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(54) Title: IDENTIFICATION OF NOVEL SPLICE VARIANTS OF THE HUMAN CATALYTIC SUBUNIT C β OF cAMP-DEPENDENT PROTEIN KINASE AND THE USE THEREOF

(57) Abstract: The present invention demonstrates that the C β gene encodes at least 6 different gene products, designated C β 1, C β 2, C β 3, C β 4, C β 4ab and C β 4abc. As is the case with the murine and bovine splice variants, all the human C β splice variants vary in the N-terminal part preceding the part encoded by exon 2. Homologues to all C β splice variants identified in mouse and bovine were identified in human (C β 1, C β 2, C β 3, and C β 4) in addition to two novel C β splice variants (C β 4ab and C β 4abc), that have previously not been identified in any other species. The present invention includes in this respect genomic DNA- and cDNA sequences encoding said splice variants and comprises the nucleotide sequences shown in SEQ ID NO: 1, 2, 3, 4, 5 and 6 respectively. Wherein the said proteins are new splice variants of the C β protein. The present invention is further directed to vectors comprising said cDNA sequences. The invention also includes proteins characterised by the specific amino acid C β splice variant proteins shown in SEQ ID NO: 7, 8 and 9 respectively. The invention includes further use of the said C β splice variant proteins and DNA sequences in preparation of pharmaceuticals for diagnostic- and therapeutic purposes.



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IDENTIFICATION OF NOVEL SPLICE VARIANTS OF THE HUMAN CATALYTIC SUBUNIT C β OF cAMP-DEPENDENT PROTEIN KINASE AND THE USE THEREOF

FIELD OF THE INVENTION

The present invention relates to genomic- and complementary DNA sequences encoding the 6 different gene products, designated C β 1, C β 2, C β 3, C β 4, C β 4ab and C β 4abc which are novel splice variants of C β . The present invention also relates to vectors comprising said DNA sequences and is also directed to said proteins in diagnosis and treatment.

BACKGROUND OF THE INVENTION

Cyclic 3', 5'-adenosine monophosphate (cAMP) is a key intracellular signalling molecule, which main function is to activate the cAMP-dependent protein kinases (PKA) [1]. PKA consists of a heterotetramere, with a regulatory (R) subunit dimer and two catalytic (C) subunits. The holoenzyme is activated when four molecules of cAMP bind to the R subunit dimer, two to each R subunit, releasing two free active C subunits [2]. In man, four different R subunits (RI α , RI β , RII α , RII β), and four different C subunits (C α , C β , C γ and PrKX) have been identified [3]. The C α and C β subunits are expressed in most tissues, while the C γ subunit, which is transcribed from an intron-less gene and represents a retroposon derived from the C α subunit [4], is only expressed in human testis [5]. PrKX is an X chromosome-encoded protein kinase, and was recently identified as a PKA C subunit since it is inhibited by both PKI and RI α and the RI α /PrKX complex is activated by cAMP [6].

Splice variants of both C α and C β have been identified. The splice variants of C α have been termed C α 1 (previously named C α [7]), C α 2 [8] and C α -s [9]. Originally C α 2 was isolated from interferon-treated cells and identified as a C-terminally truncated C α 1 subunit. However,

recently a novel $C\alpha 2$ splice variant was reported [10]. The novel $C\alpha 2$ variant was shown to be identical to the previously identified $C\alpha$ splice variant, $C\alpha$ -s. Moreover, $C\alpha$ -s which was originally isolated and characterized from ovine sperm [9], has later been cloned from a human testis cDNA library and identified in human sperm [11]. Both $C\alpha$ -s/ $C\alpha 2$ are encoded with a truncated N-terminal end when compared to $C\alpha 1$. The variable parts of $C\alpha 1$ and $C\alpha$ -s are located upstream of exon 2 in the murine $C\alpha$ gene, implying that the variation in the N-terminal end of the $C\alpha 1$ and $C\alpha$ -s/ $C\alpha 2$ are due to alternative use of different first exons. In bovine, two splice variants of $C\beta$ have been identified, termed bovine $C\beta 1$ [12] and bovine $C\beta 2$ [13]. The bovine splice variants contain variable N-terminal ends in which the non-identical sequences are most probably encoded by different forms of exon 1. Bovine $C\beta 2$ is expressed at low levels in most tissues with the highest expression in the spleen, thymus, and kidney and to some extent brain. Furthermore, in the mouse, three splice variants of $C\beta$ have been identified and are designated mouse $C\beta 1$, m $C\beta 2$ and mouse $C\beta 3$ [14]. Whereas mouse $C\beta 1$ is ubiquitously expressed, mouse $C\beta 2$ and mouse $C\beta 3$ have so far only been identified in the brain. The mouse $C\beta 1$ and bovine $C\beta 1$ are similar in the entire sequence, demonstrating that they represent orthologous protein sequences. However, neither mouse $C\beta 3$ nor mouse $C\beta 4$ were similar to bovine $C\beta 2$ in the N-terminal part, indicating that their N-terminals are encoded by unrelated exons. Previous to this study, only a single splice variant of human $C\beta$ had been identified ($C\beta 1$), homologous to mouse $C\beta 1$ and bovine $C\beta 1$.

SUMMARY OF THE INVENTION

The present invention demonstrates that the $C\beta$ gene encodes at least 6 different gene products, designated $C\beta 1$, $C\beta 2$, $C\beta 3$, $C\beta 4$, $C\beta 4ab$ and $C\beta 4abc$. As is the case with the murine and bovine splice variants, all the human $C\beta$ splice variants vary in the N-terminal part preceding

the part encoded by exon 2. Homologues to all C β splice variants identified in mouse and bovine were identified in human (C β 1, C β 2, C β 3 and C β 4) in addition to two novel C β splice variants (C β 4ab and C β 4abc), that have previously not been identified in any other species. The present invention includes in this respect genomic DNA- and cDNA sequences encoding said splice variants and comprises the nucleotide sequences shown in SEQ ID NO: 1,2,3,4,5 and 6 respectively. Wherein the said proteins are new splice variants of the C β protein. The present invention is further directed to vectors comprising said cDNA sequences. The invention also includes proteins characterised by the specific amino acid C β splice variant proteins C β 2, C β 4ab and C β 4abc shown in SEQ ID NO: 7,8 and 9. The invention includes further use of the said C β splice variant proteins and DNA sequences in preparation of pharmaceuticals for diagnostic- and therapeutic purposes.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1: A: Identification of cDNAs encoding human C β splice variants. Schematic representation of the protein-encoding sequences of the various C β splice variants found in human. Human cDNAs from total fetus and brain were amplified using primers complementary to the C β cDNA, subcloned and sequenced. The resulting cDNAs were identical to the previously published C β cDNA (C β 1) downstream of nucleotide 46 (constant region). However, five novel cDNA sequences, designated C β 2, C β 3, C β 4, C β 4ab and C β 4abc, could be identified based on differences in the 5'-ends of the sequences (variable region).

Figure 2: A: Structure of the human genomic region encoding the novel C β splice variants. Primers were made based on exon 2 and the most 5'-end of the different C β cDNAs, and used to amplify human genomic DNA by PCR. Two overlapping PCR products of 14 and

17 kb, respectively, were identified and mapped by Southern blotting and hybridization to oligonucleotides corresponding to the different cDNAs. As derived from the 14 and 17 kb PCR products, exon 1-2 1-3, 1-4 and exon a, b and c are located 31, 14.1, 14, 8.1, 5.4 and 4.4 kb upstream of exon 2. Based on restriction mapping of the PAC clone RPCI-6-228E23, exon 1-1 is located approximately 60 kb upstream of exon 1-2. Exon 1-1 is specific for the splice variant, which encodes CB1. The exons are indicated as vertical lines. The introns are drawn to scale as indicated. **B: Nucleotide sequence of genomic regions encoding novel splice variants of C β .** Protein encoding sequences are in capital letters, intron and 5'-untranslated sequences are in lower case letters. Translation initiation codons are underlined. Only the 5'-end of exon 2 is included. **C: Schematic representation of how the various human C β exons 5'to exon 2 may be spliced.** The upper panel describes a potential model in which four variants of exon 1 designated exon 1-1, 1-2, 1-3 and 1-4 may alternatively splice with exon 1 to encode the splice variant specific sequence in C β 1, C β 2, C β 3 and C β 4. The lower panel describes a model in which the exons a, b and c may splice with exon 1-4 and 1-3 upstream of exon 2 to encode the splice variant-specific sequences in C β 4ab, C β 4abc and C β 3ab.

Figure 3: Deduced amino acid sequence of C β splice variants. The amino acid sequences of the amino terminal parts of C β 1 and five new splice variants, designated C β 2, C β 3, C β 4, C β 4ab and C β 4abc according to the cDNA clones shown in figure 1A. The amino acid sequences are shown in the one letter code and demonstrate that six novel C β exons give rise to five different cDNAs as a result of alternative promoter use and alternative splicing. The myristylation motive G-N previously identified in C β 1 is boxed. A PKA autophosphorylation motive that has previously been identified in C β 1, is underlined and Ser10 which is potentially

phosphorylated, is labeled by an asterisk. Note that there is a PKA autophosphorylation motif, encoded by exon a, present in C β 4ab and C β 4abc.

Figure 4: Tissue distribution of different C β splice variants. Northern blots containing various human tissues were hybridized using probes specific for C β 1, C β 2, C β 4, exon a+b and a probe common to all C β splice variants (C β common). For comparison, the same blots were hybridized using a GAPDH cDNA (GAPDH). All C β mRNAs had the same apparent length (4.4 kb).

Figure 5: A: Species distribution of C β 2. A Southern blot containing EcoRI digested genomic DNA from various species was hybridized using a DNA probe corresponding to exon 1-2 (C β 2 specific). A single hybridizing band identifying genomic sequence homologous to human exon 1-2 was identified in mammals such as monkey, dog, rabbit and human except mouse and rat. **B: C β 2 is not expressed in the mouse.** A Northern blot containing total RNA (20 μ g pr. lane) isolated from wild type (+/+) mouse brain and spleen (lane 1 and 3), brain and spleen of mice ablated (-/-) for C β 1 (lane 2 and 4) and human peripheral blood leukocytes (lane 5) was probed with a C β probe expected to recognize all known C β splice variants (C β Common, upper panel) and a C β probe specific for the C β 2 splice variant (C β 2, lower panel). Messenger RNA recognized by the two probes is indicated as 4.4 kb.

DETAILED DESCRIPTION OF THE INVENTION

The present invention demonstrate that the human C β gene encodes five novel C β splice variants, designated C β 2, C β 3, C β 4, C β 4ab and C β 4abc, in addition to the previously identified splice variant C β 1 [12]. All the C β splice variants contained a unique N-terminal end, and showed tissue specific expression. As we found no evidence of an additional exon upstream of exon 1-1 and all the cDNA characterized had unique 5'-ends, it is reasonable to assume that the exon 1-1, 1-2, 1-3 and 1-4 each contain a separate promoter, and that the resulting mRNA products are due to alternative use of different promoters. Despite this, we can not rule out the possibility that two or more of these splice variants share a common promoter used to alternatively splice the different exons. Furthermore, we found two C β variants, C β 4ab and C β 4abc, that were the results of alternative splicing of either exon a and b, or exon a, b and c, between exon 1-4 and exon 2. The presence of the corresponding mRNA was confirmed by hybridizing a Northern blot with a probe complimentary to the sequences found in exons a and b. This probe and the probe specific for C β 4 bound to an RNA with the same apparent length located in human brain. The location of the exons a, b and c may suggest that they generate splice variants of C β in addition to those demonstrated here. Indeed, a short cDNA from human infant brain have been sequenced and demonstrated to contain a combination of exons 1-3, a, b and 2 (Accession no. AA351487, see Fig. 2C). We were unable to produce such a cDNA, which could be due to low level expression of C β 3 in adult brain.

The two splice variants C α 1 and C β 1 are highly conserved in the parts encoded by exon 1, differing in only 2 of the first 16 amino acids [7;12]. It is therefore tempting to suggest that this region serve a specific role in the function of these splice variants. Thus, the fact that we have identified several C β splice variants with variable N-terminal ends could suggest that the N-terminal domain might reflect specific functional features associated with each splice

variant. This is supported by studies of the mouse C β 1 KO mouse, which displayed impaired hippocampal plasticity [16]. However, to what extent N-terminal differences influence catalytic activity is not known since it was shown that the N-terminally truncated C β splice variants in mouse, C β 2 and C β 3 were catalytically active, an activity that was inhibited both by PKI and the R subunit *in vivo* [14]. In addition, a study by Herberg et al [17] showed that deleting amino acids 1-14 in the C α isoform did not influence catalytic activity, demonstrating that the N-terminal specific for the C α 1/C β 1 is not necessary for catalytic activity.

The N-terminal of C α 1 and C β 1 contain two sites for post-translational modification, a myristylation site and an autophosphorylation site [5;18;19]. In C α 1, C β 1 and C β 3 the N-terminal amino acid is G (Gly) which has been shown as an absolute requirement for myristylation [20]. Despite this, it was previously demonstrated in the mouse that C β 3 does not undergo myristylation *in vivo* [14]. This phenomena may be explained based on a recent study, demonstrating that the amino acid C-terminal to G must be N if myristylation shall occur. This because deamination of N to yield D is an absolute requirement [21]. Because the amino acid C-terminal to G is L in both mouse and human C β 3, it explains why mouse C β 3 is not myristylated and suggests that the human C β 3 may not be myristylated *in vivo*.

The fact that several human C β splice variants (C β 2, C β 3, C β 4, C β 4ab and C β 4abc) lack the ability to become myristylated *in vivo*, question the role of this post translational modification. Based on the C α crystal structure it appears that the myristyl group serves to fill and shade a hydrophobic pocket in the large lobe [22], suggesting that this N-terminal modification serves to solubilize the C subunit. This is supported by two independent observations. Firstly, expression of an N-terminally truncated form of C α 1 revealed a C subunit tightly associated with the particulate fraction [23]. Secondly, the C α -s/C α 2 which is a naturally occurring N-terminally truncated splice variant is tightly associate with sub cellular

structures in both ovine- [9;24] and human [11] sperm. This taken together with a recent report, which demonstrated that the myristyl group serves to increase the lipophilic properties of the C subunit when binding the RII- but not the RI subunit [25], suggests that the N-terminal amino acids of C α 1 together with myristylation serves to influence C subunit solubility. Thus, the sequence similarity between C α 1 and C β 1 and the difference in solubility of C α 1 and C α -s/C α 2, may imply comparable difference in solubility between C β 1 and the truncated C β forms.

Previously a consensus autophosphorylation motif (-KKGS¹⁰ -) was identified in C α 1 and C β 1 [12;26], that is phosphorylated when C α 1 is expressed in bacteria [18;23]. In the study by Yonemoto et al. (1993) mutation of S¹⁰ yielded an insoluble enzyme that appeared inactive. Thus, the N-terminal domain may also have implications for catalytic activity by an unknown mechanism. However, like the human C β 2, C β 3, C β 4, the mouse C β 2 and C β 3 lack S¹⁰, yet these splice variants are soluble and catalytically active *in vivo* [14]. This suggests that the human homologues most probably are active and may imply that S¹⁰ phosphorylation is not crucial for C subunit catalysis. Interestingly, we identified a potential autophosphorylation site (-RKSS⁶-) in C β 4ab and C β 4abc that was encoded by exon a. To what extent this site represents a true autophosphorylation site that will influence C β 4ab and C β 4abc properties, remains to be seen.

The human C β 2 splice variant was similar to the previously identified bovine C β 2 splice variant, but we have been unable to identify a similar splice variant in mice. Interestingly, the human C β 2 splice variant is expressed only in peripheral tissues, while no detectable C β 2 mRNA signal is found in human brain. However, no C β can be detected outside the brain in mice lacking the C β 1 splice variant [14;16]. In addition, we were unable to detect any signal

when hybridizing mouse DNA using a human C β 2 specific probe. Thus, it is likely that mice do not contain a homologue of the human and bovine C β 2 splice variants.

Interestingly, C β 2 is the most atypical of the C β splice variants. This subunit is encoded with an extended N-terminal domain, which do not resemble any of the other C β splice variants. The unique domain together with the fact that C β 2 lacks the myristylation- as well the autophosphorylation site, and that C β 2 is the only C β splice variant not identified in the brain, may suggest specific and unique features associated with this splice variant in other tissues that will await further studies.

The inventors suggest that tissue-specific expression of various C β splice variants when complexed with R subunits may imply novel PKA holoenzymes with specific functional features that may be important as mediators of cAMP effects.

The present invention includes in this respect genomic DNA- and cDNA sequences encoding splicevariants C β 1, C β 2, C β 3, C β 4, C β 4ab and C β 4abc and comprises the nucleotide sequence shown in SEQ ID NO: 1,2,3,4,5 and 6 respectively. Wherein the said proteins are new splice variants of the C β protein. The present invention is further directed to vectors comprising said cDNA sequences. The invention also includes proteins characterised by the specific amino acid C β splice variant proteins; C β 2, C β 4ab and C β 4abc shown in SEQ ID NO: 7,8and 9 respectively. The invention includes further use of the said C β splice variant proteins and DNA sequences in preparation of pharmaceuticals for diagnostic- and therapeutic in order to identify, characterize and produce pharmacological compositions .

C β 2 is an enzyme that is expressed in lymphoid cells, whereby its function is to mediate the regulatory effects of cAMP on T cell activation. Thus, altered levels, location and/or activity of C β 2 will according to the inventors results, have impact on the regulation and normal function

of receptors and enzymes which are important for T cell activation and are regulated by cAMP. This knowledge can be used to diagnose hyperreactive and dysfunctional T cells associated with various immune diseases.

1) *Malfunctioned T cells:* It is well known that T cells isolated from patients suffering from T cell-dependent common variable immune deficiency (CVI) and acquired immune deficiency syndrome (AIDS) do not respond to antigen. Furthermore, T cells isolated from patients suffering from certain types of rheumatoid arthritis and other auto immune diseases are hyper sensitive to foreign antigens. In both cases these situations evoke abnormal immune responses that may involve malfunctioned C β 2. This may either be monitored as constitutively activated C β 2, sub-normal activity or dislocation of C β 2.

1.1) *Improving T cell dysfunction:* Present invention makes it possible to identify, characterize and produce pharmacological compositions after high through put screening that specifically will inhibit the enzymatic activity of C β 2. These compositions should be developed such that they can be introduced orally or intra venously to enter the blood system reaching the dysfunctional T cells.

Furthermore, dislocation of C β 2 protein from the T cell membrane will short cut the regulatory effects of C β 2 on relevant receptors. Thus, the present invention makes it possible to identify, characterize and produce pharmacological composition after high through put screening that will specifically and irreversibly block C β 2 interaction with the T cell membrane. These compositions should be developed such that they can be introduced orally or intra venously to enter the blood system reaching the T cell.

1.2) *Down regulation of hyper active T cells:* Present invention makes it possible to identify, characterize and produce pharmacological compositions after high through put screening that specifically will activate the enzymatic activity of C β 2. These compositions should be

developed such that they can be introduced orally or intra venously to enter the blood system reaching the dysfunctional T cells.

1.3) Kits for diagnosing C β 2 mutations: T cell malfunction caused by mal function or - localization of C β 2 enzyme activity may be caused by mutation(s) in the C β 2 protein. Present invention makes it possible to develop kits, which would diagnostically facilitate if mutated C β 2 is present. Such kits should be developed with C β 2 specific DNA probes.

Present invention makes it possible to develop a method for inspection and screening of patient T cells for the presence and location of C β 2 comprising:

- a) collection and washing in buffer of isolated peripheral blood T lymphocytes according to [27];
- b) preparing for identification of C β 2 protein by immunofluorescence, T cells are let to settle onto poly L-lysine coated cover slips following detergent-dependent lysis;
- c) incubation with primary antibody (Ab), either irrelevant Ab or C β 2 specific Ab, Ab overshoot will be removed by washing buffer and T cells incubated with secondary anti-IgG Ab conjugated with a fluorescent;
- d) inspection of T cells under fluorescent microscopy.

Present invention makes it further possible to develop a method of screening patient T cells for membrane associated C β 2 catalytic activity comprising:

- a) collection and washing in buffer of isolated peripheral blood T lymphocytes according to [27];
- b) preparation of T cells by lysing in detergent buffer;
- b) monitoring C β 2 specific catalytic activity by established assay, C β 1 activity is used as an internal control to determine relative activity.

Present invention makes it also possible to screen patients for mutations in the C β 2 gene and mRNA comprising:

- a) collection and washing in buffer of isolated peripheral blood T lymphocytes according to [27];

b) isolation of total RNA and genomic DNA according to established methods followed by RT-PCR using C β 2 specific primers according to cDNA sequence of C β 2 specific nucleotides or the C β 2 specific exon, designated exon 1-2.

Materials and methods.

General protocols

Complementary DNA probes were radiolabeled using the Megaprime random priming kit and α -[32P]dCTP (Amersham) as instructed by the manufacturers to a specific activity of at least 1×10^9 cpm. Synthetic oligonucleotides were radiolabeled using T4 polynucleotide kinase (Pharmacia) and γ -[32P]ATP as instructed by the manufacturer.

DNA was either sequenced manually using Thermo Sequenase radioabeled terminator cycle sequencing kit (Amersham, Buckinghamshire, UK) or by Medigenomix (Martinsried, Germany). Sequences were analyzed using the Wisconsin University GCG program package (UWGCG) and the basic local alignment and search tool (BLAST) [15].

Identification of cDNAs

The 5'-end of human C β cDNA was amplified from human total fetus and brain Marathon RACE-ready cDNAs (Clontech) using the Advantage KlenTaq Polymerase Mix (Clontech) as described by the manufacturer. Amplification was performed using adapter primer 1 (Clontech) and four different primers complementary to the human C β cDNA sequence (5'-CAACCCAAAGAGAAGTAAGAAAGTGGTCTA-3',
5'-TTGGTTGGTCTGCAAAGAATGGGGGATAGC-3',
5'-TTTTCTCATTCAAAGTATGCTCTATTTGC-3' and
5'-AGAATAATGCCGACTTGAAGATTTTGAAA-3').

Five cycles were performed with 45 sec 94 °C, 2 min 72 °C, five cycles 45 sec 94 °C, 2 min 70 °C, 25 cycles 45 sec 94 °C, 2 min 68 °C, and a final extension of 10 min at 72°C. The resulting products were separated by gel electrophoresis, subcloned to pCR2.1TOPO (Invitrogen) as instructed by the manufacturer and sequenced.

Amplification of C β gene fragments.

A genomic fragment was amplified using an oligonucleotide corresponding to exon 1-3 (5'-GTTTAGGTGCAATCATTCTGCTGTTTG-3') and a primer complementary to sequences in exon 2 (5'-AAAAAGTCTTCTTTGGCTTTGGCTAGA-3'). Another genomic fragment was amplified using a primer corresponding to exon 1-2 (5'-TGGCAGCTTATAGAGAACCACCTT-3') and a primer complementary to sequence found in exon 1-3 (5'-CAATCCCATGTTGAACCTGGCA-3'). PCR reactions were performed using the Boehringer-Mannheim Expand Long Template PCR kit as instructed by the manufacturer using buffer 2. PCR was performed using human genomic DNA (Boehringer-Mannheim) as template with 1 min at 92 °C, 30 cycles of 10 sec 94 °C, 30 sec 60 °C and 10 min (extended with 20 sec per cycle from cycle 11 to cycle 30) 68 °C, and a final incubation of 7 min at 68 °C. Products were separated by agarose gel electrophoresis and analyzed by Southern blotting using radiolabeled cDNAs and synthetic oligonucleotides corresponding to the different exons.

Screening of PAC library and subcloning of exon-containing sequences.

The human P1-derived Artificial Chromosome (PAC) library, RPCI-6 was screened and the isolated bacterial clone was grown in liquid culture and plasmid DNA was isolated using ion-exchange columns as described by the manufacturer (Qiagen, Hilden, Germany). Exon-containing DNA restriction fragments were identified by Southern blotting using radio labeled

cDNAs and synthetic oligonucleotides. Exon-containing fragments were excised from the gel and subcloned to the pZERO2.1 vector (Invitrogen) as instructed by the manufacturer.

Generation of splice variant specific probes, Northern blotting and Southern blotting.

DNA fragments corresponding to the splice variant-specific parts of the cDNAs were amplified by PCR. The following primers were used for the different splice variants:

C β 1: 5'-GCTCTCCACCTCGCTGCCTTTCTT-3' and primer

5'-CCAGCCCCCTTCCCTTCCCTGAC-3',

C β 2: primer 5'-TGGCAGCTTATAGAGAACCACCTT-3' and primer

5'-ATTGATCTGTCCATAAGGCAGTAT-3',

C β 3: primer 5'-TCACAGCTAGCAGTAAGAGCTG-3' and primer

5'-CAATCCCATGTTGAACCTGGCA-3',

C β 4: primer 5'-TCTCCAGTGTGTGTGTTTACAC-3' and primer

5'-ATGATGAAAACCAACCTTTCCA-3'.

The primers were used for amplification of the fragments from cloned RACE-products using Taq DNA polymerase (Perkin-Elmer) as described by the manufacturer. For generation of a probe specifically recognizing exon a and b, the primers

5'-GATATTTCTGAAGAGGAGCAAGCAGATGCATCTGATGATTTGCGTG-3' and 5'-

CACGCAAATCATCAGATGCATCTGCTTGCTCCTCTTCAGAAATATC-3' were

annealed, phosphorylated and ligated. A 1.5 kb fragment of C β cDNA [5] was used for recognizing the parts of the C β mRNA common to all splice variants. Two similar Northern blots containing RNA from various human sources were purchased from Clontech. One blot was hybridized using a probe specific for C β 2, while the other blot was probed in succession with probes specific for C β 3, C β 4, exon a and b, and the 1.5 kb C β cDNA. Both blots were hybridized using GAPDH cDNA as control. As an almost identical pattern of hybridization was

obtained using GAPDH on both blots, only one GAPDH blot is shown (Fig. 4). All probes were hybridized in ExpressHyb hybridization solution (Clontech) as described by the manufacturer. A Southern blot containing EcoRI-digested DNA from various species (Clontech) and Southern blots containing human and mouse DNA digested with various enzymes were hybridized using the probe specific for C β 2. The filters were prehybridized in 5 X Denhardt's solution, 5 X SSC, 50 mM sodium phosphate buffer, pH 6.8, 0.1 % SDS, 250 μ g/ml single stranded salmon sperm DNA, and 50 % (v/v) formamide at 42 °C for 3 h, and hybridized for 16 h in a similar solution containing the radiolabeled C β common or C β 2 probe. The membranes were washed four times in 2 X SSC, 0.1 % SDS for 5 min at room temperature, followed by two washes using 0.5 X SSC, 0.1 % SDS at 50°C for 30 min. Autoradiography was performed at -70°C using Amersham Hyperfilm MP and intensifying screens.

In order that this invention may be better understood, the following examples are set forth. These examples are for the purposes of illustration only, and are not to be construed as limiting the scope of the invention in any manner.

Examples

Example 1

Identification of exons encoding novel splice variants of human C β .

The 5'-ends of human C β cDNAs were amplified from human brain and total fetus RACE-ready cDNA using four different oligonucleotide primers complementary to the previously published human C β cDNA sequence, in combination with an anchor primer. The resulting PCR products were subcloned, sequenced and compared to the previously published human C

β cDNA sequence which is now designated C β 1 (Fig. 1). All clones sequenced were shown to lack the 46 first protein-encoding nucleotides in the human C β 1 cDNA sequence. Instead 5 novel stretches of protein encoding sequences were identified (Fig. 1, variable region). Each of the clones contained a translation initiation codon and one or more in-frame upstream stop codons. The five novel cDNA sequences were designated C β 2, C β 3, C β 4, C β 4ab and C β 4abc.

All the C β cDNAs were similar from nucleotide 47 and down stream in the C β 1 cDNA, which corresponds to the start of exon 2 in the murine C β gene. The identification of novel protein-encoding sequences upstream of exon 2, indicated the presence of several different exons upstream of exon 2. Thus, human genomic DNA was amplified using a combination of primers corresponding to exon 2 (antisense orientation) and the 5'-ends of the different novel cDNAs (sense and antisense orientation) in different combinations. A 17 kb PCR product was the result of an amplification using a primer corresponding to the 5'-end of C β 2 cDNA (sense orientation) and the 5'-end of C β 3 (antisense orientation). Furthermore, a 14 kb PCR product was the result of an amplification using a primer corresponding to the 5'-end of C β 3 cDNA (sense orientation) and a primer corresponding to exon 2 (antisense orientation). These clones enabled us to physically map six novel exons in the C β gene that were designated 1-2, 1-3, 1-4, a, b and c, and which were located 31, 14.1, 14, 8.1, 5.4 and 4.4 kb upstream of exon 2, respectively (Fig. 2A). Furthermore, a PAC library was screened using the 5' ends of C β 1 and C β 2 cDNAs as probes. One of the clones identified, RPCI-6-228E23, contained both exon 1-2 and an exon containing the entire splice variant-specific part of the C β 1 cDNA, which we termed exon 1-1. This PAC clone was selected for detailed restriction mapping using CpG cutters. The digested PAC DNA was separated by pulsed-field gel electrophoresis (PFGE), transferred to Southern blot membranes and hybridized with exon 1-1 and 1-2, as well as Sp6 and T7 oligonucleotide probes. These results revealed a distance of

approximately 60kb between exon 1-1 and 1-2 (Fig. 2A). All nucleotide sequences found in the different C β cDNAs could be identified in a continuous stretch of human genomic DNA, thereby supporting the notion that these cDNAs are products of the same gene. Exon 1-1 was shown to be homologous to the previously identified exon 1A of the murine C β gene. As shown in Fig. 2B, exon 1-2 contains the entire C β 2 specific sequence, and exon 1-3 contains the sequence specific for C β 3 which is homologous to the previously identified exon 1B in the mouse C β gene. Finally, exon 1-4 was shown to contain the sequence specific for the human C β 4 splice variant, and to be homologous to the murine exon 1C, which encodes the N-terminal end in the murine C β 2 splice variant. Based on the C β 4ab and C β 4abc cDNA sequences, the exons a, b and c (Fig. 2B), were demonstrated to be alternatively spliced in between exon 1-4 and exon 2, with either exons 1-4, a, b and 2 or exons 1-4, a, b, c and 2 (Fig. 2C, lower panel). These cDNA sequences represent novel C β splice variants not identified in any other species.

Example 2

Deduced amino acid sequence of novel C β splice variants.

The N-terminal parts of the deduced amino acid sequences of the previously published C β 1-sequence and the 5 novel C β splice variants are illustrated in Fig. 3 (upper and lower panels). The splice variants were identical starting from the sequence encoded by exon 2 (amino acid 17 in C β 1) to the C-terminus, while the N-termini varied both in length and sequence composition. The C β 2 splice variant contains a 63 amino acid sequence substituting the first 16 amino acids in C β 1, and is homologous to the previously identified bovine C β 2 [13]. Furthermore, the human C β 3 splice variant contains four amino acids in the N-terminal substituting the first 16 amino acids in C β 1, and is similar to the previously identified murine

C β 3 [14]. The human C β 4 contains three amino acids substituting the first 16 amino acids in C β 1, and is similar to the murine C β 2 [14]. Finally, the splice variants C β 4ab and C β 4abc contain 18 and 21 amino acids, respectively, that substitute the first 16 amino acids of C β 1. These splice variants show no homology to the N-terminus of any other C subunits identified thus far.

Example 3

Tissue distribution of C β splice variants.

To examine the tissue distribution of C β splice variants, exon specific DNA probes and a DNA probe common to all C β splice variants were hybridized to two similar Northern blots containing RNA from various human tissues. For comparison the blots were hybridized to a cDNA encoding glycer-aldehyde 3-phosphate dehydrogenase (GAPDH). In Fig. 4 (panel C β 1) we show that C β 1 is predominantly expressed in brain and kidney with low level expression in several other tissues as well. C β 2 is expressed at high levels in thymus, spleen and kidney in addition to a weak signal in other tissues (Fig. 4, panel C β 2). In contrast to C β 2 the exon 1-4 and exon a and b containing mRNAs appeared to be present exclusively in brain (Fig. 4, panels C β 4 and exon a+b). Finally, probing the Northern blot with a probe common to all the C β splice variants, we observed ubiquitous expression of C β with the strongest signal in brain and a somewhat weaker signal in spleen and thymus, when compared to the GAPDH signal (Fig. 4, panel C β common). Hybridization using a DNA fragment corresponding to the C β 3 specific cDNA resulted in an almost undetectable signal in the brain and no detectable signals in any other tissues (data not shown).

Example 4**The human C β 2 splice variant is not present in the mouse.**

Previously we have identified three splice variants of C β in the mouse, C β 1, C β 2 and C β 3 [14]. Based on the present work, it is apparent that mouse C β 2 is not homologous to either bovine or the human C β 2. Instead, mouse C β 2 is homologous to what we now have designated human C β 4. Thus, we investigated whether a C β splice variant similar to human C β 2 was present in the mouse genome. A Zoo-blot containing genomic DNA isolated from human, monkey, rat, mouse, dog, cow, rabbit, chicken and yeast was hybridized using a DNA fragment corresponding to exon 1-2 of human C β . In Fig. 5 (panel A, lanes 1 to 9) we show that a DNA fragment was detected using C β 2 specific probe in man, monkey, dog, cow, and rabbit. In contrast, the C β 2 specific probe did not recognize any fragments in the rat and mouse suggesting that the C β 2 specific exon is not present in the murine genome. To further substantiate this observation we isolated total RNA from human, wild type mice and mice that are ablated (knockout, KO) for exon 1A of the C β gene [16]. The RNA was isolated from immune tissues and brain since we observed high level expression of C β 2 in human thymus, spleen and peripheral blood leukocytes and high level of the other C β splice variants in the brain (Fig. 4). The Northern blots were probed with a C β cDNA probe (expected to recognize all known C β splice variants) and a C β 2 specific probe (see material and methods). In Fig. 5B (upper panel) we demonstrate that C β is present in the brain of wild type and C β exon 1 KO (lanes 1 and 2) and in human peripheral blood leukocytes (lane 5). The mouse spleen did not contain C β mRNA (lanes 3 and 4). When probing the same filter with the C β 2 specific probe

(Fig. 5, lower panel) C β 2 message was only detected in human peripheral blood leukocytes (lane 5) whereas all the mouse tissues were negative for C β 2 mRNA (lanes 1 to 4).

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Patent Claims.

1.

The genomic DNA sequence encoding the C β 1, C β 2, C β 3, C β 4, C β 4ab and C β 4abc splice variant proteins respectively comprising the nucleotide sequence of SEQ ID NO: 1, wherein said proteins are new splice variants of the catalytic sub unit of c-AMP dependent protein kinase termed C β .

2.

The cDNA sequence encoding the C β 1, C β 2, C β 3, C β 4, C β 4ab and C β 4abc splice variant proteins respectively comprising the nucleotide sequences of SEQ ID NOs: 2,3,4,5 and 6, wherein said proteins are new splice variants of the catalytic sub unit of c-AMP dependent protein kinase termed C β .

3.

The vectors comprising the DNA sequences according to claims 1 or 2.

4.

The specific amino acid sequences of SEQ IDNOs 7,8 and 9 of C β 2, C β 4ab and C β 4abc respectively.

5.

A protein encoded by the nucleotide sequences according to claims 1 or 2.

6.

A protein encoded by the specific DNA sequences according to claims 1 or 2 comprising the specific amino acid sequence of SEQ ID NO: 7,8 and 9.

7.

A kit comprising C β 2 specific DNA probes.

8.

The use of the C β 2, C β 4, C β 4ab and C β 4abc proteins for the preparation of pharmaceuticals.

9.

The use of C β 2 protein for the preparation of a medicament for inhibition of the enzymatic activity of C β 2.

10.

The use of C β 2 protein for the preparation of a medicament that will specifically and irreversibly block C β 2 interaction.

11.

The use of C β 2 protein for the preparation of a medicament that will activate the enzymatic activity of C β 2.

12.

The use of the DNA sequences which is complementary to the C β 1, C β 2, C β 3, C β 4, C β 4ab and C β 4abc DNA according to claims 1 or 2 for the preparation of an anti sense drug.

13.

A Method for inspection and screening of patient T cells for the presence and location of C β 2 comprising:

- a) collecting and washing in buffer of isolated peripheral blood T lymphocytes;
- b) preparing for identification of C β 2 protein by immunofluorescence, T cells are let to settle onto poly L-lysine coated cover slips following detergent-dependent lysis;
- c) incubating with primary antibody (Ab), either irrelevant Ab or C β 2 specific Ab, Ab overshoot will be removed by washing buffer and T cells incubated with secondary anti-IgG Ab conjugated with a fluorescent;
- d) Inspection of T cells under fluorescent microscopy.

14.

A method of screening patient T cells for membrane associated C β 2 catalytic activity comprising:

- a) collecting and washing in buffer of isolated peripheral blood T lymphocytes;
- b) preparing of T cells by lysing in detergent buffer;
- b) monitoring C β 2 specific catalytic activity by established assay, C β 1 activity is used as an internal control to determine relative activity.

15.

A method for screening of patients for mutations in the C β 2 gene and mRNA comprising:

- a) collecting and washing in buffer of isolated peripheral blood T lymphocytes;
- b) isolating of total RNA and genomic DNA according to established methods followed by RT-PCR using C β 2 specific primers according to cDNA sequence of C β 2 specific nucleotides or the C β 2 specific exon, designated exon 1-2.

16.

A product produced by the method, according to claim 13, 14 and 15.

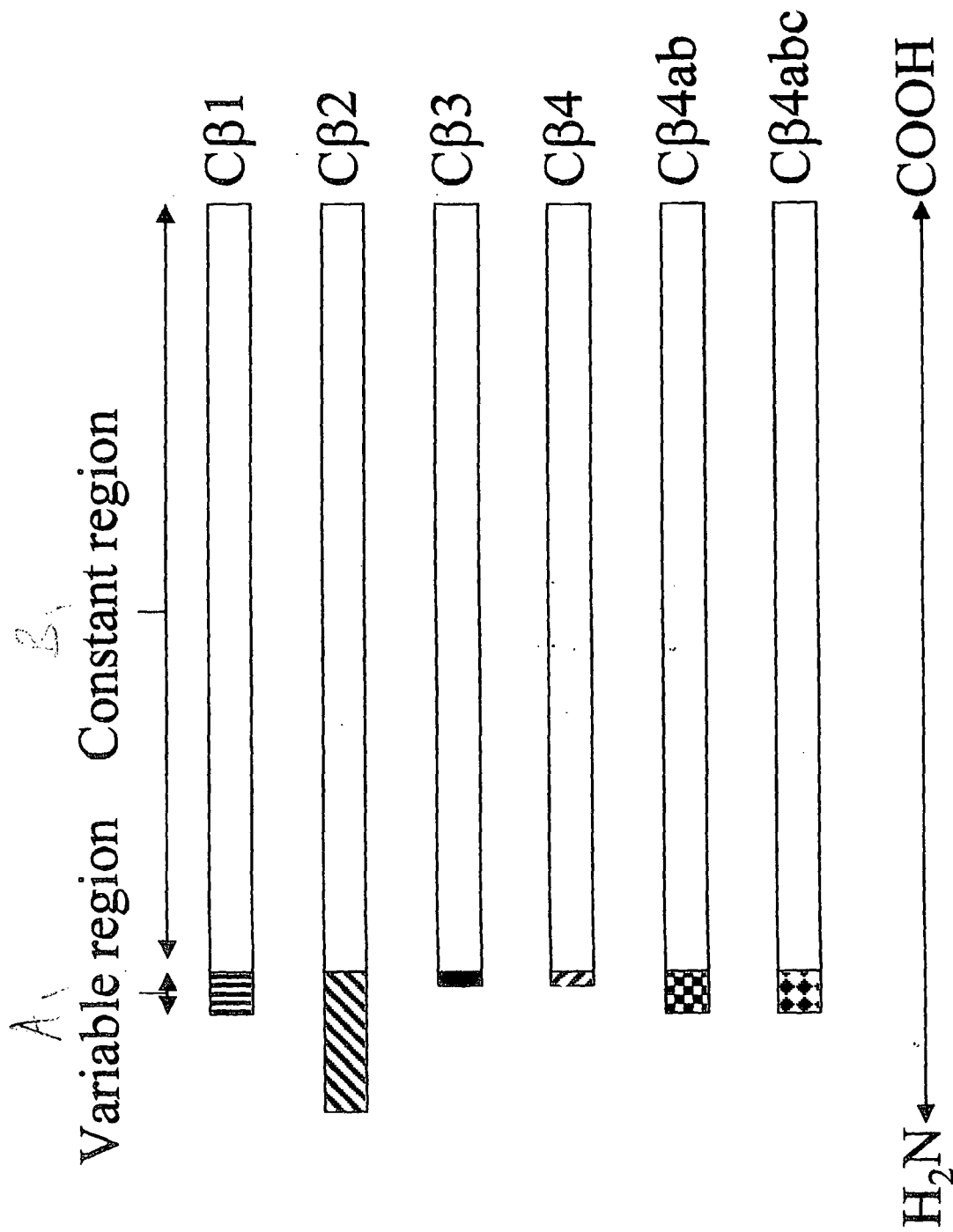
17.

A test system for screening for inhibitory- or activating molecules of the C β 2 protein.

18.

The product from the screening method according to claim 17.

FIGURE 1



A

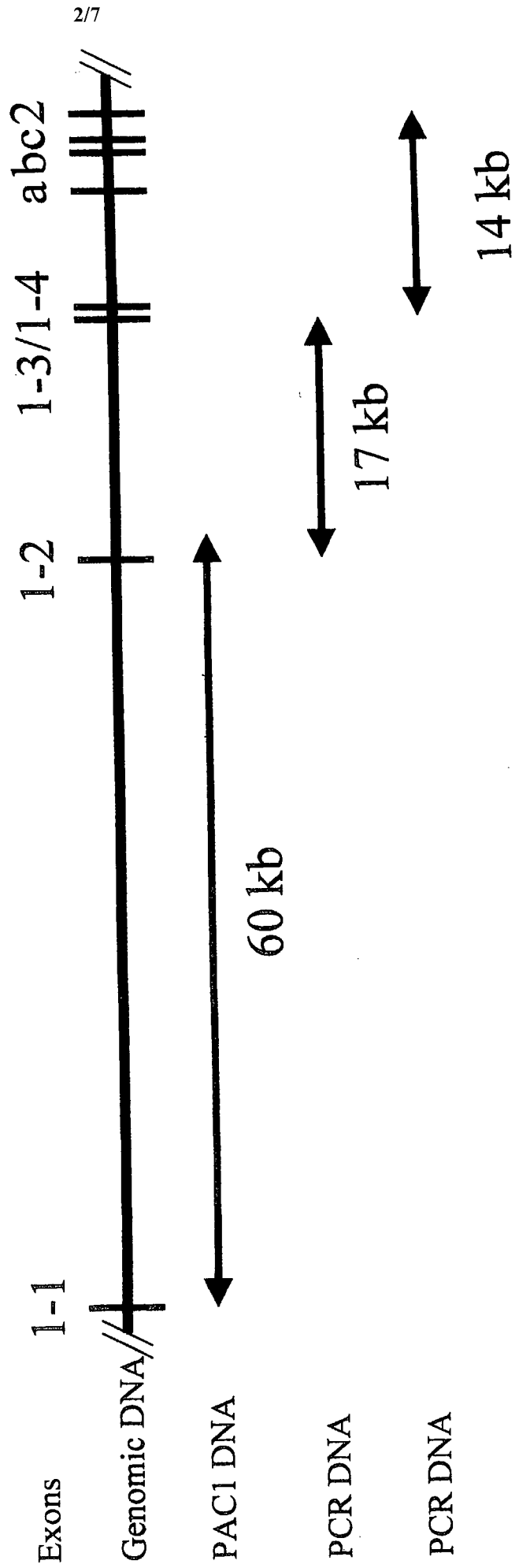


FIGURE 2B

B**Exon 1-1:**

ccagcccccttcccttccctgacccttcttgccatcgccccagacATGGGGAACGCGGCGACCG
CCAAGAAAGGCAGCGAGgtggagagcg

Exon 1-2:

agctttatatttaaatgctctcattagcctatatattaatattaaaacacccaaacataaagccttt
 taggcagatattgcaagtttttaaactcctcaactctagctgaaaagtgttttgctaagaaaagct
 cagtaatgtgctgttttatattaacaggaaacagaacagcagtagtggttgaataccctgcaaac
 aggaagtttgacacatgcatagctcttagcttctgtgtaagaagtgtgtagctccttctggaaaca
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CCAGTATACAGGTACAACCTACAGCTCTTCAGAAATTGGAAGGTTTTGCTAGCCGGTTATTTTCATAG
ACACTCTAAAGGTACTGCACATGATCAGAAAACAGCTCTGGAAAATGACAGCCTTCATTTCTCTGA
ACATACTGCCTTATGGGACAGATCAAgtaagttttg

Exon 1-3 and 1-4:

tgttttaggcagagttcagtggttcgtcaciaataaatgtttcttaatttggtgtttatgactgc
 tcgatttccagagccatgaaatcattgtttttgataattctgttcagcttcatagttgtttcttag
 gaagatttctgacttacttccacatcacaaaagtcctgcctcacatggcaactgttaaaatggc
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 ctgcatgctccagtggtgtgtttacaccatcggttcttctcctctagagattagcataactccc
 tttgctggttgattgttattttgagcaatagttttggaaagggttggttttcatcATGAGTGgtaa
 gtatgc

Exon a

cttgatcaag**CACGCAAATCATCAGATGCATCTG**gtaggaaaac

Exon b

tggacacaag**CTTGCTCCTCTTCAGAAATATCTG**gtaggcaagt

Exon c

gaacatgtag**ATTCCTTTG**gtatgctcat

Exon 2

atattttcag**TGAAAGAGTT**...

FIGURE 2C

C

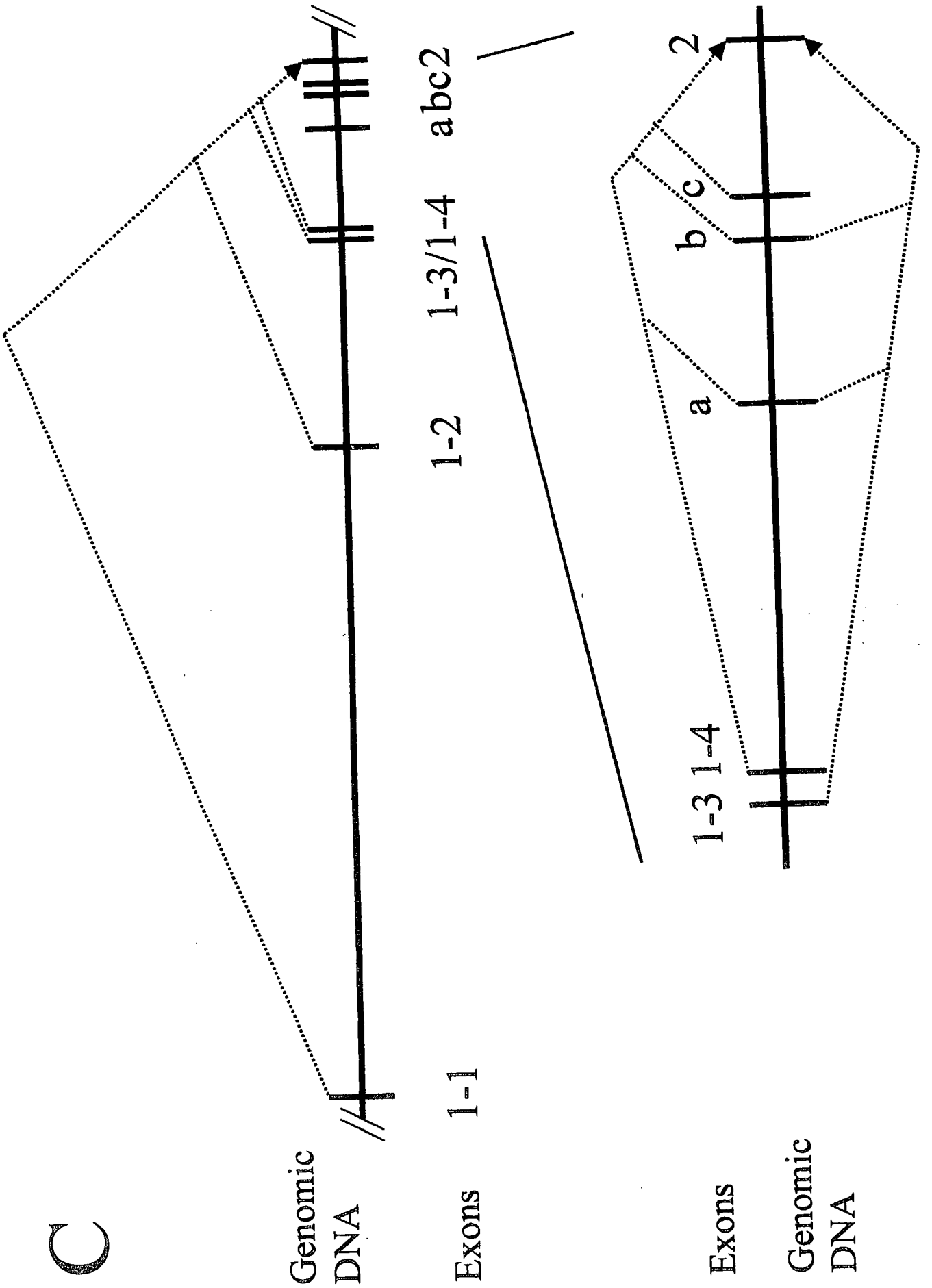
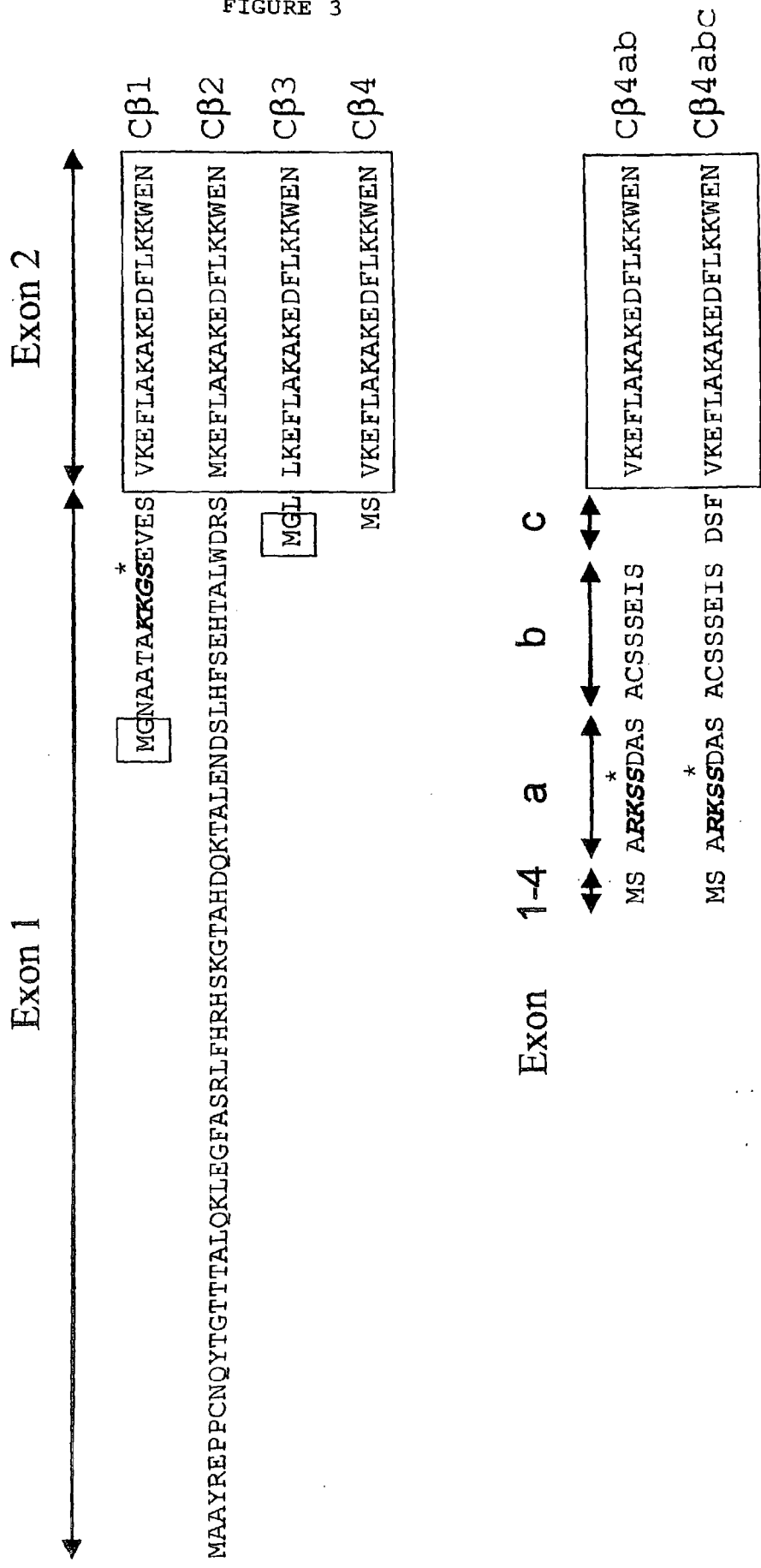
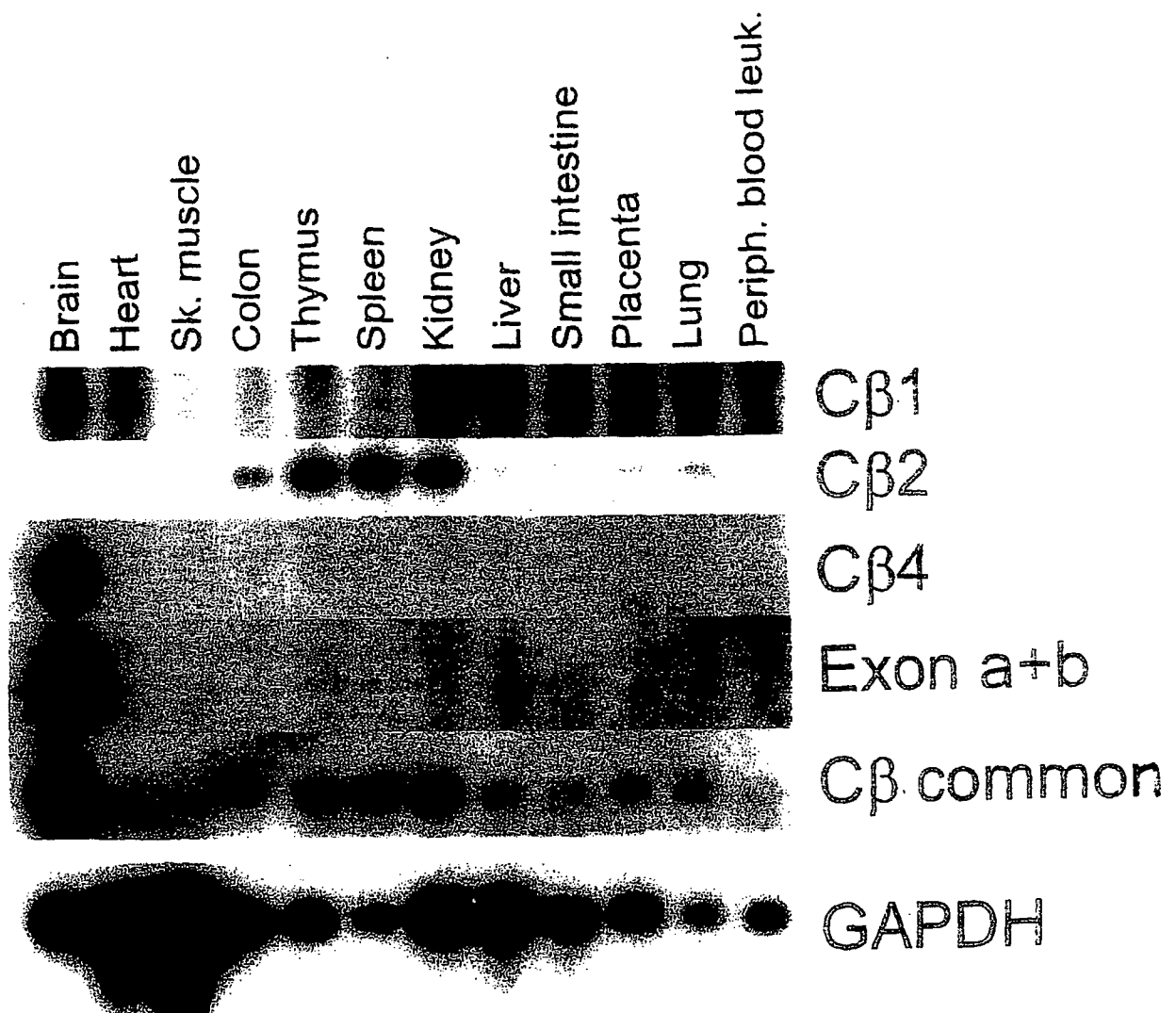


FIGURE 3



FIGUR 4



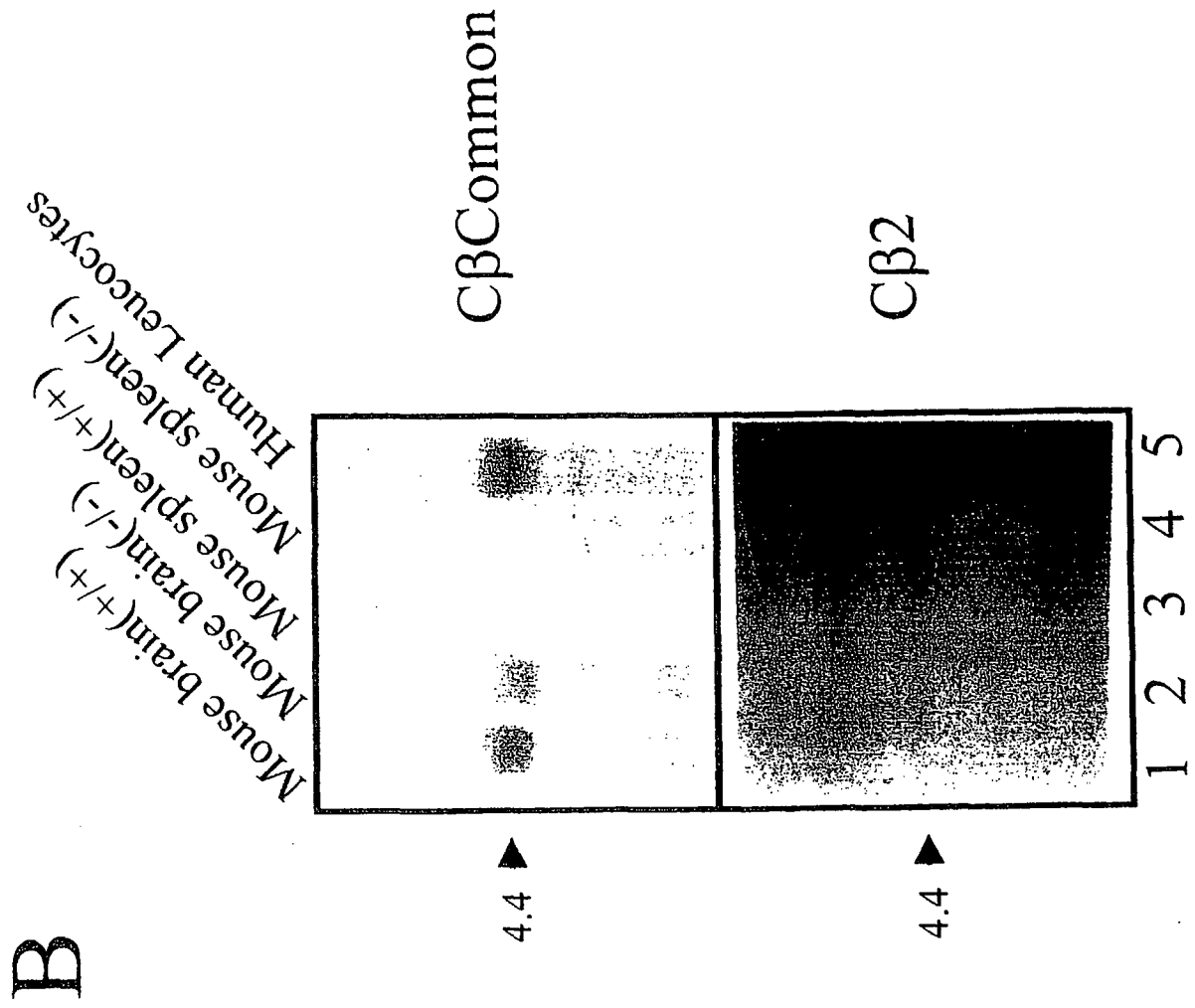


FIGURE 5B

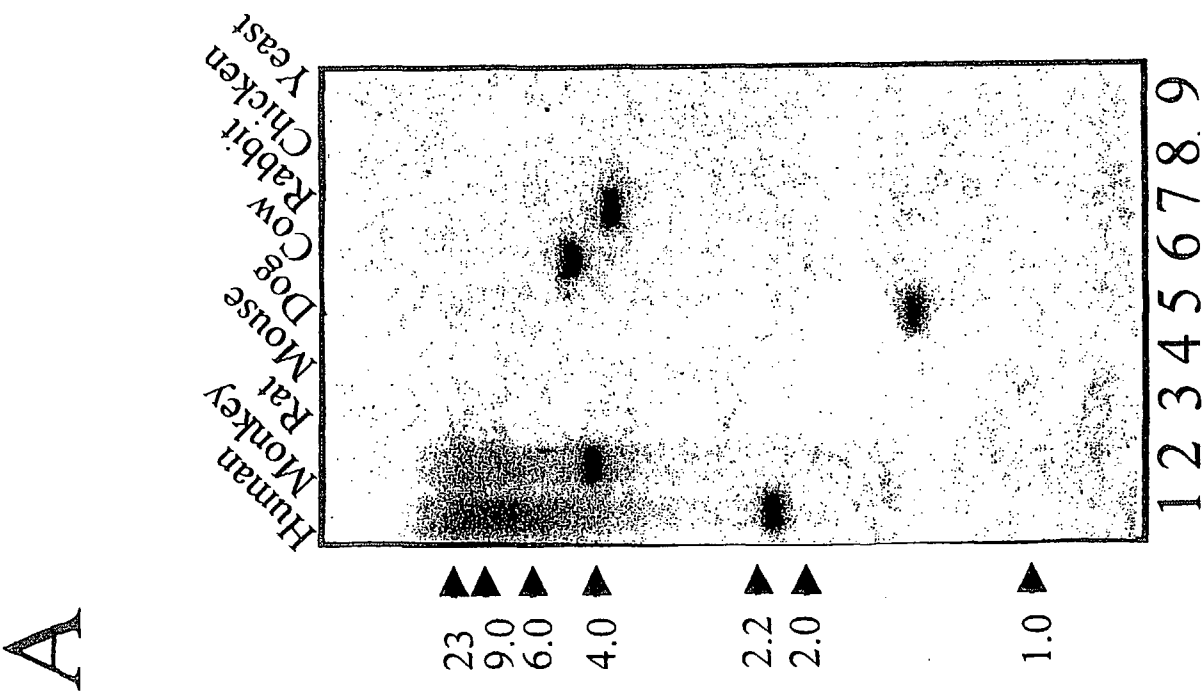


FIGURE 5A

SEQUENCE LISTING

SEQ ID NO: 1

<110> Sigurd Ørstavik, Nils Reinton, Eirik Frengen, Bjørn Tore Langeland, Tore Jahnsen og Bjørn S. Skålhegg

<120> **Identification of novel splice variants of the human catalytic subunit C β of cAMP-dependent protein kinase and the use thereof**

<160>9

<210>1

<211>

<212>genomic DNA

<213>Homo sapiens

<400>

Genomic sequence of exons and exon intron bouderies located upstream of exon 2 in the human C β gene.

Exon 1-1:

ccagccccc ttcccttccc tgacccttc ttgccatcgc cccagacATG
GGGAACGCGG CGACCGCCAA GAAAGGCAGC GAGGTGGAGA GCGgtgagtt gaaggccg

Exon 1-2:

agctttatat ttaatgctct cattagccta tatattaata ttaaaacacc
 caaacataaa gccttttagg cagatattgc aagtttttaa aatcctcaac
 tctagctgaa aagtgttttg ctaagaaaag ctcagtaatg tgctgtttta
 tattaacagg aaacagaaca gcagtagtgg tttgaatacc ctgcaaacag
 gaagtttgac acatgcatag ctcttagctt ctgtgtaaga agttgtgagc
 tccttctgga aacatttgca gttacattaa gtaaagtgta aatgcacatg
 aATGGCAGCT **TATAGAGAAC CACCTTGTA** **CCAGTATACA GGTACA****ACTA**
CAGCTCTTCA GAAATTGGAA GGTTTTGCTA GCCGGTTATT TCATAGACAC
TCTAAAGGTA CTGCACATGA TCAGAAAACA GCTCTGGAAA ATGACAGCCT
TCATTTCTCT GAACATACTG CCTTATGGGA CAGATCAAgt aagttttg

Exon 1-3 and 1-4:

tgtttttagg cagagttcag tggttcgtca caaataaatg tttcttaatt
 tgttgtttat gactgctcga tttccagagc catgaaatca tgtttttga
 taattctggt cagcttcata gttgtttctt aggaagattt cctgacttac
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attctgctgt ttgctccttg ccaggttcaa **cATGGGATTG** Tgtgagtatt
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 aacaactcat gctgatataa ttgagaacat cttatacatc ctggttcgaa
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 tgtgctgcat gctccagtgt gtgtgtttac accatcgggt cttctccctc
 tagagattag cataactccc tttgctgttg gattgttatt ttgagcaata
 tgttttggaa aggttggttt tcatc**ATGAG** TGgtaagtat gc

Exon a

cttgatcaag **CACGCAAATC ATCAGATGCA TCTG**gtagga aaac

Exon b

tggacacaag **CTTGCTCCTC TTCAGAAATA TCTG**gtaggc aagt

Exon c

gaacatgtag **ATTCCTTTG**g tatgctcat

Exon 2

atattttcag **TGAAAGAGTT**...

SEQ ID NO:2

<110> Sigurd Ørstavik, Nils Reinton, Eirik Frengen, Bjørn Tore Langeland, Tore Jahnsen og Bjørn S. Skålhegg

<120> **Identification of novel splice variants of the human catalytic subunit C β of cAMP-dependent protein kinase and the use thereof**

<160>9

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<212>cDNA

<213>Homo sapiens

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SEQ ID NO:3

<110> Sigurd Ørstavik, Nils Reinton, Eirik Frengen, Bjørn Tore Langeland, Tore Jahnsen og Bjørn S. Skålhegg

<120> **Identification of novel splice variants of the human catalytic subunit C β of cAMP-dependent protein kinase and the use thereof**

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<211>2944 nt

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<213>Homo sapiens

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SEQ ID NO:4

<110> Sigurd Ørstavik, Nils Reinton, Eirik Frengen, Bjørn Tore Langeland, Tore Jahnsen og Bjørn S. Skålhegg

<120> **Identification of novel splice variants of the human catalytic subunit C β of cAMP-dependent protein kinase and the use thereof**

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<213>Homo sapiens

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<110> Sigurd Ørstavik, Nils Reinton, Eirik Frengen, Bjørn Tore Langeland, Tore Jahnsen og Bjørn S. Skålhegg

<120> **Identification of novel splice variants of the human catalytic subunit C β of cAMP-dependent protein kinase and the use thereof**

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<110> Sigurd Ørstavik, Nils Reinton, Eirik Frengen, Bjørn Tore Langeland, Tore Jahnsen og Bjørn S. Skålhegg

<120> Identification of novel splice variants of the human catalytic subunit C β of cAMP-dependent protein kinase and the use thereof

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1201 ATGACATCTG AGCTCACACT CAGTGTTTGC ACTCTGTTGA GAGATAAGGT
1251 AGAGCTGAGA CCGTCCTTGT TGAAGCAGTT ACCTAGTTCC TTCATTCCAA
1301 CGACTGAGTG AGGTCTTTAT TGCCATCATC CGTGTGCGCA CTCTGCATCC
1351 ACCTATGTAA CAAGGCACCG CTAAGCAAGC ATTGTCTGTG CCATAACACA
1401 GTACTAGACC ACTTTCTTAC TTCTCTTTGG GTTGTCTTTC TCCTCTCCTA
1451 CATCCATTTT TTCCTTTTCA ATTTTATTGG TTTTCTCTAA ACAGTGCTCC
1501 ATTTTATTTT GTTGGTGTTT CAGATGGGCA GTGTTATGGC TACGTGATAT
1551 TTGAAGGGAA GGATAAGTGT TGCTTTCAGT AGTTATTGCC AATATTGTTG
1601 TTGGTCAATG GCTTGAAGAT AACTTTTCTA ATAATTATTA TTTCTTTGAG
1651 TAGCTCAGAC TTGGTTTTGC CAAAACCTTT GGTAATTTTT GAAGATAGAC
1701 TGTCTTATCA CCAAGGAAAT TTATACAAAT TAAGACTAAC TTTCTTGGA
1751 TTCACTATTC TGGCAATAAA TTTTGGTAGA CTAATACAGT ACAGCTAGAC
1801 CCAGAAATTT GGAAGGCTGT AGATCAGAGG TTCTAGTTCC CTTTCCCTCC
1851 TTTTATATCC TCCTCTCCTT GAGTAATGAA GTGACCAGCC TGTGTAGTGT
1901 GACAAACGFG TCTCATTAG CAGGAAAAAC TAATGATATG GATCATCACC
1951 CAGATTCTCT CACTTGGTAC CAGCATTCTT GTAGGTATTA GAGAAGAGTT
2001 CTAAGTTTTT TAAACCTTAA CTGTTCTTAA AGGATTTTAG CCAGTATTTT
2051 AATAGAACAT GATTAATGAA AGTGACAAAT TTTAAATTTT CTCTAATAGT
2101 CCTCATCATA AACTTTTTTAA AGGAAAATAA GCAAACATAA AAGAACATTG
2151 GTTTAGATAA AACTTTTATA TTTGCAAAGT CAAAATGGC TTGATTTTTG
2201 GAAACAATAT AGAGGTATTC ATATTTAAAT GAGGGTTTAC ATTTGTTTTG
2251 TTTTGTAACC GTTAAAAAGA AGTTGTTTCC AGCTAATTAT TGTGGTGTAC
2301 TATATTTGTG AGCCTAGGGT AGGGGCACTG CTGCAACTTC TGCTTTCATC
2351 CCATGCCTCA TCAATGAGGA AAGGGAACAA AGTGTATAAA ACCTGCCACA
2401 ATTGTATTTT AATTTTGAGG TATGATATTT TCAGATATTT CATAATTTCT
2451 AACCTCTGTT CTCTCAGTAA ACAGAATGTC TGATCGATCA TGCAGATACA
2501 ATGTTGGTAT TTGAGAGGTT AGTTTTTTTC CTACACTTTT TTTTGCCAAC
2551 TGACTIONACA ACATTGCTGT CAGGTGGAAA TTTCAAGCAC TTTTGCACAT
2601 TTAGTTCAGT GTTTGTTGAG AATCCATGGC TTAACCCACT TGTTTTGCTA
2651 TTTTTTCTT TGCTTTTAAAT TTTCCCATC TGATTTTATC TCTGCGTTTC

2701 AGTGACCTAC CTTAAAACAA CACACGAGAA GAGTTAAACT GGGTTCATTT
2751 TAATGATCAA TTTACCTGCA TATAAAATTT ATTTTAAATC AAGCTGATCT
2801 TAATGTATAT AATCATTCTA TTTGCTTTAT TATCGGTGCA GGTAGGTCAT
2851 TAACACCACT TCTTTTCATC TGTACCACAC CCTGGTGAAA CCTTTGAAGA
2901 CATAAAAAAA ACCTGTCTGA GATGTTCTTT CTACCAATCT ATATGTCTTT
2951 CGGTTATCAA GTGTTTCTGC ATGGTAATGT CATGTAAATG CTGATATTGA
3001 TTTCCTGGT CCATCTATAT TTAAAACGTG C

SEQ ID NO:7

<110> Sigurd Ørstavik, Nils Reinton, Eirik Frengen, Bjørn Tore Langeland, Tore Jahnsen og Bjørn S. Skålhegg

<120> **Identification of novel splice variants of the human catalytic subunit C β of cAMP-dependent protein kinase and the use thereof**

<160>9

<210>

<211>

<212>amino acid

<213>Homo sapiens

<400> **Proteinsequence C β 2**

Met-Ala-Ala-Tyr-Arg-Glu-Pro-Pro-Cys-Asn-Gln-Tyr-Thr-Gly-Thr-Thr-Thr-Ala-Leu-Gln-Lys-Leu-Glu-Gly-Phe-Ala-Ser-Arg-Leu-Phe-His-Arg-His-Ser-Lys-Gly-Thr-Ala-His-Asp-Gln-Lys-Thr-Ala-Leu-Glu-Asn-Asp-Ser-Leu-His-Phe-Ser-Glu-His-Thr-Ala-Leu-Trp-Asp-Arg-Ser-Met-Lys-Glu-Phe-Leu-Ala-Lys-Ala-Lys-Glu-Asp-Phe-Leu-Lys-Lys-Trp-Glu-Ser-Pro-Ala-Gln-Asn-Thr-Ala-His-Leu-Asp-Gln-Phe-Glu-Arg-Ile-Lys-Thr-Leu-Gly-Thr-Gly-Ser-Phe-Gly-Arg-Val-Met-Leu-Val-Lys-His-Lys-Glu-Thr-Gly-Asn-His-Tyr-Ala-Met-Lys-Ile-Leu-Asp-Lys-Gln-Lys-Val-Val-Lys-Leu-Lys-Gln-Ile-Glu-His-Thr-Leu-Asn-Glu-Lys-Arg-Ile-Leu-Gln-Ala-Val-Asn-Phe-Pro-Phe-Leu-Val-Lys-Leu-Glu-Phe-Ser-Phe-Lys-Asp-Asn-Ser-Asn-Leu-Tyr-Met-Val-Met-Glu-Tyr-Val-Pro-Gly-Gly-Glu-Met-Phe-Ser-His-Leu-Arg-Arg-Ile-Gly-Arg-Phe-Ser-Glu-Pro-His-Ala-Arg-Phe-Tyr-Ala-Ala-Gln-Ile-Val-Leu-Thr-Phe-Glu-Tyr-Leu-His-Ser-Leu-Asp-Leu-Ile-Tyr-Arg-Asp-Leu-Leu-Lys-Pro-Glu-Asn-Leu-Leu-Ile-Asp-Gln-Gln-Gly-Tyr-Ile-Gln-Val-Thr-Asp-Phe-Gly-Phe-Ala-Lys-Arg-Val-Lys-Gly-Arg-Thr-Trp-Thr-Leu-Cys-Gly-Thr-Pro-Glu-Tyr-Leu-Ala-Pro-Glu-Ile-Ile-Leu-Ser-Lys-Gly-Tyr-Asn-Lys-Ala-Val-Asp-Trp-Trp-Ala-Leu-Gly-Val-Leu-Ile-Tyr-Glu-Met-Ala-Ala-Gly-Tyr-Pro-Pro-Phe-Phe-Ala-Asp-Gln-Pro-Ile-Gln-Ile-Tyr-Glu-Lys-Ile-Val-Ser-Gly-Lys-Val-Arg-Phe-Pro-Ser-Ser-His-Phe-Ser-Ser-Asp-Leu-Lys-Asp-Leu-Leu-Arg-Asn-Leu-Leu-Gln-Val-Asp-Leu-Thr-Lys-Arg-Phe-Gly-Asn-Leu-Lys-Asn-Gly-Val-Asn-Asp-Ile-Lys-Asn-His-Lys-Trp-Phe-Ala-Thr-Thr-Asp-Trp-Ile-Ala-Ile-Tyr-Gln-Arg-Lys-Val-Glu-Ala-Pro-Phe-Ile-Pro-Lys-Phe-Lys-Gly-Pro-Gly-Asp-Thr-Ser-Asn-Phe-Asp-Asp-Tyr-Glu-Glu-Glu-Glu-Ile-Arg-Val-Ser-Ile-Asn-Glu-Lys-Cys-Gly-Lys-Glu-Phe-Ser-Glu-Phe

SEQ ID NO:8

<400> **Protein sequence C β 4ab**

Met-Ser-Ala-Arg-Lys-Ser-Ser-Asp-Ala-Ser-Ala-Cys-Ser-Ser-Ser-Glu-Ile-Ser-Val-Met-Lys-Glu-Phe-Leu-Ala-Lys-Ala-Lys-Glu-Asp-Phe-Leu-Lys-Lys-Trp-Glu-Ser-Pro-Ala-Gln-Asn-Thr-Ala-His-Leu-Asp-Gln-Phe-Glu-Arg-Ile-Lys-Thr-Leu-Gly-Thr-Gly-Ser-Phe-Gly-Arg-Val-Met-Leu-Val-Lys-His-Lys-Glu-Thr-Gly-Asn-His-Tyr-Ala-Met-Lys-Ile-Leu-Asp-Lys-Gln-Lys-Val-Val-Lys-Leu-Lys-Gln-Ile-Glu-His-Thr-Leu-Asn-Glu-Lys-Arg-Ile-Leu-Gln-Ala-Val-Asn-Phe-Pro-Phe-Leu-Val-Lys-Leu-Glu-Phe-Ser-Phe-Lys-Asp-Asn-Ser-Asn-

Leu-Tyr-Met-Val-Met-Glu-Tyr-Val-Pro-Gly-Gly-Glu-Met-Phe-Ser-His-Leu-Arg-Arg-Ile-Gly-Arg-Phe-Ser-Glu-Pro-His-Ala-Arg-Phe-Tyr-Ala-Ala-Gln-Ile-Val-Leu-Thr-Phe-Glu-Tyr-Leu-His-Ser-Leu-Asp-Leu-Ile-Tyr-Arg-Asp-Leu-Leu-Lys-Pro-Glu-Asn-Leu-Leu-Ile-Asp-Gln-Gln-Gly-Tyr-Ile-Gln-Val-Thr-Asp-Phe-Gly-Phe-Ala-Lys-Arg-Val-Lys-Gly-Arg-Thr-Trp-Thr-Leu-Cys-Gly-Thr-Pro-Glu-Tyr-Leu-Ala-Pro-Glu-Ile-Ile-Leu-Ser-Lys-Gly-Tyr-Asn-Lys-Ala-Val-Asp-Trp-Trp-Ala-Leu-Gly-Val-Leu-Ile-Tyr-Glu-Met-Ala-Ala-Gly-Tyr-Pro-Pro-Phe-Phe-Ala-Asp-Gln-Pro-Ile-Gln-Ile-Tyr-Glu-Lys-Ile-Val-Ser-Gly-Lys-Val-Arg-Phe-Pro-Ser-Ser-His-Phe-Ser-Ser-Asp-Leu-Lys-Asp-Leu-Leu-Arg-Asn-Leu-Leu-Gln-Val-Asp-Leu-Thr-Lys-Arg-Phe-Gly-Asn-Leu-Lys-Asn-Gly-Val-Asn-Asp-Ile-Lys-Asn-His-Lys-Trp-Phe-Ala-Thr-Thr-Asp-Trp-Ile-Ala-Ile-Tyr-Gln-Arg-Lys-Val-Glu-Ala-Pro-Phe-Ile-Pro-Lys-Phe-Lys-Gly-Pro-Gly-Asp-Thr-Ser-Asn-Phe-Asp-Asp-Tyr-Glu-Glu-Glu-Glu-Ile-Arg-Val-Ser-Ile-Asn-Glu-Lys-Cys-Gly-Lys-Glu-Phe-Ser-Glu-Phe

SEQ ID NO:9

Protein sequence C β 4abc

Met-Ser-Ala-Arg-Lys-Ser-Ser-Asp-Ala-Ser-Ala-Cys-Ser-Ser-Ser-Glu-Ile-Ser-Asp-Ser-Phe-Val-Met-Lys-Glu-Phe-Leu-Ala-Lys-Ala-Lys-Glu-Asp-Phe-Leu-Lys-Lys-Trp-Glu-Ser-Pro-Ala-Gln-Asn-Thr-Ala-His-Leu-Asp-Gln-Phe-Glu-Arg-Ile-Lys-Thr-Leu-Gly-Thr-Gly-Ser-Phe-Gly-Arg-Val-Met-Leu-Val-Lys-His-Lys-Glu-Thr-Gly-Asn-His-Tyr-Ala-Met-Lys-Ile-Leu-Asp-Lys-Gln-Lys-Val-Val-Lys-Leu-Lys-Gln-Ile-Glu-His-Thr-Leu-Asn-Glu-Lys-Arg-Ile-Leu-Gln-Ala-Val-Asn-Phe-Pro-Phe-Leu-Val-Lys-Leu-Glu-Phe-Ser-Phe-Lys-Asp-Asn-Ser-Asn-Leu-Tyr-Met-Val-Met-Glu-Tyr-Val-Pro-Gly-Gly-Glu-Met-Phe-Ser-His-Leu-Arg-Arg-Ile-Gly-Arg-Phe-Ser-Glu-Pro-His-Ala-Arg-Phe-Tyr-Ala-Ala-Gln-Ile-Val-Leu-Thr-Phe-Glu-Tyr-Leu-His-Ser-Leu-Asp-Leu-Ile-Tyr-Arg-Asp-Leu-Leu-Lys-Pro-Glu-Asn-Leu-Leu-Ile-Asp-Gln-Gln-Gly-Tyr-Ile-Gln-Val-Thr-Asp-Phe-Gly-Phe-Ala-Lys-Arg-Val-Lys-Gly-Arg-Thr-Trp-Thr-Leu-Cys-Gly-Thr-Pro-Glu-Tyr-Leu-Ala-Pro-Glu-Ile-Ile-Leu-Ser-Lys-Gly-Tyr-Asn-Lys-Ala-Val-Asp-Trp-Trp-Ala-Leu-Gly-Val-Leu-Ile-Tyr-Glu-Met-Ala-Ala-Gly-Tyr-Pro-Pro-Phe-Phe-Ala-Asp-Gln-Pro-Ile-Gln-Ile-Tyr-Glu-Lys-Ile-Val-Ser-Gly-Lys-Val-Arg-Phe-Pro-Ser-Ser-His-Phe-Ser-Ser-Asp-Leu-Lys-Asp-Leu-Leu-Arg-Asn-Leu-Leu-Gln-Val-Asp-Leu-Thr-Lys-Arg-Phe-Gly-Asn-Leu-Lys-Asn-Gly-Val-Asn-Asp-Ile-Lys-Asn-His-Lys-Trp-Phe-Ala-Thr-Thr-Asp-Trp-Ile-Ala-Ile-Tyr-Gln-Arg-Lys-Val-Glu-Ala-Pro-Phe-Ile-Pro-Lys-Phe-Lys-Gly-Pro-Gly-Asp-Thr-Ser-Asn-Phe-Asp-Asp-Tyr-Glu-Glu-Glu-Glu-Ile-Arg-Val-Ser-Ile-Asn-Glu-Lys-Cys-Gly-Lys-Glu-Phe-Ser-Glu-Phe

INTERNATIONAL SEARCH REPORT

International application No.

PCT/NO 00/00445

A. CLASSIFICATION OF SUBJECT MATTER		
IPC7: C12N 9/12 // C12Q 1/48 According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED		
Minimum documentation searched (classification system followed by classification symbols)		
IPC7: C12N, C12Q		
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched		
SE,DK,FI,NO classes as above		
Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	The Journal of Biological Chemistry, Volume 266, No 8, 1991, Stefan Wiemann et al, "Isoform CBeta2, an Unusual Form of the Bovine Catalytic Subunit of CAMP-independent Protein Kinase", page 5140 - page 5146, & Swissprot. Accession no. P24256, 01.03.1992, 93.5% identity in 400 aa overlap with SEQ ID No 7 --	1-18
X	The Journal of Biological Chemistry, Volume 272, No 47, 1997, Chris R. Guthrie et al, "Two Novel Brain-specific Splice Variants of the Murine C Beta Gene of caMP-dependent Protein Kinase", page 29560 - page 29565 --	1-18
<input checked="" type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.		
* Special categories of cited documents: "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier application or patent but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance: the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance: the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art "&" document member of the same patent family		
Date of the actual completion of the international search		Date of mailing of the international search report
29 May 2001		30 -05- 2001
Name and mailing address of the ISA/ Swedish Patent Office Box 5055, S-102 42 STOCKHOLM Facsimile No. +46 8 666 02 86		Authorized officer Yvonne Siösteen/BS Telephone No. +46 8 782 25 00

INTERNATIONAL SEARCH REPORT

International application No.

PCT/NO 00/00445

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	Molecular Endocrinology, Volume 4, No 3, 1990, Stephen J. Beebe et al, "Molecular Cloning of a Tissue-Specific Protein Kinase (C-Gamma) from Human Testis - Representing a Third Isoform of the Catalytic Subunit of caMP-Dependent Protein Kinase", page 465 - page 475, & EMBASE/GENBANK/DDBJ, accession no M34181, 23.06, 1990, 100% identity in 2852 bp overlap with SEQ ID No 2 --	1-18
A	The Journal of Biological Chemistry, Volume 261, No 33, 1986, Michael D. Uhlert et al, "Evidence for a Second Isoform of the Catalytic Subunit of cAMP-dependent Protein Kinase", page 15360 - page 15363 --	1-18
A	The Journal of Biological Chemistry, Volume 261, No 35, 1986, Mark OI Showers et al, "A Cloned Bovine cDNA Encodes an Alternate Form of the Catalytic Subunit of cAMP-dependent Protein Kinase", page 16288 - page 16291 --	1-18
A	Proc. Natl. Acad. Sci., Volume 89, 1992, Sven Beushausen et al, "Catalytic subunit of Aplysia neuronal cAMP-dependent protein kinase with two different N termini" page 1641 - page 1645 --	1-18
A	The Journal of Biological Chemistry, Volume 273, No 38, 1998, Jovental T. San Agustin et al, "The Catalytic Subunit of the cAMP-dependent Protein Kinase of Ovine Sperm Flagella Has a Unique Amino-terminal Sequence", page 24874 - page 24883 --	1-18

INTERNATIONAL SEARCH REPORT

International application No.
PCT/NO00/00445

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

- 1. Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

- 2. Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

- 3. Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see next sheet

- 1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
- 2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
- 3. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

- 4. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- The additional search fees were accompanied by the applicant's protest.
- No protest accompanied the payment of additional search fees.

The International Searching Authority considers that the present application contains the following six inventions which relate to different splice variants of the catalytic subunit of cAMP-dependent protein kinase:

1. The splice variant c-beta1 according to part of claims 1-3,5,6 and 12.
2. The splice variant c-beta2 according to part of claims 1-6, 8, 12,16 and claims 7,9-11,13-18.
3. The splice variant c-beta3 according to part of claims 1-3,5 and 12.
4. The splice variant c-beta4 according to part of claims 1-3,5,8 and 12.
5. The splice variant c-beta4ab according to part of claims 1-6,8 and 12.
6. The splice variant c-beta4abc according to claims 1-6,8 and 12.

The invention listed as Groups 1-6 do not relate to a single inventive concept under PCT Rule 13.1 because they lack the same or corresponding novel special technical features for the following reasons: The claims are not so linked by a special technical feature so as to form a single inventive concept. Because c-beta splice variants of the catalytic subunit of cAMP-dependent protein kinase was already known from the prior art (see description page 2, line 19 and Guthrie et al, The journal of Biological Chemistry, Vol 272, No47,1997, page 29560-29565) it cannot form basis of unity of invention.

As inventions 2-6 could be searched without effort justifying additional fee, this Authority did not invite payment of any additional fee for these inventions. Thus, inventions 1-6 have been searched.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/NO 00/00445

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	Advances in Second Messenger and Phosphoprotein Advanced, Volume 31, 1997, Kjetil Taskén et al, "Structure, Function and Regulation of Human cAMP-Dependent Protein Kinases" page 191 - page 204 -- -----	1-18

专利名称(译)	CAMP依赖性蛋白激酶人催化亚基C β 新型裂解变异体的鉴定及其应用		
公开(公告)号	EP1242585A1	公开(公告)日	2002-09-25
申请号	EP2000987849	申请日	2000-12-22
[标]申请(专利权)人(译)	ORSTAVIK SIGURD REINTON NILS FRENGEN的Eirik 朗厄兰岛比约恩撕 JAHNSEN TORE LHEGG BJ RN S		
申请(专利权)人(译)	ORSTAVIK , SIGURD REINTON , NILS FRENGEN , 的Eirik JAHNSEN , 撕		
当前申请(专利权)人(译)	ORSTAVIK , SIGURD REINTON , NILS FRENGEN , 的Eirik JAHNSEN , 撕		
[标]发明人	ORSTAVIK SIGURD REINTON NILS FRENGEN EIRIK LANGELAND BJ RN TORE JAHNSEN TORE SK LHEGG BJ RN S		
发明人	ORSTAVIK, SIGURD REINTON, NILS FRENGEN, EIRIK LANGELAND, BJ RN TORE JAHNSEN, TORE SK LHEGG, BJ RN S.		
IPC分类号	G01N33/48 A61K31/7088 A61K38/00 A61K38/45 A61K45/00 A61K48/00 A61P37/04 A61P43/00 C12N9/00 C12N9/12 C12N9/99 C12N15/09 C12Q1/02 C12Q1/48 C12Q1/68 G01N21/78 G01N33/15 G01N33/50 G01N33/53 G01N33/569		
CPC分类号	C12N9/1205 A61K38/00 C12Q1/485 C12Y207/11011 G01N33/56972		
优先权	19996423 1999-12-23 NO		
外部链接	Espacenet		

摘要(译)

本发明证明C β 基因编码至少6种不同的基因产物，命名为C β 1，C β 2，C β 3，C β 4，C β 4ab和C β 4abc。与鼠和牛剪接变体的情况一样，所有人C β 剪接变体在由外显子2编码的部分之前的N末端部分中变化。在人类中鉴定了在小鼠和牛中鉴定的所有C β 剪接变体的同源物。（C beta 1，C beta 2，C beta 3和C beta 4）以及之前未在任何其他物种中鉴定的两种新型C β 剪接变体（C beta 4ab和C beta 4abc）。在这方面，本发明包括编码所述剪接变体的基因组DNA-和cDNA序列，并且分别包含SEQ ID NO：1,2,3,4,5和6中所示的核苷酸序列。其中所述蛋白质是C β 蛋白质的新剪接变体。本发明还涉及包含所述cDNA序列的载体。本发明还包括特征在于分别如SEQ ID NO：7,8和9中所示的特定氨基酸C β 剪接变体蛋白的蛋白质。本发明还包括所述C β 剪接变体蛋白和DNA序列在制备用于诊断和治疗目的的药物中的进一步用途。

