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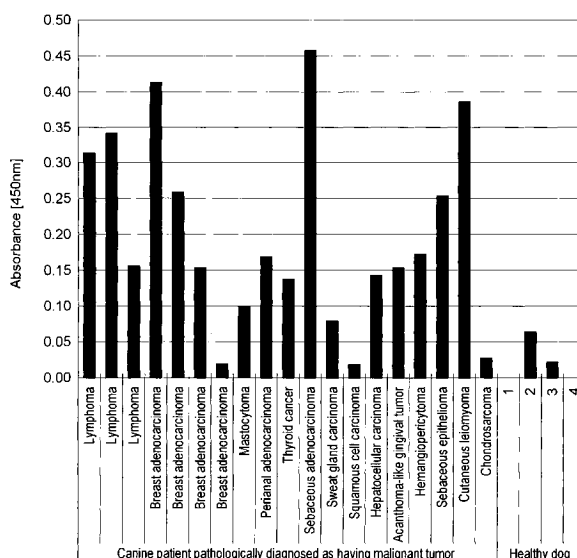
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(54) **CANCER DETECTION METHOD**

(57) The present invention relates to a method for detecting cancer, comprising measuring the expression of a polypeptide having a reactivity of binding to an antibody against a CAPRIN-1 protein having an amino acid sequence shown in any one of the even-numbered SEQ ID NOS: 2-30 in the Sequence Listing via an antigen-

antibody reaction in a sample separated from a living organism, and, a reagent for detecting a cancer comprising the CAPRIN-1 protein or a fragment thereof, an antibody against the CAPRIN-1 protein or a fragment thereof, or a polynucleotide encoding the CAPRIN-1 protein or a fragment thereof.

Fig. 3



Description

TECHNICAL FIELD

5 **[0001]** The present invention relates to a method for detecting cancer using CAPRIN-1 as a tumor marker.

BACKGROUND ART

10 **[0002]** Cancer is the leading cause of death. Treatment currently performed for cancer is mainly symptomatic therapy that mostly consists of surgical therapy with a combination of radiation therapy and chemotherapy. Owing to advancements in medical technology, cancer is now almost a curable disease if it can be detected early. Hence, a method for detecting cancer, by which detection can be conveniently performed using serum, urine, or the like without imposing physical or economic burdens on cancer patients, is now required.

15 **[0003]** As a cancer diagnostic method using blood or urine, a method for measuring a tumor product such as a tumor marker has recently become popular. The term "tumor product" refers to a tumor-associated antigen, an enzyme, a specific protein, a metabolite, a tumor gene, a tumor gene product, a tumor suppressor gene, and the like. Carcinoembryonic antigen CEA, glycoprotein CA19-9, CA125, prostate-specific antigen PSA, calcitonin, which are peptide hormones produced in the thyroid and the like are used as tumor markers for diagnosis of some cancer types. However, tumor markers useful for cancer diagnosis are absent for many cancer types. Also, most currently known tumor markers
20 are present in only trace amounts (on roughly a pg/mL order) in body fluids. Therefore, highly sensitive measurement methods or special techniques are required for detecting such tumor markers. Under the current circumstances, it is expected that provision of a new cancer testing means capable of detecting various types of cancer with high sensitivity involving a convenient procedure creates diagnostic applications for various types of cancer.

25 **[0004]** Also, such cancer testing means is very useful if it is capable of not only detecting cancer but also diagnosing cancer having developed in a location invisible to the naked eye, the extent of cancer, the malignancy or postoperative course of cancer, recurrence, metastasis, and the like.

[0005] Specifically, if diagnosis of cancer that has developed in a location invisible to the naked eye becomes possible, such cancer testing means would be useful for early detection of cancer within a location such as an intraperitoneal part that is difficult to recognize. Also, a tumor that does not have a grossly visible size such as cancer that is undetectable
30 even by ultrasonography, CT (computer tomography), or MRI (nuclear magnetic resonance imaging) can be detected.

[0006] Additionally, the extent of cancer is classified based on the degree to which a tumor spreads at the primary site and the presence or the absence of metastasis to regional lymph nodes or distant organs. In general, there are 5 disease stages (each referred to as "stage"), and higher stage numbers indicate more advanced stages of the disease. Strictly, the definition of stage differs depends on organs. However, for example, cancer at stage 0 is cancer that remains
35 intraepithelial and cancer at stage IV is cancer that has metastasized to a distant location. If such extent of cancer is found, decisions about appropriate treatment courses as well as diagnosis of the therapeutic effects of an anticancer agent become possible. As specific examples of decisions about treatment courses, in the case of prostate cancer and the like, there is a type requiring no treatment because it has very low malignancy and will almost never progress. In contrast, there is a type requiring treatment because it is progressive and metastasizes to bone or the like and causes
40 patients to die painfully. Therapies such as hormone therapy and extirpative surgery are each associated with an adverse reaction. Thus, therapies should be appropriately determined and decided upon. Also, if evaluation concerning the selection of an anticancer agent can be appropriately made or if timing or the like for the termination of administration of an anticancer agent can be appropriately determined, physical and economical burdens on patients can also be reduced. Therefore, it is important to be able to diagnose the extent of cancer.

45 **[0007]** One of the characteristics of cancer cells is that they undergo blastogenesis; that is, dedifferentiation. Except for some cancer types, poorly differentiated or undifferentiated cancer cells with a low degree of differentiation rapidly grow after metastasis and result in poor prognosis after therapy. Such cancer is said to have high malignancy. Conversely, highly differentiated cancer cells with a high degree of differentiation retain the structural and functional characteristics of affected organs. Such cancer can be said to have relatively low malignancy. If the malignancy of cancer can be
50 determined, the following measures can be taken. Even if the tumor is small, a wide surgical margin can be secured upon tumor removal, when the malignancy is high. Moreover, follow-up is possible while paying attention to a wide range of peripheral tissue.

[0008] If diagnosis of postoperative courses including recurrence and metastasis is possible, diagnosis of whether or not a tumor can be completely removed by surgery becomes possible. Incomplete tumor removal likely results in recur-
55 rence. Hence, such diagnosis can provide criteria for determining to more carefully perform follow-up at short intervals or to perform early reoperation if necessary. Also, if recurrence takes place, there is a high possibility of early detection. Detection is often delayed when distant metastasis takes place. However, if diagnosis of metastasis becomes possible, it becomes possible to provide criteria by which the range of testing can be broadened to include areas other than the

site of removal and the periphery thereof.

[0009] It is known that dogs grow old 7 times faster than humans. Recently, companion animals are being raised as family members and often have lifestyle habits similar to those of their owners. Therefore, it is predictable that an owner's risk of developing cancer would be high when his or her companion animal develops cancer. If convenient and precise cancer diagnosis becomes possible for companion animals, it would be expected to provide clues for preventing cancer of owners.

[0010] Currently, the number of domestic dogs in Japan is said to be about 6,700,000, and the same figure for the U.S. is said to be about 17,640,000. Quintuple, septuple, and octuple combined vaccines and the like have become prevalent, in addition to rabies shots, and thereby highly lethal infectious diseases have decreased, such as canine parvovirus infection, canine distemper virus infection, canine parainfluenza (kennel cough), canine adenovirus-2 infection (kennel cough), infectious canine hepatitis, canine coronavirus infection, and leptospirosis. Therefore, the average life span of dogs has increased. Elderly dogs, which are seven years old or older, account for 35.5% of all domestic dogs. Causes of death of domestic dogs are also similar to those of humans, such as cancer, hypertension, and cardiac disease, which are on the rise. In the U.S., about 4,000,000 dogs are diagnosed with cancer annually. Also in Japan, it is said that about 1,600,000 dogs are potentially affected with tumors.

[0011] However, convenient cancer diagnostic agents for animals have been absent. Furthermore, in animal medical care, testing methods that involve photographing or filming using X-rays, CT scans, MRI scans, or the like have not been prevalent. After palpation, a simple blood test, and testing using X-ray photography are performed, diagnosis currently depends significantly on the experience of veterinarians. Testing methods using serum have been partially begun, but the methods use human tumor markers since no canine tumor marker has been discovered.

[0012] Precise cancer diagnosis requires abdominal surgery that imposes significant physical burdens on dogs and cost burdens on owners. If cancer diagnosis can be conveniently made for companion animals such as dogs and cats, it would lead to early detection or precise diagnosis of cancer and would be expected to be useful for cancer therapy for companion animals. Also, if such convenient cancer diagnosis using serum becomes possible, it would be expected not only to enable cancer diagnosis but also to significantly contribute to periodic health examinations, preoperative diagnosis, and decisions about therapeutic strategy.

[0013] Health examination for companion animals, unlike the case of humans, is not prevalent. Hence, detection of cancer often occurs too late, such that an owner finds out the disease and then comes to a hospital only after the tumor has become large in many cases. If such tumor that has increased in size is malignant, it often results in treatment that is too late, even when surgical therapy such as surgery or medication using an anticancer agent or the like is performed. Hence, when a veterinarian determines that the tumor is malignant, anticancer agent treatment is generally performed without surgery. If surgery is performed, measures during surgery, such as determination of the size of margin to be secured, determination of the amount of blood required during surgery, and measures against cell scattering should also be strictly taken. It is desired that anticancer agent treatment is initiated immediately after surgery and that follow-up is performed at short intervals. Incorporation of the above cancer diagnosis into dog health checkups that are recently increasingly prevalent and are referred to as complete medical checkups for dogs is expected to lead to early cancer detection.

[0014] On the other hand, in the case of a benign tumor, surgery can be advised even if a tumor is large. After surgery, only resected areas need care without requiring any expensive anticancer agent treatment and without any need for apprehensions concerning follow-ups.

[0015] Under the current situation, provision of a convenient means for detecting cancer with high sensitivity, which is applicable to cancer diagnosis for animals, enables precise and efficient treatment and results in a number of advantages for both owners and veterinarians.

[0016] Cytoplasmic-and proliferation-associated protein 1 (CAPRIN-1) is an intracellular protein that is expressed when normal cells in resting phase are activated or undergo cell division. CAPRIN-1 is also known to be involved in mRNA transport through intracellular formation of intracellular stress grains with RNA and translation control, for example. Meanwhile, CAPRIN-1 has many different names. Examples of such names include GPI-anchored membrane protein 1 and membrane component surface marker 1 protein (M11S1), as if the protein has been known to be a membrane protein. These different names are derived from a report (J Biol Chem. 270: 20717-20723 (1995)) that the gene sequence of CAPRIN-1 originally has a GPI-binding region and CAPRIN-1 is a membrane protein expressed in large bowel-derived cell lines. It has been later reported that: the CAPRIN-1 gene sequence in this report is an error; frame shift takes place by deletion of 1 nucleotide from the CAPRIN-1 gene sequence currently registered with GenBank or the like, so that 80 amino acids are deleted from the C terminus and the resulting artifact (74 amino acids) corresponds to the GPI binding portion of the previous report; and an error is also present on the 5' side of the gene sequence and deletion of 53 amino acids from the N terminus has been proven (J Immunol. 172: 2389-2400 (2004)). Also, it has been reported that a protein encoded by the CAPRIN-1 gene sequence currently registered with GenBank or the like is not a cell membrane protein (J Immunol. 172: 2389-2400 (2004)).

[0017] In addition, based on the report of J Biol Chem. 270: 20717-20723 (1995) that CAPRIN-1 is a cell membrane

protein, US2008/0075722 and WO2005/100998 disclose that CAPRIN-1 under the name of M11S1 can be a target for cancer therapy as a cell membrane protein (not mentioned in the Examples). However, as reported in J Immunol. 172: 2389-2400 (2004), it has been accepted from the time of filing of US2008/0075722 and WO2005/100998 up to now that CAPRIN-1 is not expressed on cell surfaces. It is obvious that the content of US2008/0075722 and WO2005/100998 based only on misinformation to the effect that CAPRIN-1 is a cell membrane protein should not be understood as technical commonsense of persons skilled in the art. Moreover, it has never been reported that CAPRIN-1 is expressed at higher levels in breast cancer cells or the like than in normal cells.

SUMMARY OF THE INVENTION

PROBLEM TO BE RESOLVED BY THE INVENTION

[0018] An object of the present invention is to provide a means for detecting cancer that is useful for cancer diagnosis.

MEANS FOR RESOLVING THE PROBLEM

[0019] As a result of intensive studies, the present inventors have obtained cDNA encoding a protein that binds to an antibody existing in cancer-bearing living organism-derived serum by a SEREX method using a canine testis-derived cDNA library and the serum of a cancer-bearing dog, and thus they have prepared canine CAPRIN-1 proteins having the amino acid sequences shown in SEQ ID NOS: 6, 8, 10, 12, and 14 based on the cDNA. Also, the present inventors have prepared human CAPRIN-1 proteins having the amino acid sequences shown in SEQ ID NOS: 2 and 4 based on human genes homologous to the obtained genes. The present inventors have further discovered that: genes encoding these proteins are specifically expressed in canine and human testes and malignant cancer cells (see Example 1 described later); recombinant polypeptides prepared based on the amino acid sequences of these proteins specifically react only with sera from cancer-bearing living organisms; and CAPRIN-1 can be specifically detected from a cancer-bearing living organism using antibodies prepared using the recombinant polypeptides. Thus, the present inventors have completed that present invention.

[0020] Specifically, the present invention provides a method for detecting cancer comprising measuring CAPRIN-1 expression, which is performed for samples separated from living organisms. Also, the present invention provides a reagent for detecting cancer comprising an antibody that is induced *in vivo* against CAPRIN-1 and a polypeptide that undergoes an antigen-antibody reaction. Furthermore, the present invention provides a reagent for detecting cancer comprising an antibody that undergoes an antigen-antibody reaction with CAPRIN-1 or an antigen-binding fragment thereof. Furthermore, the present invention provides a reagent for detecting cancer comprising a polynucleotide that specifically hybridizes to a partial sequence of 15 or more nucleotides, preferably 20 to 25 or more nucleotides, and more preferably 30 or more nucleotides in the nucleotide sequence shown in SEQ ID NO: 1, 3, 5, 7, 9, 11, 13, or the like in the Sequence Listing.

[0021] Specifically, the present invention has the following characteristics.

(1) A method for detecting a cancer, comprising measuring the expression of a polypeptide having a reactivity of binding via an antigen-antibody reaction to an antibody against a CAPRIN-1 protein having any one of the amino acid sequences shown in the even-numbered SEQ ID NOS: 2-30 in the Sequence Listing, in a sample separated from a living organism.

(2) The method according to (1) above, wherein the polypeptide to be measured is a CAPRIN-1 protein having any one of the amino acid sequences shown in the even-numbered SEQ ID NOS: 2-30 (i.e., SEQ ID NOS: 2, 4, 6, 8,... 30) or a polypeptide having 85% or more sequence identity with the CAPRIN-1 protein.

(3) The method according to (1) or (2) above, wherein the living organism is a human, a dog, or a cat.

(4) The method according to (3) above, wherein the living organism is a dog and the polypeptide to be measured has an amino acid sequence shown in any one of the even-numbered SEQ ID NOS: 2-30.

(5) The method according to (4) above, wherein the living organism is a dog and the polypeptide to be measured has the amino acid sequence shown in SEQ ID NO: 6, 8, 10, 12, or 14.

(6) The method according to (3) above, wherein the living organism is a human and the polypeptide to be measured has the amino acid sequence shown in SEQ ID NO: 2 or 4.

(7) The method according to any one of (1) to (6) above, wherein the expression of the polypeptide is measured by immunoassay of an antibody that can be contained in the sample and is induced *in vivo* against the polypeptide to be measured.

(8) The method according to any one of (1) to (7) above, wherein the samples is serum, blood plasma, ascite, or pleural effusion.

(9) The method according to any one of (1) to (6) above, wherein the expression of the polypeptide is measured by

measuring mRNA encoding the polypeptide, which is contained in the sample.

(10) The method according to (9) above, comprising examining the existing amount of the mRNA in the sample using a polynucleotide that specifically hybridizes to a partial sequence of 15 or more nucleotides, preferably 20 to 25 or more nucleotides, and more preferably 30 or more nucleotides in the nucleotide sequence of the above mRNA.

(11) The method according to (10) above, wherein the above living organism is a dog and the above polynucleotide is a polynucleotide specifically hybridizing to a partial sequence of 15 or more nucleotides, preferably 20 to 25 or more nucleotides, and more preferably 30 or more nucleotides in the nucleotide sequence shown in SEQ ID NO: 5, 7, 9, 11, or 13.

(12) The method according to (10) above, wherein the above living organism is a human and the above polynucleotide is a polynucleotide specifically hybridizing to a partial sequence of 15 or more nucleotides, preferably 20 to 25 or more nucleotides, and more preferably 30 or more nucleotides in the nucleotide sequence shown in SEQ ID NO: 1 or 3.

(13) The method according to any one of (9) to (12) above, wherein the above sample is a tissue or a cell.

(14) The method according to any one of (1) to (13) above, wherein the cancer is at least one type of cancer selected from the group consisting of brain tumor, squamous cell carcinoma of the head, neck, lung, uterus, or esophagus, melanoma, adenocarcinoma of the lung or uterus, renal cancer, malignant mixed tumor, hepatocellular carcinoma, basal cell carcinoma, acanthoma-like gingival tumor, tumor of the oral cavity, perianal adenocarcinoma, anal sac tumor, anal sac apocrine adenocarcinoma, sertoli cell carcinoma, cancer of vaginal vestibule, sebaceous adenocarcinoma, sebaceous epithelioma, sebaceous adenoma, sweat gland carcinoma, intranasal adenocarcinoma, nasal adenocarcinoma, thyroid cancer, large-bowel cancer, bronchial adenocarcinoma, adenocarcinoma, ductal carcinoma, breast adenocarcinoma, composite type breast adenocarcinoma, malignant mammary mixed tumor, intraductal papillary adenocarcinoma, fibrosarcoma, hemangiopericytoma, osteosarcoma, chondrosarcoma, soft tissue sarcoma, histiocytic sarcoma, myxosarcoma, undifferentiated sarcoma, lung cancer, mastocytoma, cutaneous leiomyoma, intraperitoneal leiomyoma, leiomyoma, chronic lymphocytic leukemia, lymphoma, gastrointestinal lymphoma, digestive lymphoma, small-cell-to-medium-cell lymphoma, adrenomedullary tumor, granulosa cell tumor, and pheochromocytoma.

(15) The method according to any one of (1) to (14) above, comprising further detecting the malignancy of cancer based on the fact that the malignancy of a cancer is high when the expression level of the above polypeptide is higher than that of a control.

(16) The method according to any one of (1) to (15) above, comprising further detecting the progression of cancer on the basis of the indicator that the extent of cancer is advanced when the expression level of the above polypeptide is higher than that of a control.

(17) A reagent for detecting a cancer, comprising a polypeptide that has a reactivity of binding via an antigen-antibody reaction to an antibody that is induced *in vivo* against a CAPRIN-1 protein having any one of the amino acid sequences shown in the even-numbered SEQ ID NOS: 2-30 in the Sequence Listing.

(18) A reagent for detecting a cancer, comprising an antibody or an antigen-binding fragment thereof that undergoes an antigen-antibody reaction with a polypeptide, wherein the polypeptide has a reactivity of binding via an antigen-antibody reaction to an antibody against a CAPRIN-1 protein having any one of the amino acid sequences shown in the even-numbered SEQ ID NOS: 2-30 in the Sequence Listing and is produced *in vivo* (or in a living body).

(19) The reagent for detecting cancer according to (18), wherein the antibody or antigen-binding fragment thereof that undergoes an antigen-antibody reaction with the polypeptide is an antibody or antigen-binding fragment thereof that binds to the surface of a cancer cell.

(20) The reagent for detecting cancer according to (18) or (19), wherein the antibody or antigen-binding fragment thereof that undergoes an antigen-antibody reaction with the polypeptide has an immunological reactivity with:

a polypeptide comprising an amino acid sequence of 7 or more continuous amino acid residues within the region of amino acid residue Nos. 50-98 or amino acid residue Nos. 233-305 in any one of the amino acid sequences shown in the even-numbered SEQ IDS NO: 2-30 excluding SEQ ID NO: 6 and SEQ ID NO: 18 or a polypeptide comprising the polypeptide as a partial sequence.

(21) The reagent for detecting a cancer according to any one of (18) to (20), wherein the antibody or antigen-binding fragment thereof that undergoes an antigen-antibody reaction with the polypeptide is an antibody or antigen-binding fragment thereof which binds to SEQ ID NO: 43, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 44 and 45, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 44 and 46, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 44 and 47, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 44 and 48, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 49 and 50, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 51 and 52, a monoclonal

antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 53 and 54, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 55 and 56, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 57 and 58, or a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 59 and 60.

(22) A reagent for detecting a cancer, comprising a polynucleotide that specifically hybridizes to a partial sequence of 15 or more nucleotides, preferably 20 to 25 or more nucleotides, and more preferably 30 or more nucleotides in any one of the nucleotide sequences shown in the odd-numbered SEQ ID NOS: 1-29 (i.e., SEQ ID NOS: 1, 3, 5, 7,...29) in the Sequence Listing.

ADVANTAGE OF THE INVENTION

[0022] According to the present invention, a new method for detecting a cancer is provided. As specifically described in Examples given later, a recombinant polypeptide prepared based on the amino acid sequence of CAPRIN-1 (or also referred to as Caprin-1) reacts with an antibody that specifically exists in the serum of a patient with cancer. Therefore, the cancer existing in a living body can be detected by measuring the antibody in a sample by the method of the present invention. Also, the cancer existing in a living body can be detected by measuring CAPRIN-1 itself. According to the method of the present invention, small-size cancer invisible to the naked eye or cancer in a deep part *in vivo* can be detected. Hence, the method of the present invention is useful for early detection of cancer at the time of health examination or the like. Furthermore, recurrent cancer can be detected early by the use of the method of the present invention for the follow-up of a patient after cancer treatment. Moreover, according to the method of the present invention, the extent of cancer can also be diagnosed, such as tumor increase, infiltration to the peripheral tissue, and cancer metastasis to a lymph node and a distant organ. Also, the serum antibody level is higher in a patient with highly malignant cancer than in a patient with low-malignant cancer. According to the method of the present invention, the malignancy of cancer can also be diagnosed. Also, as described in Examples below, mRNA encoding CAPRIN-1 is specifically expressed at high levels in testes and cancer cells. Therefore, cancers can also be detected by measuring the mRNA.

BRIEF DESCRIPTION OF THE DRAWINGS

[0023]

Fig. 1 shows the expression patterns of the gene encoding a CAPRIN-1 protein in normal tissues and tumor cell lines. Reference No. 1 indicates the expression patterns of the gene encoding the CAPRIN-1 protein. Reference No. 2 indicates the expression patterns of the GAPDH gene.

Fig. 2 shows the results of detecting by Coomassie staining the canine CAPRIN-1-derived polypeptide that is an example of polypeptides to be used in the present invention, which were produced and purified using *Escherichia coli* in the Examples. Reference No. 3 indicates the band of a canine CAPRIN-1-derived polypeptide.

Fig. 3 shows some of the results of cancer diagnosis for cancer-bearing dogs using the canine CAPRIN-1-derived polypeptides prepared in the Examples.

Fig. 4 shows some of the results of detailed cancer diagnosis for cancer-bearing dogs using the canine CAPRIN-1-derived polypeptides prepared in the Examples.

BEST MODE OF CARRYING OUT THE INVENTION

[0024] According to the method of the present invention, CAPRIN-1 expression is measured using a sample separated from a living organism. Examples of a method for measuring CAPRIN-1 expression include a method (1st method) that involves immunoassay for an antibody against CAPRIN-1 contained in a sample, a method (2nd method) that involves immunoassay for CAPRIN-1 itself contained in a sample, and a method (3rd method) that involves measurement of mRNA encoding CAPRIN-1 contained in a sample. In the method of the present invention, CAPRIN-1 expression may be measured by any of these methods. In the present invention, the term "measurement" refers to any of detection, qualitative determination, quantitative determination, and semi-quantitative determination.

[0025] The amino acid sequence shown in SEQ ID NO: 6, 8, 10, 12, or 14 is the amino acid sequence of canine CAPRIN-1. Canine CAPRIN-1 having the amino acid sequence was identified as a polypeptide binding to an antibody specifically existing in the cancer-bearing dog-derived serum by the SEREX method using a canine testis-derived cDNA library and the serum of a cancer-bearing dog (see Example 1). Specifically, an antibody against CAPRIN-1 having the amino acid sequence shown in SEQ ID NO: 6, 8, 10, 12, or 14 is specifically induced *in vivo* in a cancer-bearing dog. Therefore, canine cancer can be detected by measuring the above antibody against CAPRIN-1 having the amino acid sequence shown in SEQ ID NO: 6, 8, 10, 12, or 14 using the above 1st method (see Examples 3 and 4). Canine cancer

can also be detected by measuring CAPRIN-1 itself as an antigen shown in SEQ ID NO: 6, 8, 10, 12, or 14 using the above 2nd method (see Examples 5 and 6). Also, canine cancer can be detected, as described in the following Examples, by measuring mRNA encoding CAPRIN-1 since the mRNA is expressed at significantly high levels in testes and cancer cells (see Example 1).

[0026] The term "having an amino acid sequence" as used herein refers to amino acid residues being aligned in the relevant order. Therefore, for example, the expression "polypeptide having the amino acid sequence shown in SEQ ID NO: 2" refers to a polypeptide having 709 amino acid residues, which consists of the amino acid sequence of Met Pro Ser Ala · · · (abbreviated) · · Gln Gln Val Asn shown in SEQ ID NO: 2. Also, for example, the expression "polypeptide having the amino acid sequence shown in SEQ ID NO: 2" may also be abbreviated as "the polypeptide of SEQ ID NO: 2." "The same applies to the expression "having a/the nucleotide sequence." In this case, the term "having" may be substituted with the expressions "consisting of."

[0027] Also, the term "polypeptide" as used herein refers to a molecule that is formed via peptide bonding of a plurality of amino acids. Examples of such molecule include not only polypeptide molecules with a large number of constituent amino acids, but also low-molecular-weight molecules (oligopeptides) with small number of amino acids and full-length proteins. The present invention further encompasses full-length CAPRIN-1 proteins each having an amino acid sequence shown in an even-numbered sequence ID from among SEQ ID NOS: 2-30.

[0028] In the method of the present invention, not only canine CAPRIN-1 of SEQ ID NO: 6, 8, 10, 12, or 14, but also CAPRIN-1 of other mammals (hereinafter, may also be referred to as "homolog" for canine CAPRIN-1. When simply referred to as "CAPRIN-1," CAPRIN-1 from not only a dog but also from another mammal is also encompassed herein) are also subjected to measurement. As specifically described in the following Examples, mRNA encoding human CAPRIN-1 is significantly expressed at a high level in human testis and cancer cells, as in the case of canine CAPRIN-1 of SEQ ID NO: 6, 8, 10, 12 or 14. However, no antibody against the human CAPRIN-1 is detected in a healthy human body. Also, an antibody against feline CAPRIN-1 is not detected in a healthy cat body, but is detected in a cancer-bearing cat alone. Therefore, cancer of a mammal other than a dog can be detected by measuring CAPRIN-1 expression in the mammal. Examples of CAPRIN-1 of mammals other than dogs, which are measurement subjects in the method of the present invention, include, but are not limited to, human CAPRIN-1 and feline CAPRIN-1. A nucleotide sequence encoding human CAPRIN-1 and the amino acid sequence thereof are as separately shown in SEQ ID NO: 1 and 3, and 2 and 4, respectively, in the Sequence Listing. Sequence identity with canine CAPRIN-1 is 94% in terms of nucleotide sequence and is 98% in terms of amino acid sequence. Even dogs and humans which are genetically distant mammals share as very high as 98% sequence identity in terms of the amino acid sequence of CAPRIN-1. Therefore, it is thought that a dog and a mammal other than a human; that is, canine CAPRIN-1 and homolog thereof share sequence identity as high as about 85% or more. Therefore, CAPRIN-1, the expression of which is measured in the method of the present invention, has preferably 85% or more and more preferably 95% or more sequence identity with the amino acid sequence of canine CAPRIN-1 shown in SEQ ID NO: 6, 8, 10, 12, or 14. However, such examples are not particularly limited thereto.

[0029] In the 1st method above, the above antibody that can be present in a sample can be easily measured by immunoassay using an antigenic substance that undergoes an antigen-antibody reaction with the antibody. Immunoassay itself is a known conventional method as specifically described below. As an antigenic substance for immunoassay, the canine CAPRIN-1 of SEQ ID NO: 6, 8, 10, 12, or 14 that causes the induction of the antibody within a cancer-bearing dog body can be used. Furthermore, an antibody has cross-reactivity. Thus, even a molecule other than an antigenic substance actually having served as an immunogen can bind to an antibody induced against the immunogen via an antigen-antibody reaction, as long as a structure analogous to the epitope of the immunogen is present on the molecule. In particular, a protein from a mammal and homolog thereof from another mammal share high amino acid sequence identity and often have epitope structures analogous to each other. As specifically described in the following Examples, the canine CAPRIN-1 of SEQ ID NO: 6, 8, 10, 12, or 14 undergoes an antigen-antibody reaction not only with an antibody induced against the canine CAPRIN-1 within a cancer-bearing dog body, but also with an antibody induced against feline CAPRIN-1 within a cancer-bearing cat body. Moreover, human CAPRIN-1 undergoes an antigen-antibody reaction with the above antibody induced within cancer-bearing dog or cancer-bearing cat bodies. Accordingly, in the 1st method of the present invention, CAPRIN-1 from any mammal can be used as an antigen for immunoassay.

[0030] In general, when an antigenic substance is a protein or the like having a complicated structure and high molecular weight, a plurality of sites having different structures are present on the molecule. Therefore, a plurality of types of antibody capable of recognizing and binding to different sites of such antigenic substances are produced *in vivo*. Specifically, an antibody that is produced *in vivo* against an antigenic substance such as protein is a polyclonal antibody that is a mixture of a plurality of types of antibody. An antibody discovered by the present inventors is also a polyclonal antibody. It is specifically present in cancer-bearing living organism-derived serum and specifically binds to a recombinant CAPRIN-1 protein via an antigen-antibody reaction. The term "polyclonal antibody" used in the present invention refers to an antibody that exists in serum from a living organism containing an antigenic substance therein and is induced *in vivo* against the antigenic substance.

[0031] In Examples described later, polypeptides of SEQ ID NO: 6 and SEQ ID NO: 8 (canine CAPRIN-1) and the

polypeptide of SEQ ID NO: 2 (human CAPRIN-1) were prepared as antigens for immunoassay of specific antibodies in the cancer-bearing living animals. Then reactivity between these polypeptides and the above antibody in serum from a cancer-bearing living organism was confirmed. However, the above antibody is a polyclonal antibody, so that it naturally binds to a polypeptide consisting of the homolog of SEQ ID NO: 6, 8, or 2. Even in the case of a fragment of said polypeptides, it can bind to the above antibody contained in serum from a cancer-bearing living organism, since the polyclonal antibody can contain an antibody capable of recognizing the structure of the relevant fragment. That is, either a polypeptide (that is, full-length CAPRIN-1 protein) of the homolog of SEQ ID NO: 6, 8, or 2 or a fragment thereof can be similarly used for measurement of the above polyclonal antibody contained specifically in serum of a cancer-bearing living organism and is useful for cancer detection. Therefore, examples of a polypeptide to be used as an antigen for immunoassay in the 1st method of the present invention include, not only a polypeptide that consists of the full-length region of CAPRIN-1 (e.g., SEQ ID NO: 6, 8, or 2), but also a polypeptide fragment that consists of continuous 7 or more, preferably continuous 8 or more, 9 or more, or 10 or more amino acids in the amino acid sequence of CAPRIN-1 and undergoes an antigen-antibody reaction with a polyclonal antibody against CAPRIN-1 (hereinafter, may be conveniently referred to as "a specifically reactive partial polypeptide"). It is known in the art that a polypeptide of about 7 or more amino acid residues exerts antigenicity. However, if the number of amino acid residues constituting a polypeptide is too low, such polypeptide highly likely cross-reacts with antibodies, which exist in the sample, against proteins other than CAPRIN-1. Accordingly, in view of increasing the accuracy of immunoassay, the desirable number of amino acid residues of a polypeptide fragment may be preferably 30 or more or 50 or more, further preferably 100 or more or 150 or more, further preferably 300 or more, even more preferably 600 or more, and further preferably 1000 or more and 1500 or more.

[0032] Specific preferable examples of the polypeptides to be used as antigens are the polypeptides of the even-numbered SEQ ID NOS: 2-30 or fragments thereof.

[0033] Nucleotide sequences of polynucleotides encoding proteins consisting of the amino acid sequences of the even-numbered SEQ ID NOS: 2-30 (that is, SEQ ID NOS: 2, 4, 6 ... 28, 30) are shown in the odd-numbered SEQ ID NOS: 1-29 (that is, SEQ ID NOS: 1, 3, 5 ... 27, 29).

[0034] In general, it is broadly known by persons skilled in the art concerning protein antigens such that even when few amino acid residues have been substituted, deleted, added, or inserted in the amino acid sequence of the protein, the resultant may retain antigenicity almost equivalent to that of the original protein. Therefore, a polypeptide: having a sequence that has a substitution, a deletion, and/or an insertion of a few (preferably one or several) amino acid residues with respect to the amino acid sequence of CAPRIN-1 and has 80% or more, 85% or more, preferably 90% or more, more preferably 95% or more, and further preferably 98% or more sequence identity with the original sequence; and specifically binding to a polyclonal antibody against CAPRIN-1 via an antigen-antibody reaction (hereinafter, may be conveniently referred to as "specifically reactive modified polypeptide") can be used for cancer detection in a manner similar to that for the above polypeptides. Preferably, the specifically reactive modified polypeptide has an amino acid sequence that has a substitution, a deletion, an addition, and/or an insertion of one or several amino acid residues with respect to the amino acid sequence of CAPRIN-1. The term "several" as used herein refers to an integer of 2-10, preferably an integer of 2-6, and further preferably an integer of 2-4.

[0035] The term "sequence identity (of amino acid sequences)" as used herein is obtained by aligning two amino acid sequences to be compared so that amino acid residues match as many as possible, subtracting the number of amino acid residues that have matched from the total number of amino acid residues, and then expressing the result in percentage form. Upon the above alignment, if necessary, gaps are appropriately inserted into one of or both sequences to be compared. Such sequence alignment can be performed using a known program such as BLAST, FASTA, or CLUSTAL W (Karlin and Altschul, Proc. Natl. Acad. Sci. U.S.A., 87: 2264-2268, 1993; Altschul et al., Nucleic Acids Res., 25: 3389-3402, 1997).

[0036] Twenty types of amino acid constituting natural proteins can be grouped into neutral amino acids having side chains with low polarity (Gly, Ile, Val, Leu, Ala, Met, and Pro), neutral amino acids having hydrophilic side chains (Asn, Gln, Thr, Ser, Tyr, and Cys), acidic amino acids (Asp and Glu), basic amino acids (Arg, Lys, and His), and aromatic amino acids (Phe, Tyr, Trp, and His) in which the members of each group have properties analogous to each other. It is known that substitution among these amino acids (that is, conservative substitution) rarely alters the properties of the resulting polypeptide. Therefore, when amino acid residues of CAPRIN-1 are substituted, substitution is performed between members of the same group so that a possibility of maintaining binding with the corresponding antibody becomes higher. However, in the present invention, the above variant may involve non-conservative substitution, as long as immune-inducing activity equivalent to or almost equivalent to that of a non-variant is imparted.

[0037] A polypeptide (hereinafter, may conveniently be referred to as "specifically reactive addition polypeptide") that contains as a partial sequence the above polypeptide to be used in the present invention (that is, prepared by addition of another (poly)peptide to one end or both ends of a polypeptide to be used in the present invention) and specifically binds to a polyclonal antibody against CAPRIN-1 via an antigen-antibody reaction can also be used for cancer detection in a manner similar to that for the above polypeptides.

[0038] The above polypeptides to be used in the present invention can be synthesized according to a chemical

synthesis method such as an Fmoc method (fluorenylmethyloxycarbonyl method) and a tBoc method (t-butyloxy-carbonyl method) (Ed., The Japanese Biochemical Society, Seikagaku Jikken Koza (Biochemical Experimental Lecture Series) 1, Protein Chemistry IV, Chemical Modification and Peptide Synthesis, TOKYO KAGAKU DOZIN CO., LTD (Japan), 1981). Also, the polypeptides can also be synthesized by a conventional method using various commercially available peptide synthesizers. Alternatively, the polypeptides can be easily prepared using known genetic engineering techniques (Sambrook et al., Molecular Cloning, 2nd Edition, Current Protocols in Molecular Biology (1989), Cold Spring Harbor Laboratory Press, Ausubel et al., Short Protocols in Molecular Biology, 3rd Edition, A Compendium of Methods from Current Protocols in Molecular Biology (1995), John Wiley & Sons, and the like). For example, from RNA extracted from a tissue expressing a gene encoding the human CAPRIN-1 of SEQ ID NO: 2 or a homolog thereof, cDNA of the gene is prepared by RT-PCR. The full-length sequence or a desired partial sequence of the cDNA is incorporated into an expression vector and then the vector is introduced into host cells, so that a polypeptide of interest can be obtained. The nucleotide sequences of cDNAs encoding canine CAPRIN-1 of SEQ ID NOS: 6, 8, 10, 12, and 14 are shown in SEQ ID NOS: 5, 7, 9, 11, and 13, respectively. The human factors homolog thereof; that is, the nucleotide sequences of cDNAs encoding human CAPRIN-1 of SEQ ID NOS: 2 and 4 are shown in SEQ ID NOS: 1 and 3, respectively. Hence, primers to be used for RT-PCR can be easily designed in reference to these nucleotide sequences. Also, as described later, a gene encoding CAPRIN-1 of a non-human mammal can be amplified using primers designed in reference to the nucleotide sequences of the odd-numbered SEQ ID NOS: 1-29. For example, cDNA encoding feline CAPRIN-1 can be easily prepared by techniques similar to the above techniques. RNA extraction, RT-PCR, cDNA incorporation into a vector, and introduction of a vector into host cells can be performed by known methods as described below, for example. Also, vectors and host cells to be used herein are also known and various vectors and host cells are commercially available. **[0039]** The above host cells may be any cells, as long as they can express the above polypeptides. Examples of prokaryotic host cells include *Escherichia coli* and the like. Examples of eukaryotic host cells include mammalian cultured cells such as monkey kidney cells (COS1), Chinese hamster ovary cells (CHO), the human embryonic kidney cell line (HEK293), and the mouse embryonic skin cell line (NIH3T3), budding yeast, fission yeast, silkworm cells, and *Xenopus* oocytes.

[0040] When prokaryotic cells are used as host cells, an expression vector having a replication origin in prokaryotic cells, a promoter, a ribosome-binding site, a multi-cloning site, a terminator, a drug-resistance gene, an auxotrophic complementary gene, and the like are used. As expression vectors for *Escherichia coli*, pUC vectors, pBluescriptII, pET expression systems, pGEX expression systems, and the like can be exemplified. A DNA encoding the above polypeptide is incorporated into such an expression vector, prokaryotic host cells are transformed with the vector, and then the thus obtained transformant is cultured, so that the polypeptide encoded by the DNA can be expressed in the prokaryotic host cells. At this time, the polypeptide can also be expressed as a fusion protein with another protein. A DNA encoding the above polypeptide can be obtained by preparing a cDNA by RT-PCR as described above, for example. Moreover, such DNA encoding the above polypeptide can be also synthesized by a conventional method using a commercially available nucleic acid synthesizer as described below. The nucleotide sequences of cDNAs of the genes encoding CAPRIN-1 of SEQ ID NOS: 2 and 4 are shown in SEQ ID NOS: 1 and 3, respectively, in the Sequence Listing.

[0041] When eukaryotic cells are used as host cells, expression vectors for eukaryotic cells having a promoter, a splicing region, a poly(A) additional site, and the like are used. Examples of such expression vectors include pKA1, pCDM8, pSVK3, pMSG, pSVL, pBK-CMV, pBK-RSV, EBV vector, pRS, pcDNA3, and pYES2. Similarly to the above, a DNA encoding a polypeptide to be used in the present invention is incorporated into such an expression vector, eukaryotic host cells are transformed with the vector, and then the thus obtained transformant is cultured, so that the polypeptide encoded by the above DNA can be expressed in eukaryotic host cells. When pIND/V5-His, pFLAG-CMV-2, pEGFP-N1, pEGFP-C1, or the like is used as an expression vector, the above polypeptide can be expressed as a fusion protein with various tags, such as a His tag (e.g., (His)₆ to (His)₁₀), a FLAG tag, a myc tag, a HA tag, and GFP.

[0042] For introduction of an expression vector into a host cell, known methods can be employed such as electroporation, a calcium phosphate method, a liposome method, a DEAE dextran method, microinjection, viral infection, lipofection, and binding with a cell-membrane-permeable peptide.

[0043] Isolation and purification of a polypeptide of interest from host cells can be performed using known isolation techniques in combination. Examples of such known techniques include treatment using a denaturing agent such as urea or a surfactant, ultrasonication, enzymatic digestion, salting-out, solvent fractionation and precipitation, dialysis, centrifugation, ultrafiltration, gel filtration, SDS-PAGE, isoelectric focusing, ion exchange chromatography, hydrophobic chromatography, affinity chromatography, and reverse phase chromatography.

[0044] Polypeptides obtained by the above methods include polypeptides in the form of fusion proteins with any other proteins. An example of such a fusion protein include a fusion protein with glutathione-S-transferase (GST), a His tag, or the like. Polypeptides in the form of such fusion proteins are also examples of the above-described specifically reactive addition polypeptides and can be used for the 1st detection method of the present invention. Furthermore, a polypeptide expressed in transformed cells may be subjected to various types of modification within cells after translation. Such polypeptide that is modified after translation can be used in the 1st detection method of the present invention, as long

as it is capable of binding to a polyclonal antibody against CAPRIN-1. Examples of such post-translation modification include the removal of N-terminal methionine, N-terminal acetylation, glycosylation, limited proteolysis by intracellular protease, myristoylation, isoprenylation, and phosphorylation.

[0045] An antibody in a sample can be easily measured by immunoassay using the above polypeptide as an antigen. Immunoassay itself is known in the art. Immunoassay is classified into a sandwich method, a competition method, an agglutination method, Western blot method, and the like based on types of reaction. Also, immunoassay is classified based on labels into radioimmunoassay, fluorescence immunoassay, enzyme immunoassay, and biotin immunoassay, for example. Immunoassay of the above antibody can be performed using any of these methods. Sandwich ELISA or the agglutination method are preferably applicable as an immunoassay technique for the above antibody in the method of the present invention, since the procedures of these methods are convenient and require no extensive apparatus and the like. But the techniques are not limited to them. When an enzyme is used as a label for an antibody, such enzyme is not particularly limited, as long as it satisfies conditions such that: the turn over number is high; it remains stable even if it is bound to an antibody, it specifically causes the color development of the substrate, and the like. Examples of enzymes that can be used for general enzyme immunoassay include peroxidase, β -galactosidase, alkaline phosphatase, glucose oxidase, acetylcholine esterase, glucose-6-phosphorylation dehydrogenase, and malic acid dehydrogenase. Also, enzyme-inhibiting substances, coenzymes, and the like can be used. Binding of these enzymes with antibodies can be performed by known methods using a cross-linking agent such as a maleimide compound. As a substrate, a known substance can be used depending on the type of an enzyme to be used. For example, when peroxidase is used as an enzyme, 3,3',5,5'-tetramethylbenzidine can be used. Also when alkaline phosphatase is used as an enzyme, para-nitrophenol or the like can be used. As a radio isotope, a radio isotope that is generally used for radioimmunoassay, such as ^{125}I and ^3H can be used. As a fluorescent dye, a fluorescent dye that is used for general fluorescent antibody techniques, such as fluorescence isothiocyanate (FITC) and tetramethylrhodamine isothiocyanate (TRITC) can be used.

[0046] There is no need to explain the above immunoassay techniques in the Description, since they are well-known. However, when these immunoassay techniques are briefly described, the sandwich method involves immobilizing the above polypeptide to be used as an antigen to a solid phase, reacting it with a sample such as serum, washing, reacting with an appropriate secondary antibody, washing, and then measuring the secondary antibody bound to the solid phase, for example. An unbound secondary antibody can be easily removed by immobilization of an antigen polypeptide to a solid phase. Hence, this is preferable as an embodiment of the method for detecting cancer of the present invention. As a secondary antibody, an anti-canine IgG antibody can be used if a sample is derived from a dog. A secondary antibody is labeled in advance with a labeling substance exemplified above, so that the secondary antibody binding to a solid phase can be measured. The thus measured amount of the secondary antibody corresponds to the amount of the above antibody in the serum sample. When an enzyme is used as a labeling substance, the amount of the antibody can be measured by adding a substrate that is digested to develop color by enzymatic action and then optically measuring the amount of the substrate degraded. When a radio isotope is used as a labeling substance, the amount of radiation from the radio isotope can be measured using a scintillation counter or the like.

[0047] In the 2nd method of the present invention, CAPRIN-1 that can be contained in a sample from a living organism is measured. As described above, among cancer patients, the amount of an antibody that undergoes an antigen-antibody reaction with CAPRIN-1 of a dog, a human, or the like is significantly high. This indicates that the amount of CAPRIN-1 accumulated as an antigen is significantly high in cancer cells. Cancer can also be detected by directly measuring CAPRIN-1, as specifically described in Examples below. Therefore, cancer can be *detected in vivo* by measuring CAPRIN-1 itself similarly to the 1st method above.

[0048] A polypeptide in a sample can be easily measured by well-known immunoassay techniques. Specifically, for example, an antibody or an antigen-binding fragment thereof, which undergoes an antigen-antibody reaction with CAPRIN-1, is prepared, immunoassay is performed using the antibody or its antigen-binding fragment thereof, and then CAPRIN-1 that may be present in the sample can be measured. As described above, an antibody has cross-reactivity. Hence, for example, through the use of an antibody or the antigen-binding fragment thereof, which undergoes an antigen-antibody reaction with the canine CAPRIN-1 of SEQ ID NO: 6, not only the canine CAPRIN-1 of SEQ ID NO: 6, but also its homolog in other mammals (e.g., the human CAPRIN-1 of SEQ ID NO: 2 or 4 and feline CAPRIN-1) can be measured. An immunoassay technique itself is a known conventional technique as described above.

[0049] This examination revealed that CAPRIN-1 is a cell membrane protein that is expressed on the surfaces of cancer cells. A living organism with cancer contains many kinds of proteases. Specifically, in a living organism with cancer, an extracellularly expressed portion of the CAPRIN-1 sequence is separated from the cancer cells by degradation, so that such portion exists at a level higher than an intracellularly expressed portion of the CAPRIN-1 sequence. Therefore, when an antibody against CAPRIN-1 or an antigen-binding fragment thereof to be used in this measurement, which binds to the surface of the cancer cell, is used, CAPRIN-1 can be detected at higher levels and cancer can be diagnosed with higher sensitivity. Therefore, in the present invention, antibodies binding to a portion of the CAPRIN-1 protein existing on the surfaces of cancer cells, are preferably used. An example of a partial peptide of the CAPRIN-1 protein existing on the surfaces of cancer cells, is a polypeptide comprising a sequence of continuous 7 or more amino acid residues

within the region of amino acid residue Nos. (aa) 50-98 or amino acid residue Nos. (aa) 233-305 in the amino acid sequences shown in the even-numbered SEQ ID NOS: 2-30 in the Sequence Listing excluding SEQ ID NO: 6 and SEQ ID NO: 18. A specific example thereof is the amino acid sequence shown in SEQ ID NO: 43 or SEQ ID NO: 61 (in the amino acid sequence shown in SEQ ID NO: 61, a region of the amino acid sequence shown in SEQ ID NO: 62 or SEQ ID NO: 63 is preferred) or an amino acid sequence having 80% or more, preferably 85% or more, more preferably 90% or more, further preferably 95% or more sequence identity with the relevant amino acid sequence. Examples of an antibody to be used in the present invention include all antibodies binding to these peptides. Specific examples of the antibody include an antibody or antigen-binding fragment thereof which binds to SEQ ID NO: 43, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 44 and 45, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 44 and 46, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 44 and 47, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 44 and 48, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 49 and 50, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 51 and 52, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 53 and 54, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 55 and 56, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 57 and 58, or a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 59 and 60.

[0050] The term "antigen-binding fragment" as used herein refers to an antibody fragment capable of binding to an antigen such as a Fab fragment and a $F(ab')_2$ fragment contained in an antibody molecule. An antibody to be used herein may be a polyclonal antibody or a monoclonal antibody. For immunoassay and the like, a monoclonal antibody with high reproducibility is preferable. A method for preparing a polyclonal antibody and a monoclonal antibody using a polypeptide as an immunogen is known and can be easily performed by a conventional method. For example, CAPRIN-1 is bound to a carrier protein such as keyhole limpet hemocyanin (KLH), casein, or serum albumin and then an animal is immunized with the resultant as an immunogen together with an adjuvant, and thereby an antibody against CAPRIN-1 can be induced. Antibody-producing cells such as splenocytes or lymphocytes collected from the immunized animal are fused to myeloma cells to prepare hybridomas, and then hybridomas producing an antibody that binds to CAPRIN-1 are selected and then grown, so that a monoclonal antibody, whose the corresponding antigen is CAPRIN-1, can be obtained from the cultured supernatant. The above method is a known conventional method.

[0051] In the 3rd method of the present invention, mRNA encoding CAPRIN-1 that can be contained in a sample obtained from a living organism is measured. As specifically described in Examples below, mRNA encoding the canine CAPRIN-1 of SEQ ID NO: 6, 8, 10, 12, or 14 or human CAPRIN-1 of SEQ ID NO: 2 or 4 is expressed at a significantly high level in cancer cells. Therefore, cancer can be detected *in vivo* by measuring such mRNA in a sample.

[0052] mRNA in a sample can be quantitatively determined by a conventional method such as real-time detection RT-PCR using the mRNA as a template, for example. Such mRNA can generally be quantitatively determined based on staining intensity or the like in Northern blot that is a conventional method. The cDNA sequences encoding CAPRIN-1 polypeptides of the even-numbered SEQ ID NOS: 2-30 are shown in the odd-numbered SEQ ID NOS: 1-29, respectively. Hence, based on these sequences, a polynucleotide specifically hybridizing to a partial region in the nucleotide sequence shown in any of the odd-numbered SEQ ID NOS: 1-29 (hereinafter, referred to as "polynucleotide for cancer detection") is prepared and then the amount of the mRNA in a sample can be measured using the polynucleotide as a probe or a primer for a nucleic acid amplification method. As described later, if it is a polynucleotide specifically hybridizing to a partial region in the nucleotide sequence shown in any of the odd-numbered SEQ ID NOS: 1-29, mRNA encoding CAPRIN-1 in mammals other than dogs and humans can also be determined. In addition, in the present invention, a polynucleotide may be RNA or DNA.

[0053] The term "specifically hybridizing to" as used herein refers to that under general hybridization conditions, a subject hybridizes to only a target partial region, but does not substantially hybridize to the other regions.

[0054] The term "(under) general hybridization conditions" as used herein refers to conditions employed for annealing in general PCR or detection using a probe. For example, in the case of PCR using Taq polymerase, the term refers to conditions under which a reaction is performed at an appropriate annealing temperature ranging from about 54°C to 60°C using a general buffer such as 50 mM KCl, 10 mM Tris-HCl (pH8.3-9.0), and 1.5 mM MgCl₂. Also, in the case of Northern hybridization, for example, the term refers to conditions under which a reaction is performed using a general hybridization solution such as 5 × SSPE, 50% formamide, 5 × Denhardt's solution, and 0.1% SDS-0.5% SDS, or 0.1-5 × SSC and 0.1-0.5% SDS at an appropriate hybridization temperature ranging from about 42°C to 65°C. Furthermore, after hybridization, washing is performed with 0.1-0.2 × SSC and 0.1% SDS, for example. However, appropriate annealing temperatures or hybridization temperatures are not limited to the above examples, and are determined based on T_m value for a polynucleotide for cancer detection, which is used as a primer or a probe, and the empirical rule of experimenters. Persons skilled in the art can easily determine such temperature range.

[0055] The expression "does not substantially hybridize to" as used herein refers to that a subject does not really

hybridize to a target partial region or a subject hybridizes to a target partial region in a significantly low amount; that is, in a relatively negligibly-small amount, even when it hybridizes to the target partial region. An example of a polynucleotide specifically hybridizing under such conditions is a polynucleotide having sequence identity at a level or more with the nucleotide sequence of a target partial region. A specific example of such polynucleotide has 70% or more, preferably 80% or more, 85% or more, more preferably 90% or more, further preferably 93% or more, further preferably 95% or more, and further more preferably 98% or more sequence identity. Most preferably, the polynucleotide has a nucleotide sequence identical to the nucleotide sequence of a target partial region. Sequence identity is defined in the same manner as that for the sequence identity of the above amino acid sequence. Even if a terminus of a polynucleotide for cancer detection contains a region not hybridizing to a subject, in the case of a probe, it can be used for detection as long as a hybridizing region occupies as much as about a half or more of the entire probe. Also, in the case of a primer, it can be used for detection as long as a hybridizing region occupies as much as about a half or more of the entire primer and is located on the 3' terminal side, since normal annealing and extension reaction can take place. As described above, when a terminus of a polynucleotide for cancer detection contains a non-hybridizing region, sequence identity with a target nucleotide sequence is calculated focusing on only a hybridizing region without taking non-hybridizing region into consideration.

[0056] The term "partial sequence" in the present invention refers to a partial sequence in the nucleotide sequences shown in the odd-numbered SEQ ID NOS: 1-29, specifically the partial sequence having a sequence of continuous 15 or more nucleotides, preferably continuous 18 or more nucleotides, more preferably continuous 20 or more nucleotides or 25 or more nucleotides, and further preferably continuous 30, 40, or 50 or more nucleotides. The expression "the nucleotide sequence shown in SEQ ID NO: 5" as used herein refers to, in addition to the nucleotide sequence actually shown in SEQ ID NO: 5, a sequence complementary to the sequence. Therefore, for example, the expression "a polynucleotide having the nucleotide sequence shown in SEQ ID NO: 5" refers to a single-stranded polynucleotide having the nucleotide sequence actually shown in SEQ ID NO: 5, a single-stranded polynucleotide having a nucleotide sequence complementary to that shown in SEQ ID NO: 5, and a double-stranded polynucleotide comprising them. When a polynucleotide to be used in the present invention is prepared or a polynucleotide encoding a polypeptide to be used in the present invention is prepared, any one nucleotide sequence is appropriately selected and this selection can be easily performed by persons skilled in the art.

[0057] The number of nucleotides in a polynucleotide for cancer detection is preferably 18 or more nucleotides in view of ensuring specificity. When used as a probe, the size of the polynucleotide is preferably 18 or more nucleotides, is further preferably 20 or more nucleotides and the full-length or less of the coding region. When used as a primer, the size of the polynucleotide is preferably 18 or more nucleotides and 50 or less nucleotides. A preferred example of the polynucleotide for cancer detection is a polynucleotide comprising continuous 18 or more nucleotides in a nucleotide sequence shown in any of the odd-numbered SEQ ID NOS: 1-29.

[0058] It is obvious for persons skilled in the art who refer this Description that: a polynucleotide specifically hybridizing to a partial region in SEQ ID NO: 5, 7, 9, 11, or 13 is used for measurement of the amount of mRNA encoding the canine CAPRIN-1 of SEQ ID NO: 6, 8, 10, 12, or 14, respectively; and a polynucleotide specifically hybridizing to a partial region in SEQ ID NO: 1 or 3 is used for measurement of the amount of mRNA encoding the human CAPRIN-1 of SEQ ID NO: 2 or 4, respectively. However, a protein from a mammal and a homolog thereof from another mammal generally share high sequence identity even at the nucleotide sequence level. Thus, the sequence identity among the sequences of the odd-numbered SEQ ID NOS: 1-13 also is as very high as 94% to 100%. Accordingly, a polynucleotide specifically hybridizing to a partial region of the sequence of SEQ ID NO: 5 can also specifically hybridize to a partial region corresponding to the relevant partial region of any of the odd-numbered SEQ ID NOS: 1-29.

[0059] Actually as described in Examples below, a pair of primers having the nucleotide sequences shown in SEQ ID NO: 33 and 34, respectively, specifically hybridizes to both a partial region of any of the sequences of the odd-numbered SEQ ID NOS: 1-29 and a partial region of the sequence of SEQ ID NO: 5, so that both mRNA encoding the canine CAPRIN-1 of SEQ ID NO: 6 and mRNA encoding a homolog thereof can be measured, for example. Accordingly, for example, with the use of a polynucleotide specifically hybridizing to a partial region of the sequence of SEQ ID NO: 5, not only mRNA encoding the canine CAPRIN-1 of SEQ ID NO: 6, but also mRNA encoding the human CAPRIN-1 of SEQ ID NO: 2 or 4 can be measured. Similarly, a mRNA encoding CAPRIN-1 of another mammal such as a cat can also be measured. When a polynucleotide for cancer detection is designed, it is desirable to select partial regions having a specifically high sequence identity between the SEQ ID numbers (odd-numbered SEQ ID NOS: 1-29) (preferably, the nucleotide sequences are the same). If a partial region having particularly high sequence identity between a dog and a human is present, a region having very high sequence identity with the region is expected to be present in a homologous gene of another animal species. Through selection of such partial regions, accuracy for measuring mRNA encoding CAPRIN-1 of an animal species other than dogs and humans can be increased.

[0060] A method itself for measuring a test nucleic acid using a polynucleotide specifically hybridizing to a partial region of the test nucleic acid as a primer or a probe for a nucleic acid amplification method such as PCR is well-known. Examples of such method include, in addition to RT-PCR that is specifically described in Examples below, Northern blot

and In situ hybridization. When the amount of mRNA is measured in the present invention, all of these known measuring methods can be employed.

[0061] A nucleic acid amplification method itself such as PCR is well-known in the art and thus reagent kits and apparatuses therefor are commercially available, so that the method can be easily performed. Specifically, for example, denaturation, annealing, and extension steps are each performed using a test nucleic acid (e.g., the cDNA of a gene encoding a protein having an amino acid sequence shown in any of the even-numbered SEQ ID NOS: 2-30) as a template and a pair of polynucleotides (primers) for cancer detection in a known buffer in the presence of thermostable DNA polymerase such as Taq polymerase or Pfu polymerase and dNTP (here, N = A, T, C, or G) by varying the temperature of the reaction solution. In general, the denaturation step is performed at 90°C-95°C, the annealing step is performed at or near the T_m of the template and the primers (preferably within $\pm 4^\circ\text{C}$), and the extension step is performed at 72°C which is an optimum temperature for thermostable DNA polymerase such as Taq polymerase or Pfu polymerase or a temperature near the optimum temperature. Each step is performed for about 30 seconds to 2 minutes, as appropriately selected. This heating cycle is repeated about 25 to 40 times, for example, so that the template nucleic acid region flanked by a primer pair is amplified. A nucleic acid amplification method is not limited to PCR and any other nucleic acid amplification methods known in the art can be employed herein. As described above, when a nucleic acid amplification method is performed using a pair of polynucleotides for cancer detection as primers and a test nucleic acid as a template, the test nucleic acid is amplified. However, if no test nucleic acid is contained in a sample, amplification does not take place. Hence, through detection of amplification products, the presence or the absence of the test nucleic acid in a sample can be confirmed. An amplification product can be detected by a method that involves subjecting a reaction solution after amplification to electrophoresis, and then staining the band with ethidium bromide or the like or a method that involves immobilizing an amplification product after said electrophoresis onto a solid phase such as a nylon membrane, performing hybridization with a labeling probe that specifically hybridizes to a test nucleic acid, washing, and then detecting the label. Also, namely real-time detection PCR is performed using a quencher fluorescent dye and a reporter fluorescent dye, and thereby the amount of a test nucleic acid in a specimen can be quantitatively determined. Since kits for real-time detection PCR are commercially available, real-time detection PCR can be easily performed. Furthermore, semi-quantitative determination of a test nucleic acid is also possible based on electrophoresis band intensity. A test nucleic acid may be either mRNA or cDNA resulting from mRNA via reverse transcription. When mRNA is amplified as a test nucleic acid, a NASBA method (3SR method or TMA method) using the above primer pair can also be employed. The NASBA method itself is well-known and kits for the method are also commercially available, so that the method can be easily performed using the above primer pair.

[0062] As a probe, a labeled probe that is prepared by labeling a polynucleotide for cancer detection with a fluorescent label, a radiolabel, a biotin label, or the like can be used. A method for labeling a polynucleotide itself is well-known. The presence or the absence of a test nucleic acid in a sample can be examined by immobilizing a test nucleic acid or an amplification product thereof, performing hybridization with a labeled probe, washing, and then measuring the label bound to the solid phase. Alternatively, a polynucleotide for cancer detection is immobilized, a test nucleic acid is hybridized thereto, and then the test nucleic acid bound to the solid phase can be detected using the labeled probe or the like. In such a case, a polynucleotide for cancer detection bound to a solid phase is also referred to as a probe. A method for measuring a test nucleic acid using a polynucleotide probe is also known in the art. The method can be performed by causing, in a buffer, a polynucleotide probe to come into contact with a test nucleic acid at T_m or near T_m (preferably, within $\pm 4^\circ\text{C}$) for hybridization, washing, and then measuring the labeled probe that has hybridized or the template nucleic acid bound to the solid-phase probe. Examples of such method include well-known methods such as Northern blot, in situ hybridization, and Southern blot methods. In the present invention, any well-known method is applicable.

[0063] It is determined by the detection method of the present invention whether or not a subject animal has cancer based on the expression level of CAPRIN-1 measured as described above. Cancer can be detected only by measuring CAPRIN-1 expression in a subject animal. However, it is preferable in view of enhancing detection accuracy to examine the expression levels (antibody level, polypeptide level, or mRNA level) of CAPRIN-1 in one or a plurality of samples of healthy subjects so as to obtain a standard value of healthy subjects and then to compare the measured value of a subject animal with the standard value obtained from healthy subjects. To further enhance detection accuracy, CAPRIN-1 expression levels are examined for samples obtained from many patients found to have cancer so as to obtain a standard value of cancer patients and then the measured value of a subject animal may be compared with both the standard value of healthy subjects and the standard value of cancer patients. The above standard values can be determined by quantifying the CAPRIN-1 expression level in each sample and then calculating the mean value thereof, for example. A standard value of healthy subjects and the same of cancer patients can be determined in advance by examining CAPRIN-1 expression levels in many healthy subjects and cancer patients. Therefore, when comparison with a standard value is performed in the method of the present invention, a standard value determined in advance may be used.

[0064] In the detection method of the present invention, diagnosis based on other cancer antigens or cancer markers

may be used in combination. Accordingly, cancer detection accuracy can be further increased. For example, when an antibody specifically existing in cancer patients is measured by the method of the present invention, another polypeptide that is often expressed in a cancer tissue can be used in combination as an antigen in a manner similar to that for polypeptides above. Also, the method of the present invention and diagnosis using a previously known cancer marker may be performed in combination.

[0065] Cancer can be detected *in vivo* according to the detection method of the present invention. Particularly, as described in Examples below, even a small-size tumor, which is invisible to the naked eye, or a tumor in a deep part *in vivo* can be detected according to the method of the present invention. Thus, the method of the present invention is useful for early cancer detection. Also, through application of the detection method of the present invention for a patient during follow-up after treatment of cancer, cancer can be detected early if a cancer recurrence has taken place.

[0066] Also, in a cancer-bearing living organism, as the number of cancer cells expressing CAPRIN-1 measured in the present invention increases, the amounts of the protein and its mRNA accumulated in the living organism increase and the production amount of the antibody against CAPRIN-1 in serum increases. Meanwhile, as the number of cancer cells decreases, the amounts of the protein and its mRNA accumulated *in vivo* decrease and the amount of the antibody against CAPRIN-1 in serum decreases. Therefore, when the expression level of CAPRIN-1 is higher than that of a control, it can be determined that a tumor increase or a cancer metastasis is occurring; that is, the extent of cancer is advanced. Actually, as specifically described in the Examples below, an increase in the above serum antibody level in a cancer-bearing living organism was observed in association with cancer progression (malignant) such as tumor increase and metastasis. As described above, the extent of cancer can also be detected by the method of the present invention.

[0067] Also, as described in Examples below, among tumors of the same type, the above antibody levels in malignant type tumors were significantly higher than those in benign type tumors. Accordingly, when the expression level of CAPRIN-1 is high, it can be determined that cancer malignancy is higher. Specifically, cancer malignancy can also be detected by the method of the present invention.

[0068] Cancer to be subjected to the method for detecting cancer of the present invention is cancer expressing CAPRIN-1. Examples of such cancer include, but are not limited to, brain tumor, squamous cell carcinoma of the head, neck, lung, uterus or esophagus, melanoma, adenocarcinoma of the lung or uterus, renal cancer, malignant mixed tumor, hepatocellular carcinoma, basal cell carcinoma, acanthoma-like gingival tumor, tumor of the oral cavity, perianal adenocarcinoma, anal sac tumor, anal sac apocrine adenocarcinoma, sertoli cell carcinoma, cancer of the vaginal vestibule, sebaceous adenocarcinoma, sebaceous epithelioma, sebaceous adenoma, sweat gland carcinoma, intranasal adenocarcinoma, nasal adenocarcinoma, thyroid cancer, large-bowel cancer, bronchial adenocarcinoma, adenocarcinoma, ductal carcinoma, breast adenocarcinoma, composite type breast adenocarcinoma, malignant mammary mixed tumor, intraductal papillary adenocarcinoma, fibrosarcoma, hemangiopericytoma, osteosarcoma, chondrosarcoma, soft tissue sarcoma, histiocytic sarcoma, myxosarcoma, undifferentiated sarcoma, lung cancer, mastocytoma, cutaneous leiomyoma, intraperitoneal leiomyoma, leiomyoma, chronic lymphocytic leukemia, lymphoma, gastrointestinal lymphoma, digestive lymphoma, small-cell-to-medium-cell lymphoma, adrenomedullary tumor, granulosa cell tumor, and pheochromocytoma. Also, a living organism to be subjected to the method of the present invention is a mammal and is preferably a human, a dog, or a cat.

[0069] Examples of a sample to be subjected to the method of the present invention include body fluids such as blood, serum, blood plasma, ascites, and pleural effusion, tissues, and cells. In particular, in the 1st method and the 2nd method above, serum, blood plasma, ascites, and pleural effusion can be preferably used and in the 3rd method above for measurement of mRNA, tissue samples and cell samples are preferable.

[0070] The above polypeptides to be used as antigens for immunoassay in the 1st method (that is, the canine CAPRIN-1 of SEQ ID NO: 2 and a homolog thereof, a specifically reactive partial polypeptide, a specifically reactive modified polypeptide, and a specifically reactive addition polypeptide) can be provided as reagents for cancer detection. The reagent may consist of only the above polypeptide or may contain various additives or the like, for example, useful for stabilization of the polypeptide. Also, the reagent can be provided in a form immobilized onto a solid phase such a plate or a membrane. Preferable examples of the polypeptide are as described above.

[0071] An antibody that undergoes an antigen-antibody reaction with CAPRIN-1 or an antigen-binding fragment thereof, which is used for immunoassay of CAPRIN-1 itself in the 2nd method, can also be provided as a reagent for cancer detection. The reagent for cancer detection in this case may also consist of only the above antibody or an antigen-binding fragment thereof or may contain various additives or the like useful for stabilization and the like of the antibody or an antigen-binding fragment thereof. Also, the antibody or an antigen-binding fragment thereof may be in a form binding to a metal such as manganese or iron. When such metal-bound antibody or antigen-binding fragment thereof is administered to the body of a living organism, the metal-bound antibody or antigen-binding fragment thereof is accumulated at an increased level at a site where the antigen protein is present at a higher level. Therefore, the metal is measured by MRI or the like, and thereby the presence of cancer cells producing the antigen protein can be detected.

[0072] Furthermore, the above polynucleotide for cancer detection to be used for mRNA measurement in the 3rd method can also be provided as a reagent for cancer detection. The reagent for cancer detection in this case may also

consist of only the polynucleotide or may contain various additives and the like useful for stabilization and the like of the polynucleotide. The polynucleotide for cancer detection contained in the reagent is preferably a primer or a probe. Conditions and preferable examples of the polynucleotide for cancer detection are as described above.

5 EXAMPLES

[0073] The present invention will be described in more detail with reference to the examples set forth below; however, the technical scope of the present invention is not limited to the examples.

10 Example 1: Obtainment of new cancer antigen protein by SEREX method

(1) Construction of cDNA library

[0074] Total RNA was extracted from a testis tissue of a healthy dog by an Acid guanidium-Phenol-Chloroform method and then a polyA RNA was purified using Oligotex-dT30 mRNA purification Kit (Takara Shuzo Co., Ltd.) according to protocols included with the kit.

[0075] A canine testis cDNA phage library was synthesized using the thus obtained mRNA (5 μ g). The cDNA phage library was constructed using a cDNA Synthesis Kit, a ZAP-cDNA Synthesis Kit, and a ZAP-cDNA GigapackIII Gold Cloning Kit (STRATAGENE) according to protocols included with the kits. The size of the thus constructed cDNA phage library was 7.73×10^5 pfu/ml.

(2) Screening of cDNA library using serum

[0076] Immunoscreening was performed using the above constructed canine testis cDNA phage library. Specifically, host *Escherichia coli* (XL1-Blue MRF') was infected with the phage on an NZY agarose plate ($\Phi 90 \times 15$ mm) so as to obtain 2210 clones. *E. coli* cells were cultured at 42°C for 3 to 4 hours to form plaques. The plate was covered with a nitrocellulose membrane (Hybond C Extra: GE Healthcare Bio-Science) impregnated with IPTG (isopropyl- β -D-thiogalactoside) at 37°C for 4 hours, so that the protein was induced, expressed, and then transferred to the membrane. Subsequently, the membrane was collected and then immersed in TBS (10 mM Tris-HCl, 150 mM NaCl, and pH 7.5) containing 0.5% powdered skim milk, followed by overnight shaking at 4°C, thereby suppressing nonspecific reaction. The filter was reacted with a 500-fold diluted serum of a canine patient at room temperature for 2 to 3 hours.

[0077] As the above serum of a canine patient, a serum collected from a canine patient with breast cancer was used. These sera were stored at -80°C and then subjected to pre-treatment immediately before use. A method for pretreatment of serum is as follows. Specifically, host *Escherichia coli* (XL1-Blue MRF') was infected with a λ ZAP Express phage in which no foreign gene had been inserted and then cultured overnight on a NZY plate medium at 37°C. Subsequently, buffer (0.2 M NaHCO_3 and pH 8.3) containing 0.5 M NaCl was added to the plate, the plate was left to stand at 4°C for 15 hours, and then a supernatant was collected as an *Escherichia coli*/phage extract. Next, the thus collected *Escherichia coli*/phage extract was applied to an NHS-column (GE Healthcare Bio-Science), so that an *Escherichia coli*-phage-derived protein was immobilized. The serum of a canine patient was applied to the protein-immobilized column for reaction and then *Escherichia coli* and an antibody adsorbed to the phage were removed from the serum. The serum fraction that had passed through the column was diluted 500-fold with TBS containing 0.5% powdered skim milk. The resultant was used as an immunoscreening material.

[0078] A membrane onto which the treated serum and the above fusion protein had been blotted was washed 4 times with TBS-T (0.05% Tween20/TBS) and then caused to react with goat anti-canine IgG (Goat anti-Dog IgG-h+I HRP conjugated (BETHYL Laboratories)) diluted 5000-fold with TBS containing 0.5% powdered skim milk as a secondary antibody for 1 hour at room temperature. Detection was performed via an enzyme coloring reaction using an NBT/BCIP reaction solution (Roche). Colonies that matched sites positive for a coloring reaction were collected from the NZY agarose plate ($\Phi 90 \times 15$ mm) and then suspended in 500 μ l of an SM buffer (100 mM NaCl, 10 mM MgClSO_4 , 50 mM Tris-HCl, 0.01% gelatin, and pH 7.5). Until colonies positive for coloring reaction were unified, secondary screening and tertiary screening were repeated by a method similar to the above, so that 30,940 phage clones reacting with serum IgG were screened. Thus, 5 positive clones were isolated.

(3) Homology search for isolated antigen gene

[0079] For nucleotide sequence analysis of the 5 positive clones isolated by the above method, a procedure for conversion from phage vectors to plasmid vectors was performed. Specifically, 200 μ l of a solution was prepared to contain host *Escherichia coli* (XL1-Blue MRF') so that absorbance OD_{600} was 1.0. The solution was mixed with 250 μ l of a purified phage solution and then with 1 μ l of an ExAssist helper phage (STRATAGENE), followed by 15 minutes of

reaction at 37°C. Three milliliters of LB medium was added and then culture was performed at 37°C for 2.5 to 3 hours. Immediately after culture, the temperature of the solution was kept at 70°C by water bath for 20 minutes, centrifugation was performed at 4°C and 1000 × g for 15 minutes, and then the supernatant was collected as a phagemid solution. Subsequently, 200 µl of a solution was prepared to contain phagemid host *Escherichia coli* (SOLR) so that absorbance

OD₆₀₀ was 1.0. The solution was mixed with 10 µl of a purified phage solution, followed by 15 minutes of reaction at 37°C. The solution (50 µl) was seeded on LB agar medium containing ampicillin (to a final concentration of 50 µg/ml) and then cultured overnight at 37°C. Transformed SOLR single colonies were collected and then cultured in LB medium containing ampicillin (final concentration: 50 µg/ml) at 37°C. A plasmid DNA containing an insert of interest was purified using a QIAGEN plasmid Miniprep Kit (QIAGEN).

[0080] The purified plasmid was subjected to analysis of the full-length sequence by a primer walking method using the T3 primer according to SEQ ID NO: 31 and the T7 primer according to SEQ ID NO: 32. As a result of sequence analysis, the gene sequences according to SEQ ID NOS: 5, 7, 9, 11, and 13 were obtained. A homology search program, BLAST search (<http://www.ncbi.nlm.nih.gov/BLAST/>), was performed using the nucleotide sequences of the genes and amino acid sequences (SEQ ID NOS: 6, 8, 10, 12, and 14) of the proteins encoded by the genes. As a result of this homology search with known genes, it was revealed that all of the 5 obtained genes encoded CAPRIN-1. Regarding regions to be translated to proteins, the sequence identity among the 5 genes was 100% in terms of nucleotide sequence and 99% in terms of amino acid sequence. Also, regarding regions to be translated to proteins, the sequence identity between the genes and genes encoding human homolog thereof was 94% in terms of nucleotide sequence and 98% in terms of amino acid sequence. The nucleotide sequences of the human homolog are shown in SEQ ID NOS: 1 and 3 and the amino acid sequences of the same are shown in SEQ ID NOS: 2 and 4. Also, regarding regions to be translated to proteins, the sequence identity between the obtained canine genes and a gene encoding a cattle homolog was 94% in terms of nucleotide sequence and 97% in terms of amino acid sequence. The nucleotide sequence of the cattle homolog is shown in SEQ ID NO: 15 and the amino acid sequence of the same is shown in SEQ ID NO: 16. Regarding regions to be translated to proteins, the sequence identity between the genes encoding the human homolog and the gene encoding the cattle homolog was 94% in terms of nucleotide sequences and ranged from 93% to 97% in terms of amino acid sequence. Also, regarding regions to be translated to proteins, the sequence identity between the obtained canine genes and a gene encoding an equine homolog was 93% in terms of nucleotide sequence and 97% in terms of amino acid sequence. The nucleotide sequence of the equine homolog is shown in SEQ ID NO: 17 and the amino acid sequence of the same is shown in SEQ ID NO: 18. Regarding regions to be translated to proteins, the sequence identity between the genes encoding the human homolog and the gene encoding the equine homolog was 93% in terms of nucleotide sequence and 96% in terms of amino acid sequence. Also, regarding regions to be translated to proteins, the sequence identity between the obtained canine genes and genes encoding mouse homolog ranged from 87% to 89% in terms of nucleotide sequence and ranged from 95% to 97% in terms of amino acid sequence. The nucleotide sequences of the mouse homolog are shown in SEQ ID NOS: 19, 21, 23, 25, and 27 and the amino acid sequences of the same are shown in SEQ ID NOS: 20, 22, 24, 26, and 28. Regarding regions to be translated to proteins, the sequence identity between the genes encoding the human homolog and the genes encoding the mouse homolog ranged from 89% to 91% in terms of nucleotide sequence and ranged from 95% to 96% in terms of amino acid sequence. Also, regarding regions to be translated to proteins, the sequence identity between the obtained canine genes and a gene encoding a chicken homolog was 82% in terms of nucleotide sequence and 87% in terms of amino acid sequence. The nucleotide sequence of the chicken homolog is shown in SEQ ID NO: 29 and the amino acid sequence of the same is shown in SEQ ID NO: 30. Regarding regions to be translated to proteins, the sequence identity between the genes encoding the human homolog and the gene encoding the chicken homolog ranged from 81% to 82% in terms of nucleotide sequence and was 86% in terms of amino acid sequence.

(4) Gene expression analysis in each tissue

[0081] Expression of the genes obtained by the above method in canine and human normal tissues and various cell lines was examined by an RT-PCR (Reverse Transcription-PCR) method. A reverse transcription reaction was performed as follows. Specifically, total RNA was extracted from each tissue (50 mg to 100 mg) and each cell line (5 to 10 × 10⁶ cells) using a TRIZOL reagent (Invitrogen Corporation) according to protocols included therewith. cDNA was synthesized using the total RNA and Superscript First-Strand Synthesis System for RT-PCR (Invitrogen Corporation) according to protocols included therewith. PCR was performed as follows using primers specific to the obtained genes (according to SEQ ID NOS: 33 and 34). Specifically, PCR was performed by preparing a reaction solution adjusted to a total amount of 25 µl through addition of each reagent and an included buffer (0.25 µl of a sample prepared by reverse transcription reaction, the above primers (2 µM each), dNTP (0.2 mM each), and 0.65 U of ExTaq polymerase (Takara-baio Co., Ltd.)) and then by reacting the solution through repeating 30 times a cycle of 94°C/30 seconds, 60°C/30 seconds, and 72°C/30 seconds using a Thermal Cycler (BIO RAD). The gene-specific primers mentioned above were used to amplify the region between nucleotide 206 and nucleotide 632 in the nucleotide sequence of SEQ ID NO: 5 (canine CAPRIN-

1 gene) and the region between nucleotide 698 and nucleotide 1124 in the nucleotide sequence of SEQ ID NO: 1 (human CAPRIN-1 gene). For control, GAPDH-specific primers (according to SEQ ID NOS: 35 and 36) were used at the same time. As a result, as shown in Fig. 1, strong expression was observed in testis in the case of healthy canine tissues, while expression was observed in canine breast cancer and adenocarcinoma tissues. Furthermore, expression of the human homolog of the obtained genes was also confirmed. As a result, similarly to the case of canine CAPRIN-1 genes, expression could be confirmed only in the testis in the case of normal tissues. However, in the case of cancer cells, expression was detected in many types of cancer cell line, such as cell lines of breast cancer, brain tumor, leukemia, lung cancer, and esophageal cancer. Expression was confirmed in a particularly large number of breast cancer cell lines. Based on the results, it was confirmed that CAPRIN-1 expression was not observed in normal tissues other than those of the testis, while CAPRIN-1 was expressed in many cancer cells and particularly in breast cancer cell lines.

[0082] In addition, in Fig. 1, Reference No. 1 along the longitudinal axis indicates the expression pattern of each of the above-identified genes and Reference No. 2 along the same indicates the expression pattern of the GAPDH gene for control.

(5) Immunohistochemical staining

(5)-1 CAPRIN-1 expression in normal mouse and canine tissues

[0083] Mice (Balb/c, female) and dogs (beagle dogs, female) were exsanguinated under ether anesthesia and ketamine/isoflurane anesthesia. After laparotomy, organs (stomach, liver, eyeball, thymus gland, muscle, bone marrow, uterus, small intestine, esophagus, heart, kidney, salivary gland, large intestine, mammary gland, brain, lung, skin, adrenal gland, ovary, pancreas, spleen, and bladder) were each transferred to a 10cm dish containing PBS. Each organ was cut open in PBS and then fixed by perfusion overnight with 0.1 M phosphate buffer (pH 7.4) containing 4% paraformaldehyde (PFA). The perfusate was discarded, the tissue surface of each organ was rinsed with PBS, and then a PBS solution containing 10% sucrose was added to a 50ml centrifugal tube. Each tissue was then placed in each tube and then shaken using a rotor at 4°C for 2 hours. Each solution was substituted with a PBS solution containing 20% sucrose and then left to stand at 4°C until tissues precipitated. Each solution was substituted with a PBS solution containing 30% sucrose and then left to stand at 4°C until tissues precipitated. Each tissue was removed and a necessary portion was excised with a surgical scalpel. Next, an OCT compound (Tissue Tek) was applied and spread over each tissue surface, and then the tissues were placed on Cryomold. Cryomold was placed on dry ice for rapid freezing. Tissues were sliced into pieces 10 to 20 μ m long using a cryostat (LEICA) and then the sliced tissue pieces were air-dried on glass slides for 30 minutes using a hair dryer, so that glass slides onto which sliced tissue pieces had been applied were prepared. Next, each glass slide was placed in a staining bottle filled with PBS-T (saline containing 0.05% Tween20), so that a procedure involving exchange with PBS-T every 5 minutes was performed 3 instances. Excess water around each specimen was removed using Kimwipes and then each section was encircled using DAKOPEN (DAKO). As blocking solutions, a MOM mouse Ig blocking reagent (VECTASTAIN) was applied onto mouse tissue and PBS-T solution containing a 10% fetal calf serum was applied onto canine tissue. The resultants were left to stand in a moist chamber at room temperature for 1 hour. Next, a solution prepared with the blocking solution to a 10 μ g/ml anti-CAPRIN-1 monoclonal antibody (monoclonal antibody #8) having the heavy chain variable region of SEQ ID NO: 55 and the light chain variable region of SEQ ID NO: 56, which reacts with the cancer cell surfaces prepared in Example 3, was applied onto each slide glass and then left to stand within a moist chamber at 4°C overnight. After 3 instances of 10 minutes of washing with PBS-T, a MOM biotin-labeled anti-IgG antibody (VECTASTAIN) diluted 250-fold with the blocking solution was applied onto each glass slide and then left to stand within a moist chamber at room temperature for 1 hour. After 3 instances of 10 minutes of washing with PBS-T, an avidin-biotin ABC reagent (VECTASTAIN) was applied and then left to stand within a moist chamber at room temperature for 5 minutes. After 3 instances of 10 minutes of washing with PBS-T, a DAB staining solution (DAB 10 mg + 30% H_2O_2 10 μ l/0.05 M Tris-HCl (pH 7.6) 50 ml) was applied and then the glass slides were left to stand within a moist chamber at room temperature for 30 minutes. Glass slides were rinsed with distilled water and then a hematoxylin reagent (DAKO) was applied. After being left to stand at room temperature for 1 minute, the glass slides were rinsed with distilled water. The glass slides were immersed in 70%, 80%, 90%, 95%, and 100% ethanol solutions in such order for 1 minute each and then left to stand in xylene overnight. The glass slides were removed, coverslipped with Glycergel Mounting Medium (DAKO), and then observed. As a result, CAPRIN-1 expression was observed to a slight degree within cells in all salivary gland, kidney, colon, and stomach tissues, but CAPRIN-1 expression was never observed on cell surfaces. Also, absolutely no CAPRIN-1 expression was observed in tissues from other organs.

(5)-2 CAPRIN-1 expression in canine breast cancer tissue

[0084] With the use of 108 frozen canine breast cancer tissue specimens from dogs diagnosed by pathological diag-

nosis as having malignant breast cancer, frozen section slides were prepared by a method similar to the above and immunohistochemical staining was performed using the monoclonal antibody #8 prepared in Example 3. As a result, CAPRIN-1 expression was confirmed in 100 out of the 108 specimens (92.5%). CAPRIN-1 was particularly strongly expressed on the surfaces of highly atypical cancer cells.

(5)-3 CAPRIN-1 expression in human breast cancer tissue

[0085] Immunohistochemical staining was performed using 188 breast cancer tissue specimens of a paraffin-embedded human breast cancer tissue array (BIOMAX). After 3 hours of treatment at 60°C, the human breast cancer tissue array was immersed into a staining bottle filled with xylene and then xylene replacement every 5 minutes was performed 3 instances. Next, a similar procedure was performed using ethanol and PBS-T instead of xylene. The human breast cancer tissue array was immersed into a staining bottle filled with 10 mM citrate buffer (pH6.0) containing 0.05% Tween20, treated for 5 minutes at 125°C, and then left to stand at room temperature for 40 minutes or more. Excess water around each specimen was removed from the array using Kimwipes, each section was encircled using DAKOPEN (DAKO), and then an appropriate amount of Peroxidase Block (DAKO) was added dropwise onto the array. The array was left to stand at room temperature for 5 minutes and then immersed into a staining bottle filled with PBS-T. PBS-T replacement every 5 minutes was performed 3 instances. As a blocking solution, a PBS-T solution containing 10% FBS was applied onto the array and then the array was left to stand within a moist chamber at room temperature for 1 hour. Next, the monoclonal antibody #8 prepared in Example 3 adjusted to 10 µg/ml using a PBS-T solution containing 5% FBS was applied and then the array was left to stand overnight within a moist chamber at 4°C. After 3 instances of 10 minutes of washing with PBS-T, an appropriate amount of Peroxidase Labeled Polymer Conjugated (DAKO) was added dropwise onto the array, and then the array was left to stand at room temperature for 30 minutes within a moist chamber. After 3 instances of 10 minutes of washing with PBS-T, a DAB staining solution (DAKO) was applied onto the array and then the array was left to stand at room temperature for 10 minutes. The DAB staining solution was discarded from the array and then 10 minutes of washing was performed with PBS-T for 3 instances. The array was rinsed with distilled water and then immersed in 70%, 80%, 90%, 95%, and 100% ethanol solutions in order for 1 minute each and then left to stand in xylene overnight. The array was removed, coverslipped with Glycergel Mounting Medium (DAKO), and then observed. As a result, strong CAPRIN-1 expression was observed for 138 (73%) out of the total 188 breast cancer tissue specimens. (5)-4 CAPRIN-1 expression in human malignant brain tumor

[0086] With the use of 247 malignant brain tumor tissue specimens of paraffin-embedded human malignant brain tumor tissue arrays (BIOMAX), immunohistochemical staining was performed by a method similar to that in (5)-3 above using the monoclonal antibody #8 prepared in Example 3. As a result, strong CAPRIN-1 expression was observed in 227 (92%) out of the total 247 malignant brain tumor tissue specimens.

(5)-5 CAPRIN-1 expression in human breast cancer metastatic lymph node

[0087] With the use of 150 tissue specimens of human breast cancer metastatic lymph nodes of paraffin-embedded human breast cancer metastatic lymph node tissue arrays (BIOMAX), immunohistochemical staining was performed by a method similar to that in (5)-3 above using the monoclonal antibody #8 prepared in Example 3. As a result, strong CAPRIN-1 expression was observed in 136 (90%) out of the total 150 tissue specimens of human breast cancer metastatic lymph nodes. Specifically, it was revealed that CAPRIN-1 is also strongly expressed in a cancer tissue that has metastasized from breast cancer.

Example 2: Preparation of new canine and human cancer antigen proteins

(1) Preparation of recombinant protein

[0088] A recombinant protein was prepared by the following method based on the gene of SEQ ID NO: 5 obtained in Example 1. PCR was performed by preparing a reaction solution adjusted to a total amount of 50 µl through addition of each reagent and an included buffer (1 µl of a vector prepared from the phagemid solution obtained in Example 1 and then subjected to sequence analysis, 2 types of primer (0.4 µM each; according to SEQ ID NOS: 37 and 38) containing *Nde* I and *Kpn* I restriction enzyme cleavage sequences, 0.2 mM dNTP, 1.25 U PrimeSTAR HS polymerase (Takara-baio Co., Ltd.)) and then by reacting the solution through repeating 30 times a cycle of 98°C/10 seconds and 68°C/1.5 minutes using a Thermal Cycler (BIO RAD). The above 2 types of primer were used to amplify the region encoding the full-length amino acid sequence of SEQ ID NO: 6 (P47). After PCR, the thus amplified DNA was subjected to 1% agarose gel electrophoresis and then a DNA fragment of about 1.4 kbp was purified from the gel using a QIAquick Gel Extraction Kit (QIAGEN).

[0089] The purified DNA fragment was ligated to a pCR-Blunt cloning vector (Invitrogen Corporation). The vector was

transformed into *Escherichia coli* and then the plasmid was collected. It was confirmed based on the sequence that the amplified gene fragment matched the target sequence. The plasmid that matched the sequence of interest was treated with *Nde* I and *Kpn* I restriction enzymes and then the resultant was purified using a QIAquick Gel Extraction Kit. Then the gene sequence of interest was inserted into a pET30b expression vector (Novagen) for *Escherichia coli* treated with *Nde* I and *Kpn* I restriction enzymes. A His tag-fused recombinant protein can be produced using the vector. The plasmid was transformed into *Escherichia coli* BL21 (DE3) for expression and then expression induction was performed using 1 mM IPTG, so that the target protein was expressed within *Escherichia coli*.

[0090] Also, the recombinant protein of a canine homologous gene was prepared by the following method based on the gene of SEQ ID NO: 7. PCR was performed by preparing a reaction solution adjusted to a total amount of 50 μ l through addition of each reagent and an included buffer (1 μ l of cDNA from among cDNAs of various tissues and/or cells constructed in Example 1, for which the expression could be confirmed by an RT-PCR method, 2 types of primer (0.4 μ M each; according to SEQ ID NOS: 39 and 40) containing *Nde* I and *Kpn* I restriction enzyme cleavage sequences, 0.2 mM dNTP, 1.25 U PrimeSTAR HS polymerase (Takara-baio Co., Ltd.)) and then by reacting the solution through repeating 30 times a cycle of 98°C/10 seconds and 68°C/2.5 minutes using a Thermal Cycler (BIO RAD). The above 2 types of primer were used to amplify the region encoding the full-length amino acid sequence of SEQ ID NO: 8. After PCR, the thus amplified DNA was fractionated with 1% agarose gel electrophoresis and then a DNA fragment of about 2.2 kbp was purified using a QIAquick Gel Extraction Kit (QIAGEN).

[0091] The purified DNA fragment was ligated to pCR-Blunt cloning vector (Invitrogen Corporation). The vector was transformed into *Escherichia coli*, and then the plasmid was collected. It was then confirmed based on the sequence that the amplified gene fragment matched the sequence of interest. The plasmid that matched the sequence of interest was treated with *Nde* I and *Kpn* I restriction enzymes and then the resultant was purified using a QIAquick Gel Extraction Kit. Then the gene sequence of interest was inserted into a pET30b expression vector (Novagen) for *Escherichia coli* treated with *Nde* I and *Kpn* I restriction enzymes. A His tag-fused recombinant protein can be produced using the vector. The plasmid was transformed into *Escherichia coli* BL21 (DE3) for expression and then expression induction was performed using 1 mM IPTG, so that the protein of interest was expressed within *Escherichia coli*.

[0092] Also, the recombinant protein of a human homologous gene was prepared by the following method based on the gene of SEQ ID NO: 1. PCR was performed by preparing a reaction solution adjusted to a total amount of 50 μ l through addition of each reagent and an included buffer (cDNA (1 μ l) from among cDNAs of various tissues and/or cells constructed in Example 1, for which the expression could be confirmed by an RT-PCR method, 2 types of primer (0.4 μ M each; according to SEQ ID NOS: 41 and 42) containing *Sac* I and *Xho* I restriction enzyme cleavage sequences, 0.2 mM dNTP, 1.25 U PrimeSTAR HS polymerase (Takara-baio Co., Ltd.)) and then by reacting the solution through repeating 30 times a cycle of 98°C/10 seconds and 68°C/2.5 minutes using a Thermal Cycler (BIO RAD). The above 2 types of primer were used to amplify the region encoding the full-length amino acid sequence of SEQ ID NO: 2. After PCR, the thus amplified DNA was subjected to 1% agarose gel electrophoresis and then a DNA fragment of about 2.1 kbp was purified using a QIAquick Gel Extraction Kit (QIAGEN).

[0093] The purified DNA fragment was ligated to a cloning vector pCR-Blunt (Invitrogen Corporation). The vector was transformed into *Escherichia coli*, and then the plasmid was collected. It was then confirmed based on the sequence that the amplified gene fragment matched the sequence of interest. The plasmid that matched the sequence of interest was treated with *Sac* I and *Xho* I restriction enzymes and then the resultant was purified using a QIAquick Gel Extraction Kit. Then the gene sequence of interest was inserted into a pET30a expression vector (Novagen) for *Escherichia coli* treated with *Sac* I and *Xho* I restriction enzymes. A His tag-fused recombinant protein can be produced using the vector. The plasmid was transformed into *Escherichia coli* BL21 (DE3) for expression and then expression induction was performed using 1 mM IPTG, so that the protein of interest was expressed within *Escherichia coli*.

(2) Purification of recombinant protein

[0094] The above-obtained recombinant *Escherichia coli* expressing SEQ ID NO: 1, 5, or 7 was cultured at 37°C in LB medium containing 30 μ g/ml kanamycin until the absorbance at 600 nm reached around 0.7. Then isopropyl- β -D-1-thiogalactopyranoside was added to a final concentration of 1 mM, followed by 4 hours of culture at 37°C. Subsequently, cells were collected by 10 minutes of centrifugation at 4800 rpm. The cell pellet was suspended in phosphate buffered saline and then centrifuged at 4800 rpm for 10 minutes for washing cells.

[0095] The cells were suspended in phosphate buffered saline and then subjected to ultrasonication on ice. The thus ultrasonicated *Escherichia coli* solution was centrifuged at 6000 rpm for 20 minutes. The thus obtained supernatant was used as a soluble fraction and the thus obtained precipitate was used as an insoluble fraction.

[0096] The soluble fraction was added to a nickel chelate column (carrier: Chelating Sepharose (TradeMark) Fast Flow (GE Healthcare), column capacity: 5 mL, 50 mM hydrochloric acid buffer (pH 8.0) as equilibrated buffer)) prepared according to a conventional method. The unbound fraction was washed with 50 mM hydrochloric acid buffer (pH 8.0) in an amount 10 times the capacity of the column and 20 mM phosphate buffer (pH 8.0) containing 20 mM imidazole.

Immediately after washing, 6 beds were eluted with 20 mM phosphate buffer (pH8.0) containing 100 mM imidazole. After the elution of the protein of interest had been confirmed by Coomassie staining, an elution fraction of 20 mM phosphate buffer (pH8.0) containing 100 mM imidazole was added to a strong anion exchange column (carrier: Q Sepharose (TradeMark) Fast Flow (GE Healthcare), column capacity: 5 mL, and 20 mM phosphate buffer (pH8.0) as equilibrated buffer). The unbound fraction was washed with 20 mM phosphate buffer (pH7.0) in an amount 10 times the column capacity and 20 mM phosphate buffer (pH7.0) containing 200 mM sodium chloride. Immediately after washing, 5 beds were eluted using 20 mM phosphate buffer (pH7.0) containing 400 mM sodium chloride. Thus, purified fractions of proteins each having the amino acid sequence shown in SEQ ID NO: 2, 6, or 8 were obtained. These purified fractions were then used as materials for an administration test. Fig. 2 shows the result of the protein of SEQ ID NO: 2 fractionated by electrophoresis and detected by Coomassie staining.

[0097] 200 μ l of each purified preparation obtained by the above method was dispensed into 1 ml of reaction buffer (20 mM Tris-HCl, 50 mM NaCl, 2 mM CaCl_2 pH7.4) and then 2 μ l of enterokinase (Novagen) was added. The preparation was left to stand at room temperature overnight for reaction, His tag was cleaved, and then purification was performed according to included protocols using an Enterokinase Cleavage Capture Kit (Novagen). Next, 1.2 ml of each purified preparation obtained by the above method was substituted with physiological phosphate buffer (Nissui Pharmaceutical Co., Ltd.) using ultrafiltration NANOSEP 10K OMEGA (PALL). Sterilized filtration was performed using 0.22 μ m HT Tuffryn Acrodisc (PALL) and then the resultants were used for the following experiments.

Example 3: Preparation of antibody against CAPRIN-1

(1) Preparation of polyclonal antibody against CAPRIN-1-derived peptide

[0098] To obtain an antibody binding to CAPRIN-1, CAPRIN-1-derived peptide (Arg-Asn-Leu-Glu-Lys-Lys-Lys-Gly-Lys-Leu-Asp-Asp-Tyr-Gln (SEQ ID NO: 43)) was synthesized. One milligram of the peptide as an antigen was mixed with an incomplete Freund's adjuvant (IFA) solution in an amount equivalent to the peptide. The mixture was subcutaneously administered to a rabbit 4 times every 2 weeks. Subsequently, blood was collected, so that an antiserum containing a polyclonal antibody was obtained. Furthermore, the antiserum was purified using a protein G carrier (GE Healthcare Bio-Sciences) and then a polyclonal antibody against the CAPRIN-1-derived peptide was obtained. Next, the reactivity of the obtained polyclonal antibody to the breast cancer cell surface was examined. Specifically, 10^6 cells of the MDA-MB-231 V human breast cancer cell line were subjected to centrifugation in a 1.5 ml microcentrifugal tube. A PBS solution supplemented with 0.1% fetal calf serum (FBS) containing the polyclonal antibody was added to the tube. The solution was left to stand on ice for 1 hour. After washing with PBS, an FITC-labeled goat anti-mouse IgG antibody (Invitrogen Corporation) diluted 500-fold with PBS containing 0.1% FBS was added to the solution, and then the solution was left to stand on ice for 1 hour. After washing with PBS, fluorescence intensity was measured using a FACS Calibur (Becton, Dickinson and Company). Meanwhile, a procedure similar to the above was performed so that a control was prepared by adding PBS containing 0.1% FBS instead of the polyclonal antibody. As a result, it was revealed that fluorescence intensity was found to be stronger in cells treated with the polyclonal antibody than that in control cells. Therefore, it was demonstrated that the obtained polyclonal antibody binds to the breast cancer cell surface.

(2) Preparation of monoclonal antibody against CAPRIN-1 protein

[0099] The antigen protein (human CAPRIN-1) (100 μ g) shown in SEQ ID NO: 2 prepared in Example 2 was mixed with a MPL+TDM adjuvant (Sigma) in an amount equivalent to that of the antigen protein. The mixture was used as an antigen solution per mouse. The antigen solution was administered intraperitoneally to a 6-week-old Balb/c mouse (Japan SLC Inc.) and then further administered 3 instances every week. Spleen was removed on day 3 after the final immunization and then ground in between two sterilized glass slides. The resultant was washed with PBS (-) (Nissui) and then centrifuged at 1500 rpm for 10 minutes, so that a procedure to remove supernatants was repeated 3 instances. Thus, spleen cells were obtained. The thus obtained spleen cells were mixed with mouse myeloma cells SP2/0 (purchased from ATCC) at a ratio of 10 : 1. The PEG solution prepared by mixing 200 μ l of RPMI1640 medium containing 10% FBS heated at 37°C and 800 μ l of PEG1500 (Boehringer) was added to the cells. The solution was left to stand for 5 minutes for cell fusion. Centrifugation was performed at 1700 rpm for 5 minutes to remove supernatants. Cells were suspended in 150 ml of RPMI1640 medium (HAT selective medium) containing 15% FBS, to which 2% equivalent of HAT solution (Gibco) had been added and then seeded onto fifteen 96-well plates (Nunc) at 100 μ l per well. Cells were cultured for 7 days under conditions of 37°C and 5% CO_2 , so that hybridomas resulting from fusion of spleen cells to myeloma cells were obtained.

[0100] Hybridomas were selected using as an index the binding affinity of the antibody produced by the thus prepared hybridomas for the CAPRIN-1 protein. The CAPRIN-1 protein solution (1 μ g/ml) prepared in Example 2 was added at 100 μ l per well of 96-well plates and then left to stand at 4°C for 18 hours. Each well was washed 3 instances with PBS-

T, and then 0.5% Bovine Serum Albumin (BSA) solution (Sigma) was added at 400 μ l per well, and then the plates were left to stand at room temperature for 3 hours. The solution was removed and then each well was washed 3 instances with 400 μ l of PBS-T. Each culture supernatant of the hybridomas obtained above was added at 100 μ l per well and then left to stand at room temperature for 2 hours. Each well was washed 3 instances with PBS-T, an HRP-labeled anti-mouse IgG (H+L) antibody (Invitrogen Corporation) diluted 5000-fold with PBS was added at 100 μ l per well and then left to stand at room temperature for 1 hour. Each well was washed 3 instances with PBS-T. A TMB substrate solution (Thermo) was added at 100 μ l per well and then left to stand for 15-30 minutes, so that a color reaction was performed. After color development, 1N sulfuric acid was added at 100 μ l per well to stop the reaction. Absorbance was measured at 450 nm and 595 nm using an absorption spectrometer. As a result, a plurality of hybridomas producing antibodies with high absorbances were selected.

[0101] The thus selected hybridomas were added at 0.5 hybridomas per well of 96-well plates and then cultured. After 1 week, hybridomas forming single colonies in wells were observed. Cells in these wells were further cultured. Hybridomas were selected using as an index the binding affinity of the antibody produced by cloned hybridomas for the CAPRIN-1 protein. The CAPRIN-1 protein solution (1 μ g/ml) prepared in Example 2 was added at 100 μ l per well of 96-well plates and then left to stand at 4°C for 18 hours. Each well was washed 3 instances with PBS-T. A 0.5% BSA solution was added at 400 μ l per well, and then left to stand at room temperature for 3 hours. The solution was removed and then each well was washed 3 instances with 400 μ l of PBS-T. Each culture supernatant of the hybridomas obtained above was added at 100 μ l per well and then left to stand at room temperature for 2 hours. Each well was washed 3 instances with PBS-T. An HRP-labeled anti-mouse IgG (H+L) antibody (Invitrogen Corporation) diluted 5000-fold with PBS was added at 100 μ l per well and then left to stand at room temperature for 1 hour. Each well was washed 3 instances with PBS-T, a TMB substrate solution (Thermo) was added at 100 μ l per well and then left to stand for 15-30 minutes, so that a color reaction was performed. After color development, 1N sulfuric acid was added at 100 μ l per well to stop the reaction. Absorbance was measured at 450 nm and 595 nm using an absorption spectrometer. As a result, a plurality of hybridoma cell lines producing monoclonal antibodies exerting reactivity to the CAPRIN-1 protein were obtained. Culture supernatants of hybridomas were purified using a protein G carrier, so that 150 monoclonal antibodies binding to the CAPRIN-1 protein were obtained.

[0102] Next, from among these monoclonal antibodies, monoclonal antibodies exerting reactivity to the surfaces of breast cancer cells expressing CAPRIN-1 were selected. Specifically, 10⁶ cells of the MDA-MB-231 V human breast cancer cell line were subjected to centrifugation with a 1.5 ml microcentrifugal tube. The supernatant (100 μ l) of each hybridoma above was added and then left to stand on ice for 1 hour. After washing with PBS, an FITC-labeled goat anti-mouse IgG antibody (Invitrogen Corporation) diluted 500-fold with PBS containing 0.1% fetal calf serum was added and then left to stand on ice for 1 hour. After washing with PBS, fluorescence intensity was measured using FACS Calibur (Becton, Dickinson and Company). Meanwhile, a procedure similar to the above was performed so that a control supplemented with a medium instead of the antibody was prepared. As a result, 10 monoclonal antibodies (#1-#10) having fluorescence intensity stronger than that of the control; that is, reacting with the surfaces of breast cancer cells were selected. The heavy chain variable regions and the light chain variable regions of these monoclonal antibodies were shown in SEQ ID NOS: 44-60. The above monoclonal antibody #1 comprises the heavy chain variable region of SEQ ID NO: 44 and the light chain variable region of SEQ ID NO: 45, the monoclonal antibody #2 comprises the heavy chain variable region of SEQ ID NO: 44 and the light chain variable region of SEQ ID NO: 46, the monoclonal antibody #3 comprises the heavy chain variable region of SEQ ID NO: 44 and the light chain variable region of SEQ ID NO: 47, the monoclonal antibody #4 comprises the heavy chain variable region of SEQ ID NO: 44 and the light chain variable region of SEQ ID NO: 48, the monoclonal antibody #5 comprises the heavy chain variable region of SEQ ID NO: 49 and the light chain variable region of SEQ ID NO: 50, the monoclonal antibody #6 comprises the heavy chain variable region of SEQ ID NO: 51 and the light chain variable region of SEQ ID NO: 52, the monoclonal antibody #7 comprises the heavy chain variable region of SEQ ID NO: 53 and the light chain variable region of SEQ ID NO: 54, the monoclonal antibody #8 comprises the heavy chain variable region of SEQ ID NO: 55 and the light chain variable region of SEQ ID NO: 56, the monoclonal antibody #9 comprises the heavy chain variable region of SEQ ID NO: 57 and the light chain variable region of SEQ ID NO: 58, and the monoclonal antibody #10 comprises the heavy chain variable region of SEQ ID NO: 59 and the light chain variable region of SEQ ID NO: 60.

(3) Identification of a peptide in CAPRIN-1 protein, to which an antibody against CAPRIN-1 reacting to cancer cell surface binds

[0103] With the use of monoclonal antibodies #1-#10 against CAPRIN-1, reacting with the surfaces of cancer cells obtained above, partial sequences in the CAPRIN-1 protein to be recognized by these monoclonal antibodies were identified.

[0104] First, DTT (Fluka) was added to 100 μ l of a recombinant CAPRIN-1 protein solution adjusted to contain the protein at a concentration of 1 μ g/ μ l with PBS to a final concentration of 10 mM, followed by 5 minutes of reaction at

95°C, so that reduction of disulfide bonds within the CAPRIN-1 protein was performed. Next, iodoacetamide (Wako Pure Chemical Industries, Ltd.) with a final concentration of 20 mM was added and then an alkylation reaction was performed for thiol groups at 37°C for 30 minutes under shading conditions. Fifty microgram each of monoclonal antibodies #1-#10 against CAPRIN-1 was added to 40 µg of the thus obtained reduced-alkylated CAPRIN-1 protein. The volume of the mixture was adjusted to 1 mL of 20 mM phosphate buffer (pH7.0), and then the mixture was left to react overnight at 4°C while stirring and mixing each mixture.

[0105] Next, trypsin (Promega) was added to a final concentration of 0.2 µg. After 1 hour, 2 hours, 4 hours, and then 12 hours of reaction at 37°C, the resultants were mixed with protein A-glass beads (GE) subjected in advance to blocking with PBS containing 1% BSA (Sigma) and washing with PBS in 1 mM calcium carbonate and NP-40 buffer (20 mM phosphate buffer (pH7.4), 5 mM EDTA, 150 mM NaCl, and 1% NP-40), followed by 30 minutes of reaction.

[0106] The reaction solutions were each washed with 25 mM ammonium carbonate buffer (pH8.0) and then antigen-antibody complexes were eluted using 100 µl of 0.1% formic acid. LC-MS analysis was conducted for eluates using Q-TOF Premier (Waters-MicroMass) according to protocols included with the instrument.

[0107] As a result, the polypeptide of SEQ ID NO: 61 was identified as a partial sequence of CAPRIN-1, which was recognized by all of the monoclonal antibodies #1-#10 against CAPRIN-1. Furthermore, the peptide of SEQ ID NO: 62 was identified as a partial sequence in the polypeptide of SEQ ID NO: 61 above, which was recognized by the monoclonal antibodies #1-#4, #5-#7, and #9. It was further revealed that the monoclonal antibodies #1-#4 recognized the peptide of SEQ ID NO: 63 that was a partial sequence peptide thereof.

Example 4: Cancer diagnosis using CAPRIN-1 polypeptide

(1) Canine cancer diagnosis

[0108] Blood was collected from 342 canine patients confirmed to have malignant or benign tumors and 6 healthy dogs, and serum was separated. With the use of the canine CAPRIN-1 polypeptide (SEQ ID NO: 8) and the anti-canine IgG antibody prepared in Example 2, the titer of the serum IgG antibody specifically reacting with the polypeptide was measured by an ELISA method.

[0109] Immobilization of the thus prepared polypeptide was performed by adding a recombinant protein solution diluted to 5 µg/mL with phosphate buffered saline to 96-well immobilizer amino plates (Nunc) at 100 µl/well and then leaving the plates to stand at 4°C overnight. Blocking was performed by adding a 50 mM sodium bicarbonate buffer solution (pH 8.4) (hereinafter, blocking solution) containing 0.5% BSA (bovine serum albumin) (Sigma Aldrich Japan) at 100 µl/well and then shaking the solution at room temperature for 1 hour. Serum diluted 1000-fold with the blocking solution was added at 100 µl/well and then the mixture was shaken at room temperature for 3 hours for reaction. The reaction solutions were washed 3 instances with phosphate buffered saline (hereinafter, PBS-T) containing 0.05% Tween20 (Wako Pure Chemical Industries, Ltd.). An HRP modified canine IgG antibody (Goat anti-Dog IgG-h+I HRP conjugated: BETHYL Laboratories) diluted 3000-fold with the blocking solution was added at 100 µl/well, followed by 1 hour of reaction at room temperature while shaking the solution. After 3 instances of washing with PBS-T, HRP substrate TMB (1-Step Turbo TMB (tetramethylbenzidine), PIERCE) was added at 100 µl/well and then an enzyme-substrate reaction was conducted at room temperature for 30 minutes. Subsequently, a 0.5 M sulfuric acid solution (Sigma Aldrich Japan) was added at 100 µl/well to stop the reaction. Absorbance at 450 nm was measured using a microplate reader. As controls, a specimen in connection with which no recombinant protein prepared had been immobilized and a specimen in connection with which a reaction with the serum of a cancer-bearing dog had not been conducted were similarly subjected to the above treatment and comparison.

[0110] As a result of pathologic diagnosis using excised tumor tissue, definitive diagnosis was made indicating that 215 out of the total 342 specimens used for the cancer diagnosis were malignant.

[0111] Specifically, specimens were diagnosed as having cancer such as malignant melanoma, malignant mixed tumor, hepatocellular carcinoma, basal cell carcinoma, acanthoma-like gingival tumor, tumor of oral cavity, perianal adenocarcinoma, anal sac tumor, anal sac apocrine adenocarcinoma, Sertoli cell carcinoma, cancer of vaginal vestibule, sebaceous adenocarcinoma, sebaceous epithelioma, sebaceous adenoma, sweat gland carcinoma, intranasal adenocarcinoma, nasal adenocarcinoma, thyroid cancer, large-bowel cancer, bronchial adenocarcinoma, adenocarcinoma, ductal carcinoma, breast adenocarcinoma, composite type breast adenocarcinoma, malignant mammary mixed tumor, intraductal papillary adenocarcinoma, fibrosarcoma, hemangiopericytoma, osteosarcoma, chondrosarcoma, soft tissue sarcoma, histiocytic sarcoma, myxosarcoma, undifferentiated sarcoma, lung cancer, mastocytoma, cutaneous leiomyoma, intraperitoneal leiomyoma, leiomyoma, squamous cell carcinoma, chronic lymphocytic leukemia, lymphoma, gastrointestinal lymphoma, digestive lymphoma, small-cell-to-medium-cell lymphoma, adrenomedullary tumor, granulosa cell tumor, and pheochromocytoma.

[0112] The sera from the living bodies of these cancer-bearing dogs were found to have significantly high antibody titers against the recombinant protein as shown in Fig. 3. When the reference value as malignant cancer regarding the

diagnostic method was determined to be twice or more the average value for healthy dogs, it was demonstrated that malignancy could be diagnosed for 108 specimens, which accounted for accounting for 50.2% of all the specimens. The cancer types of these 108 specimens are as follows. Although development of a plurality of types of cancer had indicated for some specimens, the following numerical values are cumulative total values for each cancer type:

6 cases of malignant melanoma, 11 cases of lymphoma, 1 case of suppurative inflammation, 1 case of granulosa cell tumor, 4 cases of hepatocellular carcinoma, 3 cases of malignant testicular tumor, 3 cases of tumor of oral cavity, 7 cases of perianal adenocarcinoma, 12 cases of sarcoma, 35 cases of breast adenocarcinoma, 1 case of lung cancer, 6 cases of ductal carcinoma, 2 cases of sebaceous adenocarcinoma, 5 cases of mastocytoma, 1 case of smooth muscle sarcoma, 3 cases of squamous cell carcinoma, 2 cases of malignant mixed tumor, 1 case of hemangiopericytoma, 1 case of transitional epithelial cancer, 1 case of hemangiopericytoma, 1 case of hemangiopericytoma, and 1 case of sebaceous epithelioma.

[0113] As a result of similar diagnosis using pleural effusions and ascites collected from canine patients with terminal cancer, values similar to the results obtained by the diagnostic method using serum could be detected and cancer diagnosis could be made.

[0114] Also, it was demonstrated that the use of the diagnostic method enables diagnosis of cancer in a location invisible to the naked eye, the extent of cancer, malignancy or postoperative course of cancer, recurrence, metastasis, and the like. Several specific examples of detailed diagnosis shown in Fig. 4 are as described below.

(2)-1 Cancer diagnosis for tumor invisible to the naked eye

[0115] On June 7, 2007, no tumor mass was confirmed for canine patient 1 (flat coated retriever). However, about 20 days later, on June 24, 2007, a peduncular tumor mass with a diameter of 2 mm was found in the gum at the root of the maxillary left cuspid tooth of canine patient 1. On the day when the mass was found, the peduncular portion was ligated and excised. Absorbance at 450 nm was found to be 0.06 before the tumor mass could be visually confirmed, and this figure was almost the same as 0.04, which was determined when the tumor was found. It was also demonstrated by the result that diagnosis of cancer in a location invisible to the naked eye, such as intraperitoneal cancer, is possible with the use of this technique.

[0116] In addition, it can be said that a warning sign of tumor development was successfully detected, since a rise in the aforementioned level could be confirmed before the tumor could be confirmed with the naked eye. Hence, it was confirmed that the technique is also useful for health examinations such as routine health checkups.

(2)-2 Diagnosis of the extent of cancer

[0117] The extent of cancer is determined based on tumor size, tumor depth, how the tumor affects the peripheral tissue, and the presence or the absence of metastasis. It was revealed that a higher value was detected when metastasis had occurred or cancer had progressed.

(2)-3 Diagnosis of cancer malignancy

[0118] Basal cell tumors include malignant basal cell tumors and benign basal cell tumors. In recent year, malignant basal cell tumors have tended to be classified as examples of basal cell carcinoma and benign basal cell tumors tend to be classified as examples of trichoblastoma according to the new WHO.

[0119] Canine patient 2 (Beagle) diagnosed as having basal cell carcinoma (malignant) was subjected to serodiagnosis upon surgery, so that the absorbance at 450 nm was found to be 0.04. Meanwhile, canine patient 3 (mongrel) diagnosed as having trichoblastoma (benign) was subjected to serodiagnosis upon surgery, so that the absorbance at 450 nm was found to be 0, indicating no detection. Therefore, it was demonstrated that different types of basal cell tumor, i.e., malignant basal cell carcinoma and benign trichoblastoma, can be diagnosed, even if they are classified as basal cell tumors.

[0120] Next, examples of mammary gland tumors are as follows. Mammary gland tumors are classified as malignant tumors such as breast adenocarcinoma and malignant mammary mixed tumor and benign mammary gland tumors exhibiting no malignant findings.

[0121] Canine patient 4 (Shetland Sheepdog) underwent extirpative surgery on July 17, 2007, for breast adenocarcinoma. Canine patient 4 had 3 tumors. Pathologic diagnosis using isolated tissue resulted in the same diagnosis. Strongly atypical and invasive mammary gland tissue experienced somewhat widespread papillary-adenoid growth, and vascular invasion was also confirmed for the specimens. Thus, canine patient 4 was diagnosed as having highly malignant breast cancer. As a result of serodiagnosis using blood collected upon surgery, absorbance at 450 nm was found to be 0.41.

[0122] Meanwhile, canine patient 5 (toy poodle) had extirpative surgery on October 9, 2007, for a mammary gland tumor. Pathologic diagnosis using isolated tissues at this time revealed that: whereas tumors were formed in which mammary gland epithelial cells and myoepithelial cells grew, myoepithelial cell components were uniform spindle cells and no malignancy was detected; and the mammary gland epithelial cell component exhibited a slight difference in size and dyskaryosis as observed. Hence, canine patient 5 was diagnosed as having a benign mammary gland tumor for which no malignancy was detected. As a result of blood collection and serodiagnosis upon surgery thereof, absorbance at 450 nm was found to be 0.

[0123] The above results for the 2 specimens revealed that the malignancy of a highly malignant tumor is greater than that of a benign low-malignant tumor.

[0124] Also, distribution of the diagnoses for 54 malignant tumor (breast cancer) specimens, such as breast adenocarcinoma or malignant mammary mixed tumor specimens and 21 benign mammary gland tumor specimens exhibiting no malignancy, were examined. Whereas benign mammary gland tumor specimens showed a distribution similar to that in the case of healthy dogs, breast cancer specimens showed a distribution of high values.

(2)-4 Diagnosis of postoperative course

[0125] Canine patient 6 (mongrel) visited the hospital because of mastocytoma and had extirpative surgery on May 23, 2005. As a result of serodiagnosis made at this time, absorbance at 450 nm was found to be 0.10. Mastocytoma is a tumor that repeatedly undergoes recurrence or metastasis when resected incompletely. Hence, whether or not complete tumor resection can be achieved by surgery is important. At the follow-up on December 19, 2006, absorbance at 450 nm was found to be 0.05, so that a decreased antibody titer was confirmed. At this time, no recurrence was confirmed. Hence, in the case of canine patient 6, it can be said that since the tumor could be completely resected, the serodiagnosis results were lower than those upon surgery.

[0126] Canine patient 7 (Beagle) had extirpative surgery on February 14, 2008, for mastocytoma. As a result of serodiagnosis performed at this time, absorbance at 450 nm was found to be 0.17. As a result of histopathological diagnosis using excised tissues, invasive hyperplasia was observed and Canine patient 7 was diagnosed as having mastocytoma corresponding to the moderately differentiated type (Patnaik II type). Canine patient 7 visited again for follow-up on March 10, 2008 and was subjected to serodiagnosis again. As a result, absorbance at 450 nm was found to be 0.07. At this time, neither metastasis nor recurrence was confirmed. Thus, in the case of canine patient 7, it can be said that the serodiagnosis results were lower than those upon surgery since the tumor could be completely resected.

(2)-5 Recurrence diagnosis

[0127] Canine patient 8 (Husky) had extirpative surgery on May 8, 2007, for breast adenocarcinoma. As a result of serodiagnosis performed at this time, absorbance at 450 nm was found to be 0.05. As a result of pathologic diagnosis using excised tissue, strongly atypical epithelial cells grew mainly forming a tubular structure. Thus, canine patient 8 was diagnosed as having adenocarcinoma from the primary mammary gland. At this time, the presence of many cancer cells in lymph ducts had already been confirmed, indicating a high risk of metastasis to or recurrence at the lymph nodes or distant sites. On June 28, 2007, (about 1 and a half months after surgery), recurrence was confirmed at the same site. The result of serodiagnosis at this time was 0.09, and thus an increased value was confirmed. In the case of canine patient 8, it was revealed that because of incomplete tumor resection or recurrence thereof, the diagnostic results were higher in late June than in early May.

(2)-6 Diagnosis of metastasis

[0128] Canine patient 9 (Scottish terrier) experienced multiple metastases and recurrences, including a mammary gland tumor in February 2003, intraoral malignant melanoma in August 2003, labial malignant melanoma in January 2005, and intraoral melanoma on April 13, 2005. All of these tumors had been resected by surgery. Canine patient 9 revisited the hospital on December 17, 2006, for follow-up after the recurrence of intraoral melanoma on April 2005 and was subjected to serodiagnosis. As a result, absorbance at 450 nm was found to be 0.09. Half a year later, canine patient 9 revisited the hospital on June 20, 2007 because of cervical lymphoid and popliteal lymphoid hyperplasia. In the case of lymphoma, the lymph nodes swell up systemically. Canine patient 9 had swelling lymph nodes at only two sites. Hence, canine patient 9 was clinically diagnosed as likely to have lymphoma due to metastasis. Diagnosis made by this technique also revealed that absorbance at 450 nm was increased to 0.10, indicating that the lymphoma was caused by metastasis from the previous tumor.

[0129] Canine patient 10 (Shiba inu) underwent tumorectomy on March 11, 2006, because of intraoral malignant melanoma of the right lip. Canine patient 10 had a history of treatment with an anticancer agent (cyclophosphamide) from June 10, 2006, to September 26, 2006, and had been under medication with BIREMO S having organic germanium

as a major ingredient since May 23, 2006. Serodiagnosis was made on March 20, 2007, upon the removal of a tumor thought to have resulted from metastasis of the previous tumor, so that the absorbance at 450 nm was found to be almost 0.03, indicating almost no detection. Pathologic diagnosis was made for the tissue excised at this time so that the disease was diagnosed as metastatic malignant melanoma. However, metastasis occurred again on June 27, 2007, 3 months after surgery for metastatic melanoma. A tumor developed at the right portion of the cervix on March 20, 2007, and further tumor development occurred on the side opposite to such portion on June 27, 2007. The tumors formed black masses analogous to those of the previous findings. Tumor size was 3.1 x 3.2 x 0.8 cm, and the tumors were clinically diagnosed as metastatic tumors. As a result of serodiagnosis at this time, absorbance at 450 nm was confirmed to have increased to 0.23, suggesting that the tumors resulted from metastasis of previous tumors.

(2)-7 Cancer diagnosis using human CAPRIN-1-derived polypeptide

[0130] With the use of the polypeptide (SEQ ID NO: 2) of human CAPRIN-1 prepared in Example 2, the titer of canine serum IgG antibody reacting with the polypeptide was measured in a manner similar to that used above. As a result of examination using serum of a healthy dog, almost no absorbance was detected at 450 nm, similarly to the case above.

[0131] Meanwhile, canine patient 11 (Shih tzu) had extirpative surgery for breast adenocarcinoma on June 21, 2007. As a result of pathologic diagnosis using excised tissues, canine patient 11 was diagnosed as having breast adenocarcinoma of moderate malignancy, wherein strongly atypical and invasive mammary gland tissues underwent adenoid-tubular-papillary growth so as to form large and small masses, in addition to the presence of somewhat diffuse hyperplasia of fibrillar connective tissues. The absorbance at 450 nm for canine patient 11 was found to be 0.26.

(3) Feline cancer diagnosis

[0132] Next, cancer-bearing cats and healthy cats were diagnosed. With the use of the polypeptide of canine CAPRIN-1 (used above) and an anti-feline IgG antibody, the titer of feline serum IgG antibody specifically reacting with the polypeptide was measured, in a manner similar to the above. As a secondary antibody, an HRP modified anti-feline IgG antibody (PEROXIDASE-CONJUGATED GOAT IgG FRACTION TO CAT IgG (WHOLE MOLECULE): CAPPEL RESEARCH REAGENTS) was diluted 8000-fold with a blocking solution and then used.

[0133] Feline patient 1 (mongrel) had tumor extirpative surgery for breast adenocarcinoma on May 8, 2007. The absorbance at 450 nm for feline patient 1 was found to be 0.21. Also, in the case of feline patient 2 (Himalayans) that had extirpative surgery for ductal carcinoma on October 17, 2006, the absorbance at 450 nm was found to be 0.18. On the other hand, no absorbance was detected in the case of healthy cats.

[0134] Also, with the use of the polypeptide (SEQ ID NO: 2) of human CAPRIN-1 prepared in Example 2, the titer of feline serum IgG antibody reacting with the polypeptide was measured in a manner similar to the above. As a result, in the case of healthy cats, almost no absorbance was detected at 450 nm when the polypeptide had been immobilized. Meanwhile, feline patient 3 (American Shorthair) had extirpative surgery for breast adenocarcinoma on April 15, 2008. As a result of pathologic diagnosis using excised tissues, feline patient 3 was diagnosed as having highly malignant breast adenocarcinoma associated with large and small dead tissues, wherein strongly atypical and invasive mammary gland tissues underwent sheet-like growth into large and small masses. Also in the case of feline patient 3, the absorbance at 450 nm was found to be 0.12.

[0135] Therefore, it was demonstrated that cancer diagnosis is also possible for cats by this technique, similarly to dogs, since values were detected for specimens from cats with cancer, but none was detected for specimens from healthy cats.

(4) Human cancer diagnosis

[0136] With the use of the polypeptide (SEQ ID NO: 2) of human CAPRIN-1 prepared in Example 2 and an anti-human IgG antibody, the titer of a healthy human serum IgG antibody specifically reacting with the polypeptide was measured. Immobilization of the prepared polypeptide was performed by adding a recombinant protein solution diluted to 100 µg/mL with phosphate buffered saline to 96-well immobilizer amino plates (Nunc) at 100 µL/well and then leaving the plates to stand overnight at 4°C. Blocking was performed as follows. Four gram of Block Ace powder (DS PHARMA BIOMEDICAL Co., Ltd.) was dissolved in 100 ml of purified water and then the solution was diluted 4-fold with purified water. Then the solution (hereinafter, blocking solution) was added at 100 µL/well and then shaken at room temperature for 1 hour. Serum diluted 1000-fold with the blocking solution was added at 100 µL/well and then shaken at room temperature for 3 hours for reaction. After washing 3 instances with phosphate buffered saline (hereinafter, PBS-T) containing 0.05% Tween20 (Wako Pure Chemical Industries, Ltd.), an HRP-modified anti-human IgG antibody (HRP-Goat Anti-Human IgG (H+L) Conjugate: Zymed Laboratories) diluted 10000-fold with the blocking solution was added at 100 µL/well and then shaken at room temperature for 1 hour for reaction. After 3 instances of washing with PBS-T, HRP substrate TMB (1-Step Turbo

TMB (tetramethylbenzidine, PIERCE) was added at 100 μ l/well and then an enzyme-substrate reaction was performed at room temperature for 30 minutes. Subsequently, a 0.5 M sulfuric acid solution (Sigma Aldrich Japan) was added at 100 μ l/well to stop the reaction and then absorbance at 450 nm was measured using a microplate reader. An ovalbumin antigen adjusted to 50 μ g/ml with phosphate buffered saline was immobilized and then used as a positive control. As a result, absorbance at 450 nm was found to be as high as 0.45 on average as the results for 7 healthy subjects in the case of the ovalbumin antigen, but no absorbance (0) was detected in the case of the above polypeptide.

[0137] In a manner similar to the above, 17 serum specimens (purchased from ProMedDx) from patients with malignant breast cancer were further subjected to measurement of the titer of serum IgG antibody specifically reacting with the human-derived cancer antigen protein (the amino acid sequence of SEQ ID NO: 3). As a result, absorbance at 450 nm was found to be as high as 0.48 in the case of the above polypeptide, on average as the results for 17 breast cancer patients.

[0138] Also, with the use of the polypeptide (SEQ ID NO: 8) of canine CAPRIN-1 prepared in Example 2 and an anti-human IgG antibody, the titer of human serum IgG antibody specifically reacting with the polypeptide was measured in a manner similar to that above. As a result, whereas the average of the results for 7 healthy subjects was 0.04, the average of the results for 17 breast cancer patients was as high as 0.55.

[0139] Based on the above results, it was demonstrated that cancer in humans can also be detected by this technique.

Example 5: Cancer diagnosis through measurement of antigen polypeptide

[0140] With the use of the polyclonal antibody against CAPRIN-1-derived peptide (SEQ ID NO: 43) obtained in Example 3 (1) and each monoclonal antibody against the CAPRIN-1 protein obtained in Example 3 (2) in combination, the antigen polypeptide itself contained in specimens (cancer-bearing living organism-derived serum) reacted positive upon cancer diagnosis using the polypeptide of CAPRIN-1 in Example 4 (1)-(3) was detected by Sandwich ELISA. The polyclonal antibody was used as a primary antibody and each monoclonal antibody was used as a secondary antibody. The serum protein level of the protein specifically reacting with each of the above antibodies was measured.

[0141] The primary antibody was immobilized by adding the polyclonal antibody diluted to a concentration of 5 μ g/ml with phosphate buffered saline to 96-well immobilizer amino plates (Nunc) at 100 μ l/well and then shaking the plates at room temperature for 2 hours. Blocking was performed by adding a 50 mM sodium bicarbonate buffer solution (pH 8.4) (hereinafter, blocking solution) containing 0.5% BSA (bovine serum albumin, Sigma Aldrich Japan) at 100 μ l/well and then shaking at room temperature for 1 hour. Subsequently, a cancer-bearing living organism-derived serum diluted using the blocking solution was added at 100 μ l/well and then the resultants were shaken at room temperature for 3 hours for reaction. The dilution rate at this time was adjusted with 10-fold (10-1000-fold) dilution series. After 3 instances of washing with phosphate buffered saline (hereinafter, PBS-T) containing 0.05% Tween20 (Wako Pure Chemical Industries, Ltd.), each monoclonal antibody as a secondary antibody diluted to a concentration of 1 μ g/ml with the blocking solution was added at 100 μ l/well and then the resultants were shaken at room temperature for 1 hour for reaction. After 3 instances of washing with PBS-T, an HRP-labeled anti-mouse IgG (H+L) antibody (Invitrogen Corporation) as a tertiary antibody diluted 5000-fold with the blocking solution was added at 100 μ l per well and then left to stand at room temperature for 1 hour. After 3 instances of washing of wells with PBS-T, a TMB substrate solution (Thermo) was added at 100 μ l per well and then left to stand for 15-30 minutes for color reaction. After color development, 1 N sulfuric acid was added at 100 μ l per well to stop the reaction and then absorbance at 450 nm was measured using an absorption spectrometer.

[0142] As a result, when the #1-#10 monoclonal antibodies reacting with the surfaces of cancer cells were used as secondary antibodies, absorbance values (polypeptide levels) of 0.3 or higher were detected for all specimens from cancer-bearing dogs and cancer-bearing cats with breast cancer, malignant melanoma, and the like, but no absorbance was detected for healthy dogs and healthy cats. On the other hand, when monoclonal antibodies reacting with the CAPRIN-1 protein itself but not reacting with the surfaces of cancer cells were used as secondary antibodies, polypeptide levels were detected for all specimens, but absorbance values were all 0.05 or less, which were lower than the results for combinations of antibodies reacting with the surfaces of cancer cells.

[0143] Therefore, cancer can also be diagnosed by this technique that involves detection of antigen polypeptides using antibodies against CAPRIN-1.

INDUSTRIAL APPLICABILITY

[0144] The present invention is industrially useful for diagnosis or detection of cancer.

[0145] This description includes part or all of the contents as disclosed in the description and/or drawings of Japanese Patent Application No. 2008-202320, which is a priority document of the present application. Also, all publications, patents, and patent applications cited herein are incorporated herein by reference in their entirety.

SEQUENCE LISTING FREE TEXT

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 30 ttatggagtt aacggggagg aagaccctc aggaaaacga aagtaaattg ttaaggetca 5469
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Gly Ala Ala Ala Pro Ala Ser Gln His Pro Ala Thr Gly Thr Gly Ala
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10
Val Gln Thr Glu Ala Met Lys Gln Ile Leu Gly Val Ile Asp Lys Lys
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Leu Arg Asn Leu Glu Lys Lys Lys Gly Lys Leu Asp Asp Tyr Gln Glu
65 70 75 80

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Arg Met Asn Lys Gly Glu Arg Leu Asn Gln Asp Gln Leu Asp Ala Val
85 90 95

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Ser Lys Tyr Gln Glu Val Thr Asn Asn Leu Glu Phe Ala Lys Glu Leu
100 105 110

30
Gln Arg Ser Phe Met Ala Leu Ser Gln Asp Ile Gln Lys Thr Ile Lys
115 120 125

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Lys Thr Ala Arg Arg Glu Gln Leu Met Arg Glu Glu Ala Glu Gln Lys
130 135 140

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Arg Leu Lys Thr Val Leu Glu Leu Gln Tyr Val Leu Asp Lys Leu Gly
145 150 155 160

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Asp Asp Glu Val Arg Thr Asp Leu Lys Gln Gly Leu Asn Gly Val Pro
165 170 175

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Ile Leu Ser Glu Glu Glu Leu Ser Leu Leu Asp Glu Phe Tyr Lys Leu
180 185 190

55

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10	His Ala Ser Ile His Leu Trp Asp Leu Leu Glu Gly Lys Glu Lys Pro	210	215	220
15	Val Cys Gly Thr Thr Tyr Lys Val Leu Lys Glu Ile Val Glu Arg Val	225	230	235
20	Phe Gln Ser Asn Tyr Phe Asp Ser Thr His Asn His Gln Asn Gly Leu	245	250	255
25	Cys Glu Glu Glu Glu Ala Ala Ser Ala Pro Ala Val Glu Asp Gln Val	260	265	270
30	Pro Glu Ala Glu Pro Glu Pro Ala Glu Glu Tyr Thr Glu Gln Ser Glu	275	280	285
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40	Phe Thr Ser Gly Glu Lys Glu Gln Val Asp Glu Trp Thr Val Glu Thr	305	310	315
45	Val Glu Val Val Asn Ser Leu Gln Gln Gln Pro Gln Ala Ala Ser Pro	325	330	335
50	Ser Val Pro Glu Pro His Ser Leu Thr Pro Val Ala Gln Ala Asp Pro	340	345	350
55	Leu Val Arg Arg Gln Arg Val Gln Asp Leu Met Ala Gln Met Gln Gly			

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355

360

365

5

Pro Tyr Asn Phe Ile Gln Asp Ser Met Leu Asp Phe Glu Asn Gln Thr
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Leu Asp Pro Ala Ile Val Ser Ala Gln Pro Met Asn Pro Thr Gln Asn
385 390 395 400

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Met Asp Met Pro Gln Leu Val Cys Pro Pro Val His Ser Glu Ser Arg
405 410 415

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Leu Ala Gln Pro Asn Gln Val Pro Val Gln Pro Glu Ala Thr Gln Val
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Pro Leu Val Ser Ser Thr Ser Glu Gly Tyr Thr Ala Ser Gln Pro Leu
435 440 445

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Tyr Gln Pro Ser His Ala Thr Glu Gln Arg Pro Gln Lys Glu Pro Ile
450 455 460

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Asp Gln Ile Gln Ala Thr Ile Ser Leu Asn Thr Asp Gln Thr Thr Ala
465 470 475 480

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Ser Ser Ser Leu Pro Ala Ala Ser Gln Pro Gln Val Phe Gln Ala Gly
485 490 495

45

Thr Ser Lys Pro Leu His Ser Ser Gly Ile Asn Val Asn Ala Ala Pro
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Phe Gln Ser Met Gln Thr Val Phe Asn Met Asn Ala Pro Val Pro Pro
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55

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20	Pro Asp Gln Ser His Gln Val Thr Gly Asn His Gln Gln Pro Pro Gln	580	585	590
25	Gln Asn Thr Gly Phe Pro Arg Ser Asn Gln Pro Tyr Tyr Asn Ser Arg	595	600	605
30	Gly Val Ser Arg Gly Gly Ser Arg Gly Ala Arg Gly Leu Met Asn Gly	610	615	620
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40	Pro Ser Phe Ser Asn Thr Pro Asn Ser Gly Tyr Thr Gln Ser Gln Phe	645	650	655
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	gcgcgcacg atg ccc tcg gcc acc agc cac agc ggg agc ggc agc aag tcg			231
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	gga gcc ggg gcc gcc gcg ccg gct tct cag cac ccc gca acc ggc acc			327
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	ggc gct gtc cag acc gag gcc atg aag cag att ctc ggg gtg atc gac			375
50	Gly Ala Val Gln Thr Glu Ala Met Lys Gln Ile Leu Gly Val Ile Asp			
	50	55	60	
	aag aaa ctt cgg aac ctg gag aag aaa aag ggt aag ctt gat gat tac			423
55	Lys Lys Leu Arg Asn Leu Glu Lys Lys Lys Gly Lys Leu Asp Asp Tyr			

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5	cag gaa cga atg aac aaa ggg gaa agg ctt aat caa gat cag ctg gat Gln Glu Arg Met Asn Lys Gly Glu Arg Leu Asn Gln Asp Gln Leu Asp 80 85 90			471
10	gcc gtt tct aag tac cag gaa gtc aca aat aat ttg gag ttt gca aaa Ala Val Ser Lys Tyr Gln Glu Val Thr Asn Asn Leu Glu Phe Ala Lys 95 100 105 110			519
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20	ata aag aag aca gca cgt cgg gag cag ctt atg aga gaa gaa gct gaa Ile Lys Lys Thr Ala Arg Arg Glu Gln Leu Met Arg Glu Glu Ala Glu 130 135 140			615
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30	gaa acg gtt gag gtg gta aat tca ctc cag cag caa cct cag gct gca Glu Thr Val Glu Val Val Asn Ser Leu Gln Gln Gln Pro Gln Ala Ala 320 325 330	1191
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50	cag aca ctt gat cct gcc att gta tct gca cag cct atg aat cca aca Gln Thr Leu Asp Pro Ala Ile Val Ser Ala Gln Pro Met Asn Pro Thr 385 390 395	1383
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20	cca att gat cag att cag gca aca atc tct tta aat aca gac cag act Pro Ile Asp Gln Ile Gln Ala Thr Ile Ser Leu Asn Thr Asp Gln Thr 465 470 475	1623		
25	aca gca tca tca tcc ctt cct gct gcg tct cag cct caa gta ttt cag Thr Ala Ser Ser Ser Leu Pro Ala Ala Ser Gln Pro Gln Val Phe Gln 480 485 490	1671		
30	gct ggg aca agc aaa cct tta cat agc agt gga atc aat gta aat gca Ala Gly Thr Ser Lys Pro Leu His Ser Ser Gly Ile Asn Val Asn Ala 495 500 505 510	1719		
35	gct cca ttc caa tcc atg caa acg gtg ttc aat atg aat gcc cca gtt Ala Pro Phe Gln Ser Met Gln Thr Val Phe Asn Met Asn Ala Pro Val 515 520 525	1767		
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35	cag cag aat ttc aag cga ggc tct ggg cag agt gga cca cgg gga gcc Gln Gln Asn Phe Lys Arg Gly Ser Gly Gln Ser Gly Pro Arg Gly Ala 675 680 685	2247
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 20 Leu Arg Asn Leu Glu Lys Lys Lys Gly Lys Leu Asp Asp Tyr Gln Glu
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 25 Arg Met Asn Lys Gly Glu Arg Leu Asn Gln Asp Gln Leu Asp Ala Val
 85 90 95
 30 Ser Lys Tyr Gln Glu Val Thr Asn Asn Leu Glu Phe Ala Lys Glu Leu
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 35 Gln Arg Ser Phe Met Ala Leu Ser Gln Asp Ile Gln Lys Thr Ile Lys
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 40 Lys Thr Ala Arg Arg Glu Gln Leu Met Arg Glu Glu Ala Glu Gln Lys
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 45 Arg Leu Lys Thr Val Leu Glu Leu Gln Tyr Val Leu Asp Lys Leu Gly
 145 150 155 160
 50 Asp Asp Glu Val Arg Thr Asp Leu Lys Gln Gly Leu Asn Gly Val Pro
 165 170 175
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Ile Leu Ser Glu Glu Glu Leu Ser Leu Leu Asp Glu Phe Tyr Lys Leu
180 185 190

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Val Asp Pro Glu Arg Asp Met Ser Leu Arg Leu Asn Glu Gln Tyr Glu
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His Ala Ser Ile His Leu Trp Asp Leu Leu Glu Gly Lys Glu Lys Pro
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Val Cys Gly Thr Thr Tyr Lys Val Leu Lys Glu Ile Val Glu Arg Val
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Phe Gln Ser Asn Tyr Phe Asp Ser Thr His Asn His Gln Asn Gly Leu
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Cys Glu Glu Glu Glu Ala Ala Ser Ala Pro Ala Val Glu Asp Gln Val
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Phe Thr Ser Gly Glu Lys Glu Gln Val Asp Glu Trp Thr Val Glu Thr
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Pro	Asp	Gln	Ser	His	Gln	Val	Thr	Gly	Asn	His	Gln	Gln	Pro	Pro	Gln	
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Gln Asp Ile Gln Lys Thr Ile Lys Lys Thr Ala Arg Arg Glu Gln Leu
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Met Arg Glu Glu Ala Glu Gln Lys Arg Leu Lys Thr Val Leu Glu Leu
25 30 35

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Gln Tyr Val Leu Asp Lys Leu Gly Asp Asp Glu Val Arg Thr Asp Leu
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Lys Gln Gly Leu Asn Gly Val Pro Ile Leu Ser Glu Glu Glu Leu Ser
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Leu Leu Asp Glu Phe Tyr Lys Leu Ala Asp Pro Glu Arg Asp Met Ser
70 75 80

55 ttg agg ttg aat gag cag tat gaa cat gct tcc att cac ctg tgg gac 345

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	Asp Leu Met Ala Gln Met Gln Gly Pro Tyr Asn Phe Ile Gln Asp Ser	
	245 250 255 260	
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15	cct cca gtt cat tct gaa tct aga ctt gct caa cct aat caa gtt cct Pro Pro Val His Ser Glu Ser Arg Leu Ala Gln Pro Asn Gln Val Pro 295 300 305	969
20	gta caa cca gaa gct aca cag gtt cct ttg gtt tca tcc aca agt gag Val Gln Pro Glu Ala Thr Gln Val Pro Leu Val Ser Ser Thr Ser Glu 310 315 320	1017
25	ggg tat aca gca tct caa ccc ttg tac cag cct tct cat gct aca gag Gly Tyr Thr Ala Ser Gln Pro Leu Tyr Gln Pro Ser His Ala Thr Glu 325 330 335 340	1065
30	caa cga cca caa aag gaa cca att gac cag att cag gca aca atc tct Gln Arg Pro Gln Lys Glu Pro Ile Asp Gln Ile Gln Ala Thr Ile Ser 345 350 355	1113
35	tta aat aca gac cag act aca gcg tca tca tcc ctt ccg gct gct tct Leu Asn Thr Asp Gln Thr Thr Ala Ser Ser Ser Leu Pro Ala Ala Ser 360 365 370	1161
40	cag cct cag gta ttc cag gct ggg aca agc aaa cca tta cat agc agt Gln Pro Gln Val Phe Gln Ala Gly Thr Ser Lys Pro Leu His Ser Ser 375 380 385	1209
45	gga atc aat gta aat gca gct cca ttc caa tcc atg caa acg gtg ttc Gly Ile Asn Val Asn Ala Ala Pro Phe Gln Ser Met Gln Thr Val Phe 390 395 400	1257
50	aat atg aat gcc cca gtt cct cct gtt aat gaa cca gaa act ttg aaa Asn Met Asn Ala Pro Val Pro Pro Val Asn Glu Pro Glu Thr Leu Lys 405 410 415 420	1305
55	caa caa aat cag tac cag gcc agt tat aac cag agc ttt tct agt cag	1353

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Gln Gln Asn Gln Tyr Gln Ala Ser Tyr Asn Gln Ser Phe Ser Ser Gln
425 430 435

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cct cac caa gta gaa caa aca gag gga tgc cgc aaa tga acactcagca 1402
Pro His Gln Val Glu Gln Thr Glu Gly Cys Arg Lys
440 445

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agtgaattaa tctgattcac aggattatgt ttaaacgccca aaaacacact ggccagtgtg 1462

ccataatatg ttaccagaag agttattatc tatttgttct cccitttcagg aaacttattg 1522

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Val Leu Glu Leu Gln Tyr Val Leu Asp Lys Leu Gly Asp Asp Glu Val
35 40 45

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Arg Thr Asp Leu Lys Gln Gly Leu Asn Gly Val Pro Ile Leu Ser Glu
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50
Glu Glu Leu Ser Leu Leu Asp Glu Phe Tyr Lys Leu Ala Asp Pro Glu
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55

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5	Arg Asp Met Ser Leu Arg Leu Asn Glu Gln Tyr Glu His Ala Ser Ile	85	90	95
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15	Thr Tyr Lys Ala Leu Lys Glu Ile Val Glu Arg Val Phe Gln Ser Asn	115	120	125
20	Tyr Phe Asp Ser Thr His Asn His Gln Asn Gly Leu Cys Glu Glu Glu	130	135	140
25	Glu Ala Ala Ser Ala Pro Thr Val Glu Asp Gln Val Ala Glu Ala Glu	145	150	155
30	Pro Glu Pro Ala Glu Glu Tyr Thr Glu Gln Ser Glu Val Glu Ser Thr	165	170	175
35	Glu Tyr Val Asn Arg Gln Phe Met Ala Glu Thr Gln Phe Ser Ser Gly	180	185	190
40	Glu Lys Glu Gln Val Asp Glu Trp Thr Val Glu Thr Val Glu Val Val	195	200	205
45	Asn Ser Leu Gln Gln Gln Pro Gln Ala Ala Ser Pro Ser Val Pro Glu	210	215	220
50	Pro His Ser Leu Thr Pro Val Ala Gln Ala Asp Pro Leu Val Arg Arg	225	230	235
55	Gln Arg Val Gln Asp Leu Met Ala Gln Met Gln Gly Pro Tyr Asn Phe			

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	245	250	255
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	260	265	270
10	Ile Val Ser Ala Gln Pro Met Asn Pro Thr Gln Asn Met Asp Met Pro		
	275	280	285
15	Gln Leu Val Cys Pro Pro Val His Ser Glu Ser Arg Leu Ala Gln Pro		
	290	295	300
20	Asn Gln Val Pro Val Gln Pro Glu Ala Thr Gln Val Pro Leu Val Ser		
	305	310	315
25	Ser Thr Ser Glu Gly Tyr Thr Ala Ser Gln Pro Leu Tyr Gln Pro Ser		
	325	330	335
30	His Ala Thr Glu Gln Arg Pro Gln Lys Glu Pro Ile Asp Gln Ile Gln		
	340	345	350
35	Ala Thr Ile Ser Leu Asn Thr Asp Gln Thr Thr Ala Ser Ser Ser Leu		
	355	360	365
40	Pro Ala Ala Ser Gln Pro Gln Val Phe Gln Ala Gly Thr Ser Lys Pro		
	370	375	380
45	Leu His Ser Ser Gly Ile Asn Val Asn Ala Ala Pro Phe Gln Ser Met		
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50	Gln Thr Val Phe Asn Met Asn Ala Pro Val Pro Pro Val Asn Glu Pro		
	405	410	415

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 Pro Pro Pro Pro Ser Gly Ser Ser Gly Ser Glu Ala Ala Ala Ala Ala
 35 20 25 30

 ggg gcg gcg ggg gcg gcg ggg gcc ggg gcg gct gcg ccc gcc tcc cag 144
 Gly Ala Ala Gly Ala Ala Gly Ala Gly Ala Ala Ala Pro Ala Ser Gln
 40 35 40 45

 cac ccc gcg acc ggc acc ggc gct gtc cag acc gag gcc atg aag cag 192
 His Pro Ala Thr Gly Thr Gly Ala Val Gln Thr Glu Ala Met Lys Gln
 45 50 55 60

 atc ctc ggg gtg atc gac aag aaa ctc cgg aac ctg gag aag aaa aag 240
 Ile Leu Gly Val Ile Asp Lys Lys Leu Arg Asn Leu Glu Lys Lys Lys
 50 65 70 75 80

 ggc aag ctt gat gat tac cag gaa cga atg aac aaa ggg gaa agg ctt 288
 Gly Lys Leu Asp Asp Tyr Gln Glu Arg Met Asn Lys Gly Glu Arg Leu
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	85	90	95	
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10	aac ttg gag ttt gca aaa gaa tta cag agg agt ttc atg gca tta agt Asn Leu Glu Phe Ala Lys Glu Leu Gln Arg Ser Phe Met Ala Leu Ser 115 120 125			384
15	caa gat att cag aaa aca ata aag aag act gca cgt cgg gag cag ctt Gln Asp Ile Gln Lys Thr Ile Lys Lys Thr Ala Arg Arg Glu Gln Leu 130 135 140			432
20	atg aga gag gaa gcg gaa caa aaa cgt tta aaa act gta ctt gag ctc Met Arg Glu Glu Ala Glu Gln Lys Arg Leu Lys Thr Val Leu Glu Leu 145 150 155 160			480
25	cag tat gtt ttg gac aaa ttg gga gat gat gaa gtg aga act gac ctg Gln Tyr Val Leu Asp Lys Leu Gly Asp Asp Glu Val Arg Thr Asp Leu 165 170 175			528
30	aag caa ggt ttg aat gga gtg cca ata ttg tct gaa gaa gaa ttg tcg Lys Gln Gly Leu Asn Gly Val Pro Ile Leu Ser Glu Glu Glu Leu Ser 180 185 190			576
35	ttg ttg gat gaa ttc tac aaa tta gca gac cct gaa cgg gac atg agc Leu Leu Asp Glu Phe Tyr Lys Leu Ala Asp Pro Glu Arg Asp Met Ser 195 200 205			624
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45	ttg ctg gaa gga aag gaa aag tct gta tgt gga aca acc tat aaa gca Leu Leu Glu Gly Lys Glu Lys Ser Val Cys Gly Thr Thr Tyr Lys Ala 225 230 235 240			720
50	cta aag gaa att gtt gag cgt gtt ttc cag tca aat tac ttt gac agc Leu Lys Glu Ile Val Glu Arg Val Phe Gln Ser Asn Tyr Phe Asp Ser 245 250 255			768
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	260 265 270	
10	gca cct aca gtt gaa gac cag gta gct gaa gct gag cct gag cca gca Ala Pro Thr Val Glu Asp Gln Val Ala Glu Ala Glu Pro Glu Pro Ala	864
	275 280 285	
15	gaa gaa tac act gaa caa agt gaa gtt gaa tca aca gag tat gta aat Glu Glu Tyr Thr Glu Gln Ser Glu Val Glu Ser Thr Glu Tyr Val Asn	912
	290 295 300	
20	aga caa ttt atg gca gaa aca cag ttc agc agt ggt gaa aag gag cag Arg Gln Phe Met Ala Glu Thr Gln Phe Ser Ser Gly Glu Lys Glu Gln	960
	305 310 315 320	
25	gta gat gag tgg acg gtc gaa aca gtg gag gtg gtg aat tca ctc cag Val Asp Glu Trp Thr Val Glu Thr Val Glu Val Val Asn Ser Leu Gln	1008
	325 330 335	
30	cag caa cct cag gct gcg tct cct tca gta cca gag ccc cac tct ttg Gln Gln Pro Gln Ala Ala Ser Pro Ser Val Pro Glu Pro His Ser Leu	1056
	340 345 350	
35	act ccg gtg gct cag gca gat ccc ctt gtg aga aga cag cga gtc cag Thr Pro Val Ala Gln Ala Asp Pro Leu Val Arg Arg Gln Arg Val Gln	1104
	355 360 365	
40	gac ctt atg gcg cag atg cag ggg ccc tat aat ttc ata cag gat tca Asp Leu Met Ala Gln Met Gln Gly Pro Tyr Asn Phe Ile Gln Asp Ser	1152
	370 375 380	
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	385 390 395 400	
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	405 410 415	
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10	ggg tat aca gca tct caa ccc ttg tac cag cct tct cat gct aca gag Gly Tyr Thr Ala Ser Gln Pro Leu Tyr Gln Pro Ser His Ala Thr Glu 450 455 460	1392		
15	caa cga cca caa aag gaa cca att gac cag att cag gca aca atc tct Gln Arg Pro Gln Lys Glu Pro Ile Asp Gln Ile Gln Ala Thr Ile Ser 465 470 475 480	1440		
20	tta aat aca gac cag act aca gcg tca tca tcc ctt ccg gct gct tct Leu Asn Thr Asp Gln Thr Thr Ala Ser Ser Ser Leu Pro Ala Ala Ser 485 490 495	1488		
25	cag cct cag gta ttc cag gct ggg aca agc aaa cca tta cat agc agt Gln Pro Gln Val Phe Gln Ala Gly Thr Ser Lys Pro Leu His Ser Ser 500 505 510	1536		
30	gga atc aat gta aat gca gct cca ttc caa tcc atg caa acg gtg ttc Gly Ile Asn Val Asn Ala Ala Pro Phe Gln Ser Met Gln Thr Val Phe 515 520 525	1584		
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40	caa caa aat cag tac cag gcc agt tat aac cag agc ttt tct agt cag Gln Gln Asn Gln Tyr Gln Ala Ser Tyr Asn Gln Ser Phe Ser Ser Gln 545 550 555 560	1680		
45	cct cac caa gta gaa caa aca gac ctt cag caa gaa cag ctt caa aca Pro His Gln Val Glu Gln Thr Asp Leu Gln Gln Glu Gln Leu Gln Thr 565 570 575	1728		
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	610 615 620	
15	ggt gct aga ggc tta atg aat gga tac agg ggc cct gcc aat gga ttc Gly Ala Arg Gly Leu Met Asn Gly Tyr Arg Gly Pro Ala Asn Gly Phe	1920
	625 630 635 640	
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	645 650 655	
25	agt ggt tat aca cag tct cag ttc agt gct ccc cgg gac tac tct ggc Ser Gly Tyr Thr Gln Ser Gln Phe Ser Ala Pro Arg Asp Tyr Ser Gly	2016
	660 665 670	
30	tat cag cgg gat gga tat cag cag aat ttc aag cga ggc tct ggg cag Tyr Gln Arg Asp Gly Tyr Gln Gln Asn Phe Lys Arg Gly Ser Gly Gln	2064
	675 680 685	
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	690 695 700	
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35 40 45

45 His Pro Ala Thr Gly Thr Gly Ala Val Gln Thr Glu Ala Met Lys Gln
50 50 55 60

55 Ile Leu Gly Val Ile Asp Lys Lys Leu Arg Asn Leu Glu Lys Lys Lys
65 70 75 80

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5 Gly Lys Leu Asp Asp Tyr Gln Glu Arg Met Asn Lys Gly Glu Arg Leu
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10 Asn Gln Asp Gln Leu Asp Ala Val Ser Lys Tyr Gln Glu Val Thr Asn
100 105 110

15 Asn Leu Glu Phe Ala Lys Glu Leu Gln Arg Ser Phe Met Ala Leu Ser
115 120 125

20 Gln Asp Ile Gln Lys Thr Ile Lys Lys Thr Ala Arg Arg Glu Gln Leu
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25 Met Arg Glu Glu Ala Glu Gln Lys Arg Leu Lys Thr Val Leu Glu Leu
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30 Gln Tyr Val Leu Asp Lys Leu Gly Asp Asp Glu Val Arg Thr Asp Leu
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35 Lys Gln Gly Leu Asn Gly Val Pro Ile Leu Ser Glu Glu Glu Leu Ser
180 185 190

40 Leu Leu Asp Glu Phe Tyr Lys Leu Ala Asp Pro Glu Arg Asp Met Ser
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45 Leu Arg Leu Asn Glu Gln Tyr Glu His Ala Ser Ile His Leu Trp Asp
210 215 220

50 Leu Leu Glu Gly Lys Glu Lys Ser Val Cys Gly Thr Thr Tyr Lys Ala
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 Glu Glu Tyr Thr Glu Gln Ser Glu Val Glu Ser Thr Glu Tyr Val Asn
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 Arg Gln Phe Met Ala Glu Thr Gln Phe Ser Ser Gly Glu Lys Glu Gln
 305 310 315 320
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 325 330 335
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 Gln Gln Pro Gln Ala Ala Ser Pro Ser Val Pro Glu Pro His Ser Leu
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 Thr Pro Val Ala Gln Ala Asp Pro Leu Val Arg Arg Gln Arg Val Gln
 355 360 365
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 Asp Leu Met Ala Gln Met Gln Gly Pro Tyr Asn Phe Ile Gln Asp Ser
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 Gln Pro Met Asn Pro Thr Gln Asn Met Asp Met Pro Gln Leu Val Cys
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10 Val Gln Pro Glu Ala Thr Gln Val Pro Leu Val Ser Ser Thr Ser Glu
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25 Leu Asn Thr Asp Gln Thr Thr Ala Ser Ser Ser Leu Pro Ala Ala Ser
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45 Gln Gln Asn Gln Tyr Gln Ala Ser Tyr Asn Gln Ser Phe Ser Ser Gln
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50 Pro His Gln Val Glu Gln Thr Asp Leu Gln Gln Glu Gln Leu Gln Thr
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Val Val Gly Thr Tyr His Gly Ser Gln Asp Gln Pro His Gln Val Thr
580 585 590

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Gly Asn His Gln Gln Pro Pro Gln Gln Asn Thr Gly Phe Pro Arg Ser
595 600 605

10

Ser Gln Pro Tyr Tyr Asn Ser Arg Gly Val Ser Arg Gly Gly Ser Arg
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Gly Ala Arg Gly Leu Met Asn Gly Tyr Arg Gly Pro Ala Asn Gly Phe
625 630 635 640

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Arg Gly Gly Tyr Asp Gly Tyr Arg Pro Ser Phe Ser Asn Thr Pro Asn
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Tyr Gln Arg Asp Gly Tyr Gln Gln Asn Phe Lys Arg Gly Ser Gly Gln
675 680 685

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EP 2 325 648 A1

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Gly Ala Ala Gly Ala Ala Gly Ala Gly Ala Ala Ala Pro Ala Ser Gln
35 40 45

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His Pro Ala Thr Gly Thr Gly Ala Val Gln Thr Glu Ala Met Lys Gln
50 55 60

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Ile Leu Gly Val Ile Asp Lys Lys Leu Arg Asn Leu Glu Lys Lys Lys
65 70 75 80

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Gly Lys Leu Asp Asp Tyr Gln Glu Arg Met Asn Lys Gly Glu Arg Leu
85 90 95

40

aat caa gat cag ctg gat gcc gta tct aag tac cag gaa gtc aca aat 336
Asn Gln Asp Gln Leu Asp Ala Val Ser Lys Tyr Gln Glu Val Thr Asn
100 105 110

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aac ttg gag ttt gca aaa gaa tta cag agg agt ttc atg gca tta agt 384
Asn Leu Glu Phe Ala Lys Glu Leu Gln Arg Ser Phe Met Ala Leu Ser
115 120 125

50

caa gat att cag aaa aca ata aag aag act gca cgt cgg gag cag ctt 432
Gln Asp Ile Gln Lys Thr Ile Lys Lys Thr Ala Arg Arg Glu Gln Leu
130 135 140

55

atg aga gag gaa gcg gaa caa aaa cgt tta aaa act gta ctt gag ctc 480

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	Gln	Tyr	Val	Leu	Asp	Lys	Leu	Gly	Asp	Asp	Glu	Val	Arg	Thr	Asp	Leu	
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	Lys	Gln	Gly	Leu	Asn	Gly	Val	Pro	Ile	Leu	Ser	Glu	Glu	Glu	Leu	Ser	
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	Leu	Arg	Leu	Asn	Glu	Gln	Tyr	Glu	His	Ala	Ser	Ile	His	Leu	Trp	Asp	
				210				215						220			
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	ttg	ctg	gaa	gga	aag	gaa	aag	tct	gta	tgt	gga	aca	acc	tat	aaa	gca	720
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	Leu	Lys	Glu	Ile	Val	Glu	Arg	Val	Phe	Gln	Ser	Asn	Tyr	Phe	Asp	Ser	
					245					250					255		
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	Thr	His	Asn	His	Gln	Asn	Gly	Leu	Cys	Glu	Glu	Glu	Glu	Ala	Ala	Ser	
				260				265						270			
40																	
	gca	cct	aca	gtt	gaa	gac	cag	gta	gct	gaa	gct	gag	cct	gag	cca	gca	864
	Ala	Pro	Thr	Val	Glu	Asp	Gln	Val	Ala	Glu	Ala	Glu	Pro	Glu	Pro	Ala	
				275				280						285			
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	gaa	gaa	tac	act	gaa	caa	agt	gaa	gtt	gaa	tca	aca	gag	tat	gta	aat	912
	Glu	Glu	Tyr	Thr	Glu	Gln	Ser	Glu	Val	Glu	Ser	Thr	Glu	Tyr	Val	Asn	
				290				295					300				
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	Arg	Gln	Phe	Met	Ala	Glu	Thr	Gln	Phe	Ser	Ser	Gly	Glu	Lys	Glu	Gln	
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55																	

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5	gta gat gag tgg acg gtc gaa aca gtg gag gtg gtg aat tca ctc cag Val Asp Glu Trp Thr Val Glu Thr Val Glu Val Val Asn Ser Leu Gln 325 330 335	1008
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35	cct cca gtt cat tct gaa tct aga ctt gct caa cct aat caa gtt cct Pro Pro Val His Ser Glu Ser Arg Leu Ala Gln Pro Asn Gln Val Pro 420 425 430	1296
40	gta caa cca gaa gct aca cag gtt cct ttg gtt tca tcc aca agt gag Val Gln Pro Glu Ala Thr Gln Val Pro Leu Val Ser Ser Thr Ser Glu 435 440 445	1344
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50	caa cga cca caa aag gaa cca att gac cag att cag gca aca atc tct Gln Arg Pro Gln Lys Glu Pro Ile Asp Gln Ile Gln Ala Thr Ile Ser 465 470 475 480	1440
55	tta aat aca gac cag act aca gcg tca tca tcc ctt ccg gct gct tct	1488

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	Leu Asn Thr Asp Gln Thr Thr Ala Ser Ser Ser Leu Pro Ala Ala Ser	
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	Gln Pro Gln Val Phe Gln Ala Gly Thr Ser Lys Pro Leu His Ser Ser	
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	gga atc aat gta aat gca gct cca ttc caa tcc atg caa acg gtg ttc	1584
	Gly Ile Asn Val Asn Ala Ala Pro Phe Gln Ser Met Gln Thr Val Phe	
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	aat atg aat gcc cca gtt cct cct gtt aat gaa cca gaa act ttg aaa	1632
	Asn Met Asn Ala Pro Val Pro Pro Val Asn Glu Pro Glu Thr Leu Lys	
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	cct cac caa gta gaa caa aca gac ctt cag caa gaa cag ctt caa aca	1728
	Pro His Gln Val Glu Gln Thr Asp Leu Gln Gln Glu Gln Leu Gln Thr	
	565 570 575	
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	gtg gtt ggc act tac cat ggt tcc cag gac cag ccc cac caa gtg act	1776
	Val Val Gly Thr Tyr His Gly Ser Gln Asp Gln Pro His Gln Val Thr	
	580 585 590	
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	ggt aac cat cag cag cct ccc cag cag aac act gga ttt cca cgt agc	1824
	Gly Asn His Gln Gln Pro Pro Gln Gln Asn Thr Gly Phe Pro Arg Ser	
	595 600 605	
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	agt cag ccc tat tac aat agt cgt ggt gtg tct cgt ggt ggt tcc cgt	1872
	Ser Gln Pro Tyr Tyr Asn Ser Arg Gly Val Ser Arg Gly Gly Ser Arg	
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15	agt gga cca cgg gga gcc cca cga ggt aat att ttg tgg tgg tga Ser Gly Pro Arg Gly Ala Pro Arg Gly Asn Ile Leu Trp Trp 690 695 700	2109
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50 Pro Pro Pro Pro Ser Gly Ser Ser Gly Ser Glu Ala Ala Ala Ala Ala
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55 Gly Ala Ala Gly Ala Ala Gly Ala Gly Ala Ala Ala Pro Ala Ser Gln

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35

40

45

5

His Pro Ala Thr Gly Thr Gly Ala Val Gln Thr Glu Ala Met Lys Gln
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10

Ile Leu Gly Val Ile Asp Lys Lys Leu Arg Asn Leu Glu Lys Lys Lys
65 70 75 80

15

Gly Lys Leu Asp Asp Tyr Gln Glu Arg Met Asn Lys Gly Glu Arg Leu
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20

Asn Gln Asp Gln Leu Asp Ala Val Ser Lys Tyr Gln Glu Val Thr Asn
100 105 110

25

Asn Leu Glu Phe Ala Lys Glu Leu Gln Arg Ser Phe Met Ala Leu Ser
115 120 125

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Gln Asp Ile Gln Lys Thr Ile Lys Lys Thr Ala Arg Arg Glu Gln Leu
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Met Arg Glu Glu Ala Glu Gln Lys Arg Leu Lys Thr Val Leu Glu Leu
145 150 155 160

40

Gln Tyr Val Leu Asp Lys Leu Gly Asp Asp Glu Val Arg Thr Asp Leu
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Lys Gln Gly Leu Asn Gly Val Pro Ile Leu Ser Glu Glu Glu Leu Ser
180 185 190

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Leu Leu Asp Glu Phe Tyr Lys Leu Ala Asp Pro Glu Arg Asp Met Ser
195 200 205

55

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15	Leu Lys Glu Ile Val Glu Arg Val Phe Gln Ser Asn Tyr Phe Asp Ser	245	250	255
20	Thr His Asn His Gln Asn Gly Leu Cys Glu Glu Glu Glu Ala Ala Ser	260	265	270
25	Ala Pro Thr Val Glu Asp Gln Val Ala Glu Ala Glu Pro Glu Pro Ala	275	280	285
30	Glu Glu Tyr Thr Glu Gln Ser Glu Val Glu Ser Thr Glu Tyr Val Asn	290	295	300
35	Arg Gln Phe Met Ala Glu Thr Gln Phe Ser Ser Gly Glu Lys Glu Gln	305	310	315
40	Val Asp Glu Trp Thr Val Glu Thr Val Glu Val Val Asn Ser Leu Gln	325	330	335
45	Gln Gln Pro Gln Ala Ala Ser Pro Ser Val Pro Glu Pro His Ser Leu	340	345	350
50	Thr Pro Val Ala Gln Ala Asp Pro Leu Val Arg Arg Gln Arg Val Gln	355	360	365
55	Asp Leu Met Ala Gln Met Gln Gly Pro Tyr Asn Phe Ile Gln Asp Ser			

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370

375

380

5

Met Leu Asp Phe Glu Asn Gln Thr Leu Asp Pro Ala Ile Val Ser Ala
385 390 395 400

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Gln Pro Met Asn Pro Thr Gln Asn Met Asp Met Pro Gln Leu Val Cys
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Pro Pro Val His Ser Glu Ser Arg Leu Ala Gln Pro Asn Gln Val Pro
420 425 430

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Val Gln Pro Glu Ala Thr Gln Val Pro Leu Val Ser Ser Thr Ser Glu
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Gly Tyr Thr Ala Ser Gln Pro Leu Tyr Gln Pro Ser His Ala Thr Glu
450 455 460

30

Gln Arg Pro Gln Lys Glu Pro Ile Asp Gln Ile Gln Ala Thr Ile Ser
465 470 475 480

35

Leu Asn Thr Asp Gln Thr Thr Ala Ser Ser Ser Leu Pro Ala Ala Ser
485 490 495

40

Gln Pro Gln Val Phe Gln Ala Gly Thr Ser Lys Pro Leu His Ser Ser
500 505 510

45

Gly Ile Asn Val Asn Ala Ala Pro Phe Gln Ser Met Gln Thr Val Phe
515 520 525

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Asn Met Asn Ala Pro Val Pro Pro Val Asn Glu Pro Glu Thr Leu Lys
530 535 540

55

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5	Gln Gln Asn Gln Tyr Gln Ala Ser Tyr Asn Gln Ser Phe Ser Ser Gln	545	550	555	560
10	Pro His Gln Val Glu Gln Thr Asp Leu Gln Gln Glu Gln Leu Gln Thr	565	570	575	
15	Val Val Gly Thr Tyr His Gly Ser Gln Asp Gln Pro His Gln Val Thr	580	585	590	
20	Gly Asn His Gln Gln Pro Pro Gln Gln Asn Thr Gly Phe Pro Arg Ser	595	600	605	
25	Ser Gln Pro Tyr Tyr Asn Ser Arg Gly Val Ser Arg Gly Gly Ser Arg	610	615	620	
30	Gly Ala Arg Gly Leu Met Asn Gly Tyr Arg Gly Pro Ala Asn Gly Phe	625	630	635	640
35	Arg Gly Gly Tyr Asp Gly Tyr Arg Pro Ser Phe Ser Asn Thr Pro Asn	645	650	655	
40	Ser Gly Tyr Thr Gln Ser Gln Phe Ser Ala Pro Arg Asp Tyr Ser Gly	660	665	670	
45	Tyr Gln Arg Asp Gly Tyr Gln Gln Asn Phe Lys Arg Gly Ser Gly Gln	675	680	685	
50	Ser Gly Pro Arg Gly Ala Pro Arg Gly Asn Ile Leu Trp Trp	690	695	700	
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 Ile Leu Gly Val Ile Asp Lys Lys Leu Arg Asn Leu Glu Lys Lys Lys
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 Gly Lys Leu Asp Asp Tyr Gln Glu Arg Met Asn Lys Gly Glu Arg Leu
 85 90 95

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 Asn Gln Asp Gln Leu Asp Ala Val Ser Lys Tyr Gln Glu Val Thr Asn
 100 105 110

 50 aac ttg gag ttt gca aaa gaa tta cag agg agt ttc atg gca tta agt 384
 Asn Leu Glu Phe Ala Lys Glu Leu Gln Arg Ser Phe Met Ala Leu Ser
 115 120 125

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10	atg aga gag gaa gcg gaa caa aaa cgt tta aaa act gta ctt gag ctc Met Arg Glu Glu Ala Glu Gln Lys Arg Leu Lys Thr Val Leu Glu Leu 145 150 155 160	480
15	cag tat gtt ttg gac aaa ttg gga gat gat gaa gtg aga act gac ctg Gln Tyr Val Leu Asp Lys Leu Gly Asp Asp Glu Val Arg Thr Asp Leu 165 170 175	528
20	aag caa ggt ttg aat gga gtg cca ata ttg tct gaa gaa gaa ttg tcg Lys Gln Gly Leu Asn Gly Val Pro Ile Leu Ser Glu Glu Glu Leu Ser 180 185 190	576
25	ttg ttg gat gaa ttc tac aaa tta gca gac cct gaa cgg gac atg agc Leu Leu Asp Glu Phe Tyr Lys Leu Ala Asp Pro Glu Arg Asp Met Ser 195 200 205	624
30	ttg agg ttg aat gag cag tat gaa cat gct tcc att cac ctg tgg gac Leu Arg Leu Asn Glu Gln Tyr Glu His Ala Ser Ile His Leu Trp Asp 210 215 220	672
35	ttg ctg gaa gga aag gaa aag tct gta tgt gga aca acc tat aaa gca Leu Leu Glu Gly Lys Glu Lys Ser Val Cys Gly Thr Thr Tyr Lys Ala 225 230 235 240	720
40	cta aag gaa att gtt gag cgt gtt ttc cag tca aat tac ttt gac agc Leu Lys Glu Ile Val Glu Arg Val Phe Gln Ser Asn Tyr Phe Asp Ser 245 250 255	768
45	act cac aac cac cag aat ggg cta tgt gag gaa gaa gag gca gcc tca Thr His Asn His Gln Asn Gly Leu Cys Glu Glu Glu Glu Ala Ala Ser 260 265 270	816
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10	gta gat gag tgg acg gtc gaa aca gtg gag gtg gtg aat tca ctc cag Val Asp Glu Trp Thr Val Glu Thr Val Glu Val Val Asn Ser Leu Gln 325 330 335	1008		
15	cag caa cct cag gct gcg tct cct tca gta cca gag ccc cac tct ttg Gln Gln Pro Gln Ala Ala Ser Pro Ser Val Pro Glu Pro His Ser Leu 340 345 350	1056		
20	act ccg gtg gct cag gca gat ccc ctt gtg aga aga cag cga gtc cag Thr Pro Val Ala Gln Ala Asp Pro Leu Val Arg Arg Gln Arg Val Gln 355 360 365	1104		
25	gac ctt atg gcg cag atg cag ggg ccc tat aat ttc ata cag gat tca Asp Leu Met Ala Gln Met Gln Gly Pro Tyr Asn Phe Ile Gln Asp Ser 370 375 380	1152		
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35	cag cct atg aat ccg aca caa aac atg gac atg ccc cag ctg gtt tgc Gln Pro Met Asn Pro Thr Gln Asn Met Asp Met Pro Gln Leu Val Cys 405 410 415	1248		
40	cct cca gtt cat tct gaa tct aga ctt gct caa cct aat caa gtt cct Pro Pro Val His Ser Glu Ser Arg Leu Ala Gln Pro Asn Gln Val Pro 420 425 430	1296		
45	gta caa cca gaa gct aca cag gtt cct ttg gtt tca tcc aca agt gag Val Gln Pro Glu Ala Thr Gln Val Pro Leu Val Ser Ser Thr Ser Glu 435 440 445	1344		
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55				

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10	tta aat aca gac cag act aca gcg tca tca tcc ctt ccg gct gct tct Leu Asn Thr Asp Gln Thr Thr Ala Ser Ser Ser Leu Pro Ala Ala Ser 485 490 495	1488
15	cag cct cag gta ttc cag gct ggg aca agc aaa cca tta cat agc agt Gln Pro Gln Val Phe Gln Ala Gly Thr Ser Lys Pro Leu His Ser Ser 500 505 510	1536
20	gga atc aat gta aat gca gct cca ttc caa tcc atg caa acg gtg ttc Gly Ile Asn Val Asn Ala Ala Pro Phe Gln Ser Met Gln Thr Val Phe 515 520 525	1584
25	aat atg aat gcc cca gtt cct cct gtt aat gaa cca gaa act ttg aaa Asn Met Asn Ala Pro Val Pro Pro Val Asn Glu Pro Glu Thr Leu Lys 530 535 540	1632
30	caa caa aat cag tac cag gcc agt tat aac cag agc ttt tct agt cag Gln Gln Asn Gln Tyr Gln Ala Ser Tyr Asn Gln Ser Phe Ser Ser Gln 545 550 555 560	1680
35	cct cac caa gta gaa caa aca gac ctt cag caa gaa cag ctt caa aca Pro His Gln Val Glu Gln Thr Asp Leu Gln Gln Glu Gln Leu Gln Thr 565 570 575	1728
40	gtg gtt ggc act tac cat ggt tcc cag gac cag ccc cac caa gtg act Val Val Gly Thr Tyr His Gly Ser Gln Asp Gln Pro His Gln Val Thr 580 585 590	1776
45	ggt aac cat cag cag cct ccc cag cag aac act gga ttt cca cgt agc Gly Asn His Gln Gln Pro Pro Gln Gln Asn Thr Gly Phe Pro Arg Ser 595 600 605	1824
50	agt cag ccc tat tac aat agt cgt ggt gtg tct cgt ggt ggt tcc cgt Ser Gln Pro Tyr Tyr Asn Ser Arg Gly Val Ser Arg Gly Gly Ser Arg 610 615 620	1872
55	ggt gct aga ggc tta atg aat gga tac agg ggc cct gcc aat gga ttc Gly Ala Arg Gly Leu Met Asn Gly Tyr Arg Gly Pro Ala Asn Gly Phe	1920

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5	aga gga gga tat gat ggt tac cgc cct tca ttc tct aac act cca aac				1968
	Arg Gly Gly Tyr Asp Gly Tyr Arg Pro Ser Phe Ser Asn Thr Pro Asn				
	645	650	655		
10	agt ggt tat aca cag tct cag ttc agt gct ccc cgg gac tac tct ggc				2016
	Ser Gly Tyr Thr Gln Ser Gln Phe Ser Ala Pro Arg Asp Tyr Ser Gly				
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	Tyr Gln Arg Gly Cys Arg Lys				
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	tggcactttt tgaaaaatat gcaacaaata tgggatgtaa tctggatggc cgcttctgta				2790
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 10 aatcataaca ctcttgggtca catgtttttc ctgcagcctg aaggttttta aaagaaaaag 3030
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 20 gtgttttaga ttgatttccc tattttaggg aaatgacaga cagtagtttc agttctgatg 3150
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<213> Canis familiaris

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Pro Pro Pro Pro Ser Gly Ser Ser Gly Ser Glu Ala Ala Ala Ala Ala
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Gly Ala Ala Gly Ala Ala Gly Ala Gly Ala Ala Ala Pro Ala Ser Gln
 35 40 45

45

His Pro Ala Thr Gly Thr Gly Ala Val Gln Thr Glu Ala Met Lys Gln
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50

Ile Leu Gly Val Ile Asp Lys Lys Leu Arg Asn Leu Glu Lys Lys Lys
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5	Gly	Lys	Leu	Asp	Asp	Tyr	Gln	Glu	Arg	Met	Asn	Lys	Gly	Glu	Arg	Leu	
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				100					105					110			
15	Asn	Leu	Glu	Phe	Ala	Lys	Glu	Leu	Gln	Arg	Ser	Phe	Met	Ala	Leu	Ser	
				115				120					125				
20	Gln	Asp	Ile	Gln	Lys	Thr	Ile	Lys	Lys	Thr	Ala	Arg	Arg	Glu	Gln	Leu	
				130				135				140					
25	Met	Arg	Glu	Glu	Ala	Glu	Gln	Lys	Arg	Leu	Lys	Thr	Val	Leu	Glu	Leu	
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30	Gln	Tyr	Val	Leu	Asp	Lys	Leu	Gly	Asp	Asp	Glu	Val	Arg	Thr	Asp	Leu	
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35	Lys	Gln	Gly	Leu	Asn	Gly	Val	Pro	Ile	Leu	Ser	Glu	Glu	Glu	Leu	Ser	
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40	Leu	Leu	Asp	Glu	Phe	Tyr	Lys	Leu	Ala	Asp	Pro	Glu	Arg	Asp	Met	Ser	
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 10 Ala Pro Thr Val Glu Asp Gln Val Ala Glu Ala Glu Pro Glu Pro Ala
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 15 Glu Glu Tyr Thr Glu Gln Ser Glu Val Glu Ser Thr Glu Tyr Val Asn
 290 295 300
 20 Arg Gln Phe Met Ala Glu Thr Gln Phe Ser Ser Gly Glu Lys Glu Gln
 305 310 315 320
 25 Val Asp Glu Trp Thr Val Glu Thr Val Glu Val Val Asn Ser Leu Gln
 325 330 335
 30 Gln Gln Pro Gln Ala Ala Ser Pro Ser Val Pro Glu Pro His Ser Leu
 340 345 350
 35 Thr Pro Val Ala Gln Ala Asp Pro Leu Val Arg Arg Gln Arg Val Gln
 355 360 365
 40 Asp Leu Met Ala Gln Met Gln Gly Pro Tyr Asn Phe Ile Gln Asp Ser
 370 375 380
 45 Met Leu Asp Phe Glu Asn Gln Thr Leu Asp Pro Ala Ile Val Ser Ala
 385 390 395 400
 50 Gln Pro Met Asn Pro Thr Gln Asn Met Asp Met Pro Gln Leu Val Cys
 405 410 415
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Pro Pro Val His Ser Glu Ser Arg Leu Ala Gln Pro Asn Gln Val Pro
 420 425 430
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Val Gln Pro Glu Ala Thr Gln Val Pro Leu Val Ser Ser Thr Ser Glu
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Gly Tyr Thr Ala Ser Gln Pro Leu Tyr Gln Pro Ser His Ala Thr Glu
 450 455 460
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Gln Arg Pro Gln Lys Glu Pro Ile Asp Gln Ile Gln Ala Thr Ile Ser
 465 470 475 480
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Leu Asn Thr Asp Gln Thr Thr Ala Ser Ser Ser Leu Pro Ala Ala Ser
 485 490 495
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Gln Pro Gln Val Phe Gln Ala Gly Thr Ser Lys Pro Leu His Ser Ser
 500 505 510
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Gly Ile Asn Val Asn Ala Ala Pro Phe Gln Ser Met Gln Thr Val Phe
 515 520 525
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Asn Met Asn Ala Pro Val Pro Pro Val Asn Glu Pro Glu Thr Leu Lys
 530 535 540
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Gln Gln Asn Gln Tyr Gln Ala Ser Tyr Asn Gln Ser Phe Ser Ser Gln
 545 550 555 560
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Pro His Gln Val Glu Gln Thr Asp Leu Gln Gln Glu Gln Leu Gln Thr
 565 570 575
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Val Val Gly Thr Tyr His Gly Ser Gln Asp Gln Pro His Gln Val Thr
 580 585 590
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5 Gly Asn His Gln Gln Pro Pro Gln Gln Asn Thr Gly Phe Pro Arg Ser
595 600 605

10 Ser Gln Pro Tyr Tyr Asn Ser Arg Gly Val Ser Arg Gly Gly Ser Arg
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15 Gly Ala Arg Gly Leu Met Asn Gly Tyr Arg Gly Pro Ala Asn Gly Phe
625 630 635 640

20 Arg Gly Gly Tyr Asp Gly Tyr Arg Pro Ser Phe Ser Asn Thr Pro Asn
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	Gly	Ala	Ala	Gly	Ala	Ala	Gly	Ala	Gly	Ala	Ala	Ala	Pro	Ala	Ser	Gln	
				35					40					45			
10																	
	cac	ccc	gcg	acc	ggc	acc	ggc	gct	gtc	cag	acc	gag	gcc	atg	aag	cag	192
	His	Pro	Ala	Thr	Gly	Thr	Gly	Ala	Val	Gln	Thr	Glu	Ala	Met	Lys	Gln	
				50					55					60			
15																	
	atc	ctc	ggg	gtg	atc	gac	aag	aaa	ctc	cgg	aac	ctg	gag	aag	aaa	aag	240
	Ile	Leu	Gly	Val	Ile	Asp	Lys	Lys	Leu	Arg	Asn	Leu	Glu	Lys	Lys	Lys	
	65						70					75				80	
20																	
	ggc	aag	ctt	gat	gat	tac	cag	gaa	cga	atg	aac	aaa	ggg	gaa	agg	ctt	288
	Gly	Lys	Leu	Asp	Asp	Tyr	Gln	Glu	Arg	Met	Asn	Lys	Gly	Glu	Arg	Leu	
							85				90				95		
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	aat	caa	gat	cag	ctg	gat	gcc	gta	tct	aag	tac	cag	gaa	gtc	aca	aat	336
	Asn	Gln	Asp	Gln	Leu	Asp	Ala	Val	Ser	Lys	Tyr	Gln	Glu	Val	Thr	Asn	
							100				105				110		
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	aac	ttg	gag	ttt	gca	aaa	gaa	tta	cag	agg	agt	ttc	atg	gca	tta	agt	384
	Asn	Leu	Glu	Phe	Ala	Lys	Glu	Leu	Gln	Arg	Ser	Phe	Met	Ala	Leu	Ser	
				115					120					125			
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	Gln	Asp	Ile	Gln	Lys	Thr	Ile	Lys	Lys	Thr	Ala	Arg	Arg	Glu	Gln	Leu	
				130					135					140			
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	145						150					155				160	
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	Gln	Tyr	Val	Leu	Asp	Lys	Leu	Gly	Asp	Asp	Glu	Val	Arg	Thr	Asp	Leu	
							165					170			175		
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	aag	caa	ggt	ttg	aat	gga	gtg	cca	ata	ttg	tct	gaa	gaa	gaa	ttg	tcg	576
	Lys	Gln	Gly	Leu	Asn	Gly	Val	Pro	Ile	Leu	Ser	Glu	Glu	Glu	Leu	Ser	
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20	cta aag gaa att gtt gag cgt gtt ttc cag tca aat tac ttt gac agc Leu Lys Glu Ile Val Glu Arg Val Phe Gln Ser Asn Tyr Phe Asp Ser 245 250 255	768
25	act cac aac cac cag aat ggg cta tgt gag gaa gaa gag gca gcc tca Thr His Asn His Gln Asn Gly Leu Cys Glu Glu Glu Ala Ala Ser 260 265 270	816
30	gca cct aca gtt gaa gac cag gta gct gaa gct gag cct gag cca gca Ala Pro Thr Val Glu Asp Gln Val Ala Glu Ala Glu Pro Glu Pro Ala 275 280 285	864
35	gaa gaa tac act gaa caa agt gaa gtt gaa tca aca gag tat gta aat Glu Glu Tyr Thr Glu Gln Ser Glu Val Glu Ser Thr Glu Tyr Val Asn 290 295 300	912
40	aga caa ttt atg gca gaa aca cag ttc agc agt ggt gaa aag gag cag Arg Gln Phe Met Ala Glu Thr Gln Phe Ser Ser Gly Glu Lys Glu Gln 305 310 315 320	960
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50	cag caa cct cag gct gcg tct cct tca gta cca gag ccc cac tct ttg Gln Gln Pro Gln Ala Ala Ser Pro Ser Val Pro Glu Pro His Ser Leu 340 345 350	1056
55	act ccg gtg gct cag gca gat ccc ctt gtg aga aga cag cga gtc cag	1104

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	Asp Leu Met Ala Gln Met Gln Gly Pro Tyr Asn Phe Ile Gln Asp Ser	
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	Met Leu Asp Phe Glu Asn Gln Thr Leu Asp Pro Ala Ile Val Ser Ala	
	385 390 395 400	
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	ggg tat aca gca tct caa ccc ttg tac cag cct tct cat gct aca gag	1392
	Gly Tyr Thr Ala Ser Gln Pro Leu Tyr Gln Pro Ser His Ala Thr Glu	
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	Gln Arg Pro Gln Lys Glu Pro Ile Asp Gln Ile Gln Ala Thr Ile Ser	
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	tta aat aca gac cag act aca gcg tca tca tcc ctt ccg gct gct tct	1488
	Leu Asn Thr Asp Gln Thr Thr Ala Ser Ser Ser Leu Pro Ala Ala Ser	
	485 490 495	
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	Gln Pro Gln Val Phe Gln Ala Gly Thr Ser Lys Pro Leu His Ser Ser	
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	gga atc aat gta aat gca gct cca ttc caa tcc atg caa acg gtg ttc	1584
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30	agt cag ccc tat tac aat agt cgt ggt gtg tct cgt ggt ggt tcc cgt Ser Gln Pro Tyr Tyr Asn Ser Arg Gly Val Ser Arg Gly Gly Ser Arg 610 615 620	1872
35	ggt gct aga ggc tta atg aat gga tac agg ggc cct gcc aat gga ttc Gly Ala Arg Gly Leu Met Asn Gly Tyr Arg Gly Pro Ala Asn Gly Phe 625 630 635 640	1920
40	aga gga gga tat gat ggt tac cgc cct tca ttc tct aac act cca aac Arg Gly Gly Tyr Asp Gly Tyr Arg Pro Ser Phe Ser Asn Thr Pro Asn 645 650 655	1968
45	agt ggt tat aca cag tct cag ttc agt gct ccc cgg gac tac tct ggc Ser Gly Tyr Thr Gln Ser Gln Phe Ser Ala Pro Arg Asp Tyr Ser Gly 660 665 670	2016
50	tat cag cgg gat gga tat cag cag aat ttc aag cga ggc tct ggg cag Tyr Gln Arg Asp Gly Tyr Gln Gln Asn Phe Lys Arg Gly Ser Gly Gln 675 680 685	2064
55	agt gga cca cgg gga gcc cca cga ggt cgt gga ggg ccc cca aga ccc	2112

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690 695 700

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aac aga ggg atg ccg caa atg aac act cag caa gtg aat taa 2154
Asn Arg Gly Met Pro Gln Met Asn Thr Gln Gln Val Asn
705 710 715

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tctgattcac aggattatgt ttaaacgccca aaaacacact ggccagtgtta ccataatatg 2214
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Pro Pro Pro Pro Ser Gly Ser Ser Gly Ser Glu Ala Ala Ala Ala Ala
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Gly Ala Ala Gly Ala Ala Gly Ala Gly Ala Ala Ala Pro Ala Ser Gln
35 40 45

45
His Pro Ala Thr Gly Thr Gly Ala Val Gln Thr Glu Ala Met Lys Gln
50 55 60

50
Ile Leu Gly Val Ile Asp Lys Lys Leu Arg Asn Leu Glu Lys Lys Lys
65 70 75 80

55
Gly Lys Leu Asp Asp Tyr Gln Glu Arg Met Asn Lys Gly Glu Arg Leu

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	85	90	95
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10	Asn Leu Glu Phe Ala Lys Glu Leu Gln Arg Ser Phe Met Ala Leu Ser		
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15	Gln Asp Ile Gln Lys Thr Ile Lys Lys Thr Ala Arg Arg Glu Gln Leu		
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20	Met Arg Glu Glu Ala Glu Gln Lys Arg Leu Lys Thr Val Leu Glu Leu		
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25	Gln Tyr Val Leu Asp Lys Leu Gly Asp Asp Glu Val Arg Thr Asp Leu		
	165	170	175
30	Lys Gln Gly Leu Asn Gly Val Pro Ile Leu Ser Glu Glu Glu Leu Ser		
	180	185	190
35	Leu Leu Asp Glu Phe Tyr Lys Leu Ala Asp Pro Glu Arg Asp Met Ser		
	195	200	205
40	Leu Arg Leu Asn Glu Gln Tyr Glu His Ala Ser Ile His Leu Trp Asp		
	210	215	220
45	Leu Leu Glu Gly Lys Glu Lys Ser Val Cys Gly Thr Thr Tyr Lys Ala		
	225	230	235
50	Leu Lys Glu Ile Val Glu Arg Val Phe Gln Ser Asn Tyr Phe Asp Ser		
	245	250	255

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5 Thr His Asn His Gln Asn Gly Leu Cys Glu Glu Glu Glu Ala Ala Ser
 260 265 270

10 Ala Pro Thr Val Glu Asp Gln Val Ala Glu Ala Glu Pro Glu Pro Ala
 275 280 285

15 Glu Glu Tyr Thr Glu Gln Ser Glu Val Glu Ser Thr Glu Tyr Val Asn
 290 295 300

20 Arg Gln Phe Met Ala Glu Thr Gln Phe Ser Ser Gly Glu Lys Glu Gln
 305 310 315 320

25 Val Asp Glu Trp Thr Val Glu Thr Val Glu Val Val Asn Ser Leu Gln
 325 330 335

30 Gln Gln Pro Gln Ala Ala Ser Pro Ser Val Pro Glu Pro His Ser Leu
 340 345 350

35 Thr Pro Val Ala Gln Ala Asp Pro Leu Val Arg Arg Gln Arg Val Gln
 355 360 365

40 Asp Leu Met Ala Gln Met Gln Gly Pro Tyr Asn Phe Ile Gln Asp Ser
 370 375 380

45 Met Leu Asp Phe Glu Asn Gln Thr Leu Asp Pro Ala Ile Val Ser Ala
 385 390 395 400

50 Gln Pro Met Asn Pro Thr Gln Asn Met Asp Met Pro Gln Leu Val Cys
 405 410 415

55 Pro Pro Val His Ser Glu Ser Arg Leu Ala Gln Pro Asn Gln Val Pro

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20	Leu Asn Thr Asp Gln Thr Thr Ala Ser Ser Ser Leu Pro Ala Ala Ser		
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25	Gln Pro Gln Val Phe Gln Ala Gly Thr Ser Lys Pro Leu His Ser Ser		
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30	Gly Ile Asn Val Asn Ala Ala Pro Phe Gln Ser Met Gln Thr Val Phe		
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35	Asn Met Asn Ala Pro Val Pro Pro Val Asn Glu Pro Glu Thr Leu Lys		
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40	Gln Gln Asn Gln Tyr Gln Ala Ser Tyr Asn Gln Ser Phe Ser Ser Gln		
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45	Pro His Gln Val Glu Gln Thr Asp Leu Gln Gln Glu Gln Leu Gln Thr		
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595 600 605

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15 Gly Ala Arg Gly Leu Met Asn Gly Tyr Arg Gly Pro Ala Asn Gly Phe
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20 Arg Gly Gly Tyr Asp Gly Tyr Arg Pro Ser Phe Ser Asn Thr Pro Asn
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25 Ser Gly Tyr Thr Gln Ser Gln Phe Ser Ala Pro Arg Asp Tyr Ser Gly
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30 Tyr Gln Arg Asp Gly Tyr Gln Gln Asn Phe Lys Arg Gly Ser Gly Gln
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Met Pro Ser Ala Thr Ser His Ser Gly Ser
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Glu Ala Gly Ala Gly Ala Ala Ala Pro Ala Ser Gln His Pro Met Thr
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Gly Thr Gly Ala Val Gln Thr Glu Ala Met Lys Gln Ile Leu Gly Val
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Ile Asp Lys Lys Leu Arg Asn Leu Glu Lys Lys Lys Gly Lys Leu Asp
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Asp Tyr Gln Glu Arg Met Asn Lys Gly Glu Arg Leu Asn Gln Asp Gln
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Leu Asp Ala Val Ser Lys Tyr Gln Glu Val Thr Asn Asn Leu Glu Phe
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Ala Lys Glu Leu Gln Arg Ser Phe Met Ala Leu Ser Gln Asp Ile Gln
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Lys Thr Ile Lys Lys Thr Ala Arg Arg Glu Gln Leu Met Arg Glu Glu
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gct gaa cag aaa cgt tta aaa aca gta ctt gag ctg cag tat gtt ttg 543
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20	caa atg cag ggg ccc tat aat ttc ata cag gat tca atg ttg gat ttt Gln Met Gln Gly Pro Tyr Asn Phe Ile Gln Asp Ser Met Leu Asp Phe 365 370 375	1215
25	gaa aac cag aca ctt gat cct gcc att gta tct gca cag ccg atg aat Glu Asn Gln Thr Leu Asp Pro Ala Ile Val Ser Ala Gln Pro Met Asn 380 385 390	1263
30	cca gca cag aac atg gac ata ccc cag ctg gtt tgc cct cca gtt cat Pro Ala Gln Asn Met Asp Ile Pro Gln Leu Val Cys Pro Pro Val His 395 400 405 410	1311
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	Phe Gln Ala Gly Thr Ser Lys Pro Leu His Ser Ser Gly Ile Asn Val							
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	Asn Ala Ala Pro Phe Gln Ser Met Gln Thr Val Phe Asn Met Asn Ala							
		510		515		520		
15	cca gtt cct cct gtt aat gaa cca gaa act tta aaa cag caa aat cag							1695
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25	gaa caa aca gag ctt cag caa gaa cag ctt caa aca gtg gtt ggc act							1791
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30	tat cat ggt tct cag gac cag ccc cat caa gtg act ggt aac cac cag							1839
	Tyr His Gly Ser Gln Asp Gln Pro His Gln Val Thr Gly Asn His Gln							
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35	cag cct cct cag cag aac act gga ttt cca cgt agc aat cag ccc tat							1887
	Gln Pro Pro Gln Gln Asn Thr Gly Phe Pro Arg Ser Asn Gln Pro Tyr							
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	Tyr Asn Ser Arg Gly Val Ser Arg Gly Gly Ser Arg Gly Ala Arg Gly							
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45	ttg atg aat gga tac aga gga cct gct aat gga ttc aga gga gga tat							1983
	Leu Met Asn Gly Tyr Arg Gly Pro Ala Asn Gly Phe Arg Gly Gly Tyr							
		620		625		630		
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15	cgg gga gcc cca cga ggt cgt gga ggg ccc cca aga ccc aac aga ggg Arg Gly Ala Pro Arg Gly Arg Gly Gly Pro Pro Arg Pro Asn Arg Gly 685 690 695	2175
20	atg ccg caa atg aac act cag caa gtg aat taa tctgattcac aggattatgt Met Pro Gln Met Asn Thr Gln Gln Val Asn 700 705	2228
25	ttaatcgcca aaaacacact ggccagtgtta ccataatatg ttaccagaag agttattatc	2288
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gtgttttaga ttgatttccc tattttaggg aaatgacagt cagtagtttc acttctgatg 3308

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35 40 45

45 Thr Glu Ala Met Lys Gln Ile Leu Gly Val Ile Asp Lys Lys Leu Arg

50 55 60

50 Asn Leu Glu Lys Lys Lys Gly Lys Leu Asp Asp Tyr Gln Glu Arg Met

65 70 75 80

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Tyr Gln Glu Val Thr Asn Asn Leu Glu Phe Ala Lys Glu Leu Gln Arg	
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Lys Thr Val Leu Glu Leu Gln Tyr Val Leu Asp Lys Leu Gly Asp Asp	
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305 310 315 320

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325 330 335

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35 Arg Arg Gln Arg Val Gln Asp Leu Met Ala Gln Met Gln Gly Pro Tyr
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40 Asn Phe Ile Gln Asp Ser Met Leu Asp Phe Glu Asn Gln Thr Leu Asp
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 Ser Leu Pro Ala Ala Ser Gln Pro Gln Val Phe Gln Ala Gly Thr Ser
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 Glu Pro Glu Thr Leu Lys Gln Gln Asn Gln Tyr Gln Ala Ser Tyr Asn
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 Gln Glu Gln Leu Gln Thr Val Val Gly Thr Tyr His Gly Ser Gln Asp
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 Gln Pro His Gln Val Thr Gly Asn His Gln Gln Pro Pro Gln Gln Asn
 580 585 590
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 610 615 620
 15 Gly Pro Ala Asn Gly Phe Arg Gly Gly Tyr Asp Gly Tyr Arg Pro Ser
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 20 Phe Ser Thr Asn Thr Pro Asn Ser Gly Tyr Thr Gln Ser Gln Phe Ser
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 25 Ala Pro Arg Asp Tyr Ser Gly Tyr Gln Arg Asp Gly Tyr Gln Gln Asn
 660 665 670
 30 Phe Lys Arg Gly Ser Gly Gln Ser Gly Pro Arg Gly Ala Pro Arg Gly
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	Arg Leu Asn Gln Asp Gln Leu Asp Ala Val Ser Lys Tyr Gln Glu Val	
	20 25 30	
15	aca aat aac ttg gag ttt gcg aaa gaa ttg cag agg agt ttc atg gcg	144
	Thr Asn Asn Leu Glu Phe Ala Lys Glu Leu Gln Arg Ser Phe Met Ala	
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	Asp Leu Lys Gln Gly Leu Asn Gly Val Pro Ile Leu Ser Glu Glu Glu	
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	Leu Ser Leu Leu Asp Glu Phe Tyr Lys Leu Ala Asp Pro Val Arg Asp	
	115 120 125	
45	atg agc ttg agg ttg aat gag cag tat gag cat gcc tcc att cac ctg	432
	Met Ser Leu Arg Leu Asn Glu Gln Tyr Glu His Ala Ser Ile His Leu	
	130 135 140	
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	145 150 155 160	
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15	acc tca gct cca aca gct gaa gac cag gga gct gaa gct gaa cct gag Thr Ser Ala Pro Thr Ala Glu Asp Gln Gly Ala Glu Ala Glu Pro Glu 195 200 205	624
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45	cag gac ctt atg gcg caa atg cag ggg ccc tat aat ttc ata cag gat Gln Asp Leu Met Ala Gln Met Gln Gly Pro Tyr Asn Phe Ile Gln Asp 290 295 300	912
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	325 330 335	
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10	cct gta caa cca gaa gct aca cag gtt cct ttg gtt tca tcc aca agt Pro Val Gln Pro Glu Ala Thr Gln Val Pro Leu Val Ser Ser Thr Ser	1104
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15	gag ggg tat aca gca tct cag ccc ttg tac cag cct tct cat gct aca Glu Gly Tyr Thr Ala Ser Gln Pro Leu Tyr Gln Pro Ser His Ala Thr	1152
	370 375 380	
20	gag caa cga ccg caa aag gaa ccg act gac cag atc cag gca aca atc Glu Gln Arg Pro Gln Lys Glu Pro Thr Asp Gln Ile Gln Ala Thr Ile	1200
	385 390 395 400	
25	tct tta aat aca gac cag act aca gca tca tca tcc ctt cct gct gct Ser Leu Asn Thr Asp Gln Thr Thr Ala Ser Ser Ser Leu Pro Ala Ala	1248
	405 410 415	
30	tct cag cct cag gtg ttc cag gct ggg aca agc aaa cct tta cac agc Ser Gln Pro Gln Val Phe Gln Ala Gly Thr Ser Lys Pro Leu His Ser	1296
	420 425 430	
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	435 440 445	
40	ttc aac atg aat gcc ccg gtt cct cct gtt aat gaa cca gaa act tta Phe Asn Met Asn Ala Pro Val Pro Pro Val Asn Glu Pro Glu Thr Leu	1392
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	485 490 495	
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15	agc agt cag ccc tat tac aac agt cgt ggt gtg tct cgt gga ggc tcc Ser Ser Gln Pro Tyr Tyr Asn Ser Arg Gly Val Ser Arg Gly Gly Ser 530 535 540	1632
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25	ttc aga gga gga tat gat ggt tac cgc cct tcg ttc tct aac act cca Phe Arg Gly Gly Tyr Asp Gly Tyr Arg Pro Ser Phe Ser Asn Thr Pro 565 570 575	1728
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35	ggc tat cag cgg gat gga tat cag cag aat ttc aag cga ggc tct ggg Gly Tyr Gln Arg Asp Gly Tyr Gln Gln Asn Phe Lys Arg Gly Ser Gly 595 600 605	1824
40	cag agt gga ccc cgg gga gcc cca cga ggt cgt gga ggg ccc cca aga Gln Ser Gly Pro Arg Gly Ala Pro Arg Gly Arg Gly Gly Pro Pro Arg 610 615 620	1872
45	ccc aac aga ggg atg ccg caa atg aac act cag caa gtg aat taa Pro Asn Arg Gly Met Pro Gln Met Asn Thr Gln Gln Val Asn 625 630 635	1917
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Leu Ser Leu Leu Asp Glu Phe Tyr Lys Leu Ala Asp Pro Val Arg Asp
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Trp Asp Leu Leu Glu Gly Lys Glu Lys Ser Val Cys Gly Thr Thr Tyr
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25	Asn	Leu	Glu	Lys	Lys	Lys	Gly	Lys	Leu	Asp	Asp	Tyr	Gln	Glu	Arg	Met	65	70	75	80
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30	Glu Glu Glu Ala Ala Ser Ala Pro Thr Val Glu Asp Gln Val Ala Glu		
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40	Ser Thr Glu Tyr Val Asn Arg Gln Phe Met Ala Glu Thr Gln Phe Ser		
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	305	310	315 320
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Arg Arg Gln Arg Val Gln Asp Leu Met Ala Gln Met Gln Gly Pro Tyr
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Pro Ala Ile Val Ser Ala Gln Pro Met Asn Pro Thr Gln Asn Met Asp
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Gln Ser Asn Gln Val Pro Val Gln Pro Glu Ala Thr Gln Val Pro Leu
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Val Ser Ser Thr Ser Glu Gly Tyr Thr Ala Ser Gln Pro Leu Tyr Gln
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Pro Ser His Ala Thr Glu Gln Arg Pro Gln Lys Glu Pro Met Asp Gln
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50 Ser Leu Pro Ala Ala Ser Gln Pro Gln Val Phe Gln Ala Gly Thr Ser
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55 Lys Pro Leu His Ser Ser Gly Ile Asn Val Asn Ala Ala Pro Phe Gln

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15	Gln Ser Phe Ser Ser Gln Pro His Gln Val Glu Gln Thr Glu Leu Gln				
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20	Gln Asp Gln Leu Gln Thr Val Val Gly Thr Tyr His Gly Ser Gln Asp				
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tctcttctcg gtctaaag atg ccc tcg gcc acc agc cac agc gga agc ggc 171
 Met Pro Ser Ala Thr Ser His Ser Gly Ser Gly

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 Ser Lys Ser Ser Gly Pro Pro Pro Pro Ser Gly Ser Ser Gly Ser Glu

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gcg gcg gcc ggg gca get gcg ccg get tet cag cat ccg gca acc ggc 267
 Ala Ala Ala Gly Ala Ala Ala Pro Ala Ser Gln His Pro Ala Thr Gly

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 Thr Gly Ala Val Gln Thr Glu Ala Met Lys Gln Ile Leu Gly Val Ile

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	Tyr Gln Glu Arg Met Asn Lys Gly Glu Arg Leu Asn Gln Asp Gln Leu												
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	Asp Ala Val Ser Lys Tyr Gln Glu Val Thr Asn Asn Leu Glu Phe Ala												
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	Lys Glu Leu Gln Arg Ser Phe Met Ala Leu Ser Gln Asp Ile Gln Lys												
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	Thr Ile Lys Lys Thr Ala Arg Arg Glu Gln Leu Met Arg Glu Glu Ala												
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	Glu Gln Lys Arg Leu Lys Thr Val Leu Glu Leu Gln Tyr Val Leu Asp												
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25	ata tct ttg aat aca gac cag act aca gca tcc tca tcc ctt cct gct Ile Ser Leu Asn Thr Asp Gln Thr Thr Ala Ser Ser Ser Leu Pro Ala	460	465	470	1563
30	gct tct cag cct caa gtg ttc cag gct ggg aca agt aaa cct ttg cac Ala Ser Gln Pro Gln Val Phe Gln Ala Gly Thr Ser Lys Pro Leu His	480	485	490	1611
35	agc agt gga atc aat gta aat gca gct cca ttc cag tcc atg caa acg Ser Ser Gly Ile Asn Val Asn Ala Ala Pro Phe Gln Ser Met Gln Thr	495	500	505	1659
40	gtg ttc aat atg aat gct cca gtc cct cct gct aat gaa cca gaa acg Val Phe Asn Met Asn Ala Pro Val Pro Pro Ala Asn Glu Pro Glu Thr	510	515	520	1707
45	tta aaa caa cag agt cag tac cag gcc act tat aac cag agt ttt tcc Leu Lys Gln Gln Ser Gln Tyr Gln Ala Thr Tyr Asn Gln Ser Phe Ser	525	530	535	1755
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	575 580 585	
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	590 595 600	
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30 Asn Leu Glu Lys Lys Lys Gly Lys Leu Asp Asp Tyr Gln Glu Arg Met
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115 120 125

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130 135 140

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145 150 155 160

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Asp Val Arg Thr Asp Leu Lys Gln Gly Leu Ser Gly Val Pro Ile Leu
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Ser Glu Glu Glu Leu Ser Leu Leu Asp Glu Phe Tyr Lys Leu Val Asp
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Pro Glu Arg Asp Met Ser Leu Arg Leu Asn Glu Gln Tyr Glu His Ala
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Gly Thr Thr Tyr Lys Ala Leu Lys Glu Ile Val Glu Arg Val Phe Gln
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Glu Glu Glu Ala Ala Ser Ala Pro Thr Val Glu Asp Gln Val Ala Glu
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Ser Gly Glu Lys Glu Gln Val Asp Glu Trp Thr Val Glu Thr Val Glu
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 45 Gln Lys Glu Pro Met Asp Gln Ile Gln Ala Thr Ile Ser Leu Asn Thr
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Thr Glu Ala Met Lys Gln Ile Leu Gly Val Ile Asp Lys Lys Leu Arg
25 50 55 60

Asn Leu Glu Lys Lys Lys Gly Lys Leu Asp Asp Tyr Gln Glu Arg Met
30 65 70 75 80

Asn Lys Gly Glu Arg Leu Asn Gln Asp Gln Leu Asp Ala Val Ser Lys
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Tyr Gln Glu Val Thr Asn Asn Leu Glu Phe Ala Lys Glu Leu Gln Arg
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Ser Phe Met Ala Leu Ser Gln Asp Ile Gln Lys Thr Ile Lys Lys Thr
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Ala Arg Arg Glu Gln Leu Met Arg Glu Glu Ala Glu Gln Lys Arg Leu
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Lys Thr Val Leu Glu Leu Gln Tyr Val Leu Asp Lys Leu Gly Asp Asp
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			240
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10	Ser Met Gln Thr Val Phe Asn Met Asn Ala Pro Val Pro Pro Ala Asn		
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15	Glu Pro Glu Thr Leu Lys Gln Gln Ser Gln Tyr Gln Ala Thr Tyr Asn		
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20	Gln Ser Phe Ser Ser Gln Pro His Gln Val Glu Gln Thr Glu Leu Gln		
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25	Gln Asp Gln Leu Gln Thr Val Val Gly Thr Tyr His Gly Ser Gln Asp		
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30	Gln Pro His Gln Val Pro Gly Asn His Gln Gln Pro Pro Gln Gln Asn		
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35	Thr Gly Phe Pro Arg Ser Ser Gln Pro Tyr Tyr Asn Ser Arg Gly Val		
	595	600	605
40	Ser Arg Gly Gly Ser Arg Gly Ala Arg Gly Leu Met Asn Gly Tyr Arg		
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45	Gly Pro Ala Asn Gly Phe Arg Gly Gly Tyr Asp Gly Tyr Arg Pro Ser		
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50	Phe Ser Asn Thr Pro Asn Ser Gly Tyr Ser Gln Ser Gln Phe Thr Ala		
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	Glu Ser Arg Leu Ala Gln Ser Asn Gln Val Pro Val Gln Pro Glu Ala				
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	Glu Pro Met Asp Gln Ile Gln Ala Thr Ile Ser Leu Asn Thr Asp Gln				
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	Thr Thr Ala Ser Ser Ser Leu Pro Ala Ala Ser Gln Pro Gln Val Phe				
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 40 attaatTTtg atagtatgat gtcacttctt attgaaatgt aagctagcgt gtaccttaga 3377
 atgtgagctc catgagagca ggtaccttgt ttgtcttcac tgctgtatct attcccaacg 3437
 45 cctcatgaca gtgcctggca catagtaggc actcaataaa tacttgttga atgaatgaaa 3497
 aaaaaaaaa a 3508
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 <210> 28
 <211> 692
 55 <212> PRT

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<213> Mus musculus

5 <400> 28

Met Pro Ser Ala Thr Ser His Ser Gly Ser Gly Ser Lys Ser Ser Gly
1 5 10 15

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Pro Pro Pro Pro Ser Gly Ser Ser Gly Ser Glu Ala Ala Ala Gly Ala
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Ala Ala Pro Ala Ser Gln His Pro Ala Thr Gly Thr Gly Ala Val Gln
35 40 45

20

Thr Glu Ala Met Lys Gln Ile Leu Gly Val Ile Asp Lys Lys Leu Arg
50 55 60

25

Asn Leu Glu Lys Lys Lys Gly Lys Leu Asp Asp Tyr Gln Glu Arg Met
65 70 75 80

30

Asn Lys Gly Glu Arg Leu Asn Gln Asp Gln Leu Asp Ala Val Ser Lys
85 90 95

35

Tyr Gln Glu Val Thr Asn Asn Leu Glu Phe Ala Lys Glu Leu Gln Arg
100 105 110

40

Ser Phe Met Ala Leu Ser Gln Asp Ile Gln Lys Thr Ile Lys Lys Thr
115 120 125

45

Ala Arg Arg Glu Gln Leu Met Arg Glu Glu Ala Glu Gln Lys Arg Leu
130 135 140

50

Lys Thr Val Leu Glu Leu Gln Tyr Val Leu Asp Lys Leu Gly Asp Asp
145 150 155 160

55

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5 Asp Val Arg Thr Asp Leu Lys Gln Gly Leu Ser Gly Val Pro Ile Leu
 165 170 175

10 Ser Glu Glu Glu Leu Ser Leu Leu Asp Glu Phe Tyr Lys Leu Val Asp
 180 185 190

15 Pro Glu Arg Asp Met Ser Leu Arg Leu Asn Glu Gln Tyr Glu His Ala
 195 200 205

20 Ser Ile His Leu Trp Asp Leu Leu Glu Gly Lys Glu Lys Pro Val Cys
 210 215 220

25 Gly Thr Thr Tyr Lys Ala Leu Lys Glu Ile Val Glu Arg Val Phe Gln
 225 230 235 240

30 Ser Asn Tyr Phe Asp Ser Thr His Asn His Gln Asn Gly Leu Cys Glu
 245 250 255

35 Glu Glu Glu Ala Ala Ser Ala Pro Thr Val Glu Asp Gln Val Ala Glu
 260 265 270

40 Ala Glu Pro Glu Pro Ala Glu Glu Tyr Thr Glu Gln Ser Glu Val Glu
 275 280 285

45 Ser Thr Glu Tyr Val Asn Arg Gln Phe Met Ala Glu Thr Gln Phe Ser
 290 295 300

50 Ser Gly Glu Lys Glu Gln Val Asp Glu Trp Thr Val Glu Thr Val Glu
 305 310 315 320

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5	Val Val Asn Ser Leu Gln Gln Gln Pro Gln Ala Ala Ser Pro Ser Val	325	330	335	
10	Pro Glu Pro His Ser Leu Thr Pro Val Ala Gln Ser Asp Pro Leu Val	340	345	350	
15	Arg Arg Gln Arg Val Gln Asp Leu Met Ala Gln Met Gln Gly Pro Tyr	355	360	365	
20	Asn Phe Ile Gln Asp Ser Met Leu Asp Phe Glu Asn Gln Thr Leu Asp	370	375	380	
25	Pro Ala Ile Val Ser Ala Gln Pro Met Asn Pro Thr Gln Asn Met Asp	385	390	395	400
30	Met Pro Gln Leu Val Cys Pro Gln Val His Ser Glu Ser Arg Leu Ala	405	410	415	
35	Gln Ser Asn Gln Val Pro Val Gln Pro Glu Ala Thr Gln Val Pro Leu	420	425	430	
40	Val Ser Ser Thr Ser Glu Gly Tyr Thr Ala Ser Gln Pro Leu Tyr Gln	435	440	445	
45	Pro Ser His Ala Thr Glu Gln Arg Pro Gln Lys Glu Pro Met Asp Gln	450	455	460	
50	Ile Gln Ala Thr Ile Ser Leu Asn Thr Asp Gln Thr Thr Ala Ser Ser	465	470	475	480
55	Ser Leu Pro Ala Ala Ser Gln Pro Gln Val Phe Gln Ala Gly Thr Ser	485	490	495	

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5 Lys Pro Leu His Ser Ser Gly Ile Asn Val Asn Ala Ala Pro Phe Gln
500 505 510

10 Ser Met Gln Thr Val Phe Asn Met Asn Ala Pro Val Pro Pro Ala Asn
515 520 525

15 Glu Pro Glu Thr Leu Lys Gln Gln Ser Gln Tyr Gln Ala Thr Tyr Asn
530 535 540

20 Gln Ser Phe Ser Ser Gln Pro His Gln Val Glu Gln Thr Glu Leu Gln
545 550 555 560

25 Gln Asp Gln Leu Gln Thr Val Val Gly Thr Tyr His Gly Ser Gln Asp
565 570 575

30 Gln Pro His Gln Val Pro Gly Asn His Gln Gln Pro Pro Gln Gln Asn
580 585 590

35 Thr Gly Phe Pro Arg Ser Ser Gln Pro Tyr Tyr Asn Ser Arg Gly Val
595 600 605

40 Ser Arg Gly Gly Ser Arg Gly Ala Arg Gly Leu Met Asn Gly Tyr Arg
610 615 620

45 Gly Pro Ala Asn Gly Phe Arg Gly Gly Tyr Asp Gly Tyr Arg Pro Ser
625 630 635 640

50 Phe Ser Asn Thr Pro Asn Ser Gly Tyr Ser Gln Ser Gln Phe Thr Ala
645 650 655

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Pro Arg Asp Tyr Ser Gly Tyr Gln Arg Asp Gly Tyr Gln Gln Asn Phe
660 665 670

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Lys Arg Gly Ser Gly Gln Ser Gly Pro Arg Gly Ala Pro Arg Gly Asn
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Ile Leu Trp Trp
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<211> 2109
20 <212> DNA
<213> Gallus gallus

<220>
25 <221> CDS
<222> (1).. (2109)
<223>

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35 ggc ccg ggc ggc aac gag cag gcc ccg gcg gcg gca gcg gcg gcc ccg 96
Gly Pro Gly Gly Asn Glu Gln Ala Pro Ala Ala Ala Ala Ala Ala Pro
20 25 30

40 cag gcg tcg ggc ggc agc atc acc tcg gtt cag acc gag gcc atg aag 144
Gln Ala Ser Gly Gly Ser Ile Thr Ser Val Gln Thr Glu Ala Met Lys
35 40 45

45 cag atc ttg gga gtg atc gac aaa aag ctc cgc aac ctc gag aag aaa 192
Gln Ile Leu Gly Val Ile Asp Lys Lys Leu Arg Asn Leu Glu Lys Lys
50 55 60

55 aag agc aaa ctt gac gat tac cag gaa cga atg aac aag ggg gaa cgt 240
Lys Ser Lys Leu Asp Asp Tyr Gln Glu Arg Met Asn Lys Gly Glu Arg
65 70 75 80

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5	cta aat caa gat caa ctg gat gca gtg tca aaa tac cag gaa gtg aca Leu Asn Gln Asp Gln Leu Asp Ala Val Ser Lys Tyr Gln Glu Val Thr	288
	85 90 95	
10	aat aac ctg gaa ttc gct aaa gaa ctg cag agg agc ttt atg gca ctg Asn Asn Leu Glu Phe Ala Lys Glu Leu Gln Arg Ser Phe Met Ala Leu	336
	100 105 110	
15	agc caa gat atc cag aaa aca ata aaa aag acg gct cgc agg gag cag Ser Gln Asp Ile Gln Lys Thr Ile Lys Lys Thr Ala Arg Arg Glu Gln	384
	115 120 125	
20	ctg atg aga gaa gag gct gag cag aag cgt tta aag act gtg cta gag Leu Met Arg Glu Glu Ala Glu Gln Lys Arg Leu Lys Thr Val Leu Glu	432
	130 135 140	
25	ctg cag ttc att ttg gac aag ttg ggt gac gat gaa gtg cgc agt gac Leu Gln Phe Ile Leu Asp Lys Leu Gly Asp Asp Glu Val Arg Ser Asp	480
	145 150 155 160	
30	ttg aaa caa gga tca aat gga gta ccg gta ctg aca gag gag gaa ctg Leu Lys Gln Gly Ser Asn Gly Val Pro Val Leu Thr Glu Glu Glu Leu	528
	165 170 175	
35	aca atg ctg gat gaa ttt tac aag cta gtt tac cct gaa agg gac atg Thr Met Leu Asp Glu Phe Tyr Lys Leu Val Tyr Pro Glu Arg Asp Met	576
	180 185 190	
40	aac atg agg ttg aat gag cag tat gag caa gca tct gtt cac ctg tgg Asn Met Arg Leu Asn Glu Gln Tyr Glu Gln Ala Ser Val His Leu Trp	624
	195 200 205	
45	gac tta ctg gaa ggg aag gaa aaa ccc gtt tgt gga aca acc tat aaa Asp Leu Leu Glu Gly Lys Glu Lys Pro Val Cys Gly Thr Thr Tyr Lys	672
	210 215 220	
50	gcc ctg aag gag gtt gtt gaa cgt att ctt caa act agt tac ttt gat Ala Leu Lys Glu Val Val Glu Arg Ile Leu Gln Thr Ser Tyr Phe Asp	720
	225 230 235 240	
55	agc acc cat aac cat cag aac ggg tta tgt gag gaa gaa gag gca gca	768

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	Ser	Thr	His	Asn	His	Gln	Asn	Gly	Leu	Cys	Glu	Glu	Glu	Glu	Ala	Ala	
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	ccc	aca	cct	gca	gta	gaa	gac	act	gta	gca	gaa	gct	gag	cct	gat	cca	816
	Pro	Thr	Pro	Ala	Val	Glu	Asp	Thr	Val	Ala	Glu	Ala	Glu	Pro	Asp	Pro	
					260					265					270		
10																	
	gca	gaa	gaa	ttt	act	gaa	cct	act	gaa	gtt	gaa	tcg	act	gag	tat	gta	864
	Ala	Glu	Glu	Phe	Thr	Glu	Pro	Thr	Glu	Val	Glu	Ser	Thr	Glu	Tyr	Val	
					275					280					285		
15																	
	aac	aga	caa	ttc	atg	gca	gag	act	cag	ttc	agc	agt	agt	gag	aag	gaa	912
	Asn	Arg	Gln	Phe	Met	Ala	Glu	Thr	Gln	Phe	Ser	Ser	Ser	Glu	Lys	Glu	
					290					295					300		
20																	
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	Gln	Val	Asp	Glu	Trp	Thr	Val	Glu	Thr	Val	Glu	Val	Val	Asn	Ser	Leu	
	305					310					315					320	
25																	
	cag	caa	caa	aca	caa	gct	aca	tct	cct	cca	gtt	cct	gaa	cct	cat	aca	1008
	Gln	Gln	Gln	Thr	Gln	Ala	Thr	Ser	Pro	Pro	Val	Pro	Glu	Pro	His	Thr	
					325						330				335		
30																	
	ctc	act	act	gtg	gct	caa	gca	gat	cct	ctt	gtt	aga	aga	cag	aga	gta	1056
	Leu	Thr	Thr	Val	Ala	Gln	Ala	Asp	Pro	Leu	Val	Arg	Arg	Gln	Arg	Val	
					340					345					350		
35																	
	cag	gac	ctt	atg	gcc	cag	atg	cag	ggc	cca	tat	aac	ttc	atg	cag	gac	1104
	Gln	Asp	Leu	Met	Ala	Gln	Met	Gln	Gly	Pro	Tyr	Asn	Phe	Met	Gln	Asp	
					355					360					365		
40																	
	tct	atg	ctg	gag	ttt	gag	aac	cag	aca	ctt	gat	cct	gcc	att	gta	tct	1152
	Ser	Met	Leu	Glu	Phe	Glu	Asn	Gln	Thr	Leu	Asp	Pro	Ala	Ile	Val	Ser	
					370					375					380		
45																	
	gca	cag	ccc	atg	aat	cca	gca	cag	aat	ttg	gac	atg	ccg	caa	atg	gtc	1200
	Ala	Gln	Pro	Met	Asn	Pro	Ala	Gln	Asn	Leu	Asp	Met	Pro	Gln	Met	Val	
	385					390					395				400		
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	tgc	cct	cca	gtt	cat	act	gag	tca	aga	ctt	gcc	cag	cct	aat	caa	gtt	1248
	Cys	Pro	Pro	Val	His	Thr	Glu	Ser	Arg	Leu	Ala	Gln	Pro	Asn	Gln	Val	
					405						410				415		
55																	

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5	cct gtg caa cca gaa gct acg cag gtt ccc ttg gtt tca tct aca agt Pro Val Gln Pro Glu Ala Thr Gln Val Pro Leu Val Ser Ser Thr Ser 420 425 430	1296
10	gag gga tat aca gcc tcc cag ccc atg tat cag cct tct cat acc aca Glu Gly Tyr Thr Ala Ser Gln Pro Met Tyr Gln Pro Ser His Thr Thr 435 440 445	1344
15	gag caa cgg cca cag aag gaa tcc att gac cag att cag gct tca atg Glu Gln Arg Pro Gln Lys Glu Ser Ile Asp Gln Ile Gln Ala Ser Met 450 455 460	1392
20	tca ctg aat gca gac cag acc ccg tca tca tca tca ctt ccc act gca Ser Leu Asn Ala Asp Gln Thr Pro Ser Ser Ser Ser Leu Pro Thr Ala 465 470 475 480	1440
25	tcc cag ccg caa gtt ttc caa gct gga tct agc aaa cct ttg cat agc Ser Gln Pro Gln Val Phe Gln Ala Gly Ser Ser Lys Pro Leu His Ser 485 490 495	1488
30	agc gga atc aat gtt aat gca gct cca ttc caa tcc atg caa aca gta Ser Gly Ile Asn Val Asn Ala Ala Pro Phe Gln Ser Met Gln Thr Val 500 505 510	1536
35	ttc aac atg aat gca cct gtt cct cct gtt aat gag cca gaa gcc ctt Phe Asn Met Asn Ala Pro Val Pro Pro Val Asn Glu Pro Glu Ala Leu 515 520 525	1584
40	aag caa caa aat cag tac cag gcc agt tac aac cag agt ttc tcc aat Lys Gln Gln Asn Gln Tyr Gln Ala Ser Tyr Asn Gln Ser Phe Ser Asn 530 535 540	1632
45	cag cca cac caa gta gaa caa tca gat ctt cag caa gaa cag ctc cag Gln Pro His Gln Val Glu Gln Ser Asp Leu Gln Gln Glu Gln Leu Gln 545 550 555 560	1680
50	aca gtg gtt ggt act tac cat ggt tct ccg gac cag acc cat caa gtg Thr Val Val Gly Thr Tyr His Gly Ser Pro Asp Gln Thr His Gln Val 565 570 575	1728
55	gca gga aac cac cag caa cct ccc cag cag aat act gga ttt cca cgc	1776

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Ala Gly Asn His Gln Gln Pro Pro Gln Gln Asn Thr Gly Phe Pro Arg
580 585 590

5

aac agt cag cct tat tac aac agt cgg gga gtg tct cgt ggt gga tca 1824
Asn Ser Gln Pro Tyr Tyr Asn Ser Arg Gly Val Ser Arg Gly Gly Ser
595 600 605

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cgt ggg act cgt gga ttg atg aat ggt tac agg gga cct gca aat gga 1872
Arg Gly Thr Arg Gly Leu Met Asn Gly Tyr Arg Gly Pro Ala Asn Gly
610 615 620

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ttt aga gga gga tat gat ggc tac cgt cct tca ttt tcc aac act ccg 1920
Phe Arg Gly Gly Tyr Asp Gly Tyr Arg Pro Ser Phe Ser Asn Thr Pro
625 630 635 640

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aac agt ggt tac acg cag ccc caa ttt aat gct cct cga gat tat tca 1968
Asn Ser Gly Tyr Thr Gln Pro Gln Phe Asn Ala Pro Arg Asp Tyr Ser
645 650 655

25

aac tac cag cgg gat gga tat cag cag aac ttc aaa cgt ggt tct gga 2016
Asn Tyr Gln Arg Asp Gly Tyr Gln Gln Asn Phe Lys Arg Gly Ser Gly
660 665 670

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caa agt ggg cct cgg gga gct cct cga ggt cgt gga ggg ccc cca aga 2064
Gln Ser Gly Pro Arg Gly Ala Pro Arg Gly Arg Gly Gly Pro Pro Arg
675 680 685

35

cca aac aga ggg atg cct caa atg aac gct cag caa gtg aat taa 2109
Pro Asn Arg Gly Met Pro Gln Met Asn Ala Gln Gln Val Asn
690 695 700

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<210> 30
<211> 702
<212> PRT
<213> Gallus gallus

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<400> 30

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Met Pro Ser Ala Thr Asn Gly Thr Met Ala Ser Ser Ser Gly Lys Ala
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10	Gln Ala Ser Gly Gly Ser Ile Thr Ser Val Gln Thr Glu Ala Met Lys	35	40	45
15	Gln Ile Leu Gly Val Ile Asp Lys Lys Leu Arg Asn Leu Glu Lys Lys	50	55	60
20	Lys Ser Lys Leu Asp Asp Tyr Gln Glu Arg Met Asn Lys Gly Glu Arg	65	70	75
25	Leu Asn Gln Asp Gln Leu Asp Ala Val Ser Lys Tyr Gln Glu Val Thr	85	90	95
30	Asn Asn Leu Glu Phe Ala Lys Glu Leu Gln Arg Ser Phe Met Ala Leu	100	105	110
35	Ser Gln Asp Ile Gln Lys Thr Ile Lys Lys Thr Ala Arg Arg Glu Gln	115	120	125
40	Leu Met Arg Glu Glu Ala Glu Gln Lys Arg Leu Lys Thr Val Leu Glu	130	135	140
45	Leu Gln Phe Ile Leu Asp Lys Leu Gly Asp Asp Glu Val Arg Ser Asp	145	150	155
50	Leu Lys Gln Gly Ser Asn Gly Val Pro Val Leu Thr Glu Glu Glu Leu	165	170	175
55	Thr Met Leu Asp Glu Phe Tyr Lys Leu Val Tyr Pro Glu Arg Asp Met			

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	180		185		190
5	Asn Met Arg Leu Asn Glu Gln Tyr Glu Gln Ala Ser Val His Leu Trp				
	195		200		205
10	Asp Leu Leu Glu Gly Lys Glu Lys Pro Val Cys Gly Thr Thr Tyr Lys				
	210		215		220
15	Ala Leu Lys Glu Val Val Glu Arg Ile Leu Gln Thr Ser Tyr Phe Asp				
	225		230		235 240
20	Ser Thr His Asn His Gln Asn Gly Leu Cys Glu Glu Glu Glu Ala Ala				
		245		250	255
25	Pro Thr Pro Ala Val Glu Asp Thr Val Ala Glu Ala Glu Pro Asp Pro				
	260		265		270
30	Ala Glu Glu Phe Thr Glu Pro Thr Glu Val Glu Ser Thr Glu Tyr Val				
	275		280		285
35	Asn Arg Gln Phe Met Ala Glu Thr Gln Phe Ser Ser Ser Glu Lys Glu				
	290		295		300
40	Gln Val Asp Glu Trp Thr Val Glu Thr Val Glu Val Val Asn Ser Leu				
	305		310		315 320
45	Gln Gln Gln Thr Gln Ala Thr Ser Pro Pro Val Pro Glu Pro His Thr				
		325		330	335
50	Leu Thr Thr Val Ala Gln Ala Asp Pro Leu Val Arg Arg Gln Arg Val				
	340		345		350
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5	Gln Asp Leu Met Ala Gln Met Gln Gly Pro Tyr Asn Phe Met Gln Asp	355	360	365
10	Ser Met Leu Glu Phe Glu Asn Gln Thr Leu Asp Pro Ala Ile Val Ser	370	375	380
15	Ala Gln Pro Met Asn Pro Ala Gln Asn Leu Asp Met Pro Gln Met Val	385	390	395
20	Cys Pro Pro Val His Thr Glu Ser Arg Leu Ala Gln Pro Asn Gln Val	405	410	415
25	Pro Val Gln Pro Glu Ala Thr Gln Val Pro Leu Val Ser Ser Thr Ser	420	425	430
30	Glu Gly Tyr Thr Ala Ser Gln Pro Met Tyr Gln Pro Ser His Thr Thr	435	440	445
35	Glu Gln Arg Pro Gln Lys Glu Ser Ile Asp Gln Ile Gln Ala Ser Met	450	455	460
40	Ser Leu Asn Ala Asp Gln Thr Pro Ser Ser Ser Ser Leu Pro Thr Ala	465	470	475
45	Ser Gln Pro Gln Val Phe Gln Ala Gly Ser Ser Lys Pro Leu His Ser	485	490	495
50	Ser Gly Ile Asn Val Asn Ala Ala Pro Phe Gln Ser Met Gln Thr Val	500	505	510
55	Phe Asn Met Asn Ala Pro Val Pro Pro Val Asn Glu Pro Glu Ala Leu			

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515 520 525

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Lys Gln Gln Asn Gln Tyr Gln Ala Ser Tyr Asn Gln Ser Phe Ser Asn
530 535 540

10

Gln Pro His Gln Val Glu Gln Ser Asp Leu Gln Gln Glu Gln Leu Gln
545 550 555 560

15

Thr Val Val Gly Thr Tyr His Gly Ser Pro Asp Gln Thr His Gln Val
565 570 575

20

Ala Gly Asn His Gln Gln Pro Pro Gln Gln Asn Thr Gly Phe Pro Arg
580 585 590

25

Asn Ser Gln Pro Tyr Tyr Asn Ser Arg Gly Val Ser Arg Gly Gly Ser
595 600 605

30

Arg Gly Thr Arg Gly Leu Met Asn Gly Tyr Arg Gly Pro Ala Asn Gly
610 615 620

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Phe Arg Gly Gly Tyr Asp Gly Tyr Arg Pro Ser Phe Ser Asn Thr Pro
625 630 635 640

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Asn Ser Gly Tyr Thr Gln Pro Gln Phe Asn Ala Pro Arg Asp Tyr Ser
645 650 655

45

Asn Tyr Gln Arg Asp Gly Tyr Gln Gln Asn Phe Lys Arg Gly Ser Gly
660 665 670

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Gln Ser Gly Pro Arg Gly Ala Pro Arg Gly Arg Gly Gly Pro Pro Arg
675 680 685

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Pro Asn Arg Gly Met Pro Gln Met Asn Ala Gln Gln Val Asn
690 695 700

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aaggtttgaa tggagtgc

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	<212> DNA	
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	<223> GAPDH primer	
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	<400> 36	
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	<211> 27	
	<212> DNA	
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	<220>	
55	<223> primer	

5	<p><400> 37 catatggcat taagtcaaga tattcag</p>	27
10	<p><210> 38 <211> 23 <212> DNA <213> Artificial</p>	
15	<p><220> <223> primer</p>	
20	<p><400> 38 ggtacctttg cggcatccct ctg</p>	23
25	<p><210> 39 <211> 21 <212> DNA <213> Artificial</p>	
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35	<p><400> 39 catatgccgt cggccaccag c</p>	21
40	<p><210> 40 <211> 22 <212> DNA <213> Artificial</p>	
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55	<p><210> 41</p>	

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 <213> Artificial

 10 <220>
 <223> primer

 <400> 41
 gagctcatgc cctcgccac cag 23

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 20 <213> Artificial

 <220>
 <223> primer
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 <400> 42
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 35 <213> Homo sapiens

 <400> 43

 40 Arg Asn Leu Glu Lys Lys Lys Gly Lys Leu Asp Asp Tyr Gln
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 45 <210> 44
 <211> 148
 <212> PRT
 50 <213> Mus musculus

 <400> 44

 55 Met Glu Trp Ser Gly Val Phe Ile Phe Leu Leu Ser Gly Thr Ala Gly

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	20	25	30
10	Pro Gly Ala Ser Val Lys Ile Ser Cys Lys Ala Ser Gly Tyr Thr Phe		
	35	40	45
15	Thr Asp Tyr Asn Met Asp Trp Val Lys Gln Ser His Gly Lys Ser Leu		
	50	55	60
20	Glu Trp Ile Gly Asp Ile Asn Pro Asn Tyr Asp Ser Thr Ser Tyr Asn		
	65	70	75
25	Gln Lys Phe Lys Gly Lys Ala Thr Leu Thr Val Asp Lys Ser Ser Ser		
	85	90	95
30	Thr Ala Tyr Met Glu Leu Arg Ser Leu Thr Ser Glu Asp Thr Ala Val		
	100	105	110
35	Tyr Tyr Cys Ala Arg Ser Arg Ser Tyr Asp Tyr Glu Gly Phe Ala Tyr		
	115	120	125
40	Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ala Ala Lys Thr Thr Pro		
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45	Pro Ser Val Tyr		
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<213> Mus musculus

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Ala Val Leu Arg Cys Ser Arg Gly Leu Leu Val Ile Trp Ile Ser Asp
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Ile Gln Leu Thr Gln Ser Pro Ser Ser Leu Ala Val Thr Ala Gly Glu
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Lys Val Thr Met Ser Cys Lys Ser Ser Gln Ser Leu Leu Trp Ser Val
35 40 45

20

Asn Gln Lys Asn Tyr Leu Ser Trp Tyr Gln Gln Lys Gln Arg Gln Pro
50 55 60

25

Pro Lys Leu Leu Ile Tyr Gly Ala Ser Ile Arg Glu Ser Trp Val Pro
65 70 75 80

30

Asp Arg Phe Thr Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile
85 90 95

35

Ser Asn Val His Ala Glu Asp Leu Ala Val Tyr Tyr Cys Gln His Asn
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His Gly Ser Phe Leu Pro Ser Arg Ser Glu Gln Val Pro Ser Trp Arg
115 120 125

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Ser Asn Asn Arg
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<210> 46

<211> 117

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<212> PRT

<213> Mus musculus

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<400> 46

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Arg Thr Thr Ser His Met Asp Ser Asp Ile Gln Leu Thr Gln Ser Pro
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Ala Ser Leu Ser Ala Ser Val Gly Glu Thr Val Thr Ile Thr Cys Arg
20 25 30

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Ala Ser Gly Asn Ile His Asn Tyr Leu Ala Trp Tyr Gln Gln Lys Gln
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Claims

1. A method for detecting a cancer, comprising measuring the expression of a polypeptide having a reactivity of binding via an antigen-antibody reaction to an antibody against a CAPRIN-1 protein having any one of the amino acid sequences shown in the even-numbered SEQ ID NOS: 2-30 in the Sequence Listing, in a sample separated from a living organism.
2. The method according to claim 1, wherein the polypeptide to be measured is a CAPRIN-1 protein having any one of the amino acid sequences shown in the even-numbered SEQ ID NOS: 2-30 or a polypeptide having 85% or more sequence identity with the CAPRIN-1 protein.
3. The method according to claim 1 or 2, wherein the living organism is a human, a dog, or a cat.
4. The method according to claim 3, wherein the living organism is a dog and the polypeptide to be measured has an amino acid sequence shown in any one of the even-numbered SEQ ID NOS: 2-30.
5. The method according to claim 4, wherein the living organism is a dog and the polypeptide to be measured has the amino acid sequence shown in SEQ ID NO: 6, 8, 10, 12, or 14.
6. The method according to claim 3, wherein the living organism is a human and the polypeptide to be measured has the amino acid sequence shown in SEQ ID NO: 2 or 4.
7. The method according to any one of claims 1 to 6, wherein the expression of the polypeptide is measured by immunoassay of an antibody that can be contained in the sample and is induced *in vivo* against the polypeptide to be measured.
8. The method according to any one of claims 1 to 7, wherein the sample is serum, blood plasma, ascite, or pleural effusion.
9. The method according to any one of claims 1 to 6, wherein the expression of the polypeptide is measured by measuring mRNA encoding the polypeptide, which is contained in the sample.
10. The method according to claim 9, comprising examining the existing amount of the mRNA in the sample using a

polynucleotide that specifically hybridizes to a partial sequence of 15 or more nucleotides in the nucleotide sequence of the mRNA.

11. The method according to claim 10, wherein the living organism is a dog and the polynucleotide is a polynucleotide specifically hybridizing to a partial sequence of 15 or more nucleotides in the nucleotide sequence shown in SEQ ID NO: 5, 7, 9, 11, or 13.

12. The method according to claim 10, wherein the living organism is a human and the polynucleotide is a polynucleotide specifically hybridizing to a partial sequence of 15 or more nucleotides in the nucleotide sequence shown in SEQ ID NO: 1 or 3.

13. The method according to any one of claims 9 to 12, wherein the sample is a tissue or a cell.

14. The method according to any one of claims 1 to 13, wherein the cancer is at least one type of cancer selected from the group consisting of brain tumor, squamous cell carcinoma of the head, neck, lung, uterus or esophagus, melanoma, adenocarcinoma of the lung or uterus, renal cancer, malignant mixed tumor, hepatocellular carcinoma, basal cell carcinoma, acanthoma-like gingival tumor, tumor of the oral cavity, perianal adenocarcinoma, anal sac tumor, anal sac apocrine adenocarcinoma, sertoli cell carcinoma, cancer of vaginal vestibule, sebaceous adenocarcinoma, sebaceous epithelioma, sebaceous adenoma, sweat gland carcinoma, intranasal adenocarcinoma, nasal adenocarcinoma, thyroid cancer, large-bowel cancer, bronchial adenocarcinoma, adenocarcinoma, ductal carcinoma, breast adenocarcinoma, composite type breast adenocarcinoma, malignant mammary mixed tumor, intraductal papillary adenocarcinoma, fibrosarcoma, hemangiopericytoma, osteosarcoma, chondrosarcoma, soft tissue sarcoma, histiocytic sarcoma, myxosarcoma, undifferentiated sarcoma, lung cancer, mastocytoma, cutaneous leiomyoma, intraperitoneal leiomyoma, leiomyoma, chronic lymphocytic leukemia, lymphoma, gastrointestinal lymphoma, digestive lymphoma, small-cell-to-medium-cell lymphoma, adrenomedullary tumor, granulosa cell tumor, and pheochromocytoma.

15. The method according to any one of claims 1 to 14, comprising further detecting the malignancy of a cancer based on the fact that the malignancy of cancer is high when the expression level of the polypeptide is higher than that of a control.

16. The method according to any one of claims 1 to 15, comprising further detecting the progression of cancer on the basis of the indicator that the extent of cancer is advanced when the expression level of the polypeptide is higher than that of a control.

17. A reagent for detecting a cancer, comprising a polypeptide that has a reactivity of binding via an antigen-antibody reaction to an antibody that is induced *in vivo* against a CAPRIN-1 protein having any one of the amino acid sequences shown in the even-numbered SEQ ID NOS: 2-30 in the Sequence Listing.

18. A reagent for detecting a cancer, comprising an antibody or antigen-binding fragment thereof that undergoes an antigen-antibody reaction with a polypeptide, wherein the polypeptide has a reactivity of binding via an antigen-antibody reaction to an antibody against a CAPRIN-1 protein having any one of the amino acid sequences shown in the even-numbered SEQ ID NOS: 2-30 in the Sequence Listing and is produced *in vivo*.

19. The reagent for detecting cancer according to claim 18, wherein the antibody or antigen-binding fragment thereof that undergoes an antigen-antibody reaction with the polypeptide is an antibody or antigen-binding fragment thereof that binds to the surface of a cancer cell.

20. The reagent for detecting cancer according to claim 18 or 19, wherein the antibody or antigen-binding fragment thereof that undergoes an antigen-antibody reaction with the polypeptide has an immunological reactivity with:

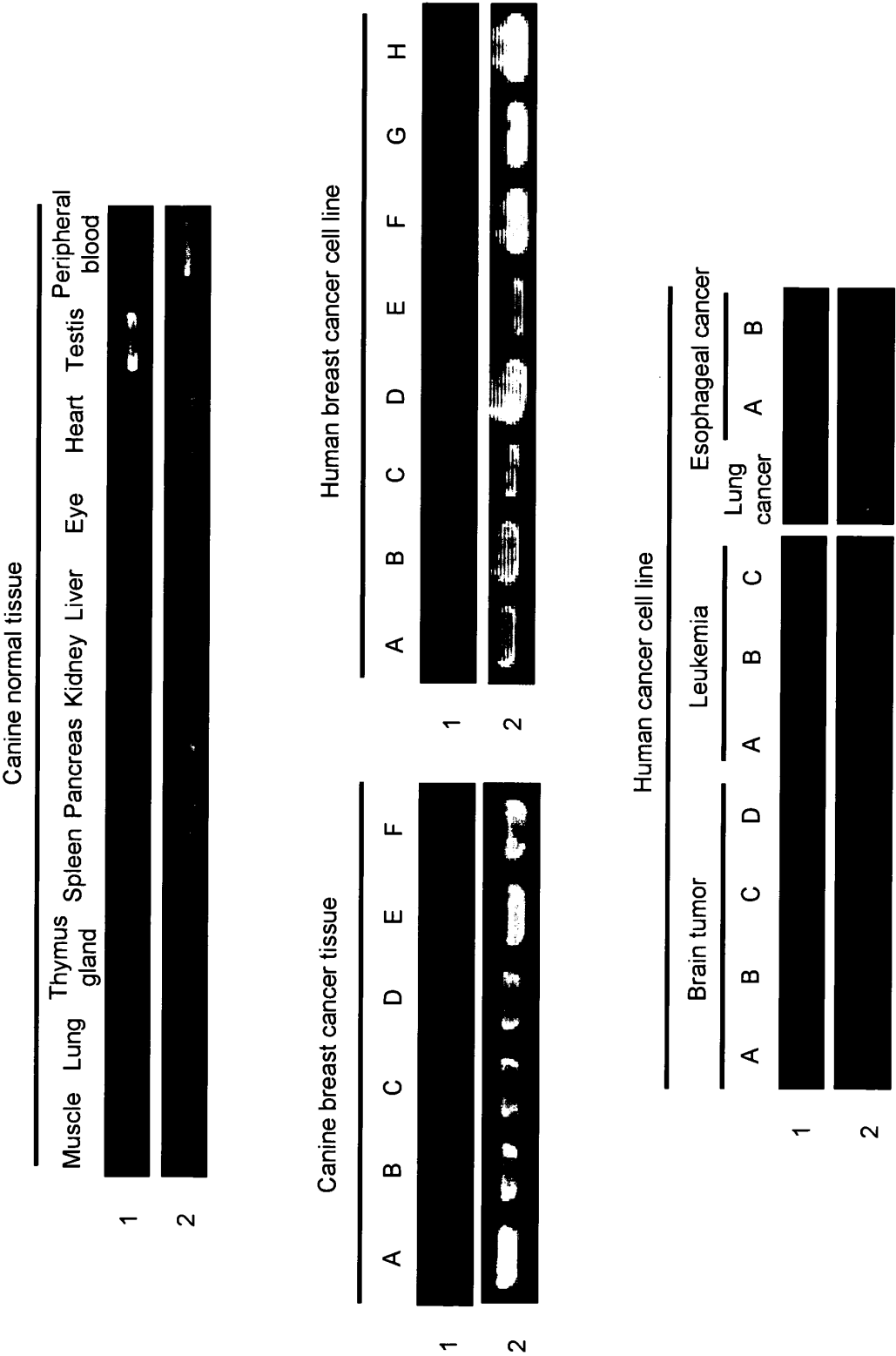
a polypeptide comprising an amino acid sequence of 7 or more continuous amino acid residues within the region of amino acid residue Nos. 50-98 or amino acid residue Nos. 233-305 in any one of the amino acid sequences shown in the even-numbered SEQ IDS NO: 2-30 excluding SEQ ID NO: 6 and SEQ ID NO: 18 or a polypeptide comprising the polypeptide as a partial sequence.

21. The reagent for detecting a cancer according to any one of claims 18 to 20, wherein the antibody or antigen-binding fragment thereof that undergoes an antigen-antibody reaction with the polypeptide is an antibody or antigen-binding

fragment thereof which binds to SEQ ID NO: 43, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 44 and 45, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 44 and 46, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 44 and 47, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 44 and 48, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 49 and 50, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 51 and 52, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 53 and 54, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 55 and 56, a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 57 and 58, or a monoclonal antibody or antigen-binding fragment thereof having the amino acid sequences of SEQ ID NOS: 59 and 60.

- 22.** A reagent for detecting a cancer, comprising a polynucleotide that specifically hybridizes to a partial sequence of 15 or more nucleotides in any one of the nucleotide sequences shown in the odd-numbered SEQ ID NOS: 1-29 in the Sequence Listing.

Fig. 1



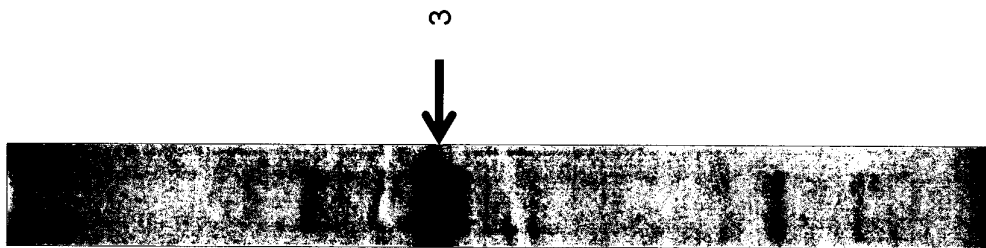


Fig. 2

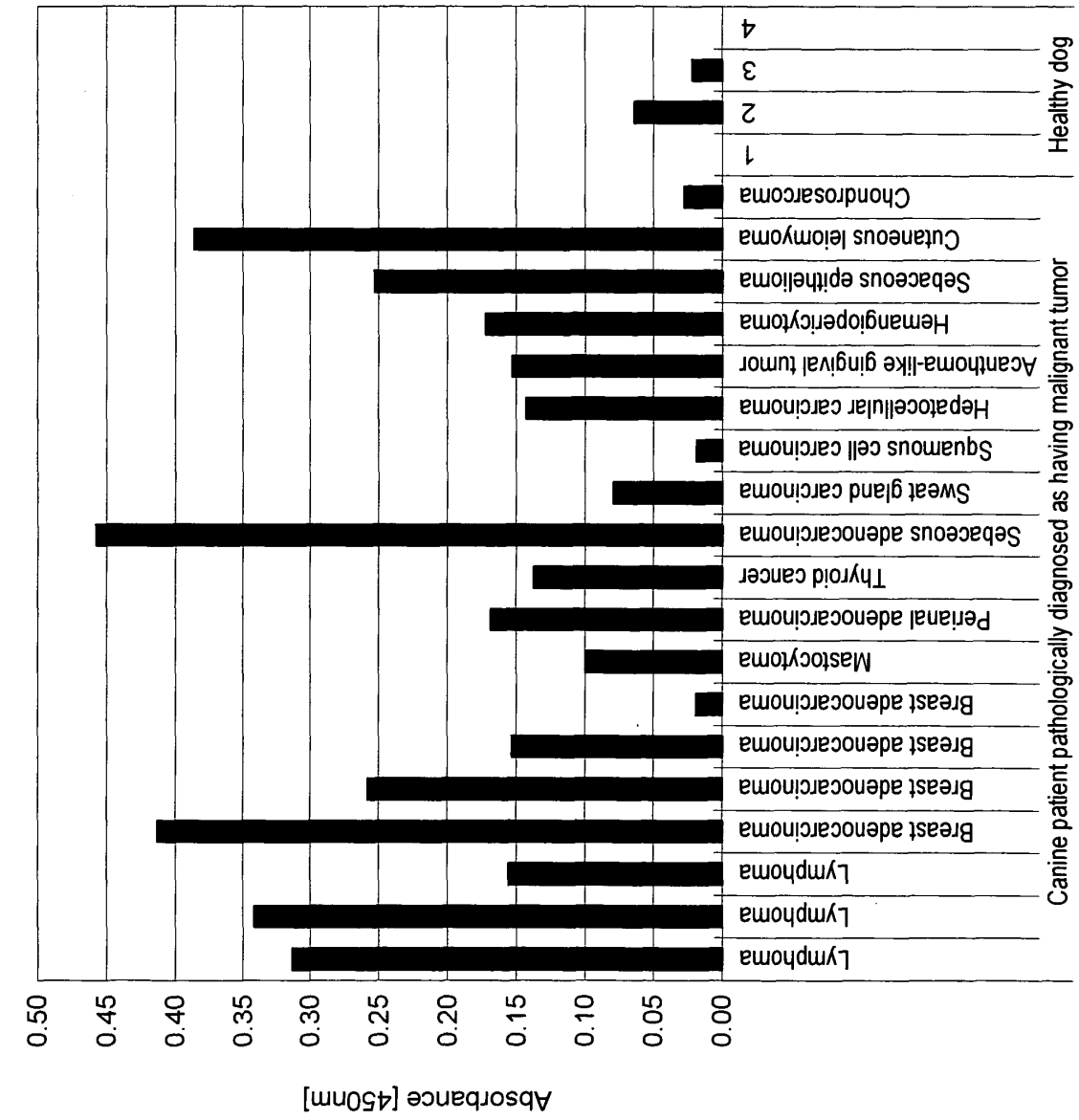
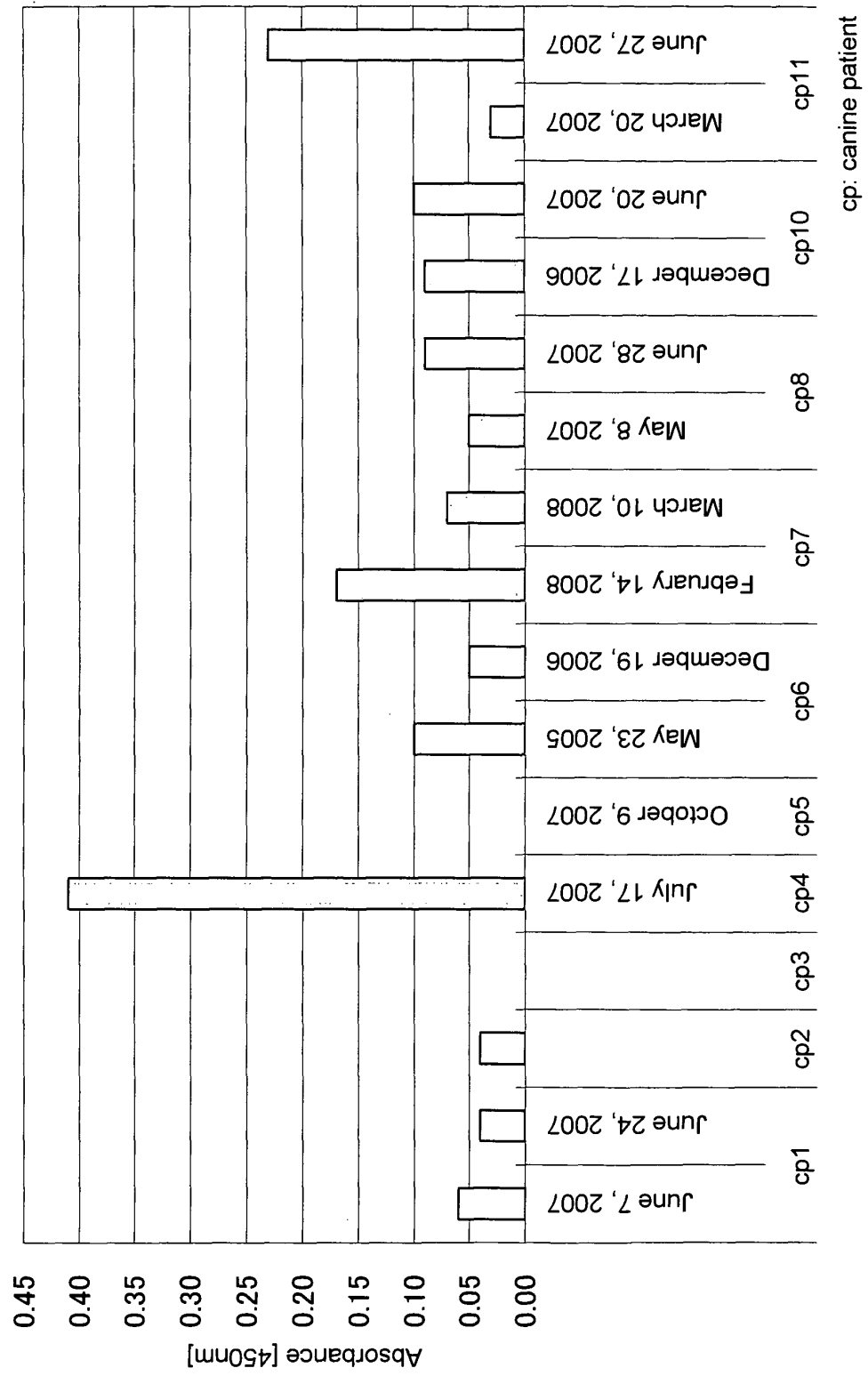


Fig. 4



INTERNATIONAL SEARCH REPORT

International application No.

PCT/JP2009/063883

A. CLASSIFICATION OF SUBJECT MATTER

G01N33/574(2006.01)i, C12N15/09(2006.01)i, C12Q1/68(2006.01)i, G01N33/53(2006.01)i

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

G01N33/574, C12N15/09, C12Q1/68, G01N33/53

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Jitsuyo Shinan Koho	1922-1996	Jitsuyo Shinan Toroku Koho	1996-2009
Kokai Jitsuyo Shinan Koho	1971-2009	Toroku Jitsuyo Shinan Koho	1994-2009

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

JSTPlus/JMEDPlus/JST7580(JDreamII), CA/MEDLINE/BIOSIS(STN)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	JP 2002-540790 A (Corixa Corp.), 03 December, 2002 (03.12.02), Abstract; Par. Nos. [0051], [0106], [0122], [0123], [0125] (example 3) sequence Nos.84, 93 & US 6444425 B1 & US 2003/0118599 A1 & US 2004/0225900 A1 & EP 1187915 A & WO 2000/060077 A2 & AU 4185100 A	1, 3, 7-20, 22
A	JP 2003-528587 A (HUMAN GENOME SCIENCES, INC.), 30 September, 2003 (30.09.03), Sequence No.159 & EP 1224285 A & WO 2001/032910 A2 & AU 1438001 A & CA 2388822 A	1-22

☒ Further documents are listed in the continuation of Box C.☐ See patent family annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

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"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search
28 August, 2009 (28.08.09)Date of mailing of the international search report
08 September, 2009 (08.09.09)Name and mailing address of the ISA/
Japanese Patent Office

Authorized officer

Facsimile No.

Telephone No.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/JP2009/063883

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 2008/088583 A2 (Immunotope Inc.), 24 July, 2008 (24.07.08), SEQ ID NO:168 & US 2008/0107668 A1 & EP 2061503 A & CA 2661651 A	1-22
A	Katsafanas, George C.; Moss, Bernard, Colocalization of transcription and translation within cytoplasmic poxvirus factories coordinates viral expression and subjugates host functions, Cell Host & Microbe, 2007, Vol.2, No.4, Page.221-228	1-22
A	Solomon, Samuel; Xu, Yaoxian; Wang, Bin; David, Muriel D.; Schubert, Peter; Kennedy, Derek; Schrader, John W., Distinct structural features of caprin-1 mediate its interaction with G3BP-1 and its induction of phosphorylation of eukaryotic translation initiation factor 2 alpha, entry to cytoplasmic stress granules, and selective interaction with a subset of mRNAs, Molecular and Cellular Biology, 2007, Vol.27, No.6, Page.2324-2342	1-22

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REFERENCES CITED IN THE DESCRIPTION

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专利名称(译)	癌症检测方法		
公开(公告)号	EP2325648A1	公开(公告)日	2011-05-25
申请号	EP2009805010	申请日	2009-08-05
[标]申请(专利权)人(译)	东丽株式会社		
申请(专利权)人(译)	TORAY INDUSTRIES , INC.		
当前申请(专利权)人(译)	TORAY INDUSTRIES , INC.		
[标]发明人	OKANO FUMIYOSHI SUZUKI KANA		
发明人	OKANO, FUMIYOSHI SUZUKI, KANA		
IPC分类号	G01N33/574 C12N15/09 C12Q1/68 G01N33/53		
CPC分类号	C12Q1/6886 C12Q2600/112 G01N33/57407 G01N33/57415 G01N33/6893 G01N33/53 G01N33/574		
优先权	2008202320 2008-08-05 JP		
其他公开文献	EP2325648B1 EP2325648A4		
外部链接	Espacenet		

摘要(译)

本发明涉及一种检测癌症的方法，包括测量具有与抗CAPRIN-1蛋白抗体结合的反应性的多肽的表达，所述CAPRIN-1蛋白具有偶数SEQ ID NO 中任一所示的氨基酸序列：序列表中的2-30通过在与生物体分离的样品中的抗原 - 抗体反应，以及用于检测包含CAPRIN-1蛋白或其片段的癌症的试剂，针对CAPRIN-1蛋白的抗体或其片段，或编码CAPRIN-1蛋白或其片段的多核苷酸。

Fig. 3

