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(54) **METHOD FOR MEASURING CYSTATIN C IN HUMAN BODY FLUID**

VERFAHREN ZUR MESSUNG VON CYSTATIN C IN MENSCHLICHEN KÖRPERFLÜSSIGKEITEN  
 PROCÉDÉ POUR MESURER LA CYSTATINE C DANS UN FLUIDE CORPOREL HUMAIN

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**Description**

## TECHNICAL FIELD

5 **[0001]** The present invention relates to a particle-enhanced immunoassay method for cystatin C in a human body fluid, and a particle-enhanced immunoassay reagent.

## BACKGROUND ART

10 **[0002]** Cystatin C is a basic low molecular weight protein (isoelectric point pH 9.3) having a molecular weight of 13 kDa, and is continually produced in all of nucleated cells in the human body, while a certain amount is secreted extracellularly without being affected by environmental changes. Therefore, the concentration of cystatin C in blood becomes constant, and is not subject to the influence such as inflammation due to other diseases, or to the influence of age, gender, exercise, meal or the like. Furthermore, cystatin C does not form composites with other plasma proteins, but is  
15 filtered at the renal glomerulus and reabsorbed in the proximal renal tubule. Therefore, it is known that if the glomerular filtration rate (GFR) decreases, the blood concentration of cystatin C increases, and accordingly, cystatin C is attracting attention as an indicator of the GFR instead of creatinine. Furthermore, since cystatin C serves as a diagnostic indicator of early renal failure in test for renal function, cystatin C is widely useful also in the areas of ambulatory care, medical examination and the like.

20 **[0003]** In regard to the method for measuring cystatin C, there have been reported automated particle-enhanced immunoassay methods for human cystatin C (Non-Patent Documents 1 to 3, and Patent Document 1). For example, in the method described in Non-Patent Document 1, use is made of an anti-cystatin C polyclonal antibody having a strong agglutinating power, which is in the form of being bound to carrier particles at a weight ratio of 5%. However, as shown  
25 in Non-Patent Document 4, cystatin C belongs to the cystatin family in which the members are similar in structure, and cystatin C is differently localized in various types of human body fluids. Accordingly, there is a very high possibility that an accurate measurement value for cystatin C may not be obtained with a polyclonal antibody which has low specificity, depending on the type of the human body fluid (for example, saliva or lacrimal fluid). Meanwhile, Patent Document 1 describes information to the effect that generally an immunoagglutinate is more easily formed when a polyclonal antibody is used (page 20, lines 21 to 27). Furthermore, it is suggested to use an anti-cystatin C monoclonal antibody having high  
30 specificity. However, it is also described to the effect that in the case the target is a monomer and low molecular weight proteins such as cystatin C, better results are obtained by using a cocktail of many different monoclonal antibodies (page 21, lines 5 to 10), which is essentially equivalent to the use of a polyclonal antibody. Moreover, in order to improve the performance, it has been necessary to bind a markedly large amount of antibodies such that the binding amount of antibodies to the total weight of antibody-coated particles is greater than 5% by weight and equal to or less than 35%  
35 by weight.

## Prior art Documents

## PATENT DOCUMENTS

40 **[0004]** Patent Document 1: WO 2007/102054 Pamphlet

## NON-PATENT DOCUMENTS

45 **[0005]**

Non-Patent Document 1: CLINICAL CHEMISTRY, Vol. 40, No. 10 (1994), 1921-1926

Non-Patent Document 2: KIDNEY INTERNATIONAL, Vol. 47 (1995), 312-318

Non-Patent Document 3: CLINICAL CHEMISTRY, Vol. 43, No. 6 (1997), 1016-1022

50 Non-Patent Document 4: METHODS IN ENZYMOLOGY, Vol. 244 (1994), 685-700

## DISCLOSURE OF INVENTION

## PROBLEM TO BE SOLVED BY THE INVENTION

55 **[0006]** There has been a problem that polyclonal antibodies that are generally used in the particle-enhanced immunoassay methods for human cystatin C have low specificity, and an assay that is specific to cystatin C cannot be carried out depending on the type of sample. As a measure against the problem, it has been designed to use monoclonal

antibodies; however, since their agglutinating power is weak, it is necessary to bind a large amount of cocktail composed of multiple types of monoclonal antibodies to insoluble carrier particles, posing a new problem that enormous efforts and costs are required. Therefore, the particle-enhanced immunoassay methods for cystatin C in a human body fluid which involve monoclonal antibodies only, have not been hitherto put to practical use. In addition, the conventional particle-enhanced immunoassay methods for human cystatin C require large amounts of antibodies such that the binding amount relative to the total weight of antibody-coated particles is greater than 5% by weight and equal to or less than 35% by weight, in order to obtain practical performance.

[0007] Therefore, it is an object of the present invention to provide a particle-enhanced immunoassay method for cystatin C in a human body fluid, which has high specificity and can be easily automated at low cost.

#### MEANS FOR SOLVING PROBLEM

[0008] The inventors of the present invention conducted a thorough investigation in order to solve the problems described above, and as a result, they found that even for monoclonal antibodies that have been conventionally regarded inappropriate for cystatin C assays, when antibody-coated particles are combined in which an anti-human cystatin C monoclonal antibody having high affinity and an anti-human cystatin C monoclonal antibody recognizing an epitope different from that of the aforementioned monoclonal antibody and having relatively low affinity are each independently bound to the antibody-coated particles at a binding amount ratio of less than 5% by weight relative to the total weight of the antibody-coated particles, a cystatin C-specific particle-enhanced immunoassay can be achieved. Thus, the inventors completed the present invention.

[0009] That is, the present invention is to provide a method for measuring cystatin C in a human body fluid, the method being a particle-enhanced immunoassay method comprising a combination of antibody-coated particles in which some insoluble carrier particles are coated with anti-human cystatin C monoclonal antibodies of one type which have high affinity, and in which some other insoluble carrier particles are coated with anti-human cystatin C monoclonal antibodies of another type which recognize an epitope different from that of the monoclonal antibodies of the first-mentioned type and have relatively low affinity, wherein the binding amount of anti-human cystatin C monoclonal antibodies of either type relative to the total weight of the particles being coated with the corresponding antibodies is less than 5% by weight.

[0010] Furthermore, the present invention is to provide a particle-enhanced immunoassay reagent specific to cystatin C in a human body fluid, the reagent comprising being a combination of antibody-coated particles in which some insoluble carrier particles are coated with anti-human cystatin C monoclonal antibodies of one type having high affinity and, in which some other insoluble carrier particles are coated with anti-human cystatin C monoclonal antibodies of another type which recognize an epitope different from that of the monoclonal antibodies of the first-mentioned type and have relatively low affinity, wherein the binding amount of anti-human cystatin C monoclonal antibodies of either type relative to the total weight of the particles being coated with the corresponding antibodies is less than 5% by weight.

#### EFFECT OF THE INVENTION

[0011] According to the present invention, it has been made possible to provide a particle-enhanced immunoassay and reagent for cystatin C in a human body fluid, which have higher specificity than the conventional methods and reagents, and can be easily applied to automatic analyzers at low cost.

#### BRIEF DESCRIPTION OF DRAWINGS

##### [0012]

FIG. 1 is a cystatin C concentration-sensitivity curve showing the sensitivity that accompanies particle-enhanced immunoagglutination when antibody-coated latex particles in which anti-human cystatin C monoclonal antibodies a, b and d were bound respectively to latex particles having particle sizes of 0.13  $\mu\text{m}$  and 0.30  $\mu\text{m}$ , were combined in the manner of a-b, a-d or b-d and with the same particle size, and cystatin C was measured;

FIG. 2 is a graph showing the specificity of an anti-human cystatin C monoclonal antibody identified by ELISA, and of an anti-human cystatin C polyclonal antibody as a reference;

FIG. 3 is a correlation diagram (sample number N = 50) for cystatin C assays between the invented reagent and the reference reagent; and

FIG. 4 is a graph showing the detection limit concentrations of the invented reagent and the reference reagent.

#### BEST MODE(S) FOR CARRYING OUT THE INVENTION

[0013] Two or more kinds of anti-human cystatin C monoclonal antibodies, which specifically react with human cystatin

C, are used in the present invention, and they shall include at least one from each kind of the following monoclonal antibodies: one has high affinity, and the other recognizes a different epitope and has relatively low affinity. The aforementioned antibodies also include an antibody fragment containing a functional site (that is, an antibody fragment containing Fab which is a human cystatin C recognition site) or a recombinant type antibody (that is, a recombinant antibody having a structure recombined at sites other than Fab).

**[0014]** There are no particular limitations on the method of obtaining the monoclonal antibody having high affinity as described above, but effective methods for obtaining an antibody having high affinity include selection of an immunization site or a hybridoma, an increase in the quantity of human cystatin C for immunization, prolongation of the immunization period, and the like. Here, the phrase "specifically react with human cystatin C" implies that the antibody causes an antigen-antibody reaction with human cystatin C, but substantially does not cause any antigen-antibody reaction with other cystatin family members (for example, human cystatin A, cystatin B, cystatin D, cystatin E/M, cystatin F, cystatin S, cystatin SA, and kininogen).

**[0015]** Among the anti-human cystatin C monoclonal antibodies used in the present invention, the anti-human cystatin C monoclonal antibody having high affinity has a dissociation constant (kd value), which is calculated by titer determination using, for example, an apparatus of "trade name: BIACORE (registered trademark), manufactured by GE Healthcare Corp. ", of less than 1 nM, and preferably 0.5 nM or less. In an analysis made by the BIACORE apparatus, the dissociation constant of a monoclonal antibody having ordinary affinity is 1 nM or greater. There are no particular limitations on the method of examining the dissociation constant as long as the method is a conventional method that is known in the pertinent art, but there is a possibility that the dissociation constant may slightly fluctuate depending on the principle of assay or assay conditions.

**[0016]** Examples of such a high-affinity monoclonal antibody include a monoclonal antibody produced by Hybridoma 75202 (FERM BP-11186), which will be described in the following Examples.

**[0017]** According to the present invention, if at least one of the high-affinity monoclonal antibody described above is used, the other anti-human cystatin C monoclonal antibody having relatively low affinity may have ordinary affinity. Among such combinations, a combination in which the ratio obtained by dividing the dissociation constant of the anti-human cystatin C monoclonal antibody having relatively low affinity by the dissociation constant of the other high-affinity monoclonal antibody is 2 or greater, is preferred. Such monoclonal antibodies can be produced according to conventional methods. In addition, when there is a difference of two or more times in the affinity (relative ratio of dissociation constants), a combination of high-affinity monoclonal antibodies each having a dissociation constant of less than 1 nM, can also be used.

**[0018]** The two or more kinds of monoclonal antibodies used in the present invention constitute a combination of monoclonal antibodies, in which the respective monoclonal antibodies recognize different sites of human cystatin C. The fact that the recognition sites are different can be identified by, for example, a sandwich ELISA.

**[0019]** In regard to the particle-enhanced immunoassay method and reagent of the present invention, there are no particular limitations on the material that is used as the insoluble carrier particles as long as the material can be used as a component for examination reagents, but specific examples include latexes, metal colloids, silica and carbon. The size of the insoluble carrier particles can be appropriately selected from 0.05  $\mu\text{m}$  to 0.5  $\mu\text{m}$  in accordance with the particle-enhanced immunoassay method of the present invention and the detection principle employed for the reagent. An average particle size used for the optical assays employed for an automatic analyzer is generally 0.1  $\mu\text{m}$  to 0.4  $\mu\text{m}$ , and preferably 0.1 to 0.2  $\mu\text{m}$ . The average particle size of the insoluble carrier particles can be examined using a particle size distribution analyzer, an electron microscope or the like.

**[0020]** The combination of monoclonal antibodies of the present invention enables a human cystatin C assay even when the binding amount of each of the antibodies relative to the total weight of the antibody-coated particles is less than 5% by weight, and the amount of antibodies is preferably equal to or greater than 1% by weight and less than 4% by weight.

**[0021]** Coating of monoclonal antibodies on insoluble carrier particles (sensitization) can be carried out by, for example, a conventional method called a physical adsorption method or a chemical binding method.

**[0022]** The sample as the object of assay for the particle-enhanced immunoassay method, reagent and kit is not particularly limited as long as it is a human body fluid and contains human cystatin C, such as blood serum, blood plasma, synovial fluid, milk, saliva, cerebrospinal fluid, seminal plasma, amniotic fluid, urine, and lacrimal fluid.

**[0023]** The particle-enhanced immunoassay method, reagent and kit of the present invention can be applied to a turbidimetric method using general-purpose automatic analyzers manufactured by Hitachi, Ltd. as well as Toshiba Corp. , JEOL, Ltd. , Olympus Corp. , and Sekisui Medical Co. , Ltd. , or using an exclusive automatic analyzer LPIA (registered trademark) (manufactured by Mitsubishi Kagaku Iatron, Inc.) which uses near-infrared region as the measurement wavelength, and to a nephelometric method using an apparatus for measuring the scattered light intensity (manufactured by Dade Behring, Inc.), and therefore, the method, reagent and kit can be easily automated.

**[0024]** The reagent for particle-enhanced immunoagglutination assay of the present invention may contain, in addition to the main component, for example, acetic acid, citric acid, phosphoric acid, Tris, glycine, boric acid, carbonic acid, and

Good's buffer, or sodium salts, potassium salts and calcium salts of the acids, as components for buffering the ionic strength, osmotic pressure and the like of the sample. Furthermore, the reagent may also contain a polymer such as polyethylene glycol, polyvinylpyrrolidone, or a phospholipid polymer, as a component for enhancing immunological agglutination. The reagent may also contain a single kind or a combination of plural kinds of the components that are

generally used for particle-enhanced immunoagglutination assays, such as proteins, amino acids, sugars, metal salts, surfactants, reducing substances or chaotropic substances, as a component that controls the formation of immunological agglutination. Furthermore, the reagent may also be prepared into a kit containing these components in combination.

**[0025]** The particle-enhanced immunoassay method for human cystatin C and the particle-enhanced immunoassay reagent and kit of the present invention can be applied to any use, as long as the use is intended for measuring human cystatin C in a sample specifically with high accuracy.

#### EXAMPLES

**[0026]** Hereinafter, an embodiment of the present invention will be described in detail by way of a Production Example, Evaluation Examples, and an Example, but the present invention is not intended to be limited thereto.

[Production Example]

**[0027]** Production of high-affinity anti-human cystatin C monoclonal antibody

100  $\mu\text{g}$  of purified human cystatin C (SCIPAC, Ltd.) was used for a single immunization. The first immunization was carried out using 200  $\mu\text{L}$  of an emulsion prepared by mixing equal amounts of human cystatin C and Freund's complete adjuvant, by injecting this to the abdominal cavities of BALB/c mice. For additional immunization, long-term immunization was carried out in which additional immunization was repeated 6 times at an interval of 2 weeks in order to acquire an antibody having high affinity, using 200  $\mu\text{L}$  of an emulsion prepared in the same manner using Freund's incomplete adjuvant. The antibody titers in the blood collected from the orbital vein of the mice were measured by ELISA, and mice which gave high antibody titer were selected and supplied to cell fusion. After two weeks from the seventh immunization, a solution prepared by dissolving 100  $\mu\text{g}$  of human cystatin C in 200  $\mu\text{L}$  of physiological saline, was injected into the abdominal cavity of each mouse, and after three days, the spleen was enucleated. The spleen was dissociated in RPMI1640 medium, and then the resultant was centrifuged at 1500 rpm to collect spleen cells. These cells were washed three or more times with bovine fetal serum-free RPMI1640 medium, and then suspended in 2 mL of RPMI1640 medium containing 15% bovine fetal serum which was added to the cells to obtain a spleen cell suspension. The spleen cells and myeloma cells SP2/0-AG14 were mixed at a ratio of 6:1, and then the cells were subjected to cell fusion in the presence of 50% polyethylene glycol. Thus, a hybridoma (fused cell) was obtained. The cells were centrifuged at 1500 rpm, and the precipitation was collected and suspended in GKN solution (a solution obtained by dissolving 2 g of glucose, 0.4 g of potassium chloride, 8 g of sodium chloride, 1.41 g of disodium hydrogen phosphate, and 0.78 g of sodium dihydrogen phosphate dihydrate in purified water, and adjusting the volume to 1 liter). The precipitation was washed by centrifugation, and the precipitation was collected. This collected precipitation was suspended in 30 mL of RPMI1640 medium containing 15% bovine fetal serum, and the suspension was dispensed into three 96-well microplates in an amount of 100  $\mu\text{L}$  per well, together with 200  $\mu\text{L}$  per well of HAT medium containing thymic cells of a BALB/c mouse in an amount of  $2.5 \times 10^6$  cells/mL as feeder cells. The hybridoma was cultured in an incubator containing 5% carbon dioxide at 37°C.

**[0028]** The presence of an anti-human cystatin C antibody in the culture supernatant was identified by ELISA in which human cystatin C was immobilized, and after 10 days, proliferation of the hybridoma was confirmed in all of the wells. Specifically, 100  $\mu\text{L}$  of 10 mM phosphate buffer solution (pH 7.2; hereinafter, abbreviated as PBS) containing 10  $\mu\text{g}/\text{mL}$  of human cystatin C and 150 mM sodium chloride was dispensed on the 96-well microplate where proliferation of the hybridoma had been confirmed. The microplate was left to stand overnight at 4°C. Subsequently, this 96-well microplate was washed three times with 300  $\mu\text{L}$  of PBS containing 0.05% Tween 20 and 1% bovine serum albumin (hereinafter, BSA), and then the culture supernatant of the respective wells was added to the microplate in an amount of 50  $\mu\text{L}/\text{well}$  and was left to stand for one hour at room temperature. Thereafter, the plate was washed three times with a PBS containing 0.05% Tween20, and then a peroxidase-labeled anti-mouse antibody (manufactured by Sekisui Medical Co., Ltd.) was added to the microplate in an amount of 50  $\mu\text{L}/\text{well}$  and was left to stand for one hour at room temperature. Thereafter, the microplate was washed three times with a PBS containing 0.05% Tween 20, and then a citrate buffer solution (pH 5) containing 0.2% ortho-phenylenediamine and 0.02% hydrogen peroxide was added to the microplate in an amount of 50  $\mu\text{L}/\text{well}$ . The microplate was left to stand for 15 minutes at room temperature, and then 4.5 N sulfuric acid was added thereto in an amount of 50  $\mu\text{L}/\text{well}$  to stop the reaction. The absorbance of each well at a wavelength of 492 nm was measured, and the wells having high absorbance were selected as positive wells.

**[0029]** The cells were rendered monoclonal by limiting dilution method. That is, on a 96-well microplate to which thymus cells of a BALB/c mouse had been dispensed in an amount of  $10^6$  cells per well as feeder cells, 0.1 mL of a solution

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obtained by diluting the hybridoma in the positive wells to a concentration of 10 cells/mL, was dispensed in an amount of 0.1 mL each. Regarding the medium, HT medium was used for the first time, while RPMI1640 medium containing 15 % bovine fetal serum was used since the second time, and the cells were cultured for 10 days in an incubator containing 5% carbon dioxide at 37°C. The monoclonalization operation based on the selection of positive wells according to ELISA and on a limiting dilution method was repeated three times, and hybridomas producing anti-human cystatin C monoclonal antibody having high reactivity were obtained. About 10<sup>5</sup> cells of each of the hybridomas were administered to the abdominal cavity of a mouse which had been pretreated with pristane, and the ascites fluids were respectively collected. Insoluble materials were removed by centrifugation from each of the ascites fluids thus collected, and an equal amount of saturated ammonium sulfate solution was added thereto. The resulting mixture was left to stand overnight while stirred, and a precipitation was collected by centrifugation. The collected precipitation was dissolved in a 20 mM Tris buffer solution (pH 8), and dialyzed into the same buffer solution. Each of the dialysis contents was separately adsorbed to a DEAE-Sepharose column which had been equilibrated with the same buffer solution, and then elution was performed at a concentration gradient of 0 to 300 mM of sodium chloride in the same buffer solution. Thus, various purified anti-human cystatin C monoclonal antibodies were obtained.

[Evaluation Example 1]

**[0030]** Selection of anti-human cystatin C monoclonal antibodies recognizing different epitopes

**[0031]** An investigation was made on combinations of anti-human cystatin C monoclonal antibodies recognizing different epitopes, which had been produced using the reactivity in sandwich ELISA as an index. Specifically, various anti-human cystatin C monoclonal antibodies were immobilized on a plate by adding a PBS containing 1 µg/mL each of the antibodies in an amount of 50 µL/well and leaving the plate to stand for 2 hours at room temperature. Subsequently, the plate was washed three times with 300 µL of PBS containing 0.05% Tween 20, and a PBS containing 0.05% Tween 20 and 1% BSA was added to the plate in an amount of 300 µL/well. The plate was left to stand for one hour at room temperature. Thereafter, the plate was washed three times with a PBS containing 0.05% Tween 20, and then a PBS containing 10 ng/mL of human cystatin C was added to the plate in an amount of 50 µL/well. The plate was left to stand for one hour at room temperature to allow a reaction to occur, and the plate was washed three times with a PBS containing 0.05% Tween 20. Then, a PBS containing 1 µg/mL of each of the anti-human cystatin C monoclonal antibodies which had been biotininated for detection, was added to the plate in an amount of 50 µL/well, and the plate was left to stand for one hour at room temperature. The plate was washed three times with a PBS containing 0.05% Tween 20, and then a peroxidase-labeled streptavidin (ZYMED Laboratories, Inc.) which was diluted to 5000 times, was added to the plate in an amount of 50 µL/well. The plate was left to stand for one hour at room temperature. The plate was washed three times with a PBS containing 0.05% Tween 20, and then a citrate buffer solution (pH 5) containing 0.2% ortho-phenylenediamine and 0.02 % hydrogen peroxide was added to the plate in an amount of 50 µL/well to allow a reaction for 15 minutes at room temperature. Then, 4.5 N sulfuric acid was added in an amount of 50 µL/well to the plate to stop the reaction, and the absorbance of each well at a wavelength of 492 nm was measured. The antibodies of the combinations which were found to have reactivity were considered the monoclonal antibodies recognizing different epitopes. As a result, it was confirmed that four kinds of anti-human cystatin C monoclonal antibodies a to d could be used to constitute combinations such as indicated in Table 1.

[Table 1]

		Immobilized antibody			
		a	b	c	d
Biotininated antibody	a		++	-	+
	b	++		-	++
	c	+	-		+
	d	++	++	-	
++: Strongly reactive +: Reactive -: Non-reactive					

**[0032]** It was confirmed that the antibody a could be combined with any of the antibodies b to d. Furthermore, in regard to the antibody c, since this antibody is not affected by the antibody that is combined therewith and does not exhibit reactivity when in a state of being immobilized, it is thought that binding to an insoluble carrier, that is, use in a particle-

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enhanced immunoassay where immobilization is required, is impossible.

[Evaluation Example 2]

5 **[0033]** Identification of particle-enhanced immunoagglutination in a combination of anti-human cystatin C monoclonal antibodies recognizing different epitopes

**[0034]** Antibody-coated latex particles to which anti-human cystatin C monoclonal antibodies a, b and d were each independently bound in small amounts, were prepared, and the formation of a particle-enhanced immunoagglutinate which was dependent on the concentration of human cystatin C and was constituted of a combination recognizing  
10 different epitopes, was confirmed.

(Preparation of antibody-coated latex particles)

**[0035]** To a 0.5% latex solution (30 mM Tris-HCl, pH 8.5 to 9.0) of particles having an average particle size of 0.13  $\mu\text{m}$  or 0.3  $\mu\text{m}$  (manufactured by Sekisui Medical Co., Ltd.), an equal amount of 0.2 mg/mL solution of each cystatin C monoclonal antibody (30 mM Tris-HCl, pH 8.5 to 9.0) was added, and the mixture was stirred for one hour at 4°C. Then, an equal amount of 1% BSA solution (30 mM Tris-HCl, pH 8.5 to 9.0) was added thereto, and the resulting mixture was stirred for 30 minutes at 4°C. The mixture was centrifuged, and the supernatant was removed. Subsequently, a precip-  
15 itation obtained therefrom was resuspended in purified water to obtain an antibody-coated latex particle solution. At this  
20 time, the binding amount of the antibody relative to the total weight of the antibody-coated particles was about 3.8% by weight.

(Preparation of first reagent)

25 **[0036]** A 30 mM CHES buffer solution (pH 8.75) containing 750 mM of potassium chloride and 1% BSA was prepared, and this was used as the first reagent.

(Preparation of second reagent)

30 **[0037]** Equal amounts of two kinds of antibody-coated latex particle solutions were mixed, and the mixture was diluted with a 5 mM MOPS buffer solution (pH7.0) such that the final absorbance was 2.0 OD. This dilution was used as the second reagent. The reagent was prepared by combining the antibody-coated latex particles of a-b, a-d or b-d, which were recognized to have strong reactivity from the results of the Evaluation Example 1, such that particles having the same particle size were combined.  
35

(Assay method)

**[0038]** The first reagent and the second reagent were combined, and the formation of a particle-enhanced immuno-  
40 agglutinate which was dependent on the concentration of human cystatin C was identified using a Hitachi 7170 type automatic analyzer. Specifically, 100  $\mu\text{L}$  of the first reagent was added to 2.5  $\mu\text{L}$  of human cystatin C solution having a concentration of 0, 0.5, 1.0, 2.0, 4.0 or 8.0 mg/L, and the mixture was warmed to 37°C for 5 minutes. Subsequently, 100  $\mu\text{L}$  of the second reagent was added thereto, and the resulting mixture was stirred. Changes in the absorbance that accompanied the formation of an agglutinate for 5 minutes after the mixing, were measured as the sensitivity at a main wavelength of 570 nm and a sub-wavelength of 800 nm.  
45

(Results)

**[0039]** A reagent prepared by combining antibody a-coated latex particles and antibody b-coated latex particles was used to analyze cystatin C solutions of various concentrations, and as shown in the cystatin C concentration-sensitivity  
50 curve described in FIG. 1, it was identified that the sensitivity increases in a cystatin C concentration-dependent manner in all of the particles, and an assay can be carried out. Particles having a size of 0.13  $\mu\text{m}$  (closed square) exhibited slightly low sensitivity in the lower concentration region but exhibited a satisfactory sensitivity increase in the higher concentration region. Particles having a size of 0.3  $\mu\text{m}$  (open square) exhibited high sensitivity in the lower concentration region but exhibited a poor sensitivity increase in the higher concentration region. Furthermore, in are agent prepared  
55 by combining antibody a-coated latex particles and antibody d-coated latex particles, the sensitivities in the lower concentration region of the particles having a size of 0. 13  $\mu\text{m}$  (closed circle) and the particles having a size of 0.3  $\mu\text{m}$  (open circle) were equal, and while the particles having a size of 0.13  $\mu\text{m}$  exhibited an increase in sensitivity up to the higher concentration region, a sensitivity increase was hardly recognized in the particles having a size of 0.3  $\mu\text{m}$ . Therefore, it

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was thought that although there are particles that cannot be utilized depending on the combination, both of the combinations can be applied to the immunoagglutination assay. On the other hand, in a reagent prepared by combining antibody b-coated latex particles and antibody d-coated latex particles, which are indicated by broken lines, cystatin C concentration-dependent increases in sensitivity were hardly recognized in both the particles having a size of 0.13  $\mu\text{m}$  (closed triangle) and the particles having a size of 0.3  $\mu\text{m}$  (open triangle), and it was thought that use of these reagents in a particle-enhanced immunoassay method was difficult.

[Evaluation Example 3]

**[0040]** Determination of the dissociation constant of anti-human cystatin C monoclonal antibody by BIACORE

**[0041]** The dissociation constants of purified anti-human cystatin C monoclonal antibodies a, b and d were calculated according to a conventional method using a BIACORE apparatus (GE Healthcare Corp.). Specifically, the anti-human cystatin C monoclonal antibodies were respectively immobilized on sensor chips using a Mouse Antibody Capture Kit (GE Healthcare Corp.), and the dissociation constants were calculated from their reactivity with human cystatin C (R&D Systems, Inc.) at concentrations of 0  $\mu\text{g}/\text{mL}$  and up to 0.0625, 0.125, 0.25, 0.5, and 1.0  $\mu\text{g}/\text{mL}$  (Table 2). The combinations a-b and a-d, which were considered to be applicable to particle-enhanced immunoassays in Evaluation Example 2, were such that an anti-human cystatin C monoclonal antibody a having high affinity (dissociation constant: 0.29 nM) and an anti-human cystatin C monoclonal antibody b (dissociation constant: 1.32 nM) or d (dissociation constant: 2.30 nM) having ordinary affinity were combined. Thus, the ratio of dissociation constants about 4.6 for the ratio b/a (1.32/0.29) and about 7.9 for the ratio d/a (2.30/0.29) showed clear differences in affinity between the antibodies combined. On the other hand, the combination b-d which was considered to be inapplicable to particle-enhanced immunoagglutination, was a combination of antibodies having ordinary affinity, and the ratio of dissociation constants was about 1.7 for d/b (2.30/1.32), by which a clear difference in affinity was not recognized. Thus, Hybridoma 75202 that produces the monoclonal antibody a, which is considered to have the high affinity and the highest general-purpose applicability among these antibodies, was selected. This hybridoma was deposited with International Patent Organism Depository, National Institute of Advanced Industrial Science and Technology (305-8566, Tsukuba Central 6, 1-1-1 Higashi, Tsukuba, Ibaraki, Japan) on November 20, 2008, under the Accession Number FERM BP-11186.

[Table 2]

Antibody	Dissociation constant, KD value (nM)
a	0.29
b	1.32
d	2.30

[Evaluation Example 4]

**[0042]** Examination of anti-human cystatin C monoclonal antibody specificity according to ELISA

**[0043]** An evaluation of the specificities of the anti-human cystatin C monoclonal antibodies a, b and d produced in this study was carried out by ELISA. Specifically, 50  $\mu\text{L}$  of PBS containing 1  $\mu\text{g}/\text{mL}$  of human cystatin C and members of the cystatin family, such as human cystatinA, cystatin B, cystatin D, cystatin E/M, cystatin F, cystatin S, cystatin SA and kininogen (all by R&D Systems, Inc.), was dispensed on a 96-well microplate, and the microplate was left to stand overnight at 4°C to immobilize the cystatins on the plate. Subsequently, this 96-well microplate was washed three times with 300  $\mu\text{L}$  of PBS containing 0.05% Tween 20, and then a PBS containing 0.05% Tween 20 and 1% BSA was added to the microplate in an amount of 300  $\mu\text{L}/\text{well}$ . The microplate was left to stand for 1 hour at room temperature. Thereafter, a PBS containing 1  $\mu\text{g}/\text{mL}$  of each of the anti-human cystatin C monoclonal antibodies produced was added to the microplate in an amount of 50  $\mu\text{L}/\text{well}$ , and the microplate was left to stand for 1 hour at room temperature. The microplate was washed three times with a PBS containing 0.05% Tween 20, and then a peroxidase-labeled anti-mouse antibody (Biosource, Inc.) which was diluted to 5000 times with PBS, was added to the microplate in an amount of 50  $\mu\text{L}/\text{well}$ . The microplate was left to stand for 1 hour at room temperature. Thereafter, the plate was washed three times with a PBS containing 0.05% Tween 20, and a citrate buffer solution (pH 5) containing 0.2% ortho-phenylenediamine and 0.02% hydrogen peroxide was added to the microplate in an amount of 50  $\mu\text{L}/\text{well}$  to allow a reaction to proceed for 15 minutes at room temperature. Subsequently, 4.5 N sulfuric acid was added thereto in an amount of 50  $\mu\text{L}/\text{well}$  to stop the reaction, and the absorbance of each well at a wavelength of 492 nm was measured.

**[0044]** As controls, an anti-human cystatin C rabbit polyclonal antibody (BioVendor, Inc.) was used in place of the anti-human cystatin C monoclonal antibody, and a peroxidase-labeled anti-rabbit antibody (Cappel, Inc.) was used in place

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of the peroxidase-labeled anti-mouse antibody, to compare specificity.

**[0045]** As shown in FIG. 2, it was confirmed that any of the monoclonal antibodies a, b and d specifically reacted only with human cystatin C. Thus, it is contemplated that cystatin C in a human body fluid can be specifically measured by using these antibodies. On the other hand, the polyclonal antibody used as a control exhibited strong reactivity toward cystatin B, cystatin D, cystatin E/M, cystatin S, cystatin SA and cystatin SN, in addition to cystatin C, to an equal extent, and also exhibited weak reactivity toward cystatin A, cystatin F and kininogen. Therefore, it was expected that in an assay using an anti-cystatin C polyclonal antibody, accurate measurement of cystatin C is impossible in a sample in which these cystatin family molecules are co-present.

**[0046]** The same scheme of Evaluation Examples 1 to 4 were repeated, and as a result, combinations 1 and 2 that use anti-human cystatin C monoclonal antibodies e, f and g, which are believed to be applicable to particle-enhanced immunoassays, were further discovered. These combinations were all combinations including at least one kind of high-affinity antibody having a dissociation constant of less than 1 nM. Combination 1 was a combination of high-affinity antibodies, and the ratio of dissociation constants was about 2.1. Combination 2 was a combination of a high-affinity antibody and an antibody having ordinary affinity, and the ratio of dissociation constants was about 15.4 (Table 3).

[Table 3]

Combination	Antibody	Dissociation constant	
		KD value (nM)	Ratio
1	a	0.29	2.07
	e	0.14	(a/e)
2	f	0.54	15.37
	g	8.30	(g/f)

[Example]

**[0047]** A reagent for cystatin C assay of the present invention, in which the latex particle size and other general conditions were optimized for the analysis by a Hitachi 7170 type automatic analyzer, was prepared using the anti-human cystatin C monoclonal antibodies a and b, and a comparison was made on the performance with an existing cystatin C assay reagent (N-Latex Cystatin C Kit manufactured by Dade Behring, Inc.), which uses an anti-cystatin C polyclonal antibody. However, an assay reagent for general-purpose automatic analyzers that use anti-human cystatin C monoclonal antibodies, was not available.

(Preparation of antibody-coated latex particles)

**[0048]** To a 0.5% latex solution (30 mM Tris-HCl, pH 8.5) of particles having an average particle size of 0.15  $\mu\text{m}$ , an equal amount of 0.15 mg/mL solution of antibody a or b (30 mM Tris-HCl, pH 8.5) was added, and the mixture was stirred for one hour at 4°C. Subsequently, an equal amount of 1% BSA solution (30 mM Tris-HCl, pH 8.5) was added thereto, and the resulting mixture was stirred for 30 minutes at 4°C. The mixture was centrifuged, the supernatant was removed, and then a precipitate produced therefrom was resuspended in purified water. The resultant was used as an anti-human cystatin C monoclonal antibody a- or b-coated latex particle solution. At this time, the binding amount of the antibody relative to the total weight of the antibody-coated particles was about 2.9% by weight.

(Preparation of the first reagent)

**[0049]** A 30 mM Tris-HCl solution at pH 8.0 containing 750 mM of potassium chloride and 1% BSA was prepared, and this solution was used as the first reagent.

(Preparation of the second reagent)

**[0050]** Equal amounts of the antibody a-coated latex particle solution and the antibody b-coated latex particle solution were mixed, and the mixture was diluted with a 5 mM MOPS buffer solution (pH 7.0) such that the final absorbance was 1.5 OD. This dilution was used as the second reagent.

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(Assay sample)

Human serum specimen

5 (Assay conditions)

**[0051]** Hitachi 7170 Automatic Analyzer: Parameter conditions

- Specimen - First reagent - Second reagent: 2.4  $\mu$ L - 120  $\mu$ L - 120  $\mu$ L
- 10 • Analysis method: Two-point end method (measurement point 18-34)
- Measurement wavelength: Main wavelength 570 nm/sub-wavelength 800 nm
- Calibration: Spline

(Assay method)

15 **[0052]** A calibration curve was produced from the measured sensitivities of human cystatin C solutions at concentrations of 0, 0.5, 1.0, 2.0, 4.0 and 10.0 mg/L, and the cystatin C concentrations in various samples were measured based on the calibration curve. Thus, correlation between samples of the present invention and reference reagents was determined (number of samples, N = 50); and samples of the present invention and reference reagents were compared on simultaneous reproducibility (number of measurements in succession, n = 10) and the detection limit concentration (the concentration at which the error bars of the average measured sensitivity + 2SD of a sample with a zero cystatin C concentration and the average measured sensitivity -2SD of a sample with a known cystatin C concentration do not overlap, is defined as the detection limit concentration).

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25 (Reference reagent and analyzer)

**[0053]**

Reference reagent: N-Latex Cystatin C Kit  
 Analyzer: BN Prospec  
 All manufactured by Dade Behring, Inc.

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(Results)

35 **[0054]** The correlation on the assay values between the reagent of the present invention and the reference reagent was satisfactory (FIG. 3). Furthermore, in regard to reproducibility, the coefficient of variation of the reagent of the present invention was as small as 1/5 or less of the value of existing reagents, and reproducibility was excellent (Table 4). In regard to the detection limit concentration, the detection limit concentration for cystatin C was 0.2 mg/L with the reference reagent, while 0.05 mg/L with the reagent of the present invention. Thus, it was confirmed that the reagent of the present invention was capable of detecting a concentration as low as 1/4 or less of the detection limit concentration of the reference reagent (FIG. 4).

40

**[0055]** The particle-enhanced immunoassay method of the present invention which uses antibody-coated particles in which some insoluble carrier particles are coated with an anti-human cystatin C monoclonal antibodies of one type which have high affinity, and in which some insoluble carrier particles are coated with anti-human cystatin C monoclonal antibodies of another type which recognize an epitope different from that the monoclonal antibodies of the first-mentioned type recognize and which have relatively low affinity, has made it possible to establish a method for measuring cystatin C in a human body fluid, which method is inexpensive but of high performance and high specificity, and in which the binding amount of the antibody relative to the total weight of the antibody-coated particles is smaller than the binding amount that has been used in conventional reagents, i.e. from greater than 5% by weight to not less than 35% by weight.

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[Table 4]

	Reagent of present invention		Reference reagent	
	Sample 1	Sample 2	Sample 1	Sample 2
	0.49	1.99	0.379	1.64
	0.49	2.00	0.379	1.69

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(continued)

	Reagent of present invention		Reference reagent	
	Sample 1	Sample 2	Sample 1	Sample 2
	0.49	2.01	0.401	1.66
	0.49	2.01	0.406	1.64
	0.48	2.01	0.424	1.75
	0.48	2.01	0.408	1.71
	0.49	2.01	0.379	1.75
	0.48	2.01	0.447	1.75
	0.49	2.01	0.428	1.71
	0.50	1.99	0.383	1.72
Average value (mg/L)	0.49	2.01	0.41	1.70
Standard deviation	0.006	0.008	0.023	0.043
Coefficient of variation (%)	1.30	0.42	5.57	2.25

INDUSTRIAL APPLICABILITY

**[0056]** The particle-enhanced immunoassay method, reagent and kit of the present invention provide a high-performance, inexpensive, and convenient method for detecting and measuring human cystatin C in a human body fluid.

**Claims**

1. A method for measuring cystatin C in a human body fluid, the method being a particle-enhanced immunoassay method using a combination of antibody-coated particles, said combination comprising

- (i) an insoluble carrier particle coated with anti-human cystatin C monoclonal antibodies which have high human cystatin C affinity, and
- (ii) an insoluble carrier particle coated with anti-human cystatin C monoclonal antibodies which have relatively low human cystatin C affinity as well as recognize an epitope different from that the first mentioned monoclonal antibodies can recognize,

wherein the binding amount of anti-human cystatin C monoclonal antibodies of either type relative to the total weight of the particles being coated with the corresponding antibodies is less than 5 % by weight, and wherein the anti-human cystatin C monoclonal antibody having high human cystatin C affinity (i) has an affinity for cystatin C that is higher than the anti-human cystatin C monoclonal antibody (ii) having low human cystatin C affinity.

2. The method for measuring cystatin C in a human body fluid according to claim 1, wherein the average particle size of the insoluble carrier particles is 0.1 to 0.4  $\mu\text{m}$ .

3. The method for measuring cystatin C in a human body fluid according to claim 1 or 2, wherein the anti-human cystatin C monoclonal antibody is an antibody fragment containing as a functional site a Fab which is a human cystatin C recognition site, or a recombinant antibody.

4. The method for measuring cystatin C in a human body fluid according to claim 1, wherein the dissociation constant (kd value) of the anti-human cystatin C monoclonal antibody having high affinity is less than 1 nM, and the ratio obtained by dividing the dissociation constant of the anti-human cystatin C monoclonal antibody having relatively low affinity by the dissociation constant of the anti-human cystatin C monoclonal antibody having high affinity, is 2 or greater.

5. The method for measuring cystatin C in a human body fluid according to any one of claims 1 to 4, wherein the human body fluid is blood serum, blood plasma, synovial fluid, milk, saliva, cerebrospinal fluid, seminal plasma, amniotic fluid, urine, or lacrimal fluid.

5 6. A particle-enhanced immunoassay reagent for cystatin C in a human body fluid, the reagent comprising  
 (i) an insoluble carrier particle coated with anti-human cystatin C monoclonal antibodies which have high human cystatin C affinity, and  
 10 (ii) an insoluble carrier particle coated with anti-human cystatin C monoclonal antibodies which have relatively low human cystatin C affinity as well as recognize an epitope different from that the first-mentioned monoclonal antibodies can recognize,

wherein the binding amount of anti-human cystatin C monoclonal antibodies of either type relative to the total weight of the particle being coated with the corresponding antibodies is less than 5%, wherein the anti-human cystatin C monoclonal antibody having high human cystatin C affinity (i) has an affinity for cystatin C that is higher than the  
 15 anti-human cystatin C monoclonal antibody (ii) having low human cystatin C affinity.

7. The assay reagent according to claim 6, wherein the average particle size of the insoluble carrier particles is 0.1 to 0.4  $\mu\text{m}$ .

**Patentansprüche**

25 1. Verfahren zur Messung von Cystatin C in einer menschlichen Körperflüssigkeit, wobei das Verfahren ein Verfahren eines partikelverstärkten Immunoassays ist, bei dem eine Kombination von mit Antikörpern beschichteten Partikeln verwendet wird, wobei die Kombination folgendes umfasst:

(i) einen unlöslichen Trägerpartikel, der mit monoklonalen anti-human Cystatin C Antikörpern beschichtet ist, die eine hohe Affinität zu humanem Cystatin C aufweisen, und  
 30 (ii) einen unlöslichen Trägerpartikel, der mit monoklonalen anti-human Cystatin C Antikörpern beschichtet ist, die eine relativ geringe Affinität zu humanem Cystatin C aufweisen sowie ein Epitop erkennen, das sich von dem unterscheidet, das die zuerst genannten monoklonalen Antikörper erkennen können, wobei die Bindungsmenge der monoklonalen anti-human Cystatin C Antikörper beider Arten relativ zu dem Gesamtgewicht der Partikel, die mit den entsprechenden Antikörpern beschichtet werden, weniger als 5 Gewichtsprozent beträgt, und wobei der monoklonale anti-human Cystatin C Antikörper, der eine hohe Affinität zu humanem Cystatin C aufweist (i) eine Affinität zu Cystatin C aufweist, die höher ist als die des monoklonalen anti-human Cystatin C Antikörpers, der eine geringe Affinität zu humanem Cystatin C aufweist (ii).

40 2. Verfahren zur Messung von Cystatin C in einer menschlichen Körperflüssigkeit nach Anspruch 1, wobei die durchschnittliche Partikelgröße der unlöslichen Trägerpartikel 0,1 bis 0,4  $\mu\text{m}$  beträgt.

45 3. Verfahren zur Messung von Cystatin C in einer menschlichen Körperflüssigkeit nach Anspruch 1 oder 2, wobei der monoklonale anti-human Cystatin C Antikörper ein Antikörper-Fragment ist, das als funktionelle Stelle ein Fab enthält, welches eine humane Cystatin Erkennungsstelle ist, oder ein rekombinanter Antikörper ist.

50 4. Verfahren zur Messung von Cystatin C in einer menschlichen Körperflüssigkeit nach Anspruch 1, wobei die Dissoziationskonstante ( $K_d$ -Wert) des monoklonalen anti-human Cystatin C Antikörpers, der eine hohe Affinität aufweist, weniger als ein 1 nM beträgt, und das Verhältnis, das durch Teilen der Dissoziationskonstante des monoklonalen anti-human Cystatin C Antikörpers, der eine relativ geringe Affinität aufweist, durch die Dissoziationskonstante des monoklonalen anti-human Cystatin C Antikörpers, der eine hohe Affinität aufweist, erhalten wird, 2 oder mehr beträgt.

55 5. Verfahren zur Messung von Cystatin C in einer menschlichen Körperflüssigkeit nach einem der Ansprüche 1 bis 4, wobei die menschliche Körperflüssigkeit Blutserum, Blutplasma, Gelenkflüssigkeit, Milch, Speichel, Zerebrospinalflüssigkeit, Samenplasma, Fruchtwasser, Urin oder Tränenflüssigkeit ist.

6. Reagens für einen partikewerstützten Immunoassay für Cystatin C in einer menschlichen Körperflüssigkeit, wobei das Reagens folgendes umfasst:

- (i) einen unlöslichen Trägerpartikel, der mit monoklonalen anti-human Cystatin C Antikörpern beschichtet ist, die eine hohe Affinität zu humanem Cystatin C aufweisen, und  
 (ii) einen unlöslichen Trägerpartikel, der mit monoklonalen anti-human Cystatin C Antikörpern beschichtet ist, die eine relativ geringe Affinität zu humanem Cystatin C aufweisen sowie ein Epitop erkennen, das sich von dem unterscheidet, das die zuerst genannten monoklonalen Antikörper erkennen können, wobei die Bindungsmenge der monoklonalen anti-human Cystatin C Antikörper beider Arten relativ zu dem Gesamtgewicht der Partikel, die mit den entsprechenden Antikörpern beschichtet werden, weniger als 5 Gewichtsprozent beträgt, wobei der monoklonale anti-human Cystatin C Antikörper, der eine hohe Affinität zu humanem Cystatin C aufweist (i), eine Affinität zu Cystatin C aufweist, die höher ist als die des monoklonalen anti-human Cystatin C Antikörpers, der eine geringe Affinität zu humanem Cystatin C aufweist (ii).

7. Reagens für einen Assay nach Anspruch 6, wobei die durchschnittliche Partikelgröße der unlöslichen Trägerpartikel 0,1 bis 0,4  $\mu\text{m}$  beträgt.

## Revendications

1. Procédé pour mesurer la cystatine C dans un fluide corporel humain, le procédé étant un immunodosage amplifié par des particules utilisant une combinaison de particules revêtues d'anticorps, ladite combinaison comprenant

- (i) une particule de support insoluble revêtue d'anticorps monoclonaux anti-cystatine C humaine qui présentent une forte affinité pour la cystatine C humaine, et  
 (ii) une particule de support insoluble revêtue d'anticorps monoclonaux anti-cystatine C humaine qui présentent une affinité relativement faible pour la cystatine C humaine et également qui reconnaissent un épitope différent de celui que peuvent reconnaître les premiers anticorps monoclonaux mentionnés,

dans lequel la quantité de liaison d'anticorps monoclonaux anti-cystatine C humaine de l'un ou l'autre type par rapport à la quantité totale des particules revêtues des anticorps correspondants est inférieure à 5 % en poids, et dans lequel l'anticorps monoclonal anti-cystatine C humaine présentant une forte affinité pour la cystatine C humaine (i) présente une affinité pour la cystatine C qui est supérieure à celle de l'anticorps monoclonal anti-cystatine C humaine (ii) présentant une faible affinité pour la cystatine C humaine.

2. Procédé pour mesurer la cystatine C dans un fluide corporel humain selon la revendication 1, dans lequel la granulométrie moyenne des particules de support insolubles est de 0,1 à 0,4  $\mu\text{m}$ .

3. Procédé pour mesurer la cystatine C dans un fluide corporel humain selon la revendication 1 ou 2, dans lequel l'anticorps monoclonal anti-cystatine C humaine est un fragment d'anticorps contenant, en tant que site fonctionnel, un Fab qui est un site de reconnaissance de la cystatine C humaine, ou un anticorps recombiné.

4. Procédé pour mesurer la cystatine C dans un fluide corporel humain selon la revendication 1, dans lequel la constante de dissociation (valeur  $K_d$ ) de l'anticorps monoclonal anti-cystatine C humaine présentant une forte affinité est inférieure à 1 nM, et le rapport obtenu par division de la constante de dissociation de l'anticorps monoclonal anti-cystatine C humaine présentant une affinité relativement faible par la constante de dissociation de l'anticorps monoclonal anti-cystatine C humaine présentant une forte affinité est de 2 ou plus.

5. Procédé pour mesurer la cystatine C dans un fluide corporel humain selon l'une quelconque des revendications 1 à 4, dans lequel le fluide corporel humain est le sérum sanguin, le plasma sanguin, le fluide synovial, le lait, la salive, le liquide céphalo-rachidien, le plasma séminal, le liquide amniotique, l'urine, ou le fluide lacrymal.

6. Réactif d'immunodosage amplifié par des particules pour la cystatine C dans un fluide corporel humain, le réactif comprenant

- (i) une particule de support insoluble revêtue d'anticorps monoclonaux anti-cystatine C humaine qui présentent une forte affinité pour la cystatine C humaine, et  
 (ii) une particule de support insoluble revêtue d'anticorps monoclonaux anti-cystatine C humaine qui présentent une affinité relativement faible pour la cystatine C humaine et également qui reconnaissent un épitope différent de celui que peuvent reconnaître les premiers anticorps monoclonaux mentionnés,

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dans lequel la quantité de liaison d'anticorps monoclonaux anti-cystatine C humaine de l'un ou l'autre type par rapport à la quantité totale des particules revêtues des anticorps correspondants est inférieure à 5 % en poids, et dans lequel l'anticorps monoclonal anti-cystatine C humaine présentant une forte affinité pour la cystatine C humaine (i) présente une affinité pour la cystatine C qui est supérieure à celle de l'anticorps monoclonal anti-cystatine C humaine (ii) présentant une faible affinité pour la cystatine C humaine.

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7. Réactif de dosage selon la revendication 6, dans lequel la granulométrie moyenne des particules de support insolubles est de 0,1 à 0,4  $\mu\text{m}$ .

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Drawings

Fig. 1

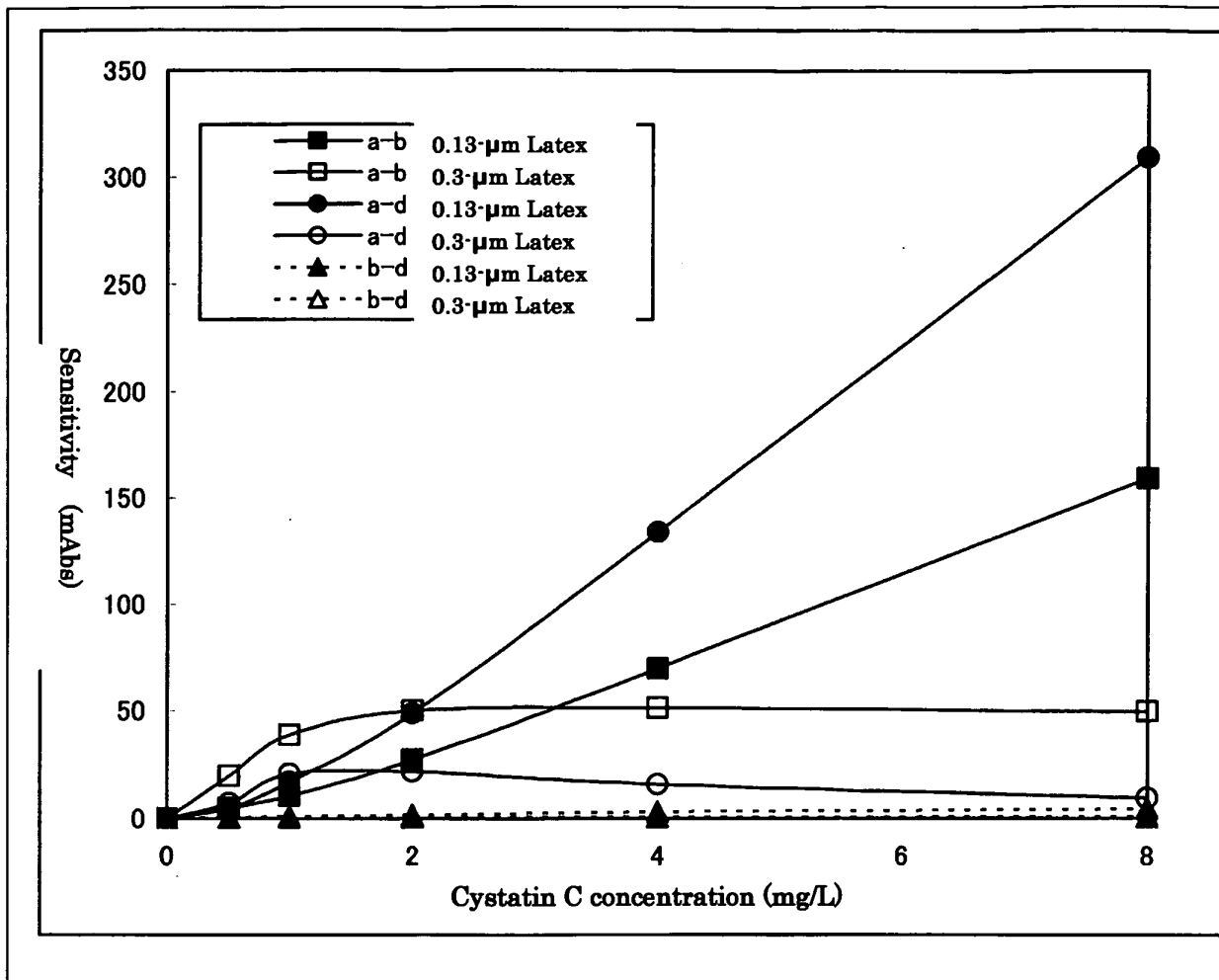


Fig. 2

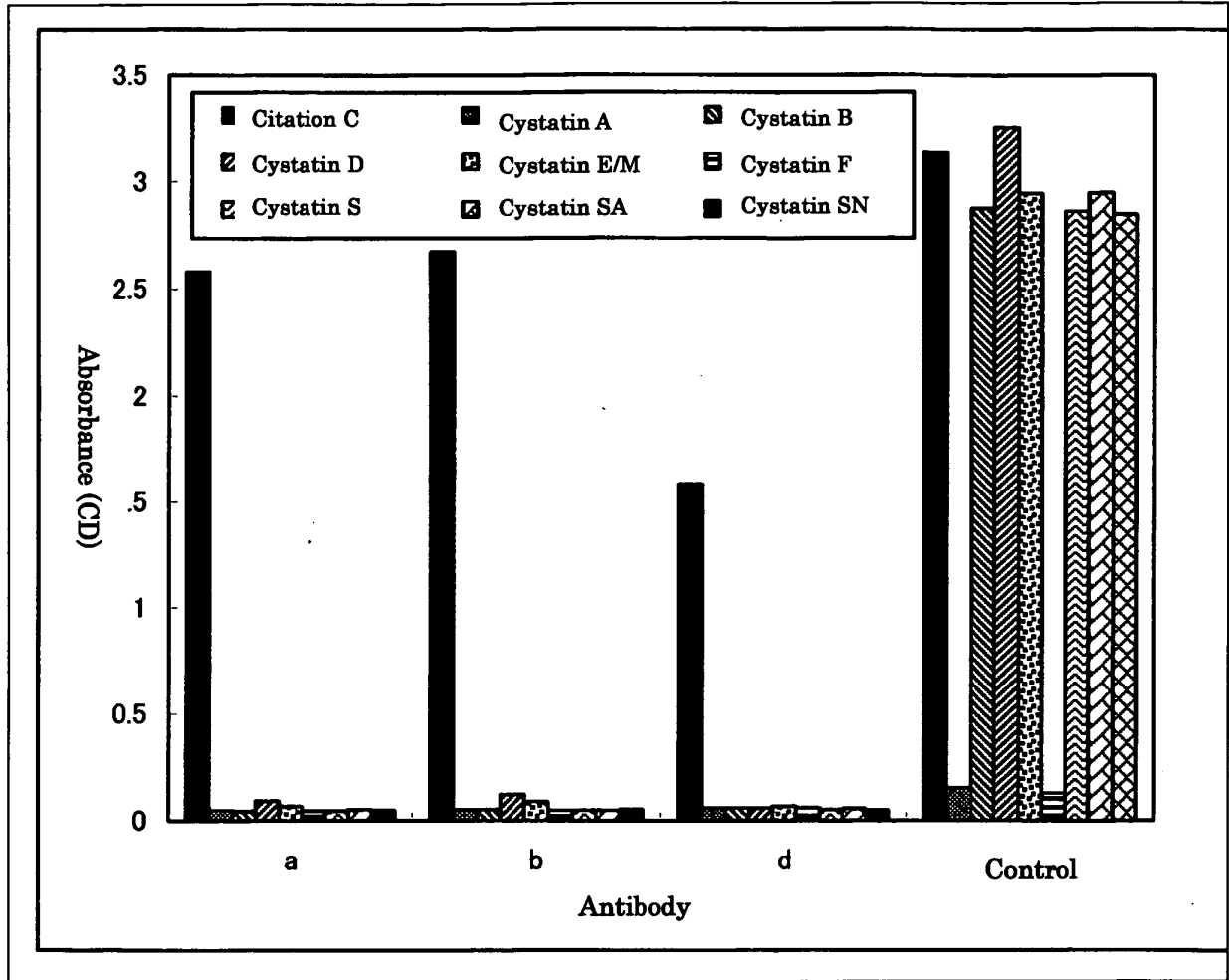


Fig. 3

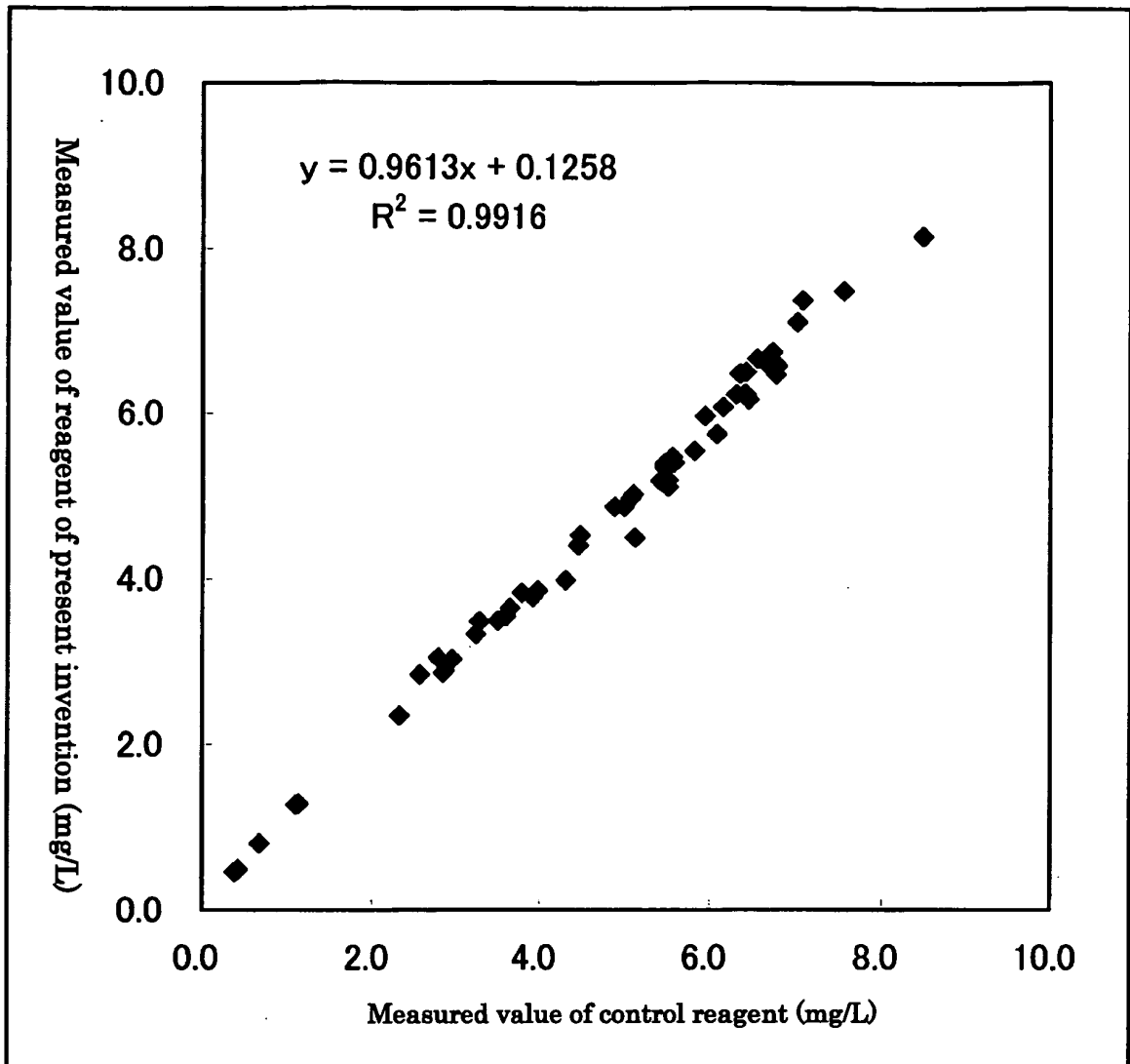
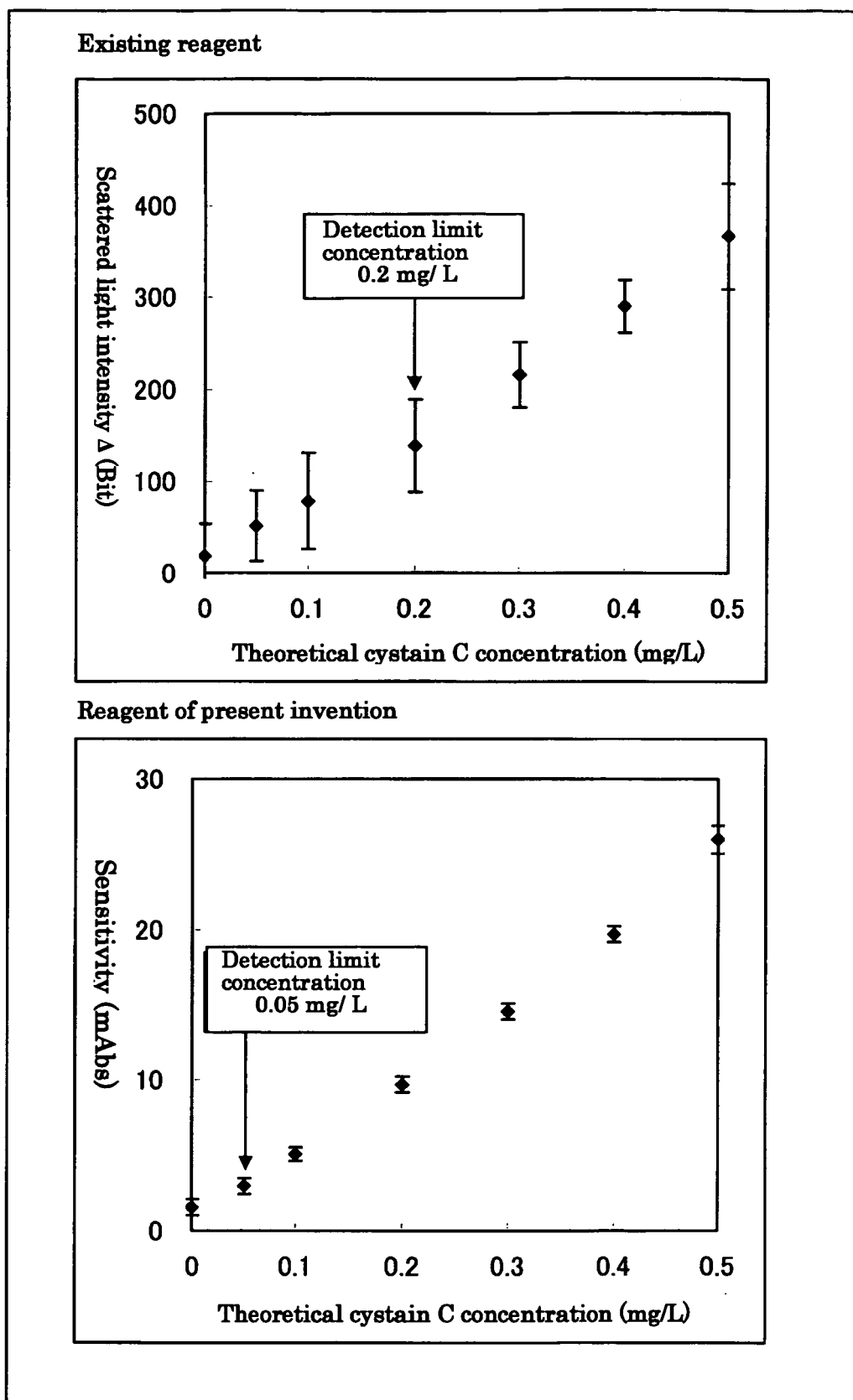


Fig. 4



**REFERENCES CITED IN THE DESCRIPTION**

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**Patent documents cited in the description**

- WO 2007102054 A [0004]

**Non-patent literature cited in the description**

- *CLINICAL CHEMISTRY*, 1994, vol. 40 (10), 1921-1926 [0005]
- *KIDNEY INTERNATIONAL*, 1995, vol. 47, 312-318 [0005]
- *CLINICAL CHEMISTRY*, 1997, vol. 43 (6), 1016-1022 [0005]
- *METHODS IN ENZYMOLOGY*, 1994, vol. 244, 685-700 [0005]

专利名称(译)	测量人体液中胱抑素c的方法		
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申请号	EP2009830205	申请日	2009-12-03
[标]申请(专利权)人(译)	积水医疗株式会社		
申请(专利权)人(译)	积水医疗CO., LTD.		
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其他公开文献	EP2357475A1 EP2357475A4		
外部链接	<a href="#">Espacenet</a>		

摘要(译)

本发明提供了一种用于人体液中胱抑素C的颗粒增强免疫测定方法，与使用大量具有低特异性的多克隆抗体或具有高单克隆抗体的常规测定方法相比，其具有更高的特异性并且易于以低成本自动化。特异性但凝集性差。本发明公开了一种特异于人体液中半胱氨酸蛋白酶抑制剂C的测定方法，该方法是利用抗体包被颗粒的组合的颗粒增强免疫测定方法，其中一些不溶性载体颗粒用一种类型的抗人胱抑素C单克隆抗体包被。其具有高亲和力并且其中一些其它不溶性载体颗粒被一种类型的抗人半胱氨酸蛋白酶抑制剂C单克隆抗体包被，其识别与第一种类型的单克隆抗体不同的表位并且具有相对低的亲和力，其中相对于用相应抗体包被的颗粒的总重量，抗人胱抑素C单克隆抗体的结合量小于5重量%。

Drawings

Fig. 1

