSCHILD KENNETH J BLOCH DONALD B		
发明人	LIM, MARK J. OSTENDORFF, HEATHER P. ROTHSCHILD, KENNETH J. BLOCH, DONALD B.		
IPC分类号	G01N33/53		
CPC分类号	G01N33/6893 G01N2800/085 G01N33/564 G01N2333/47		
优先权	61/248768 2009-10-05 US		
其他公开文献	EP3012631B1		
外部链接	Espacenet		

摘要(译)

描述了用于诊断原发性胆汁性肝硬化的方法和组合物。描述了新的自身抗原用于使用来自个体的测试样品的测定中。

