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(54) **Method for identifying type-I interferon responsive MS patients by determining TRAIL expression**

(57) This invention relates to a method for identifying an individual, in particular human, as a positive responder or a non-responder to a type-I Interferon (IFN) therapy, in particular IFN- β therapy, used in the treatment of Multiple Sclerosis by determining TRAIL gene

expression and/or TRAIL protein level. The invention further relates to the use of said method for monitoring IFN therapy in the treatment of Multiple Sclerosis and to a kit for carrying out said method.

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Description

[0001] This invention relates to a method for identifying MS patients susceptible to type I-Interferon (IFN), in particular IFN- β , and for monitoring IFN therapy in MS patients by determining TRAIL gene and/or protein expression as a marker.

[0002] Multiple Sclerosis (MS) affects approximately 1 million individuals worldwide and is the most common disease of the central nervous system (CNS) that causes prolonged and severe disability in young adults. Although its etiology remains elusive, strong evidence supports the concept that a T cell-mediated inflammatory process against self-molecules within the white matter of the brain and spinal cord underlies its pathogenesis (Martin *et al.* 1992, *Annu Rev Immunol* **10**: 153-187). Since myelin-reactive T cells are present in both MS patients and healthy individuals (Pette *et al.* 1990, *Neurology* **40**: 1770-1776; Hohlfeld *et al.* 1995, *Neurology* **45** (suppl 6): S33-S38), the primary immune abnormality in MS most likely involves failed regulatory mechanisms that lead to an enhanced T cell activation status and less stringent activation requirements (Zang *et al.* 1994, *J Exp Med* **179**: 973-984; Markovic-Plese *et al.* 2001, *J Clin Invest* **108**: 1185-1194).

[0003] Interferon-beta (IFN- β), a type-I Interferon (IFN), is a pleiotropic cytokine with immunomodulatory properties and has become a global standard in the treatment of MS. Despite the well documented efficacy in responders to this medication, a substantial number of patients fail to respond to IFN- β . Why IFN- β therapy is or is not effective with respect to MS, and how IFN- β alters the clinical course of MS remains unclear (The IFNB Multiple Sclerosis Study Group and the University of British Columbia MS/MRI Analysis Group 1995, *Neurology* **45**: 1277-1285; Jacobs *et al.* 1996, *Ann Neurol* **39**: 285-294; PRISMS Study Group 1998, *Lancet* **352**: 1498-1504). Putative mechanisms of action include the inhibition of T cell proliferation, regulation of a large number of cytokines, and blocking of blood-brain barrier opening via interference with cell adhesion, migration and matrix metalloproteinase activity (Rudick *et al.* 1996, *Ann Neurol* **40**: 618-627; Wang *et al.* 2000, *J Immunol* **165**: 548-557; Stone *et al.* 2000, *Ann Neurol* **37**: 611-619). Understanding how IFN- β works in MS, however, is imperative for a development of markers that would allow predicting an individual's chance of a positive IFN- β response. In the face of the costs of IFN- β therapy as well as possible side-effects for the patient, such a prediction might be valuable in increasing the efficacy of current and future strategies to treat patients suffering from MS.

[0004] Consequently, there is a need for identifying markers which correlate with the clinical outcome of IFN- β therapy in MS patients, and which are usable in a method for predicting therapy response of an individual patient.

[0005] Recently, a number of genes involved in innate and specific immune responses that are potential effector targets of IFN- β in MS were identified (Wandinger *et al.* 2001, *Ann Neurol* **50**: 349-357). Tumor necrosis factor (TNF)-related apoptosis inducing ligand (TRAIL; also referred to as APO2L), a novel member of the TNF/nerve growth factor superfamily (Wiley *et al.* 1995, *Immunity* **3**: 673-682), was among the genes that were consistently up-regulated by IFN- β *in vitro*. Interestingly, TRAIL has also been shown to mediate the induction of endogenous IFN- β , thereby amplifying the effects of this therapy (Kumar-Sinha *et al.* 2002, *J Biol Chem* **277**: 575-585).

[0006] Apart from its apoptosis inducing effects (Wiley *et al.* 1995, *Immunity* **3**: 673-682), TRAIL was recently shown to exert potent anti-inflammatory properties at the T cell level (Song *et al.* 2000, *J Exp Med* **191**: 1095-1104; Hilliard *et al.* 2001, *J Immunol* **166**: 1314-1319; Lünemann *et al.* 2002, *J Immunol* **168**: 4881-4888). Studies on animal models of autoimmune diseases demonstrated that systemic neutralization by TRAIL receptor 2 led to exacerbation of collagen-induced arthritis and experimental autoimmune encephalomyelitis and suggested an influence of TRAIL on T-cell growth and effector function (Song *et al.* 2000, *J Exp Med* **191**: 1095-1104; Hilliard *et al.* 2001, *J Immunol* **166**: 1314-1319). In the human immune system, Lünemann *et al.* (2002, *J Immunol* **168**: 4881-4888) have very recently shown that TRAIL fails to induce apoptosis, but inhibits the activation of autoreactive and foreign antigen-specific T cells. In particular, TRAIL negatively regulates calcium influx through store-operated calcium release-activated calcium channels which is crucial to lymphocyte activation, and moreover inhibits subsequent cell cycle progression as well as cytokine production well as cytokine production independently of the T cell's antigen-specificity or the T helper phenotype (Lünemann *et al.* 2002, *J Immunol* **168**: 4881-4888). This impact of TRAIL on human T cell effector functions suggests a role of the TRAIL system in the pathogenesis of MS in which activated myelin-specific T cells are thought to mediate the inflammatory brain damage. This notion is supported by the fact that TRAIL was found to be elevated in peripheral immune cells of MS patients during the natural disease course, and that levels were highest in patients with relapsing-remitting MS at the time of clinical remission (Huang *et al.* 2000, *Neurology* **55**: 928-934).

[0007] In addition to its immunoregulatory properties on human T cells, TRAIL was shown to induce apoptotic cell death in nontransformed human hepatocytes and brain cells (Jo *et al.* 2000, *Nat Med* **6**: 564-657; Nitsch *et al.* 2000, *Lancet* **356**: 827-828). The latter finding suggests a two edged role for TRAIL in neuroinflammation by promoting both anti-inflammatory and destructive properties. However, given the immediate effects of IFN- β on the integrity of the blood-brain barrier, the IFN- β -induced TRAIL up-regulation in the treatment of MS more likely comprises immunoregulatory mechanisms in the periphery than direct effect on the CNS (Stone *et al.* 1995, *Ann Neurol* **37**: 611-619).

[0008] Up till now it has not been possible to predict a patient's susceptibility to IFN- β treatment with a high level of confidence. Given that the IFN- β treatment is extremely expensive and places an immense financial burden on the

health system, it would be desirable to be able to make predictions about the outcome of an IFN- β therapy to a patient. Moreover, for the individual patient it is highly desirable to have such a prediction in order to weigh up the probability of benefit on the one hand and side-effects that have to be expected on the other.

[0009] Accordingly, it has been an object of the present invention to be able to predict the chances of a successful IFN- β therapy in MS affected individuals.

[0010] This object is solved by a method for identifying an individual as a positive responder or a 'negative responder, i.e. non-responder, to type-I IFN therapy used in the treatment of MS, characterized in that TRAIL expression and/or TRAIL protein level is determined.

[0011] It is also solved by the use of the method according to the present invention for monitoring type-I IFN therapy used in the treatment of MS.

[0012] It is furthermore solved by a kit for performing the method according to the present invention, comprising a nucleic acid probe and/or one or more primers specific for the TRAIL gene or a portion thereof and/or an antibody specific for the TRAIL polypeptide or a portion thereof.

[0013] Preferably, the method of the present invention is used in an IFN- β therapy.

[0014] The method of the present invention is preferably used to determine TRAIL expression and/or TRAIL protein level prior to IFN therapy.

[0015] In said method, a sample is preferably obtained from a mammalian, and more preferably is obtained from a human.

[0016] In a preferred embodiment, the sample is selected from the group comprising a bodily fluid, a fraction thereof, tissue extract, and cell extract.

[0017] In a more preferred embodiment, the bodily fluid is selected from the group comprising whole blood, blood serum, plasma, cerebrospinal fluid, synovial fluid, ascites exsudate, inflammatory exsudate, and urine.

[0018] In a particular preferred embodiment of the method according to the present invention,

(a) TRAIL expression is determined by detecting and/or measuring the amount and/or concentration of TRAIL nucleic acids encoding the TRAIL polypeptide or parts thereof, and

(b) TRAIL protein level is determined by detecting and/or measuring the amount and/or concentration of TRAIL polypeptide or parts thereof.

[0019] In one embodiment of the method according to the present invention, TRAIL protein level determination is performed by measuring the amount and/or concentration of soluble TRAIL (sTRAIL) protein in a liquid sample.

[0020] The measurement of the amount and/or concentration of TRAIL polypeptide is preferably done by an immunoassay using an anti-TRAIL antibody.

[0021] The immunoassay preferably is an enzyme-linked immunosorbent assay (ELISA) but also may be, for example, a radioimmunoassay (RIA) or a blotting technique such as Western blot.

[0022] The amount and/or concentration of TRAIL polypeptide determined preferably is compared to a limiting value discriminating positive responders and non-responders, which limiting value is about 350 ± 20 pg/ml.

[0023] Alternatively, the level of membrane bound TRAIL protein may be determined, which is preferably done by using an anti-TRAIL antibody in, for example, FACScan analysis, ELISPOT assay or an immunohistochemical technique.

[0024] In another embodiment of the present invention, the amount and/or concentration of TRAIL nucleic acid is determined by preferably measuring the amount and/or concentration of TRAIL mRNA in a sample of tissue or cell extract.

[0025] In a preferred embodiment, said sample comprises an extract selected from the group comprising an extract of blood cells, peripheral immune cells, neuronal tissue and muscle tissue, of which an extract of peripheral blood mononuclear cells (PBMC) is particularly preferred.

[0026] Measurement of the amount and/or concentration of TRAIL mRNA is preferably done with the aid of the polymerase chain reaction (PCR) using one or more TRAIL specific probes.

[0027] Alternatively, other techniques for determining nucleic acids such as Northern blot may be used. Furthermore, the amount and/or concentration of TRAIL mRNA may be determined in solid tissue or cell specimens by, for example, *in situ* hybridization using TRAIL specific probes.

[0028] The amount and/or concentration of TRAIL mRNA preferably is compared to a limiting value discriminating positive responders and non-responders. Said limiting value is at least twofold compared to the baseline.

[0029] The amount and/or concentration of TRAIL nucleic acid and/or polypeptide is preferably correlated with the amount and/or concentration of a nucleic acid and/or polypeptide, respectively, of one or more biological response markers of IFN- β therapy, such as MxA protein or any other IFN-inducible protein.

[0030] As used herein, an individual suffering from manifested MS is diagnosed according to generally accepted criteria (Poser *et al.* 1983, *Ann Neurol* **13**: 227-231; McDonald *et al.* 2001, *Ann Neurol* **50**: 121-127). Disability status (as defined by the Expanded Disability Status Scale, EDSS; Kurtzke 1983, *Neurology* **22**: 1444-1452) was assessed

by clinical parameters.

[0031] As used herein, "positive responders" are defined as patients who experienced no further relapses and no deterioration in the EDSS during IFN- β treatment, whereas patients who continue to have one or more relapses are defined as "non-responders". In another aspect, positive responders and non-responders to IFN- β treatment are defined by paraclinical activity markers, e.g. magnetic resonance imaging (MRI).

[0032] The method of the present invention shall be understood as a method wherein determination of TRAIL expression and/or TRAIL protein level is carried out extracorporally.

[0033] The term "determination" as used herein may mean both qualitatively detecting and quantifying.

[0034] As used herein, an anti-TRAIL antibody is an antibody directed against TRAIL polypeptide or parts thereof and may comprise polyclonal antiserum, monoclonal antibodies, diabodies and chimera.

[0035] In summary, TRAIL is disclosed herein as the first biological response marker of IFN- β therapy in MS that has been identified. It is demonstrated that early and sustained TRAIL gene induction is a marker of IFN- β therapy response by linking its gene and protein expression profile with clinical disease markers in long-term treated MS patients. More detailed, non-responders to IFN- β therapy can be distinguished from responsive individuals by their *in vivo* profile of TRAIL gene and protein expression. Furthermore, elevated sTRAIL protein levels in patient sera allowed to predict treatment response even before therapy was initiated. Using an ELISA for determining sTRAIL serum concentration, a limiting value of about 350 ± 20 pg/ml was estimated. Thus, concentrations above that limiting value indicate a prediction of positive response to IFN- β therapy, whereas lower concentrations point to a response failure.

[0036] The invention shall now be further described by the following examples with respect to the attached figures. All examples are provided by way of example only, without any intended limitation of the scope of the invention. All cited references are incorporated herein by reference in their entireties.

[0037] Prior to outlining the examples, reference is made to the figures, wherein

Figure 1 shows the spontaneous gene expression of MxA, TRAIL and CD95L in 62 MS patients during IFN- β -1a therapy. Samples of peripheral blood mononuclear cells (PBMC) were obtained before the beginning of IFN- β -1a therapy and at weeks 4, 26 and 52 under therapy. Total RNA was extracted and relative mRNA levels were quantified by realtime (rt)-PCR. Gene expression levels were normalized to expression of hypoxanthine phosphoryltransferase (HPRT). Induction of MxA protein gene expression was measured as a biological response marker to IFN- β therapy *in vivo*. Changes are expressed relative to gene expression levels before treatment. Data are presented as mean $\pm$ SEM. Statistical significance of changes after 4, 26 and 52 weeks of treatment compared to baseline was tested by Wilcoxon's signed-rank test. *p < 0.05.

Figure 2 shows the differential induction of TRAIL gene expression in IFN- β responders (n=20) and non-responders (n=19). In contrast to non-responders, IFN- β -1a therapy resulted in early and sustained up-regulation of TRAIL gene expression in MS patients with clinical treatment response. Up-regulation of MxA gene expression demonstrates the *in vivo* biological effectiveness of IFN- β -1a in both patient groups. CD95L was not significantly regulated by IFN- β therapy in any patient subset. *p < 0.05.

Figure 3 shows that the up-regulation of TRAIL gene expression during IFN- β treatment is abrogated in the presence of neutralizing antibodies (NAB). Spontaneous gene expression of MxA and TRAIL was measured in a third subset of patients (n=23) who developed NAB to IFN- β -1a after 1 year of treatment. Presence of NAB was confirmed after 18 months. Initial up-regulation of MxA and TRAIL gene expression was abrogated in the presence of NAB. *p < 0.05.

Figure 4 shows that elevated sTRAIL protein levels are correlated with IFN- β responsiveness in MS patients. Soluble TRAIL protein levels were measured in serum of IFN- β responders (n=12) and non-responders (n=11) before the beginning of IFN- β -1a therapy and at weeks 4, 26 and 52 under therapy. Box-whisker plots depict the median level for each group (horizontal line in the box), the 25th and 75th percentiles (upper and lower edges of box), and the 10th and 90th percentiles (lines extending above and below the box). Statistical significance of differences between the patient group was tested by the Mann-Whitney-U test. *p < 0.05.

Figure 5 shows the regulation of TRAIL gene transcription and protein expression by IFN- β . PBMC (10^6 per well) were cultured for the indicated time periods in 48 well plates in a final volume of 1 ml. Cells were incubated with IFN- β -1a (10 and 100 IU/ml) of PHA ($1\mu\text{g/ml}$) and Interleukin-2 (IL-2) (10 IU/ml). TRAIL mRNA was quantified by rtPCR. Gene expression levels were normalized to 18S rRNA (A). Cell surface expression was detected by flow cytometry. Cells were stained with an anti-TRAIL monoclonal antibody, followed by phycoerythrin (PE)-labeled goat anti-mouse monoclonal antibody (B). Concentration of sTRAIL protein in culture supernates was determined by ELISA (C). Results obtained in 2 healthy individuals are shown (mean $\pm$ SEM).

Examples

[0038] The following examples are intended to illustrate and to substantiate the present invention. In order to study an involvement of TRAIL in IFN- β therapy, patients with manifested MS were recruited. The study was approved by the local ethical committee, and informed consent was obtained from each individual.

[0039] All MS patients had clinically definite, relapsing-remitting MS. Healthy volunteers were selected from personnel of the Neuroimmunology lab. MS patients (n=62) studied longitudinally participated in the MASTER (MS Antibody Status and Therapy Evaluation with Rebif®) study, a clinical trial of IFN- β -1a. Eligible subjects for that study were patients at ages between 18 and 55 years who had relapsing-remitting MS for more than two years, with at least two relapses during the past two years, without any immunomodulatory treatment six months prior to the study, and without exacerbation 4 weeks prior to treatment initiation (Lünemann *et al.* 2001, *Neurology* **57**: 1132-1134). All patients were treated with 44 μ g IFN- β -1a (Rebif®, Serono, Unterschleißheim, Germany) by subcutaneous injection once weekly. This dose has been shown to significantly decrease the T2 activity and burden of disease as measured by MRI and represents twice the concentration of IFN- β -1a that has been demonstrated to delay the conversion to definite MS in patients after a first episode of neurological dysfunction (The Once Weekly Interferon for MS Study Group 1999, *Neurology* **53**: 679-686; Comi *et al.* 2001, *Lancet* **357**: 1576-1582). Furthermore, this dosage was found to be biologically effective as indicated by the clear and significant enhancement of MxA gene expression in PBMC after initiation of treatment (*cf.* Fig. 1). Clinical data on disability status (as defined by the EDSS) and relapses were obtained every time of blood drawing 3 days after IFN- β -1a application (Kurtzke 1983, *Neurology* **33**: 1444-1452). A clinical relapse was defined as significant worsening for preexisting symptoms or appearance of new neurological deficits in the absence of fever and lasting for more than 24 hours. Patients who experienced no further relapses and no deterioration in the EDSS during IFN- β treatment were defined as positive responders, whereas patients who continued to have one or more relapses were defined as non-responders. In some of these patients, clinical relapses were treated with a 3-day course of i.v. methylprednisolone at a dose of 1 g/day. However, all blood samples were obtained before treatment. For the purpose of this study, positive responders (n=20) and non-responders (n=19) were randomly selected (*cf.* Table 1). In addition, patients (n=23) who developed neutralizing antibodies (NAB) to Rebif® as determined by means of an MxA induction assay were included. The MxA concentration induced by Rebif® was determined as described (Kracke *et al.* 2000, *Neurology* **54**: 193-199). Results are given as Interferon Neutralizing Units (INU), i.e. the titer of serum that neutralizes 10 U/ml Rebif® activity to an apparent 1 U/ml activity as determined by a standard curve. Within this group, 17 patients fulfilled the clinical criteria for treatment response. Venous blood samples were collected prior to IFN- β treatment (baseline = week 0) as well as at weeks 4, 26 and 52 under therapy (each time 3 days after IFN- β application). Patients who developed NAB during the treatment course were followed up until week 78.

Example 1: Induction of TRAIL gene expression upon systemic IFN- β treatment

[0040] First, it was examined whether systemic IFN- β treatment of MS patients induces TRAIL gene expression *in vivo*. For this purpose, PBMC from MS patients (n=62) before and during IFN- β -1a therapy were investigated.

[0041] Samples of peripheral blood were obtained before the beginning of IFN- β -1a therapy and at weeks 4, 26 and 52 under therapy. PBMC were isolated from fresh blood by Ficoll density gradient centrifugation (Bio-Whittaker, Walkersville, MD) and cryopreserved in liquid nitrogen until analysis was done.

[0042] Total RNA was extracted and relative mRNA levels were quantified by realtime (rt)-PCR. In addition, the closely related TNF family member CD95L (also referred to as FasL) was quantified. In parallel, MxA protein gene expression was measured as an *in vivo* response marker to IFN- β -1a therapy (Wandinger *et al.* 2001, *Ann Neurol* **50**: 349-357).

[0043] Isolation of total RNA from cells was carried out using the RNeasy Kit (Qiagen, Santa Clarita, CA), reverse transcribed to cDNA with random hexamers using the TaqMan® Reverse Transcription Reagents as per manufacturer's instructions (Perkin Elmer, Foster City, CA). Quantitative rtPCR was performed on an ABI Prism 7700 Sequence Detection System (Perkin Elmer) (Wandinger *et al.* 2001, *Ann Neurol* **50**: 349-357). Amplification of hypoxanthine phosphoryltransferase (HPRT) and 18S rRNA for stimulated conditions was used for sample normalization. The amplification protocol followed the suggestions of the TaqMan® Gold RT-PCR kit. For detection of TRAIL, CD95L, MxA and HPRT transcripts, oligonucleotides were used at final concentrations of 200 nM for forward and reverse primer and 100 nM for the fluorogenic probe as follows:

TRAIL forward: 5'CAGAGGAAGAAGCAACACATTCTCT3',
 reverse: 5'TGATGATTCCCAGGAGTTTATTTTG3',
 probe: 5'FAM-ACTCCAAGAATGAAAAGGCTCTGGGCC-TAMRA3';

CD95L forward: 5'ATGCACACAGCATCATCTTTGG3',
 reverse: 5'ATGGGCCACTTTCCTCAGCT3',
 probe: 5'FAM-AAGCAAATAGGCCACCCCAGTCCACC-TAMRA3';

MxA forward: 5'CAGCACCTGATGGCCTATCAC3',
 reverse: 5'GAGCATGAAGAACTGGATGATCAA3',
 probe: 5'FAM-AGCAAGCGCATCTCCAGCCACATC-TAMRA3';

HPRT forward: 5'AGTCTGGCTTATATCCAACACTTCG3',
 reverse: 5'GACTTTGCTTTCCTTGGTCAGG3',
 probe: 5'FAM-TTTCACCAGCAAGCTTGCGACCTTGA-TAMRA3'.

[0044] Quantification of gene expression relative to HPRT or 18S rRNA was calculated by the protocol's $\Delta\Delta C_T$ method. 18S rRNA was amplified using TaqMan® Ribosomal RNA Control Reagents (Perkin Elmer). Changes in gene expression of twofold or more were considered valid for further statistical analysis.

[0045] Statistical analysis was performed using SPSS software (SPSS Software GmbH, Munich, Germany). The Mann-Whitney-U test was used to compare data between patient groups. For analysis of dependent variables within groups, Wilcoxon's matched pair signed-rank test was performed. Data obtained from experiments outlined below were evaluated using the same statistical approach.

[0046] In Fig. 1 it is shown that, compared to baseline, spontaneous gene expression of both MxA and TRAIL was significantly up-regulated in PBMC of MS patients after 3 weeks of treatment and remained elevated for a period of 1 year. In contrast, CD95L did not show any significant regulation during IFN- β therapy at each time-point tested.

Example 2: TRAIL gene expression in IFN- β responders and non-responders

[0047] In order to examine the functional relevance of TRAIL induction for the mechanism of action of IFN- β , patients were characterized with regard to their clinical response status. Patients negative for NAB who experienced no further relapses and no deterioration in the EDSS during IFN- β treatment were defined as positive drug responders, whereas patients who continued to have one or more relapses were defined as non-responders.

[0048] As illustrated in Fig. 2, IFN- β therapy resulted in early and sustained up-regulation of TRAIL gene expression in MS patients with clinical response. Conversely, TRAIL gene expression was only transiently induced after 6 months of treatment in drug non-responders. The biological response marker MxA was significantly up-regulated in both groups (Fig. 2A), indicating the *in vivo* biological effectiveness of IFN- β -1a in both patient groups. As outlined in Fig. 2A, non-responders showed even higher levels of MxA gene expression in response to systemic IFN- β treatment at each time point investigated. As for the whole group, CD95L was not significantly regulated by IFN- β therapy in the patient subsets. No differences in baseline expression levels of the genes investigated were observed between responders and non-responders (data not shown).

Example 3: TRAIL gene expression in patients developing neutralizing antibodies

[0049] Given the parallel up-regulation of MxA and TRAIL expression by IFN- β *in vivo*, it was investigated whether the occurrence of NAB in the course of IFN- β treatment influences TRAIL gene expression as reported for MxA (Deisenhammer *et al.* 1999, *Neurology* **52**: 1239-1243). Therefore, spontaneous gene expression was analyzed in a third subset of patients who developed NAB after 1 year (n=23). Presence of NAB was confirmed after 18 months (*cf.* Table 1).

[0050] Fig. 3 demonstrates that in these subjects, bioavailability of IFN- β as measured by MxA gene induction was completely inhibited in the presence of NAB. In parallel, initial up-regulation of TRAIL gene expression by IFN- β was abrogated, indicating a direct regulation of TRAIL transcription by IFN- β . The expression of CD95L did not show significant variations in the presence of NAB (data not shown).

[0051] Interestingly, the majority of these patients remained in the clinical response status in the study time. However, this clinical observation is in keeping with the results of the pivotal trial of IFN- β -1b and the extension of the PRISMS study of IFN- β -1a, where development of NAB was finally associated with a delayed loss of efficacy in an observation period between 18 and 24 months and after 24 months, respectively (The IFNB Multiple Sclerosis Group and the University of British Columbia MS/MRI Analysis Group 1996, *Neurology* **47**: 889-894; The PRISMS Study Group and The University of British Columbia MS/MRI Analysis Group. PRISMS-4, 2001, *Neurology* **56**: 1628-1636).

Example 4: Protein expression of TRAIL in IFN- β responders and non-responders

[0052] To address the question whether IFN- β non-responders differed from responders also with regard to TRAIL protein expression, serum concentrations of sTRAIL protein before and during IFN- β -1a therapy was measured.

[0053] Serum of NAB negative drug responders (n=12) and non-responders (n=11), from whom serum was available for investigation, was stored at -20°C until the assay were performed. Soluble TRAIL protein was quantified using a sandwich ELISA with a detection limit of 65 pg/ml according to the manufacturer's protocol (Trinova Biochem, Gießen, Germany).

[0054] As demonstrated in Fig. 4, responders showed significantly higher sTRAIL baseline levels already prior to beginning of treatment, indicating underlying differences in the post-transcriptional regulation-of TRAIL expression. Levels of sTRAIL above 350 pg/ml were definitely associated with response to IFN- β therapy. Thus, levels of sTRAIL above or below 350 pg/ml prior to therapy predicted the clinical outcome in 92% of the positive responders and 73% of the non-responders, respectively. For the total group of patients, this implied a correct prediction rate of 83,6%. Compared to baseline levels, sTRAIL protein accumulation in the serum was significantly enhanced after 4 weeks of IFN- β treatment in responders and non-responders. In long-term treated patients, sTRAIL protein levels remained significantly up-regulated in positive responders compared to non-responders after 6 and 12 months.

[0055] In addition to sTRAIL in blood serum, membrane bound TRAIL protein expressed on PBMC was measured using flow cytometry. PBMC from NAB negative responders and non-responders before and during IFN- β -1a therapy were prepared and 2×10^5 PBMC per sample were incubated with an anti-TRAIL monoclonal antibody (cat. no. 804-322-C100, Alexis Corporation, San Diego, CA) or control mouse IgG1 (cat. no. X0931, DAKO, Glostrup, Denmark) at a concentration of 1:100 for 30 min at 4°C. Washed cells were incubated with a phycoerythrin (PE)-labeled secondary goat-anti-mouse antibody (cat. no. R0480, DAKO) at a concentration of 1:20 for 30 min. After washing with phosphate buffered saline (PBS), cells were analyzed on a FACScan flow cytometer and data were processed using the Cellquest software (Becton-Dickinson, Mountain View, USA). Values are expressed as specific fluorescence intensity, calculated by histogram statistics and defined by the percentage of fluorescence intensity with TRAIL antibody excluding the fluorescence intensity with the appropriate control antibody.

[0056] Significant changes in TRAIL surface expression on PBMC during IFN- β treatment in any of the patient subsets were not observed (data not shown). Since blood samples were collected from patients 3 days after IFN- β application (subcutaneous injection) throughout the study, the negative findings might be explained by the time course of membrane bound TRAIL expression.

Example 5: TRAIL gene and protein expression in PBMC cell culture

[0057] PBMC were cultured in the presence of IFN- β -1a *in vitro* and the regulation of TRAIL gene transcription, cell surface expression and accumulation of soluble protein in culture supernates were investigated in comparison (*cf.* Fig. 5).

[0058] Freshly thawed PBMC were resuspended in Iscove's modified Dulbecco's medium (Gibco, Grand Island, NY) supplemented with 2 mM L-glutamine, 50 mg/ml gentamycin and 100 U/ml penicillin/streptomycin (Whittaker Bioproducts, Gaithersburg, MD) and 5% human plasma. PBMC (10^6 cells per well) were incubated in a humidified atmosphere of 5% CO₂ at 37°C. Where specified, phytohaemagglutinin (PHA) or cytokines were added to the cultures as follows:

PHA 1 µg/ml (Sigma-Aldrich, Steinheim, Germany), recombinant human IFN-β-1a, 10 and 100 IU/ml (Rebif®, Serono, Unterschleissheim, Germany), recombinant human IL-2, 10 IU/ml (Proleukin, Eurocetus, Frankfurt, Germany).

[0059] As demonstrated in Fig. 5A, incubation of PBMC with IFN-β resulted in rapid, dose-dependent up-regulation of TRAIL mRNA levels after 24 hrs, followed by a second peak after 72 hrs. TRAIL-expression was likewise rapidly induced on the surface of PBMC after 24 hrs by IFN-β (Fig. 5B). However, expression of membrane bound TRAIL was only marginally enhanced compared to baseline levels after 72 hrs. Soluble TRAIL concentrations in culture supernates increased over 24 hrs and remained elevated for 96 hrs when stimulated with IFN-β at a concentration of 10 IU/ml (Fig. 5C). Incubation with IFN-β at a concentration of 1.00 IU/ml resulted in further increase of sTRAIL over 96 hrs. In contrast to IFN-β, stimulation of cells with PHA and IL-2 resulted in delayed TRAIL gene and protein expression (Fig. 5A-C). Consistent with previously reported findings, these data indicate that IFN-β directly regulates the transcriptional control of TRAIL expression (Kayagaki *et al.* 1999, *J Exp Med* **189**:1451-1460).

[0060] The features disclosed in the foregoing description, in the claims and/or in the accompanying drawings may, both separately and in any combination thereof, be material for realizing the invention in diverse forms thereof.

SEQUENCE LISTING

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 <120> Method for identifying type-I Interferon responsive MS patients by determining
 TRAIL expression
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Claims

1. A method for identifying an individual as a positive responder or a non-responder to a type-I Interferon (IFN) therapy used in the treatment of Multiple Sclerosis, **characterized in that** TRAIL expression and/or TRAIL protein level is determined.
2. Method according to claim 1, wherein the therapy used is an IFN- β therapy.
3. Method according to claim 1 or 2, wherein TRAIL expression determination and/or TRAIL protein level determination is done prior to IFN therapy.
4. Method according to claims 1-3, wherein TRAIL expression determination and/or TRAIL protein level determination is carried out with a sample of mammalian origin.
5. Method according to claim 4, wherein the mammalian is a human.
6. Method according to claim 5, wherein the sample is selected from the group comprising a bodily fluid, a fraction thereof, tissue extract and, cell extract.
7. Method according to claim 6, wherein the bodily fluid is selected from the group comprising whole blood, a fraction thereof, blood serum, plasma, cerebrospinal fluid, synovial fluid, ascites exsudate, inflammatory exsudate, and urine.
8. Method according to any of the preceding claims, wherein
 - (a) TRAIL expression is determined by detecting and/or measuring the amount and/or concentration of TRAIL nucleic acids encoding the TRAIL polypeptide or parts thereof, and
 - (b) TRAIL protein level is determined by detecting and/or measuring the amount and/or concentration of TRAIL polypeptide or parts thereof.
9. Method according to claim 8, wherein TRAIL protein level determination is performed by measuring the amount and/or concentration of soluble TRAIL protein in a liquid sample.

10. Method according to any of claims 8 and 9, wherein measurement of the amount and/or concentration of TRAIL polypeptide is done by an immunoassay using an anti-TRAIL antibody.
11. Method according to claims 8-10, wherein the amount and/or concentration of TRAIL polypeptide determined is compared to a limiting value discriminating positive responders and non-responders.
12. Method according to claim 11, wherein the limiting value is 350 ± 20 pg/ml.
13. Method according to claim 8, wherein the amount and/or concentration of TRAIL nucleic acids is determined by measuring the amount and/or concentration of TRAIL mRNA in a sample according to claims 4-6.
14. Method according to claim 13, wherein the sample is selected from the group comprising an extract of blood cells, peripheral immune cells, neuronal tissue and muscle tissue.
15. Method according to any of claims 13 and 14, wherein measurement of the amount and/or concentration of TRAIL mRNA is performed with the aid of the polymerase chain reaction using one or more TRAIL specific probes.
16. Method according to any of claims 13-15, wherein the amount and/or concentration of TRAIL mRNA is compared to a limiting value discriminating positive responders and non-responders.
17. Method according to any of the preceding claims, wherein the amount and/or concentration of TRAIL nucleic acids is correlated with the amount and/or concentration of nucleic acids of one or more biological activity marker(s), and/or the amount and/or concentration of TRAIL polypeptide is correlated with the polypeptide amount and/or concentration of one or more biological activity marker(s).
18. Method according to claim 17, wherein the biological activity marker is chosen from the group comprising MxA protein or any other IFN-inducible protein.
19. Use of a method according to any of claims 1-18 for monitoring IFN therapy used in the treatment of Multiple Sclerosis.
20. Kit for performing a method according to any of claims 1-19, comprising a nucleic acid probe and/or primer(s) specific for the TRAIL gene or a portion thereof and/or an antibody specific for the TRAIL polypeptide or a portion thereof.

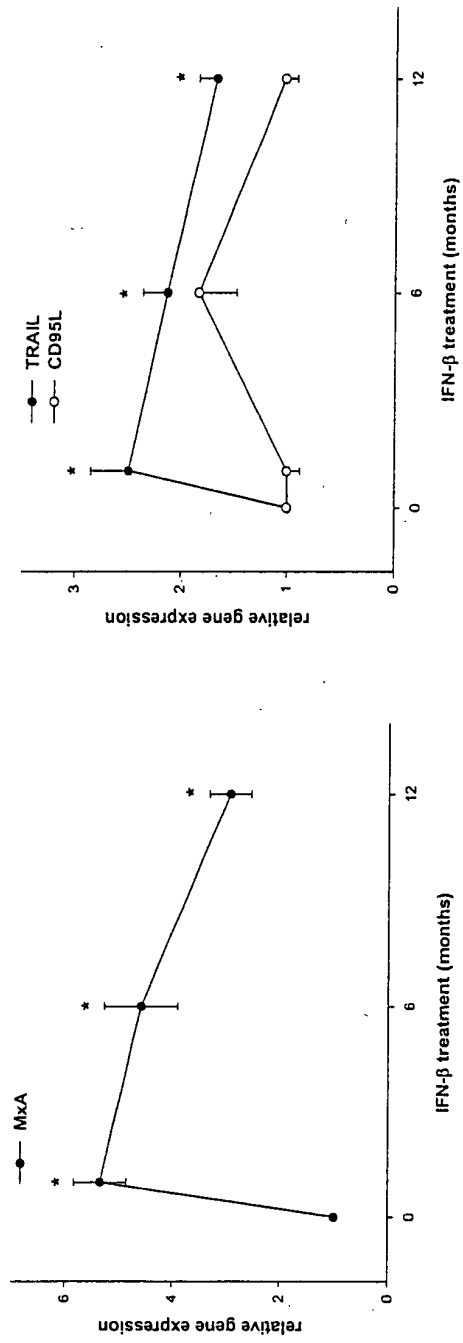
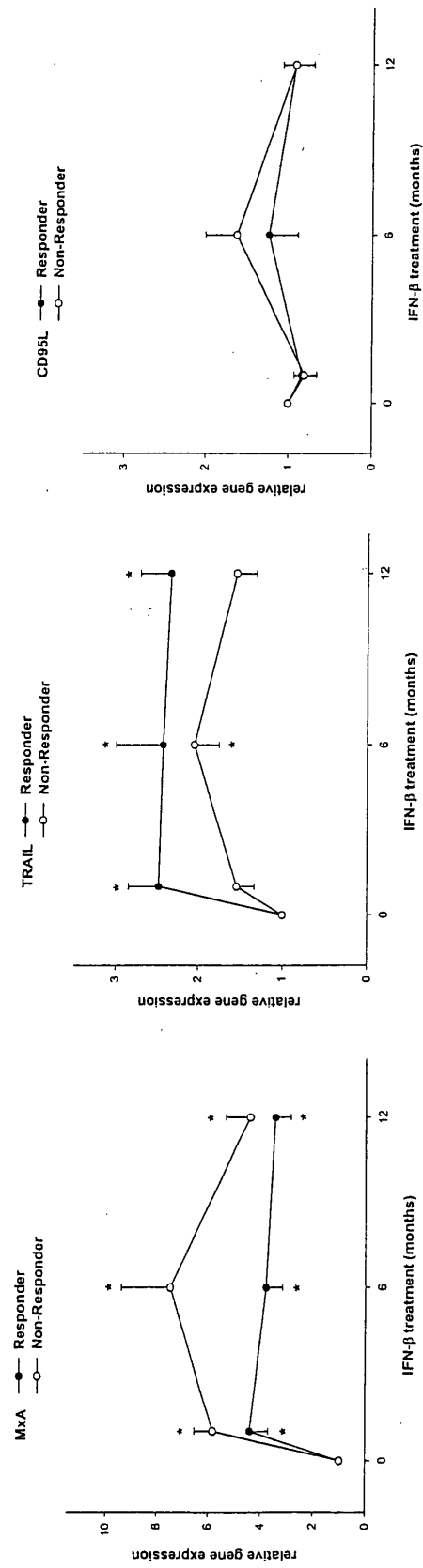


Figure 1

Figure 2



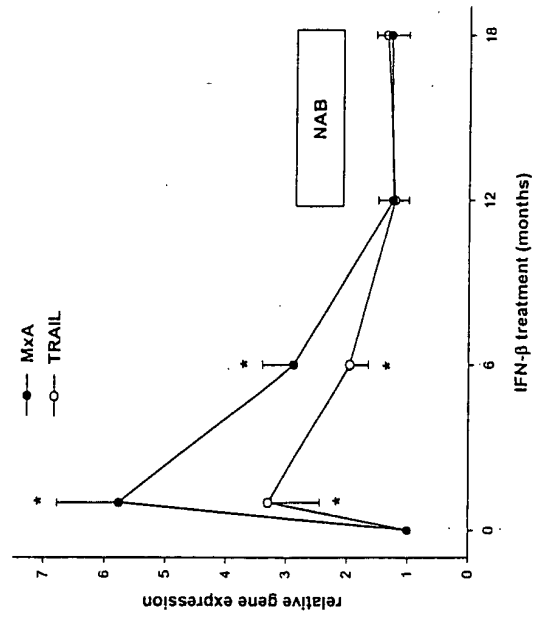


Figure 3

Figure 4

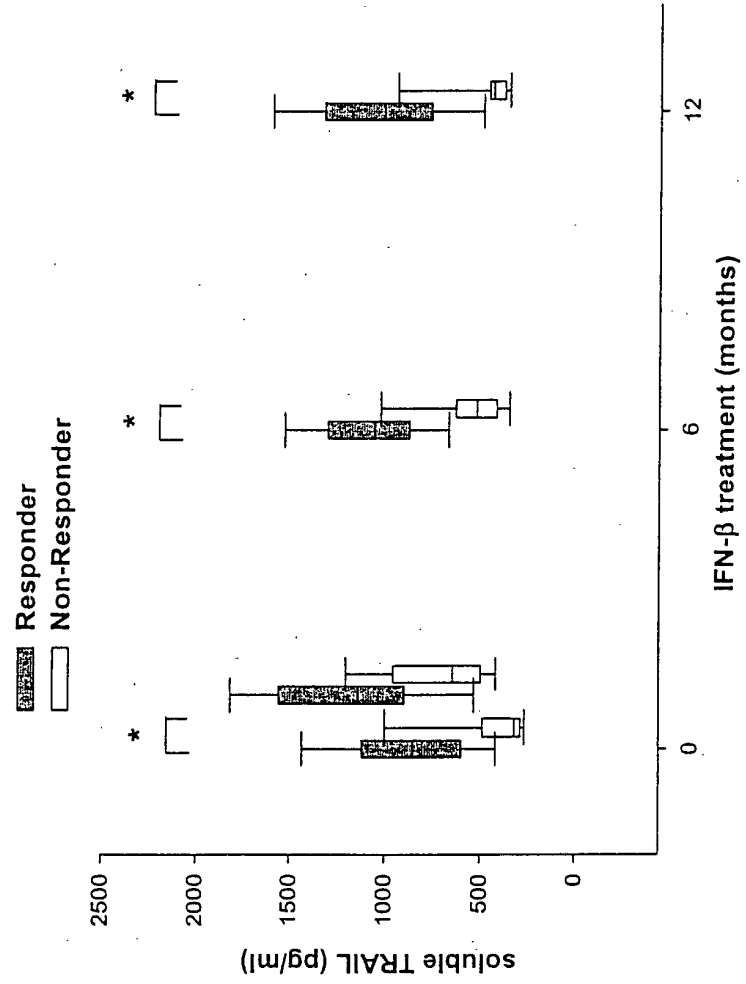
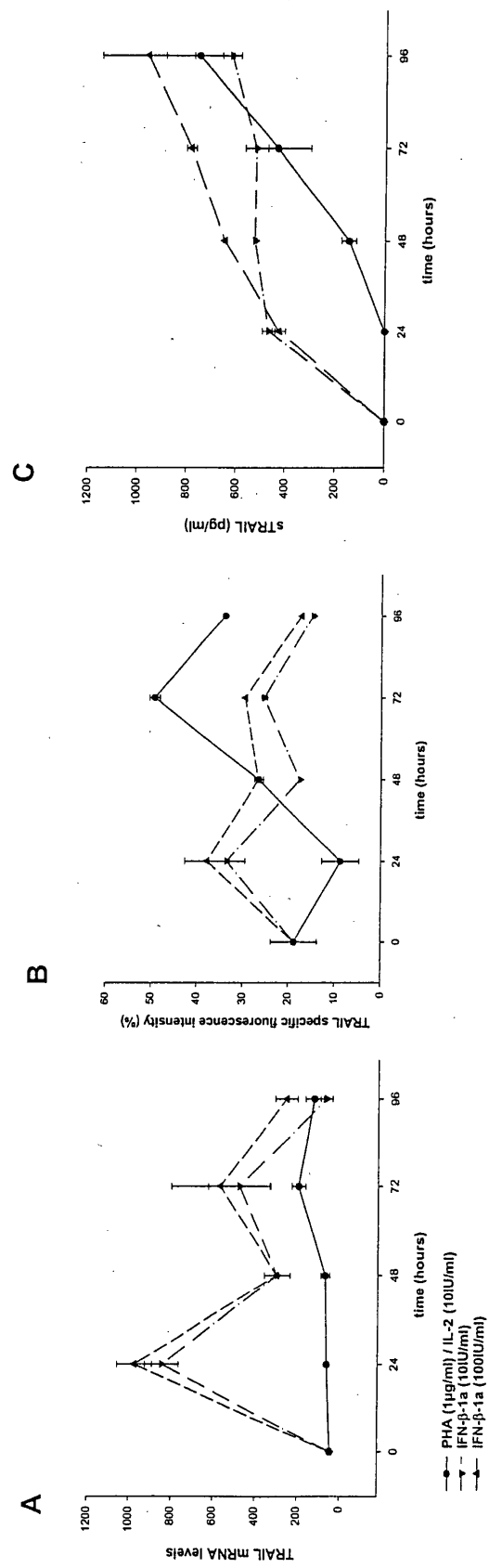


Figure 5





European Patent
Office

PARTIAL EUROPEAN SEARCH REPORT

Application Number

which under Rule 45 of the European Patent Convention shall be considered, for the purposes of subsequent proceedings, as the European search report

EP 02 01 1964

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.Cl.7)
D,X	WANDINGER KLAUS-PETER ET AL: "Complex immunomodulatory effects of interferon-beta in multiple sclerosis include the upregulation of T helper 1-associated marker genes." ANNALS OF NEUROLOGY, vol. 50, no. 3, September 2001 (2001-09), pages 349-357, XP008010479 ISSN: 0364-5134	20	G01N33/68 C07K14/705
A	* abstract; table 1 * * page 355 *	1-19	
X	WO 97 33899 A (HUMAN GENOME SCIENCES INC; RUBEN STEVEN M (US)) 18 September 1997 (1997-09-18)	20	
A	* abstract; claim 21; example 4 * * page 44, paragraph 2 *	1-19	
A	FR 2 766 713 A (BIO MERIEUX) 5 February 1999 (1999-02-05) * claims; examples *	1-20	
	--- -/--		
			TECHNICAL FIELDS SEARCHED (Int.Cl.7)
			G01N C07K G01B
INCOMPLETE SEARCH			
The Search Division considers that the present application, or one or more of its claims, does/do not comply with the EPC to such an extent that a meaningful search into the state of the art cannot be carried out, or can only be carried out partially, for these claims. Claims searched completely : Claims searched incompletely : Claims not searched : Reason for the limitation of the search: see sheet C			
Place of search		Date of completion of the search	Examiner
THE HAGUE		28 November 2002	Vadot-Van Geldre, E
CATEGORY OF CITED DOCUMENTS		T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document	
X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document			

EPO FORM 1503 03.82 (P04C07)



European Patent
Office

INCOMPLETE SEARCH
SHEET C

Application Number
EP 02 01 1964

Although claims 1-19 are directed to a method of treatment of the human/animal body (Article 52(4) EPC), the search has been carried out and based on the alleged effects of the compound/composition.

Claim(s) searched incompletely:
1-19

Reason for the limitation of the search (non-patentable invention(s)):

Article 52 (4) EPC - Method for treatment of the human or animal body by therapy
Article 52 (4) EPC - Method for treatment of the human or animal body by therapy



European Patent
Office

PARTIAL EUROPEAN SEARCH REPORT

Application Number
EP 02 01 1964

DOCUMENTS CONSIDERED TO BE RELEVANT			CLASSIFICATION OF THE APPLICATION (Int.Cl.7)
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	
A	WOSIK KAROLINA ET AL: "Interferon b (IFNb) modulates death receptor DR4 and DR5 expression on oligodendrocytes and their ligand TRAIL on T cells." NEUROLOGY, vol. 56, no. 8 Supplement 3, 24 April 2001 (2001-04-24), page A152 XP008010460 53rd Annual Meeting of the American Academy of Neurology; Philadelphia, PA, USA; May 05-11, 2001 ISSN: 0028-3878 * abstract * ---	1-19	
A	WANDINGER KLAUS-PETER ET AL: "Diminished production of type-I interferons and interleukin-2 in patients with multiple sclerosis." JOURNAL OF THE NEUROLOGICAL SCIENCES, vol. 149, no. 1, 1997, pages 87-93, XP002220790 ISSN: 0022-510X * abstract * ---	1-19	TECHNICAL FIELDS SEARCHED (Int.Cl.7)
A	JEFFERY DOUGLAS R: "Relationship between disease activity and dose-response relationships with beta interferon therapies in the treatment of multiple sclerosis." JOURNAL OF THE NEUROLOGICAL SCIENCES, vol. 178, no. 1, 1 September 2000 (2000-09-01), pages 2-9, XP002220791 ISSN: 0022-510X * page 2 - page 3, column 1 * -----	1-19	

**ANNEX TO THE EUROPEAN SEARCH REPORT
ON EUROPEAN PATENT APPLICATION NO.**

EP 02 01 1964

This annex lists the patent family members relating to the patent documents cited in the above-mentioned European search report. The members are as contained in the European Patent Office EDP file on
The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

28-11-2002

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WO 9733899	A	18-09-1997	WO	9733899 A1	18-09-1997
			AU	5711196 A	01-10-1997

FR 2766713	A	05-02-1999	FR	2766713 A1	05-02-1999
			WO	9907408 A1	18-02-1999

EPO FORM P0459

For more details about this annex : see Official Journal of the European Patent Office, No. 12/82

专利名称(译)	通过确定TRAIL表达来鉴定I型干扰素应答MS患者的方法		
公开(公告)号	EP1369695A1	公开(公告)日	2003-12-10
申请号	EP2002011964	申请日	2002-05-29
[标]申请(专利权)人(译)	UNIVSKLINIKUM CHARITE DER ZU柏林洪堡UNIV		
申请(专利权)人(译)	柏林DER的UNIVERSITÄTSKLINIKUM慈善洪堡大学		
当前申请(专利权)人(译)	CHARITE - UNIVERSITAETSMEDIZIN BERLIN		
[标]发明人	THE DESIGNATION OF THE INVENTOR HAS NOT YET BEEN FILED		
发明人	THE DESIGNATION OF THE INVENTOR HAS NOT YET BEEN FILED		
IPC分类号	G01N33/50 C07K14/705 C12N15/09 C12Q1/02 C12Q1/68 G01N33/15 G01N33/53 G01N33/68		
CPC分类号	G01N33/6863 G01N2333/70575		
其他公开文献	EP1369695B1		
外部链接	Espacenet		

摘要(译)

本发明涉及一种鉴定个体，特别是人，作为I型干扰素（IFN）疗法的阳性反应者或非反应者的方法，特别是IFN-β疗法，用于治疗多发性硬化症。确定TRAIL基因表达和/或TRAIL蛋白水平。本发明还涉及所述方法在治疗多发性硬化症中用于监测IFN疗法的用途，以及用于实施所述方法的试剂盒。

TRAIL forward: 5'CAGAGGAAGAAGCAACACATTCTCT3',
reverse: 5'TGATGATCCAGGAGTTTATTTTG3',
probe: 5'FAM-ACTCCAAGAATGAAAAGGCTCTGGGC-TAMRA3';