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(54) **Identification of a capacitative calcium channel in antigen presenting cells and uses thereof**

(57) The present invention relates to the identification of a capacitative calcium channel homologue for the immune system activation, and its use to report and to modulate the activity of immune cells *in vitro*, *ex vivo* or *in vivo*. This invention more specifically discloses that a Mln1 gene product, represents a capacitative calcium channel in immune cells such as macrophages, monocytes, T-cells, B-cells and mast-cells. This invention is a proven identification of a gene ex-

pressing a capacitative calcium channel in immune cells, and can be used in various compositions and methods for monitoring or modulating an immune response in a subject. The present invention can be used to develop biomarkers for immune system activation or inflammatory responses or to screen for specific immune system activity altering drugs.

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## Description

**[0001]** The present invention relates to the characterisation of a capacitative calcium channel homologue for the immune system activation, and its use to report or to modulate the activity of immune cells *in vitro*, *ex vivo* or *in vivo*. This invention more specifically discloses that a mlsn1 gene product represents a capacitative calcium channel in immune cells such as macrophages, monocytes, T-cells, B-cells and mast-cells. This invention is a proven identification of a gene expressing a capacitative calcium channel in immune cells, and can be used in various compositions and methods for monitoring or modulating an immune response in a subject. The present invention can be used to develop biomarkers for immune system activation or inflammatory responses or to screen for specific immune system activity altering drugs.

## BACKGROUND OF THE INVENTION

**[0002]** Regulated entry of calcium across the plasma membrane is an essential signaling mechanism, implicated in phenomena like exocytosis, contraction, gene expression, and cell differentiation (1). Store operated channels (SOCs), defined as channels that open in response to depletion of intra-cellular calcium stores, represent one of the most ubiquitous mechanisms for triggering calcium influx in non-excitabile cells.

**[0003]** A large number of cell surface receptors are coupled through G-proteins or tyrosine kinases to the activation of phospholipase C (PLC) and consequently the generation of inositol 1,4,5-triphosphate (IP3) (2). Increased IP3, acting on its receptor (3) results in an increase of intra-cellular calcium ( $[Ca^{2+}]_i$ ) through depletion of intra-cellular calcium stores. How depletion of intra-cellular calcium stores triggers  $Ca^{2+}$  entry through the cell membrane is still controversial. The simplest mechanism would involve a direct action of IP3, or of the IP3 receptor, on  $Ca^{2+}$  channels in the plasma membrane. On the other hand depletion of stores might send an activating signal of some kind to channels in the plasma membrane (4). Plasma membrane  $Ca^{2+}$  channel activation results in a small inward current in parallel with a sustained raise in  $[Ca^{2+}]_i$ . Such a current has been shown to be induced by intra-cellular dialysis with IP3 in rat mastcells (5) or by Thapsigargin-activation of Jurkat T-cells (6). Thapsigargin (TG), a plant derived sesquiterpene lactone, depletes IP3-sensitive stores through its ability to inhibit  $Ca^{2+}$ -ATPases (7). Several groups have shown that TG depletion of intracellular calcium stores in T-cells activates a  $Ca^{2+}$  influx that was similar to and not additive with that evoked by T-Cell Receptor (TCR) stimulation (8-10).

**[0004]** SOCs have a high diversity. The essential defining feature of these channels is that they are activated by a variety of stimuli that deplete intra-cellular stores such as agonists to phosphoinositide-linked receptors, intra-cellular IP3,  $Ca^{2+}$  ionophores, inhibitors of SERCA-type  $Ca^{2+}$  ATP ases, and intra-cellular dialysis with buffered solutions containing low concentrations of  $Ca^{2+}$  ( $[Ca^{2+}] < 50$  nM) (1). IP3 activation of SOCs implies that the channel is not directly coupled to the receptor and artificial store depletion activation indicates that SOC activation responds to store  $Ca^{2+}$  depletion after IP3-receptor binding than to IP3 itself. SOCs are generally described as lacking voltage dependent gating.

**[0005]** Calcium entry through Icrac has been shown to accelerate the rate of exocytosis in mast cells. It inhibits adenylyl cyclase type VI, but activates adenylyl cyclase type I. Icrac activity might be regulated either directly or indirectly through the concentration of intra-cellular  $Ca^{2+}$  (13). An increase in the intra-cellular calcium ion concentration controls a diverse range of cell functions, including adhesion, mobility, gene expression and proliferation can occur as single transients, repetitive oscillations or a sustained plateau (14). Two recent reports have suggested that the amplitude, duration and frequency of a  $Ca^{2+}$  signal in cells of the immune system control differential activation of proinflammatory transcriptional regulators NF- $\kappa$ B, JNK and NF-AT (15; 16). Thus Calcium signalling could play a crucial role in the activation of cells of the immune system. The identification of an immune system specific Calcium channel would therefore represent an important step forward in determining the pathology of Icrac in disease states. Mlsn1 and its gene products could therefore be used as a basis of screens to identify new therapeutic compounds such as immune system suppressors, including anti-allergic or anti-inflammatory compounds, or immune system activators for the treatment of immuno degenerative disorders. Mlsn1 could as well represent a powerful biomarker for onset and progression of autoimmune or inflammatory diseases (e.g. COPD, Arthritis, organ rejection).

## SUMMARY

**[0006]** The present invention discloses the characterisation of a capacitative  $Ca^{2+}$  channel in immune cells, more specifically in macrophages. MRNA expression is differentially increased in monocytes to macrophages differentiation as well as in macrophages activation.

**[0007]** More particularly, the inventors have identified the presence of a novel calcium pore sequence within the products of the mlsn1 gene, and this calcium pore is part of the major capacitative calcium channel in immune system cells, more specifically in macrophages.

**[0008]** The present invention shows that a product of the mlsn1 gene is expressed in the U937 cells (monocyte like)

specifically after 48h differentiation to a more macrophage like cell type and that mlsn1 regulates capacitative calcium entry in these cells. Expression is further increased by macrophages activation. PCR subtraction experiments show that mlsn1 gene expression is several times upregulated, which is in concordance with the previously established differential increases of the Calcium signalling phenotype during this differentiation step (18).

[0009] This invention also shows that particular mlsn1-specific anti-sense nucleic acids down regulate calcium entry in U937 cells differentiated to a macrophage phenotype. This is of particular importance since it is the first time that a sequence specific tool has succeeded to inhibit the immune cell Calcium signalling, thereby associating a given gene to the  $Ca^{2+}$  pore gene expression.

[0010] This invention represents the characterisation of a SOC channel in immuno-modulatory cells, more specifically in macrophages. This invention provides a mean 1) to use it as a modulator of immune activation 2) to use it as a biomarker for immune activation and inflammation 3) to identify new targets for the screening of immuno-modulators.

[0011] The described invention further shows, the nucleic acid sequence of the Mlsn1 transcriptional promoter elements 5' to the putative Mlsn1 start codon.

[0012] An object of the present invention resides in the use of a mlsn1 gene, gene product(s) and transcription promoters for the screening of compounds that modulate the activity of an lcrac expressing immune cell, more specifically of an antigen presenting cell, most specifically of a compound that modulates the activity of macrophages.

[0013] Further objects of the present invention include Mlsn1 peptides, nucleic acids, vectors, and recombinant cells, which can be used for therapeutic (as target) or diagnostic approaches (as biomarker).

## BRIEF DESCRIPTION OF THE FIGURES

### [0014]

Figure 1: Amino acid sequence of Mlsn1 with predicted secondary structure.

Where; <<N-TERMINUS>> represents the N-terminal domain

_TMD1 to 6	represent the transmembrane domains
--EXL1 to 3--	represents the extra-cellular loop
--INL1 and 2--	represent the intra-cellular loops
<<C-TERMINUS>>	represents the C-terminal domain

Figure 2: Represents the analysis on agarose of PCR products obtained with Mlsn1-specific primer pairs from differentiated or undifferentiated U937 macrophage cells.

Figure 3: Illustrates the Mlsn1(TrpC8) expression analysis by cDNA subtraction.

1 : dbcAMP differentiated U937 subtracted by undifferentiated U937.

2 : unsubtracted dbcAMP differentiated U937 control.

3 : undifferentiated U937 subtracted by dbcAMP differentiated U937.

4 : unsubtracted undifferentiated U937 control.

5 : 1kb ladder

PCR have been carried out on subtraction DNA fragments using "Not 1a" and "Not 2b" Mlsn1 specific primers.

Figure 4: Down regulation of calcium entry during dbcAMP differentiation of U937 cells using Mlsn1 anti-sense - Calcium flux analysis using Indo-1 and flow cytometry

a : Undifferentiated U937

b : dbcAMP differentiated U937

c : dbcAMP differentiated U937 treated with 10  $\mu$ M scrambled oligonucleotide

d : dbcAMP differentiated U937 treated with 1  $\mu$ M Mlsn1 anti-sense

e : dbcAMP differentiated U937 treated with 10  $\mu$ M Mlsn1 anti-sense

## DETAILED DESCRIPTION OF THE INVENTION

[0015] An object of the present invention resides in the characterisation of functional components of Mlsn1. The polypeptide structure of Mlsn1 has been determined. Nucleic acid sequences and peptides which encode the N- and

C-terminal regions have been identified. Also, nucleic acid sequences and peptides which encode transmembrane domains and intra- and extra-cellular loop regions have been identified. Crucially, nucleic acid and peptide sequences which encode the Mlsn1 calcium pore region have been identified. The above nucleic acid and peptide sequences offer vital target molecules for the development of therapeutic molecules to combat disorders associated with aberrant Icrac function.

**[0016]** Furthermore, the inventors have identified putative transcription promoter elements 5' to the nucleic acid encoding Methionine-1 start codon of Mlsn1.

**[0017]** Another object of this invention resides, generally, in the use of a mlsn1 gene, gene product, fragments thereof, or transcriptional promoter elements for the screening of compound(s) that modulate the activity of an immune cell, more specifically for the screening of a compound that modulates the activity of monocytes, macrophages, T-cells, B-cells and mast-cells *in vitro*, *ex vivo* or *in vivo*, most specifically for the screening of compounds that modulate the activity of macrophages *in vitro*, *ex vivo* or *in vivo*. This invention also relates to particular Mlsn1 polypeptides and nucleic acids, methods of screening of immuno-modulators.

**[0018]** Other aspects of the present invention relate to:

**[0019]** Methods of screening for immuno-modulatory compounds, comprising contacting a test compound with a Mlsn1 gene or a Mlsn1 gene product or Mlsn1 transcription promoter element(s) and determining the ability of said test compound to interact with the Mlsn1 gene or a Mlsn1 gene product or Mlsn1 transcriptional promoter element(s).

**[0020]** Methods of screening for identification of new (known or unknown) immuno-modulatory targets after modulation of the Mlsn1 gene, Mlsn1 gene product activity or transcription promoter activity.

**[0021]** Further aspects of the present invention relate to:

**[0022]** The use of mlsn1 gene, gene products or nucleic acids to report and/or to modulate the activity of Mlsn1 in immune cells *in vitro*, *ex vivo* or *in vivo*.

**[0023]** Within the context of the present invention, the term mlsn1 gene designates a nucleotide sequence encoding the Mlsn1 protein, any nucleotide sequence encoding a fragment of the Mlsn1 protein, or any variants or homologs thereof. Variants or homologs designate more particularly any naturally-existing human genes that would exhibit sequence variation(s) resulting from polymorphism(s), mutation(s), or other alteration(s) as compared to the above sequence, without depriving the property of the encoded protein to function as an Icrac channel. Homologs or variants also include any recombinant DNA molecule, such as a cDNA molecule, encoding a Mlsn1 protein. Further variants also include any sequence that would hybridize with the above nucleic acid sequences under stringent conditions and encode a functionally active Icrac channel. Nucleotide sequences encoding fragments of the Mlsn1 protein which represent a biological relevant region of this protein such as the N- and C- terminal domains, the transmembrane domains, the intra and extra-cellular loops and the predicted calcium pore as well as the polypeptides representing these biologically relevant regions are also included.

**[0024]** As an illustrative embodiment, stringent hybridization conditions can be defined as follows:

**[0025]** The hybridization step is conducted at 65°C in the presence of 6 x SSC buffer, 5 x Denhardt's solution, 0.5 % SDS and 100µg/ml of salmon sperm DNA.

**[0026]** The hybridization step is followed by four washing steps:

- two washings during 5 minutes, preferably at 65°C in a 2 x SSC and 0.1% SDS buffer;
- one washing during 30 minutes, preferably at 65°C in a 2 x SSC and 0.1% SDS buffer;
- one washing during 10 minutes, preferably at 35°C in a 0.1 x SSC and 0.1% SDS buffer,

it being understood that the hybridization conditions defined above are suitable for nucleic acids of approximately twenty nucleotides in length and that these conditions may be also adapted for shorter or longer nucleic acids, according to techniques well known in the art, for example those described by Sambrook et al. (1989).

**[0027]** A Mlsn1 gene product designates, in accordance with the present invention, polypeptides (or proteins) encoded by a mlsn1 gene as described above. The Mlsn1 gene product may comprise several polypeptides, resulting from alternative splicings of a mlsn1 gene during transcription. More specific Mlsn1 gene products of this invention are encoded by a mlsn1 gene as described above and/or comprise the amino acid sequence of Mlsn1, such as that of SEQ ID N°1.

**[0028]** Also within the context of the present invention, the term Mlsn1 transcription promoter(s) designates nucleotide sequence(s) encoding Mlsn1 transcription promoter element(s), nucleotide sequence encoding a fragment thereof, or any variants or homologs thereof. Variants or homologs designate more particularly any human transcription promoter elements that would exhibit sequence variation(s) resulting from polymorphism(s), mutation(s), or other alteration(s) as compared to the above sequence, without depriving the property of the nucleic acid sequences to function as an Icrac transcription promoter. Further variants also include any sequence that would hybridize with the above nucleic acid sequences under stringent conditions and encode a functionally active transcription promoter.

**[0029]** A Mlsn1 transcription promoter element designates, in accordance with the present invention, encoded by a

Mlsn1 nucleic acid sequence as described above. The transcription promoter elements is further characterized by being located 5' the Mlsn1 start codon. More specific Mlsn1 transcription promoter elements of this invention are encoded by Mlsn1 nucleic acid sequences of SEQ ID N°41, 42 or 43.

## NUCLEOTIDE AND AMINO ACID SEQUENCE ENCODING Mlsn1

**[0030]** The human mlsn1 cDNA has been described in US Patent 5,674,739 as FOHY030. This gene is described as being expressed specifically in certain tumor cells and not in healthy tissues and was thus characterized as involved in tumor progression. The sequence of human mlsn1 cDNA is also available from Genbank (accession number AF071787). The published nucleic acid sequence encoding the open reading frame of human Mlsn1 is predicted to be 4602 nucleic acids in length (including a stop codon). This nucleotide sequence is shown in SEQ ID N°23. This published mlsn1 gene encodes a linear polypeptide of 1533 amino acids. The predicted amino acid sequence of this Mlsn1 is shown in SEQ ID N°1 and figure 1. The present invention stems from the identification that expression of a Mlsn1 gene product is essential for a functional capacitative calcium channel in immune- cells such as monocytes, macrophages, T-cells, B-cells and mast-cells, specifically in antigen presenting cells, more specifically in macrophages. Considering the implication of calcium influx in regulating various cell activities, this invention provides additional means in designing immuno-modulatory compounds as well as modulators compounds and in reporting immune activation and inflammation. More particularly, this invention now provides a novel target for screening immuno-modulatory compounds and deciphering immune activation gene pathways.

**[0031]** Therefore, a first object of the invention is the use of any purified or isolated nucleic acid encoding a human Mlsn1 or a sequence complementary thereto, wherein, the encoding nucleic acid encodes a polypeptide having the following characteristics;

(1) the N-terminal region is encoded by a consecutive sequence of at least 75 nucleotides, preferably at least 300 nucleotides, more preferably at least 750 nucleotides, most preferably 1500 nucleotides, located 5' with respect to nucleotides encoding the transmembrane and C- terminal regions of the Mlsn1 protein. More particularly the sequence is selected within nucleotides 1 to 2268 of SEQ ID N°23,

(2) a transmembrane region encoding six membrane spanning domains, encoded by a consecutive sequence of at least 150 nucleotides, preferably at least 300 nucleotides, more preferably at least 750 nucleotides, located 3' with respect to the nucleotides encoding the N-terminal region and 5' with respect to nucleotides encoding the C-terminal region of Mlsn1. More particularly the sequence is selected within the nucleotides 2271 to 3045 of SEQ ID N°23, and

(3) a C-terminal region encoded by a consecutive sequence of at least 75 nucleotides, preferably at least 150 nucleotides, more preferably at least 600 nucleotides located 3' with respect to nucleotides encoding the N-terminal region and transmembrane region of Mlsn1. More particularly selected within nucleotides 3048 to 4599 of SEQ ID N°23, for the screening of Mlsn1 modulators, more specifically Icrac modulators. A preferred nucleic acids encoding Mlsn1 is that of SEQ ID N°23.

**[0032]** A further object of the invention consists of the use of any purified or isolated nucleic acid having at least 80%, preferably 90%, more preferably 95% and most preferably 98% nucleotide identity with the nucleotide sequence of SEQ ID N°23 or a sequence complementary thereto, for the screening of Mlsn1 modulators, more specifically Icrac modulators. Also encompassed by the present invention is the use of any nucleic acid that hybridizes, under stringent condition, with SEQ ID N°23, its complementary strand, or a portion thereof, and encodes a polypeptide with calcium channel activity.

**[0033]** A second object of the invention is the use of any purified or isolated peptide encoding a human Mlsn1 or fragment thereof, wherein, the peptide has the following characteristics;

(1) the N-terminal region comprises consecutive sequence of at least 25 amino acids, preferably at least 100 amino acids, more preferably at least 250 amino acids, most preferably 500 amino acids, located N-terminal with respect to the transmembrane and C-terminal regions of Mlsn1 protein. More particularly the sequence is selected within the amino acids 1 to 756 of SEQ ID N°1,

(2) a transmembrane region forming six membrane spanning domains, comprising a consecutive sequence of at least 50 amino acids, preferably at least 100 amino acids, more preferably at least 250 amino acids, C-terminal with respect to the N-terminal region, and N-terminal with respect to the C-terminal region of Mlsn1. More particularly the sequence is selected within amino acids 757 to 1015 of SEQ ID N°1, and

(3) a C-terminal region comprising a consecutive sequence of at least 25 amino acids, preferably at least 50 amino acids, more preferably at least 200 amino acids, located C-terminal with respect to the N-terminal and transmembrane regions of Mlsn1. More particularly the sequence is selected within amino acids 1016 to 1533 of SEQ ID N°1, for the screening of Icrac modulators, more specifically Mlsn1 modulators. Preferred amino acids encoding Mlsn1 is that of SEQ ID N°1.

**[0034]** A further object of the invention consists of the use of any purified or isolated peptide having at least 90%, preferably 95%, more preferably 98% and most preferably 99% amino acid identity with the sequence of SEQ ID N°1 or fragment thereof, for the screening of Mlsn1 modulators, more specifically Icrac modulators.

**[0035]** Another object of the invention is a purified or isolated nucleic acid sequence encoding a polypeptide, wherein said nucleic acid encodes a polypeptide having at least 90%, preferably 95%, more preferably 98% and most preferably 99% sequence identity with the polypeptide of SEQ ID N° 1.

**[0036]** Furthermore, the discovery that Icrac function is modulated by Mlsn1 provides a means to correlate the severity of an Icrac associated disorder with the levels of expression in immune cells.

**[0037]** A third object of the invention is the use of any purified or isolated nucleic acid encoding a human Mlsn1 or a sequence complementary thereto, wherein, the encoding nucleic acid encodes a polypeptide having the following characteristics;

(1) the N-terminal region is encoded by a consecutive sequence of at least 75 nucleotides, preferably at least 300 nucleotides, more preferably at least 750 nucleotides, most preferably 1500 nucleotides, located 5' with respect to nucleotides encoding the transmembrane and C- terminal regions of Mlsn1 protein. More particularly the sequence is selected within nucleotides 1 to 2268 of SEQ ID N°23,

(2) a transmembrane region encoding six membrane spanning domains, encoded by a consecutive sequence of at least 150 nucleotides, preferably at least 300 nucleotides, more preferably at least 750 nucleotides, located 3' with respect to the nucleotides encoding the N-terminal region and 5' with respect to nucleotides encoding the C-terminal region of Mlsn1. More particularly the sequence is selected within nucleotides 2271 to 3045 of SEQ ID N°23, and

(3) a C-terminal region encoded by a consecutive sequence of at least 75 nucleotides, preferably at least 150 nucleotides, more preferably at least 600 nucleotides, located 3' with respect to nucleotides encoding the N-terminal region and transmembrane region of Mlsn1. More particularly the sequence is selected within nucleotides 3048 to 4599 of SEQ ID N°23, for the diagnosis of disorders associated with aberrant Icrac function in immune cells. A preferred nucleic acid encoding Mlsn1 is that of SEQ ID N°23.

**[0038]** A further object of the invention consists of the use of any purified or isolated nucleic acid having at least 80%, preferably 90%, more preferably 95% and most preferably 98% nucleotide identity with the nucleotide sequence of SEQ ID N°23 or a sequence complementary thereto, for the diagnosis of disorders associated with aberrant Icrac function in immune cells.

**[0039]** A fourth object of the invention is the use of any purified or isolated peptide encoding a human Mlsn1 or fragment thereof, wherein, the peptide has the following characteristics;

(1) the N-terminal region comprises a consecutive sequence of at least 25 amino acids, preferably at least 100 amino acids, more preferably at least 250 amino acids, most preferably 500 amino acids, located N-terminal with respect to the transmembrane and C- terminal regions of Mlsn1 protein. More particularly the sequence is selected within amino acids 1 to 756 of SEQ ID N°1,

(2) a transmembrane region forming six membrane spanning domains, comprising a consecutive sequence of at least 50 amino acids, preferably at least 100 amino acids, more preferably at least 250 amino acids, located C-terminal with respect to the N-terminal region and N-terminal with respect to the C-terminal region of Mlsn1. More particularly the sequence is selected within nucleic acids 757 to 1015 of SEQ ID N°1, and

(3) a C-terminal region comprising a consecutive sequence of at least 25 amino acids, preferably at least 50 amino acids, more preferably at least 200 amino acids, located C-terminal with respect to the N-terminal and transmembrane regions of Mlsn1. More particularly the sequence is selected from amino acids 1016 to 1533 of SEQ ID N°1, for the diagnosis of disorders associated with aberrant Icrac function in immune cells. A preferred nucleic acids encoding Mlsn1 is that of SEQ ID N°1.

**[0040]** A further object of the invention consists of the use of any purified or isolated peptide having at least 90%, preferably 95%, more preferably 98% and most preferably 99% amino acid identity with the sequence of SEQ ID N°1 or fragment thereof, for the screening of Icrac modulators and the diagnosis of disorders associated with aberrant Icrac function in immune cells.

## STRUCTURE OF Mlsn1

**[0041]** The inventors have identified biological relevant components of the mlsn1 gene and expression products. The inventors performed gene alignment experiments with various subtypes of voltage-gated  $\text{Ca}^{2+}$  channels, using the Clustal method (MegAlign-DNAstar/Lasergene software package). In particular, the amino acid sequence of the following Trp proteins were compared, in a window of 41 residues between the predicted transmembrane segments 5 and 6 (the corresponding Genbank accession numbers are indicated): Trp, J04844; TRPL calmodulin-binding protein, M88185; TRPC7 transient receptor potential channel 7, NM 003307; Melastatin 1, AF071787 and the genome sequence of the nematode *Caenorhabditis elegans*, 277132. These experiments allowed the identification of a novel calcium pore sequence in the Mlsn1 gene. The inventors believe that the active form of Mlsn1 is a membrane spanning polypeptide. This polypeptide is predicted to possess 6 transmembrane domains (TRD 1-6), and cytoplasmic N- and C-terminal domains. The N-domain, C- domain and TMDs are all connected by 5 peptide loops, two intra-cellular loops and three extra-cellular loops. Extra-cellular loop 3 forming part of the Icrac calcium pore. The TMDs incorporate the following peptides: TRD1, amino acids 757-781 (as shown in figure 1 of the polypeptide sequence with annotations); TMD2, amino acids 790-810 (as shown in figure 1); TMD3, amino acids 828-849 (as shown in figure 1); TMD4, amino acids 859-878 (as shown in figure 1); TMD5 amino acids 892-912 (as shown in figure 1) and TMD6, amino acids 985-1015 (as shown in figure 1). The N-terminal region includes amino acids 1 to 756 as shown in SEQ ID N°2 and 3, and in figure 1. The C-terminal region includes amino acids 1016-1533 as shown in SEQ ID N°4 and 5, and in figure 1. Intra-cellular loop 1 is encoded by amino acids 811-827 as shown in SEQ ID N°6 and 7, and in figure 1 and intra-cellular loop 2 is encoded by amino acids 879-891 as shown in SEQ ID N°8 and 9 and figure 1. Extra-cellular loop 1 is encoded by amino acids 782-789 as shown in SEQ ID N°10 and 11, and in figure 1. Extra-cellular loop 2 is encoded by amino acids 850-858 as shown in SEQ ID N°12 and 13 and in figure 1. Extra-cellular loop 3 is encoded by amino acids 913-984 as shown in SEQ ID N°14 to 17 and figure 1. The inventors believe that extra-cellular loop 3 plays a role in forming the calcium channel pore. Furthermore, the inventors believe that the 23 amino acid (amino acids 922-944) peptide sequence of SEQ ID N°16 which is predicted to be part of the calcium pore is essential for the function of Icrac.

**[0042]** The inventors believe that intra-cellular and extra-cellular exposed surfaces of Mlsn1 as well as the calcium pore provide vital target molecules for the development of therapeutic compounds to modulate Icrac. These surfaces include amino acids encoding the N- and C-terminal regions, as well as amino acids which encode loops connecting the transmembrane domains. Crucially, the ascribing of Icrac function to Mlsn1 has enabled the inventors to identify a calcium pore within the transmembrane region, and an external peptide loop. This loop is thought to directly modulate calcium flux in Icrac. Both the calcium pore and associated loop hold special significance in the development of therapeutics for the treatment of aberrant Icrac function. Furthermore, peptides of Mlsn1 allow the development of diagnostic tools to assay for Icrac function and levels in immune cells.

**[0043]** Likewise the nucleic acids which encode the said peptides offer opportunities to identify modulators of Icrac, and the development of diagnostic tools.

## Peptides of Mlsn1

**[0044]** The inventors believe that the N-terminal cytoplasmic region of Mlsn1 could play a role in the translocation of the Mlsn1 polypeptide from the cytoplasm to the cell membrane.

**[0045]** Therefore, an object of the invention is a purified or isolated human Mlsn1 N-terminal region, wherein the N-terminal region comprises a consecutive sequence of at least 25 amino acids, preferably at least 100 amino acids, more preferably at least 250 amino acids and most preferably at least 500 amino acids, located N-terminal of the first transmembrane region of the Mlsn1 protein. Preferred is a consecutive sequence taken from the amino acid sequences of SEQ ID N°2 or 3. More preferred are the sequences of SEQ ID N°2 or 3.

**[0046]** A further object of the invention resides in a purified or isolated human Mlsn1 N-terminal region, wherein the N-terminal region has at least 90%, preferably 95%, more preferably 98% and most preferably 99% amino acid identity with any of the peptides of amino acid sequences SEQ ID N°2 or 3, or fragments thereof.

**[0047]** The inventors also believe that the C-terminal region of Mlsn1 plays a role in the modulation of activity of Mlsn1. Therefore, a second object of the invention consists of a purified or isolated human Mlsn1 C-terminal region, wherein the C-terminal region comprises a consecutive sequence of at least 25 amino acids, preferably at least 50 amino acids, more preferably at least 100 amino acids and most preferably at least 200 amino acids, located C-terminal of the sixth

transmembrane region of the Mlns1 protein. Preferred is a consecutive sequence taken from the amino acid sequence of SEQ ID N°4 or 5. More preferred is the sequences of SEQ ID N°4 or 5.

**[0048]** A further object of the invention resides in a purified or isolated human Mlns1 C-terminal region, wherein the C-terminal region has at least 90%, preferably 95%, more preferably 98% and most preferably 99% amino acid identity with any of the peptides of amino acid sequences SEQ ID N°4 or 5, or fragments thereof.

**[0049]** The inventors believe that peptide encoding intra-cellular loop-1 of Mlns1 play an important role in the regulation of Mlns1 and its capacity to transduce intra-cellular signals. Therefore, a third object of the invention is a purified or isolated human Mlns1 intra-cellular loop-1 region, wherein the intra-cellular loop-1 region comprises a consecutive sequence of at least 7 amino acids, preferably at least 10 amino acids, more preferably at least 15 amino acids, located C-terminal of the second transmembrane region and N-terminal to the third transmembrane region of the Mlns1 protein. Preferred is a consecutive sequence taken from the amino acid sequence of SEQ ID N°6 or 7. More preferred are the sequences of SEQ ID N°6 or 7.

**[0050]** A further object of the invention resides in a purified or isolated human Mlns1 intra-cellular loop-1 region, wherein the intra-cellular loop-1 region has at least 90%, preferably 95%, more preferably 98% and most preferably 99% amino acid identity with any of the peptides of amino acid sequences SEQ ID N°6 or 7, or fragments thereof.

**[0051]** The inventors believe that peptide encoding intra-cellular loop-2 of Mlns1 also play an important role in the regulation of Mlns1 and its capacity to transduce intra-cellular signals. Therefore, a fourth object of the invention is a purified or isolated human Mlns1 intra-cellular loop-2 region, wherein the intra-cellular loop-2 region comprises a consecutive sequence of at least 7 amino acids, preferably at least 10 amino acids, located C-terminal of the fourth transmembrane region and N-terminal of the fifth transmembrane region of the Mlns1 protein. Preferred is a consecutive sequence taken from the amino acid sequences of SEQ ID N°8 or 9. More preferred is the sequences of SEQ ID N°8 or 9.

**[0052]** A further object of the invention is a purified or isolated human Mlns1 intra-cellular loop-2 region, wherein the intra-cellular loop-2 region has at least 90%, preferably 95%, more preferably 98% and most preferably 99% amino acid identity with any of the peptides of amino acid sequences SEQ ID N°8 or 9, or fragments thereof.

**[0053]** The inventors believe that the peptides encoding extra-cellular loop 1 of Mlns1 may play a crucial role in the modulation of calcium through the Icrac channel. Therefore a fifth object of the invention is purified or isolated human Mlns1 extra-cellular loop-1 region, wherein the extra-cellular loop-1 region comprises a consecutive sequence of at least 5 amino acid located C-terminal of the first transmembrane region and N-terminal of the second transmembrane region of the Mlns1 protein. Preferred is a consecutive sequence taken from the amino acid sequences of SEQ ID N°10 or 11. More preferred is the sequence of SEQ ID N°10 or 11.

**[0054]** A further object of the invention is purified or isolated human Mlns1 extra-cellular loop-1 region, wherein the extra-cellular loop-1 region has at least 90%, preferably 95%, more preferably 98% and most preferably 99% amino acid identity with any of the peptides of amino acid sequences SEQ ID N°10 or 11, or fragments thereof.

**[0055]** The inventors believe that the peptides encoding extra-cellular loop 2 region of Mlns1 may also play a crucial role in the modulation of calcium through the Icrac channel. Therefore, a sixth object of the invention is purified or isolated human Mlns1 extra-cellular loop-2 region, wherein the extra-cellular loop-2 region comprises a consecutive sequence of at least 5 amino acids, located C-terminal of the third transmembrane region and N-terminal of the fourth transmembrane region of the Mlns1 protein. Preferred is a consecutive sequence taken from the amino acid sequence of SEQ ID N°12 or 13. More preferred is the sequence of SEQ ID N°12 or 13

**[0056]** A further object is purified or isolated human Mlns1 extra-cellular loop-2 region, wherein the extra-cellular loop-2 region has at least 90%, preferably 95%, more preferably 98% and most preferably 99% amino acid identity with any of the peptides of amino acid sequences SEQ ID N°12 or 13, or fragments thereof.

**[0057]** The inventors believe that the peptides encoding extra-cellular loop-3 region, has a function in the modulation of calcium flux in Icrac. Therefore, a seventh object of the invention is a purified or isolated human Mlns1 extra-cellular loop-3 region, wherein the extra-cellular loop-3 region comprises a consecutive sequence of at least 7 amino acids, preferably at least 10 amino acids, more preferably at least 20 amino acids, most preferably at least 50 amino acids, located C-terminal of the fifth transmembrane region and N-terminal of the sixth transmembrane region of the Mlns1 protein. Preferred is a consecutive sequence taken from the amino acid sequence of SEQ ID N°14, 15, 16 or 17. More preferred are the sequences of SEQ ID N°14, 15, 16 or 17.

**[0058]** A further object of the invention is purified or isolated human Mlns1 extra-cellular loop-3 region, wherein the extra-cellular loop-3 region has at least 90%, preferably 95%, more preferably 98% and most preferably 99% amino acid identity with any of the peptides of amino acid sequences SEQ ID N°14, 15, 16 or 17, or fragments thereof.

**[0059]** The inventors believe that peptides which contain both intra- and extra- cellular loop regions and transmembrane regions but lacking associated N-and C-terminal regions could play a role in the understanding of Icrac and the development of modulators of Icrac and diagnostic tools. Therefore, an eighth object of the invention is purified or isolated human Mlns1 peptide containing both intra- and extra-cellular loop regions, wherein said intra-and extra-cellular loop region comprises a consecutive sequence of at least 50 amino acids, preferably at least 100 amino acids

and most preferably at least 250 amino acids, located C-terminal to the N-terminal region and N-terminal to the C-terminal region of the Mln1 protein. Preferred is a consecutive sequence taken from the amino acid sequence of SEQ ID N°18 or 19. More preferred is the sequence of SEQ ID N°18 or 19.

**[0060]** A further object of the invention is purified or isolated human Mln1 peptide containing both intra-and extra-cellular loop regions, wherein the intra- and extra-cellular loop region has at least 90%, preferably 95%, more preferably 98% and most preferably 99% amino acid identity with any of the peptides of amino acid sequences SEQ ID N°18 or 19, or fragments thereof.

**[0061]** Amino acid identity may be determined using various methods known in the art. In particular, amino acid identity (or sequence homology) may be assessed by commercially available computer programs, such as CLUSTAL or BLAST (NCBI). Various search or alignment parameters may also be used. In a preferred embodiment, % amino acid (or nucleic acid) identity is determined using the CLUSTAL-W program (Compugen).

#### Nucleic acids encoding peptides of Mln1

**[0062]** This invention also relates generally to the use of nucleic acid sequences encoding said Mln1 peptides for the development of screening and diagnostic technologies.

**[0063]** An object of the invention is a purified or isolated nucleic acid sequence encoding the human Mln1 N-terminal region, wherein the N-terminal region is encoded by a consecutive sequence of at least 75 nucleotides, preferably at least 300 nucleotides, more preferably at least 750 nucleotides and most preferably at least 1500 nucleotides, located 5' with respect to nucleotides encoding the first transmembrane region of the Mln1 protein. Preferred is a consecutive sequence taken from the nucleic acid sequences of SEQ ID N°24 or 25. More preferred is the sequence of SEQ ID N°24 or 25.

**[0064]** A further object of the invention is a purified or isolated nucleic acid sequence encoding the human Mln1 N-terminal region, wherein the nucleic acid has at least 80%, preferably 90%, more preferably 95% and most preferably 98% nucleic acid identity with any of the nucleotide sequences or SEQ ID N°24 or 25, or a complementary sequences thereto.

**[0065]** Another object of the invention is a purified or isolated nucleic acid sequence encoding a polypeptide, wherein said nucleic acid encodes a polypeptide having at least 90%, preferably, 95, more preferably 98% and most preferably 99% with any of the polypeptides of SEQ ID N°2 or 3.

**[0066]** A further object of the invention is a purified or isolated nucleic acid sequence encoding the human Mln1 C-terminal region, wherein the C-terminal region is encoded by a consecutive sequence of at least 75 nucleotides, preferably at least 150 nucleotides, more preferably at least 300 nucleotides and most preferably at least 600 nucleotides, located 5' with respect to nucleotides encoding the sixth transmembrane region of the Mln1 protein. Preferred is a consecutive sequences taken from the nucleic acid sequence of SEQ ID N°26 or 27. More preferred is the sequence of SEQ ID N°26 or 27.

**[0067]** A further object of the invention is a purified or isolated nucleic acid sequence encoding the human Mln1 C-terminal region, wherein the nucleic acid has at least 80%, preferably 90%, more preferably 95% and most preferably 98% nucleic acid identity with any of the nucleotide sequences or SEQ ID N°26 or 27, or a complementary sequences thereto.

**[0068]** A further object of the invention is a purified or isolated nucleic acid sequence encoding a polypeptide, wherein said nucleic acid encodes a polypeptide having at least 90%, preferably, 95%, more preferably 98% and most preferably 99% with any of the polypeptides of SEQ ID N°4 or 5.

**[0069]** A further object of the invention resides is a purified or isolated nucleic acid sequence encoding the human Mln1 intra-cellular loop-1 region, wherein the intra-cellular loop-1 region is encoded by a consecutive sequence of at least 21 nucleotides, preferably at least 30 nucleotides, more preferably at least 45 nucleotides, located 3' with respect to nucleotides encoding the second transmembrane region and 5' with respect to nucleotides encoding the third transmembrane region of the Mln1 protein. Preferred is a consecutive sequence taken from the nucleic acid sequence of SEQ ID N°28 or 29. More preferred is the sequence of SEQ ID N°28 or 29.

**[0070]** A further object of the invention is a purified or isolated nucleic acid sequence encoding the human Mln1 loop-1 region, wherein the nucleic acid has at least 80%, preferably 90%, more preferably 95% and most preferably 98% nucleic acid identity with any of the nucleotide sequences or SEQ ID N°28 or 29, or a complementary sequences thereto.

**[0071]** Another object of the invention is a purified or isolated nucleic acid sequence encoding a polypeptide, wherein said nucleic acid encodes a polypeptide having at least 90%, preferably, 95%, more preferably 98% and most preferably 99% with any of the polypeptides of SEQ ID N°6 or 7.

**[0072]** A further object of the invention is a purified or isolated nucleic acid sequence encoding the human Mln1 intra-cellular loop-2 region, wherein the intra-cellular loops-2 region is encoded by a consecutive sequence of at least 21 nucleotides, preferably at least 30 nucleotides, located 3' with respect to nucleotides encoding the fourth transmem-

brane region and 5' with respect to nucleotides encoding the fifth transmembrane region of the Mln1 protein. Preferred is a consecutive sequence taken from the nucleic acid sequence of SEQ ID N°30 or 31. More preferred is the sequence of SEQ ID N°30 or 31.

**[0073]** A further object of the invention is a purified or isolated nucleic acid sequence encoding the human Mln1 intra-cellular loop-2 region, wherein the nucleic acid has at least 80%, preferably 90%, more preferably 95% and most preferably 98% nucleic acid identity with any of the nucleotide sequences or SEQ ID N°30 or 31, or a complementary sequences thereto.

**[0074]** Another object of the invention is a purified or isolated nucleic acid sequence encoding a polypeptide, wherein said nucleic acid encodes a polypeptide having at least 90%, preferably, 95%, more preferably 98% and most preferably 99% with any of the polypeptides of SEQ ID N°8 or 9.

**[0075]** A further object of the invention is a purified or isolated nucleic acid sequence encoding the human Mln1 extra-cellular loop-1 region, wherein the extra-cellular loops-1 region is encoded by a consecutive sequence of at least 15 nucleotides, located 3' with respect to nucleotides encoding the first transmembrane region and 5' with respect to nucleotides encoding the second transmembrane region of the Mln1 protein. Preferred is a consecutive sequence taken from the nucleic acid sequence of SEQ ID N°32 or 33. More preferred is the sequence of SEQ ID N°32 or 33.

**[0076]** A further object of the invention is purified or isolated nucleic acid sequence encoding the human Mln1 extra-cellular loop-1 region, wherein the nucleic acid has at least 80%, preferably 90%, more preferably 95% and most preferably 98% nucleic acid identity with any of the nucleotide sequences or SEQ ID N°32 or 33, or a complementary sequences thereto.

**[0077]** Another object of the invention is a purified or isolated nucleic acid sequence encoding a polypeptide, wherein said nucleic acid encodes a polypeptide having at least 90%, preferably, 95%, more preferably 98% and most preferably 99% with any of the polypeptides of SEQ ID N°10 or 11.

**[0078]** A further object of the invention is purified or isolated nucleic acid sequence encoding the human Mln1 extra-cellular loop-2 region, wherein the extra-cellular loops-2 region is encoded by a consecutive sequence of at least 15 nucleotides, located 3' with respect to nucleotides encoding the third transmembrane region and 5' with respect to nucleotides encoding the fourth transmembrane region of the Mln1 protein. Preferred is a consecutive sequence taken from the nucleic acid sequence of SEQ ID N°34 or 35. More preferred is the sequence of SEQ ID N°34 or 35.

**[0079]** A further object of the invention is purified or isolated nucleic acid sequence encoding the human Mln1 extra-cellular loop-2 region, wherein the nucleic acid has at least 80%, preferably 90%, more preferably 95% and most preferably 98% nucleic acid identity with any of the nucleotide sequences or SEQ ID N°34 or 35, or a complementary sequences thereto.

**[0080]** Another object of the invention is a purified or isolated nucleic acid sequence encoding a polypeptide, wherein said nucleic acid encodes a polypeptide having at least 90%, preferably, 95%, more preferably 98% and most preferably 99% with any of the polypeptides of SEQ ID N°12 or 13.

**[0081]** A further object of the invention is purified or isolated nucleic acid sequence encoding the human Mln1 extra-cellular loop-3 region, wherein the extra-cellular loops-3 region is encoded by a consecutive sequence of at least 21 nucleotides, preferably at least 30 nucleotides, more preferably at least 60 nucleotides and most preferably at least 150 nucleotides 3' with respect to nucleotides encoding the fifth transmembrane region and 5' with respect to nucleotides encoding the sixth transmembrane region of the Mln1 protein. Preferred is a consecutive sequence taken from the nucleic acid sequence of SEQ ID N°36, 37 or 38. More preferred is the sequence of SEQ ID N°36, 37 or 38.

**[0082]** A further object is purified or isolated nucleic acid sequence encoding the human Mln1 extra-cellular loop-3 region, wherein the nucleic acid has at least 80%, preferably 90%, more preferably 95% and most preferably 98% nucleic acid identity with any of the nucleotide sequences or SEQ ID N°36, 37 or 38, or a complementary sequences thereto.

**[0083]** Another object of the invention is a purified or isolated nucleic acid sequence encoding a polypeptide, wherein said nucleic acid encodes a polypeptide having at least 90%, preferably, 95%, more preferably 98% and most preferably 99% with any of the polypeptides of SEQ ID N°14, 15, 16 or 17.

**[0084]** An other object of the invention is purified or isolated nucleic acid sequence encoding the human Mln1 peptide containing both intra- and extra-cellular loop regions, wherein said intra- and extra-cellular loop region is encoded by a consecutive sequence of at least 150 nucleotides, preferably at least 300 nucleotides and most preferably at least 750 nucleotides 3' with respect to nucleotides encoding the N-terminal region and 5' with respect to nucleotides encoding the C-terminal regions of the Mln1 protein. Preferred is a consecutive sequence taken from the nucleic acid sequence of SEQ ID N°39 or 40. More preferred is the sequence of SEQ ID N°39 or 40.

**[0085]** Another object of the invention is purified or isolated nucleic acid sequence encoding the human Mln1 peptide containing both intra- and extra-cellular loop regions, wherein the nucleic acid has at least 80%, preferably 90%, more preferably 95% and most preferably 98% nucleic acid identity with any of the nucleotide sequences or SEQ ID N°39 or 40, or a complementary sequences thereto.

**[0086]** Another object of the invention is a purified or isolated nucleic acid sequence encoding a polypeptide, wherein

said nucleic acid encodes a polypeptide having at least 90%, preferably, 95%, more preferably 98% and most preferably 99% with any of the polypeptides of SEQ ID N°18 or 19.

**[0087]** Nucleic acid sequence identity may be determined using various methods known in the art. In particular, nucleic acid sequence identity (or sequence homology) may be assessed by commercially available computer programs, such as CLUSTAL or BLAST (NCBI). Various search or alignment parameters may also be used. In a preferred embodiment, % amino acid (or nucleic acid) identity is determined using the CLUSTAL-W program (Compugen).

#### TRANSCRIPTION PROMOTERS OF Mlsn1

**[0088]** The inventors have identified three genomic nucleic acid sequences 5' to the published codon encoding Methionine 1 of the Mlsn1 gene that encode transcription promoter sequences. The inventors believe that nucleic acids encoding transcription promoter(s) elements of Mlsn1 are targets for the development of therapeutics which modulate Icrac activity. A first preferred nucleic acid sequence is a nucleic acid sequence which encodes an Mlsn1 transcription promoter of SEQ ID N°41 or a complementary sequence thereto. A further object of the invention consists of nucleic acid sequences having at least 80%, preferably 90%, more preferably 95% and most preferably 98% nucleotide identity with the nucleotide sequence of SEQ ID N°41 or a complementary sequence thereto.

**[0089]** A second preferred nucleic acid sequence is a nucleic acid sequence which encodes an Mlsn1 transcription promoter of SEQ ID N°42 or a complementary sequence thereto. A further object of the invention consists of nucleic acid sequences having at least 80%, preferably 90%, more preferably 95% and most preferably 98% nucleotide identity with the nucleotide sequence of SEQ ID N°42 or a complementary sequence thereto.

**[0090]** A third preferred nucleic acid sequence is a nucleic acid sequence which encodes an Mlsn1 transcription promoter of SEQ ID N°43 or a complementary sequence thereto. A further object of the invention consists of nucleic acid sequence having at least 80%, preferably 90%, more preferably 95% and most preferably 98% nucleotide identity with the nucleotide sequence of SEQ ID N°43 or a complementary sequence thereto.

#### EXPRESSION OF Mlsn1

**[0091]** The inventors discovered that the Mlsn1 gene was expressed in U937 cells differentiated into a macrophage phenotype, and that the product of this gene functions as a calcium channel, more specifically as an Icrac channel. Mlsn1-specific primers were designed and used to screen for Mlsn1 gene expression in various cells. The composition of these primers are indicated in SEQ ID N°44 to 47. With said primer pairs, Mlsn1-specific PCR products were obtained from U937 cells. Furthermore, cDNA subtraction experiments were performed using DNA preparations from differentiated and undifferentiated U937 cells. The results obtained demonstrate that the expression of the Mlsn1 gene product in U937 cells is differentially increased in dbcAMP-differentiated U937 cells (macrophage-like) as compared to undifferentiated U937-cells (monocyte like). The upregulation of the mlsn1 gene correlates with the expression of the Icrac phenotype in U937 cells (18) and thus establishes that expression upregulation of Mlsn1 in these cells coincides with the increased Icrac phenotype. Sequence specific evidence for Mlsn1 involvement in the Icrac phenotype was obtained by the demonstration that Mlsn1-specific anti-sense treatment of U937 cells during dbcAMP induced differentiation could significantly decrease the calcium signal in these cells.

**[0092]** Accordingly, this invention unexpectedly demonstrates that Mlsn1 is expressed in immune cells such as macrophages and that the down-regulation of the Mlsn1 gene expression during differentiation of U937 cells to macrophage like cells decreases calcium influx in these cells. This invention also demonstrates that Mlsn1 gene product expression is up-regulated during U937 cell differentiation to macrophage like cells, which correlates with the maturation stage of these cells and their ability to exhibit the Icrac phenotype

#### PRODUCTION OF Mlsn1 POLYPEPTIDES OR NUCLEIC ACIDS

##### Recombinant expression vecotors

**[0093]** The present invention also encompasses a family of recombinant vectors comprising any one of the nucleic acids described herein. Thus, the invention further deals with a recombinant vector comprising a nucleic acid selected from the group consisting of:

- (a) a purified or isolated nucleic acid encoding Mlsn1, preferably human Mlsn1 or Mlsn1 transcription promoter element(s), and more preferably a polypeptide having at least 90% amino acid identity with a polypeptide selected from the group consisting of; nucleic acids encoding Mlsn1, such as SEQ ID N°1, nucleic acids encoding the N-terminal region of Mlsn1, such as SEQ ID N°2 and 3, nucleic acids encoding C-terminal region of Mlsn1, such as SEQ ID N°4 and 5, nucleic acids encoding Intra-cellular loops of Mlsn1, such as SEQ ID N°6 to 9, nucleic acids

encoding extra-cellular loops 1 and 2 of Mlsn1, such as SEQ ID N°10 to 13, nucleic acids encoding extra-cellular loop 3 of Mlsn1, such as SEQ ID N°14 to 17, nucleic acids encoding both intra- and extra cellular loops of Mlsn1, such as SEQ ID N°18 and 19 and nucleic acids encoding transcription promoter elements of SEQ ID N°41 to 43, or a sequence complementary thereto;

(b) a purified or isolated nucleic acid having at least 80% nucleotide identity with a polynucleotide selected from the group consisting of the nucleotide sequences of SEQ ID N°23 to 43, or a sequence complementary thereto;

(c) a purified or isolated polynucleotide comprising at least 10 consecutive nucleotides of a nucleic acid described in (a) or (b), or a sequence complementary thereto.

**[0094]** In a first preferred embodiment a recombinant vector of the invention is used to amplify the inserted polynucleotide derived from the nucleic acid encoding a Mlsn1 of the invention or Mlsn1 transcription promoter in a suitable host cell, this polynucleotide being amplified every time the recombinant vector replicates.

**[0095]** A second preferred embodiment of the recombinant vectors according to the invention consists of expression vectors comprising a nucleic acid encoding an Mlsn1 of the invention, preferably a nucleic acid encoding a human Mlsn1, and more preferably a nucleic acid encoding a polypeptide selected from the group consisting of the amino acid sequences of SEQ ID N° 1 to 22.

**[0096]** Recombinant expression vectors comprising a nucleic acid encoding the peptide fragments of an Mlsn1 that are specified in the present specification are also part of the invention.

**[0097]** Within certain embodiments, expression vectors can be employed to express Mlsn1 of the invention or a peptide fragment thereof which can then be purified and for example, be used as an immunogen in order to raise specific antibodies directed against said Mlsn1 protein or a peptide fragment thereof.

**[0098]** In another embodiment, the expression vectors are used for constructing transgenic animals and also for gene therapy, notably for anti-sense therapy.

**[0099]** Expression requires that appropriate signals are provided in the vectors, said signals including various regulatory elements such as enhancers/promoters from both viral and mammalian sources that drive expression of the genes of interest in host cells. The regulatory sequences of the expression vectors of the invention are operably linked to the nucleic acid encoding the Mlsn1 protein of interest or a peptide fragment thereof.

**[0100]** As used herein, the term "operably linked" refers to a linkage of polynucleotide elements in a functional relationship. For instance, a promoter or an enhancer is operably linked to a coding sequence if it affects the transcription of the coding sequence.

**[0101]** More precisely, two DNA molecules (such as a polynucleotide containing a promoter region and a polynucleotide encoding a desired polypeptide or polynucleotide) are said to be "operably linked" if the nature of the linkage between the two polynucleotides does not : (1) result in the introduction of a frame-shift mutation or (2) interfere with the ability of the polynucleotide containing the promoter to direct the transcription of the coding polynucleotide.

**[0102]** Generally, recombinant expression vectors will include origins of replication, selectable markers, permitting transformation of the host cell, and a promoter derived from a highly expressed gene to direct transcription of a downstream structural sequence. The heterologous structural sequence is assembled in an appropriate frame with the translation, initiation and termination sequences, and preferably a leader sequence capable of directing sequences of the translated protein into the periplasmic space or the extra-cellular medium.

**[0103]** In a specific embodiment wherein the vector is adapted for transfecting and expressing desired sequences in mammalian host cells, preferred vectors will comprise an origin of replication from the desired host, a suitable promoter and an enhancer, and also any necessary ribosome binding sites, polyadenylation site, transcriptional termination sequences, and optionally 5'-flanking non-transcribed sequences.

**[0104]** DNA sequences derived from the SV 40 viral genome, for example SV 40 origin, early promoter, enhancer, and polyadenylation sites may be used to provide the required non-transcribed genetic elements.

**[0105]** Additionally, a recombinant expression vector of the invention advantageously also comprises an untranscribed polynucleotide located at the 3' end of the coding sequence (ORF), this 3'-UTR polynucleotide being useful for stabilizing the corresponding mRNA or for increasing the expression rate of the vector insert if this 3'-UTR harbors regulation signal elements such as enhancer sequences.

**[0106]** Suitable promoter regions used in the expression vectors according to the invention are chosen taking into account the host cell in which the heterologous nucleic acids have to be expressed.

**[0107]** A suitable promoter may be heterologous with respect to the nucleic acid for which it controls the expression, or alternatively can be endogenous to the native polynucleotide containing the coding sequence to be expressed.

**[0108]** Additionally, the promoter is generally heterologous with respect to the recombinant vector sequences within which the construct promoter/coding sequence has been inserted.

**[0109]** Preferred bacterial promoters are the LacI, LacZ, T3 or T7 bacteriophage RNA polymerase promoters, the lambda PR, PL and trp promoters (a EP-0 036 776), the polyhedrin promoter, or the p10 protein promoter from *baculovirus* (kit Novagen; Smith et al., (1983); O'Reilly et al. (1992).

**[0110]** Preferred selectable marker genes contained in the expression recombinant vectors of the invention for selection of transformed host cells are preferably dehydrofolate reductase or neomycin resistance for eukaryotic cell culture, TRP1 for *S. cerevisiae* or tetracycline, rifampicin or ampicillin resistance in *E. coli*, or Levamsaccharase for *Mycobacteria*, this latter marker being a negative selection marker.

**[0111]** Preferred bacterial vectors of the invention are listed hereafter as illustrative but not limitative examples:

pQE70, pQE60, pQE-9 (Quiagen), pD10, phagescript, psiX174, p.Bluescript SK, pNH8A, pNH16A, pNH18A, pNH46A (Stratagene); pKK223-3, pKK233-3, pDR540, pRIT5 (Pharmacia); pWLNEO, pSV2CAT, pOG44, pXT1, pSG (Stratagene); pSVK3, pBPV, pMSG, pSVL (Pharmacia); pQE-30 (QIA express).

**[0112]** Preferred bacteriophage recombinant vectors of the invention are P1 bacteriophage vectors such as described by Sternberg N.L. (1992;1994).

**[0113]** A suitable vector for the expression of Mlsn1 polypeptide of the invention or a fragment thereof, is a baculovirus vector that can be propagated in insect cells and in insect cell-lines. A specific suitable host vector system is the pVL 1392/1393 *baculovirus* transfer vector (Pharmingen) that is used to transfect the SF9 cell line (ATCC N°CRL 1711) which is derived from *spodoptera frugiperda*.

**[0114]** The recombinant expression vectors from the invention may also be derived from an adenovirus such as those described by Feldman and Steig. (1996) or Ohno et al. (1994).

**[0115]** Another preferred recombinant adenovirus according to this specific embodiment of the present invention is the human adenovirus type two or five (Ad 2 or Ad 5) or an adenovirus of animal origin (French Patent Application n°FR 93 05 954).

**[0116]** Particularly preferred retrovirus as for the preparation or construction of retroviral *in vitro* or *in vivo* gene delivery vehicles of the present invention include retroviruses selected from the group consisting of Mink-Cell Focus Inducing Virus, murine sarcoma virus, and Ross Sarcoma Virus. Other preferred retroviral vectors are those described in Roth et al. (1996), in PCT Application WO 93/25 234, in PCT Application WO 94/06920, and also in Roux et al. (1989), Julan et al.(1992) and Nada et al. (1991).

**[0117]** Yet, another viral vector system that is contemplated by the invention consist in the adeno associated viruses (AAV) such as those described by Flotte et al. (1992), Samulski et al. (1989) and McLaughlin et al. (1996).

**[0118]** Thus, a further object of the invention consists of a recombinant expression vector comprising a nucleic acid encoding an Mlsn1 or a peptide fragment thereof or a variant thereof, wherein said nucleic acid is operably linked to a promoter sequence.

**[0119]** In a preferred embodiment, this nucleic acid encodes a human Mlsn1, a fragment of human Mlsn1, or Mlsn1 transcription promoter, and preferably that of SEQ ID N°23 and nucleic acid sequence encoding peptide sequence of SEQ ID N°1, or a variant or a peptide fragment thereof. Preferred fragments include those of SEQ ID N° 2-22 and 24 to 40. Preferred Mlsn1 transcription promoters include those of SEQ ID N°41 to 43.

**[0120]** In a specific embodiment of this invention, any of the identified Mlsn1 promoter elements of SEQ ID N°41 to 43 could be used to modulate the expression of a reporter molecule, as part of an expression construct. Furthermore, these Mlsn1 transcription promoter elements could be used to regulate Mlsn1 gene or gene fragment expression as part of a construct.

Host cells expressing Mlsn1

**[0121]** Host cells that endogenously express Mlsn1 or have been transformed or transfected with one of the nucleic acids described herein, or with one of the recombinant vector, particularly recombinant expression vector, described herein are also part of the present invention.

**[0122]** Also included are host cells that are transformed (prokaryotic cells) or are transfected (eukaryotic cells) with a recombinant vector such as one of those described above. Preferred host cells used as recipients for the expression vectors of the invention are the following:

(a) prokaryotic host cells: *Escherichia coli*, strains. (i.e. DH5- $\alpha$ , strain) *Bacillus subtilis*, *Salmonella typhimurium* and strains from species like *Pseudomonas*, *Streptomyces* and *Staphylococcus*,

(b) eukaryotic host cells: T-cell lines (ECACC U937, 85011440; ECACC J.CaMI.6, 96060401 ; ECACC Jurkat E6.1, 88042803 and ECACC J45.01, 93031145), HeLa cells (ATCC N°CCL2; N°CCL2.1; N°CCL2.2), Cv 1 cells (ATCC N°CCL70), COS cells (ATCC N°CRL 1650; N°CRL 1651), Sf-9 cells (ATCC N°CRL 1711), C127 cells (ATCC N°CRL-1804), 3T3 cells (ATCC N°CRL-6361), CHO cells (ATCC N°CCL-61), human kidney 293 cells (ATCC N° 45504; N°CRL-1573), BHK (ECACC N°84100 501; N°84111301), PC12 (ATCC N° CRL-1721), NT2, SHSY5Y (ATCC N° CRL-2266), NG108 (ECACC N°88112302) and F11, SK-N-SH (ATCC N° CRL-HTB-11), SK-N-BE(2) (ATCC N° CRL-2271), IMR-32 (ATCC N° CCL-127). A preferred system to which the gene of the invention can be expressed are cell lines such as T-cell lines, B-cell lines, mast cell lines, jurkat cell lines, U937 cell lines, KU-812 cell lines, COS cells, 3T3 cells, HeLa cells, 292 cells and CHO cells. A most preferred system for the efficient

expression of Mlsn1 involves the use of T-cell lines. The gene can be expressed through an endogenous promoter of native T-cells, or through an exogenous promoter. Suitable exogenous promoters include such as SV40 and CMV, or perhaps a eukaryotic promoter such as the tetracycline promoter. The preferred promoter when Mlsn1 is endogenously expressed is an endogenous promoter. A preferred promoter in a recombinant cell line would be CMV.

**[0123]** In a specific embodiment of the host cells described above, these host cells have also been transfected or transformed with a polynucleotide or a recombinant vector allowing the expression of a natural ligand of Mlsn1 or a modulator of Mlsn1.

**[0124]** The present invention also concerns a method for producing one of the Mlsn1 polypeptides or peptides described herein and especially a polypeptide selected from the group consisting the amino acid sequences of SEQ ID N°1 to 22, wherein said method comprises the steps of:

(a) inserting the nucleic acid encoding the desired Mlsn1 polypeptide or peptide fragment thereof in an appropriate vector;

(b) culturing, in an appropriate culture medium, a host cell previously transformed or transfected with the recombinant vector of step (a);

(c) harvesting the culture medium thus conditioned or lyse the host cell, for example by sonication or by an osmotic shock;

(d) separating or purifying, from said culture medium, or from the pellet of the resultant host cell lysate, the thus produced Mlsn1 polypeptide of interest.

**[0125]** In a first preferred embodiment of the above method, the nucleic acid to be inserted in the appropriate vector has previously undergone an amplification reaction, using a pair of primers.

**[0126]** Preferred primers used for such an amplification reaction are primer pairs which amplify nucleic acids encoding the Mlsn1 open reading frame and fragments thereof.

**[0127]** In a second preferred embodiment of the above method, the polypeptide thus produced is further characterized, for example by binding onto an immuno-affinity chromatography column on which polyclonal or monoclonal antibodies directed to Mlsn1 polypeptide or a peptide fragment thereof, have previously been immobilised.

**[0128]** Purification of the recombinant Mlsn1 proteins according to the present invention or a peptide fragment thereof may be carried out by passage onto a nickel or copper affinity chromatography column.

**[0129]** In another embodiment, the Mlsn1 polypeptides or peptide fragments thus obtained may be purified, for example, by high performance liquid chromatography, such as reverse phase and/or cationic exchange HPLC, as described by Rougeot et al. (1994).

**[0130]** The reason to prefer this kind of peptide or protein purification is the lack of byproducts formed in the elution samples which renders the resultant purified protein or peptide more suitable for therapeutic use.

#### Production of Mlsn1 or a fragment thereof

**[0131]** The Mlsn1 protein or fragments thereof can be prepared using recombinant technology, cell lines or chemical synthesis. Recombinant technology and chemical synthesis of the Mlsn1 or fragments thereof can allow the modification of the gene encoding the Mlsn1 to include such features as recognition tags, cleavage sites and modifications of the Mlsn1 polypeptide or fragments thereof. For efficient polypeptide production, the endogenous expression system or recombinant expression system should allow the Mlsn1 polypeptide to be expressed at and transported to the cell surface in a functional form or allow production of Mlsn1 fragments which can be purified. Preferred cell lines are those which allow high levels of expression of Mlsn1 or fragments thereof. Such cell lines include cell lines which naturally express Mlsn1 or common mammalian cell lines expressing Mlsn1 such as CHO cells and COS cells, etc., or more specific immune cell lines such as T-cell lines. However, other cell types which are commonly used for recombinant protein production such as insect cells, amphibian cells such as oocytes, yeast and procaryotic cell lines such as *E. coli* can also be considered.

**[0132]** The Mlsn1 or fragments thereof can be utilised in a ligand screen either as a purified protein, as a protein chimera such as those described in phage display, as a cell membrane (lipid or detergent) preparation, or in intact cells.

**[0133]** The Mlsn1 polypeptide or fragment thereof can be utilised in a functional screen format or ligand binding screen format. Examples of both screening formats are provided below.

#### SCREENING FOR Mlsn1 LIGANDS

**[0134]** The present invention also concerns methods for screening ligand substances or molecules that are able to modulate the biological activity of the Mlsn1 gene or gene product of the invention. For example diseases where there

is a down-regulation of the immune system and conditions associated with virus and micro-organism infection, enhancement of the activity of Mlsn1 would be beneficial. In conditions such as asthma, inflammation and auto-immune disease it would be beneficial to identify ligands which can down-regulate the activity of the Mlsn1 gene.

**[0135]** In this regard, this invention now discloses the use of a Mlsn1 gene product for the screening of compound (s) that modulate the activity of immune cells, in particular antigen presenting cells, more specifically for the screening of compound(s) that modulate the activity of macrophages, *in vitro*, *ex vivo* or *in vivo*.

**[0136]** This invention also discloses the use of Mlsn1 promoter elements for the screening of compounds that modulate the activity of immune cells, in particular antigen presenting cells, more specifically for the screening of compounds that modulate the activity of macrophages, *in vitro*, *ex vivo* or *in vivo*.

**[0137]** Several screening assays can be used within the instant invention, such as binding assays or functional assays. In a particular embodiment, this invention more specifically relates to methods of screening for immuno-modulatory compounds, comprising;

a) contacting a test compound with Mlsn1 gene, an Mlsn1 gene product or Mlsn1 promoter element and,

b) measuring the ability of said test compound to interact with the Mlsn1 gene, Mlsn1 gene product or transcription promoter element (i.e., binding assay).

**[0138]** In another embodiment, the screening method comprises;

a) contacting a test compound with an Mlsn1 expressing cell, more particularly an antigen presenting cell (or a macrophage) and,

b) determining the ability of said test compound to affect Mlsn1 gene or an Mlsn1 gene product activity within the cells.

**[0139]** In a further embodiment, the screening method comprises contacting a test compound with a cell expressing a reporter gene under the control of Mlsn1 transcription promoter elements, more particularly an antigen presenting cell (or a macrophage), expressing the said promoter/reporter construct and determining the ability of the said test compound to modulate reporter expression.

**[0140]** Ligand binding screens to the Mlsn1 gene, polypeptide, a fragment thereof or Mlsn1 transcription promoter element can allow rapid identification of ligands which are capable of interacting with the gene or product(s). Further testing may be required to determine the therapeutic activity of these ligands. Development of a functional screen, whereby the activity of Mlsn1 is measured (or reporter activity measured) upon ligand binding allows ligands to be rapidly identified which modulate the activity of Mlsn1.

**[0141]** This invention also relates to the use of the Mlsn1 gene product to screen for any molecule involved in the Mlsn1 gene pathway, i.e. for any protein or peptide that interact with Mlsn1. Such targets (or Mlsn1 partners or signaling molecules) can be screened by protein-protein interaction analysis techniques (double-hybrid system for instance), or any other method known to the skilled artisan.

Ligand binding screening method (phage display, flashplate)

**[0142]** Ligand binding screening comprises, preferably, contacting a test compound with an Mlsn1 gene product or Mlsn1 transcription promoter element(s) and determining the ability of said test compound to interact with the Mlsn1 gene product or transcription promoter element(s). In a typical embodiment, the ligand binding screen comprises contacting one or several (in parallel) test compounds with a Mlsn1 polypeptide (e.g., protein or a fragment thereof) or transcription promoter element(s), in the presence (or absence, as a reference) of a Mlsn1 ligand or transcription promoter ligand, and assessing the ability of the test compound(s) to bind Mlsn1 transcription polypeptide or promoter element(s) by assessing ligand binding.

**[0143]** The Mlsn1 polypeptide can be part of an intact cell, membrane preparation or a purified polypeptide, optionally attached to a support, such as beads, column plate, etc. The polypeptide is preferably characterized by comprising an amino acid sequence selected from SEQ ID N°1 to 22.

**[0144]** The Mlsn1 transcription promoter element(s) can preferably but not exclusively be a purified nucleic acid, optionally attached to a support such as beads, column, plate etc.. The nucleic acid encoding the promoter elements is preferably characterized by comprising nucleic acid sequences selected from SEQ ID N°41, 42 or 43.

**[0145]** The test compound can be a peptide or a protein or an antibody or chemical entity, either alone or in combination(s) or in a mixture with any other substance. The test compound may even represent a library of compounds. Optionally, excess non Mlsn1 bound ligand can be removed by separation. Separation can take the form of washing /

filtering or centrifugation (to pellet the Mlsn1 protein). In this latter case, the supernatant can then be removed and the Mlsn1 re-suspended in buffer.

**[0146]** The binding is preferably performed in the presence of a Mlsn1 ligand or transcription promoter element ligand, to allow an assessment of the binding activity of each test compound. In particular, in a preferred embodiment, a labeled Mlsn1 ligand or promoter element ligand is used and the binding activity of the test compound is determined by assessing any modulation of the ligand binding.

**[0147]** The ligand may be contacted with the Mlsn1 polypeptide or transcription promoter element either before, simultaneously or after the test compound.

**[0148]** The ligand should be detectable and/or quantifiable. To achieve this, the ligand can be labeled in a number of ways, such as with a chromophore, radioactive, fluorescent, phosphorescent, enzymatic or antibody label, for instance. If the ligand is not directly detectable it must be amenable to detection and quantification by secondary detection, which may employ the above technologies. Preferably (in the case of certain detection methods), unbound ligand is removed from the mixture prior to detecting ligand binding, as described above.

**[0149]** Alternatively the Mlsn1 polypeptide or fragment thereof of Mlsn1 transcription promoter element(s) can be detectable or quantifiable. This can be achieved in a similar manner to that described above.

**[0150]** Preferred examples of such Mlsn1 ligands that can be used in the instant invention include any product that is known to interact with Mlsn1, such as an antibody, physiological ligand or receptor, or synthetic ligands or Mlsn1 inhibitors. Specific examples of such ligands include pyrazole compounds as described in WO99/19303, which exhibit calcium release-dependent calcium channel inhibitory effect. These compounds may be labeled according to various techniques, including radioactivity or fluorescence. More specific compounds are compounds SEW04225, KM02940, KM03000 and GK02421 as listed in the Maybridge Co. Trial Drug Catalogue (UK, Cornwall, published August 95).

**[0151]** Preferred examples of Mlsn1 transcription promoter element ligands that can be used in this instant invention include any product that is known to interact with Mlsn1 promoter elements, such as polymerases, antibodies, other naturally occurring ligands or synthetic ligands.

**[0152]** Binding of the test compound modifies the interaction of the ligand with the binding site and changes the affinity or binding of ligand for/to its binding site. The difference between the observed amount of ligand bound relative to the theoretical maximum amount of ligand bound (or to the ligand bound in the absence of a test compound under the same conditions) is a reflection of the binding ability (and optionally the amount and/or affinity) of a test compound to bind Mlsn1 or its transcription promoter elements.

**[0153]** Alternatively, the amount of test compound bound to the Mlsn1 protein or a fragment thereof, or transcription promoter element(s) can be determined by a combination of chromatography and mass spectroscopy. The amount of test compound bound to the Mlsn1 protein or a fragment thereof or transcription promoter element(s) can also be determined by direct measurement of the change in mass upon compound or ligand binding to Mlsn1 or transcription promoter element(s). This can be achieved with technologies such as Biacore (Amersham Pharmacia). Alternatively, the Mlsn1 protein or a fragment thereof, or promoter element(s), the compound or the ligand can be fluorescently labeled and association of Mlsn1 protein or promoter element(s) with the compound can be followed by changes in Fluorescence Energy Transfer (FRET).

#### Functional screening Method

**[0154]** In addition to, or in replacement of the binding screen, functional screens may be used to identify or characterize immuno-modulators or Mlsn1 modulators. Such a functional screen is advantageously transferable to high throughput, to allow the screening of large numbers of test compounds. The functional screen essentially comprises contacting a Mlsn1-expressing cell with a test compound, and assessing the ability of said test compound to affect Mlsn1 gene pathway or Mlsn1 gene product activity within the cells. This can be performed by measuring calcium fluxes, activation potential (or profile) or signal transduction pathway (e.g., selected cellular gene expression level) within said cells.

**[0155]** In a particular embodiment, the present invention thus resides in a method of screening (or identifying or selecting or characterizing) immuno-modulators, comprising (i) exposing a Mlsn1-expressing cell to one or several test compounds and (ii) detecting the compound that induce a modulation of the calcium flux or the activation potential in said cells. More preferably, the cells are recombinant cells expressing a Mlsn1 polypeptide, or immune cells, in particular antigen-presenting cells, more preferably macrophages.

**[0156]** In a first preferred variant of this embodiment, the functional screening method comprises the following steps:

- (1) obtaining a recombinant cell expressing Mlsn1 gene or gene product,
- (2) exposing said recombinant cell to a substance or molecule to be tested; and
- (3) measuring the change in calcium flux or activation potential within the exposed recombinant cell.

**[0157]** In a preferred embodiment, the cell express a functional Mlsn1 polypeptide comprising all or part of SEQ ID N°1, in particular a polypeptide comprising a sequence selected from SEQ ID N°1 to 22.

#### Calcium Flux

**[0158]** Calcium Flux may be measured according to various techniques, such as for instance using sodium sensitive dyes such as Fura II. In a particular embodiment, compounds are selected for their ability to cause an increase in calcium concentration within the cell. In an other embodiment, compounds are selected for their ability to cause a decrease in calcium concentration within the cell. Alternatively, compounds are selected for their ability to alter the rate of recovery of the calcium channel.

#### Electrophysiology

**[0159]** In another embodiment of the functional screening method, substances or molecules of interest are selected for their ability to induce changes in the activation potential, the inactivation time, or the rate of recovery of the sodium channel. Preferred molecules or substances are those inducing a decrease in the inactivation potential, and/or a decrease in the rate of inactivation, and/or which decreases the rate of recovery from inactivation, as compared with the same measures performed in the absence of the substance of molecule to be tested.

**[0160]** In an other preferred variant, the functional screen is based on a measure of selected gene expression or activity within the cells. In this regard, the present invention now proposes, for the first time, to select Icrac modulators based on their ability to modify the expression pattern of particular genes. More preferably, the selected genes are selected from cytokine genes (in particular interleukin-2, interleukin-8, etc.) and chemokine genes (in particular MIPa). Accordingly, a preferred screening method of this invention comprises contacting Mlsn1-expressing cells with one or several test compounds and identifying the compounds that cause a modification in expression or activity of a gene selected from a cytokine or a chemokine gene, within said exposed cells.

**[0161]** Modification in gene expression or activity may be determined by assessing the presence or (relative) level of the gene product in the cell, in particular of the mRNA or polypeptide. Polypeptides may be detected or dosed using affinity reagents, such as antibodies (or fragments or derivatives thereof), either within the cells or at the cell surface, or in the cell culture medium (or supernatant). mRNA levels may be evaluated using conventional techniques (primers, probes, etc). In a specific embodiment, the exposed cells (or any preparation derived therefrom such as membranes, pellets, RNAs, etc) are contacted with a support to which particular affinity reagents are attached (antibodies or nucleic acids).

#### Transcription promoter-reporter

**[0162]** In a further embodiment, a promoter-reporter assay can be used to screen for compounds that modulate Mlsn1 expression. A reporter gene encoding a reporter molecule (i.e. a molecule(s) that can be directly or indirectly detected) is placed 3' with respect to the nucleic acids encoding Mlsn1 transcription promoter element(s). The position and orientation of which is such that reporter gene expression is initiated from the Mlsn1 transcription promoter. Preferable transcription promoter elements are chosen from nucleic acids encoded by SEQ ID N°41, 42 or 43. Preferred reporter genes could be those that encode functional  $\beta$ -Galactosidase, Green fluorescence Protein or Luciferase. However, many other reporter molecules could be used, the identity and characteristics of which are known to those skilled in the art.

**[0163]** Further optimization of reporter expression could be achieved with the inclusion of additional nucleic acid elements, as described previously.

**[0164]** The action of test compounds can be measured by exposure of these compounds to cell lines expressing the promoter-reporter construct. Compounds which down regulate the action of the Mlsn1 promoter elements will manifest by reducing the amount or activity of the reporter in the cell relative to cells not compound treated cells. Compounds which up regulate the action of the Mlsn1 promoter elements will manifest by increasing the amount or activity of the reporter in the cell relative to cells not compound treated cells.

**[0165]** Compounds which modulate the amount of reporter or activity by other means could be distinguished from those that do not by secondary screening with similar cell lines expressing reporter with non Mlsn1 transcription promoter elements. Compounds which modulate the reporter in the first screen and not in the second screen could be considered to mediate its action either directly or indirectly through the Mlsn1 transcription promoter elements.

**[0166]** An assay would comprise the following steps;

- a) obtaining a recombinant cell expressing a reporter molecule under the control of an Mlsn1 transcription promoter Mlsn1 gene,

b) exposing said recombinant cell to a substance or molecule to be tested ; and Measuring the change in reporter expression or activity in said recombinant cell.

According to a further embodiment, the active compounds are evaluated by their in vitro activity. This activity is determined for example by IC 50 measurement.

The choice of compounds (called in-after hit) from which series of compounds are developed should take into account parameters such as IC 50 measurement, structural features of the hits, the possibility to alter the substituents, the physico-chemical properties such as solubility as well as physiological features such as half-life and bioavailability.....

Therefore another embodiment of the instant invention is the use of an active compound (hit) as identified, selected or characterized by the screening method described previously for the preparation of pharmaceutical compounds or compositions that bind and/or modulate the biological activity of Mlsn1.

## DIAGNOSTICS ASSAYS AND KITS

**[0167]** The inventors believe that the ability to detect and measure the amount of Mlsn1 in immune cells would be a valuable tool for the prognosis of aberrant Mlsn1 and Icrac conditions in humans. The inventors believe that detection of Mlsn1 in immune cells could be achieved by measuring the amount of expressed Mlsn1 nucleic acid in immune cells. This could be achieved using a multi-step process:

- (a) obtaining a (small) blood sample from a patient (of known volume),
- (b) optionally separating immune cells from other blood components. This could be achieved by centrifugation and/or affinity chromatography.
- (c) reverse transcribe cellular RNA to cDNA using protocols similar to those described in examples,
- (d) amplify Mlsn1 nucleic acid obtained in step c) by PCR with a plurality of Mlsn1 specific oligonucleotides capable of hybridizing, under stringent conditions to Mlsn1 nucleic acids. Such oligonucleotides and amplification conditions have been outlined in examples,
- (e) characterize the PCR product. One way to identify the presence of an Mlsn1 specific PCR product would be to electrophorese an aliquote of amplified nucleic acid in agarose. Such technology has been outlined in examples.

**[0168]** This test could be used to determine if Mlsn1 is being expressed in immune cells, therefore providing a valuable indication in the prognosis of disorders associated with aberrant Icrac function. Furthermore, an ability to accurately quantitate the level of expression of Mlsn1 in immune cells would help in the prognosis of the severity of an Icrac associated disorder. Quantitation could be achieved with the inclusion of further step(s). This step(s) would require the assay of a known quantity of nucleic acid which encodes Mlsn1.

**[0169]** Co-electrophoresis of samples which have not been subjected to blood products but have been "spiked" with known concentrations of nucleic acids encoding human Mlsn1 such as SEQ ID N°23 or fragments thereof, or complementary sequences thereto with the test sample, would allow identification and quantitation of levels of Mlsn1 in the blood sample.

**[0170]** Other aspects and advantages of the present invention will be disclosed in the following experimental section, which should be regarded as illustrative and not limiting the scope of protection.

## EXAMPLES

### 1. A Mlsn1 gene product is expressed in human U937 cells

**[0171]** cDNA libraries were prepared from U937 cells (these cells being referred to as undifferentiated cells) and dbcAMP differentiated U937 cells (48 hours) using the Clontech SMART system (K1052-1). In brief, U937 cells were cultured in suspension in cell culture media (RPMI 1640 plus 10% heat inactivated FBS) at 37°C in 5 % CO<sub>2</sub>, in the presence and absence of differentiating agent (dbcAMP Conc 1µM) (48 hours). Differentiated and undifferentiated cells were harvested by centrifugation at 1,200g for 10 minutes, the supernatant removed from the cell pellet and cellular RNA extracted using FastTrack 2.0 kit (Invitrogen). A SMART library was prepared from the differentiated and undifferentiated cells according to Clontech protocols. These libraries were used to determine the relative abundance of Mlsn1 cDNA in the libraries.

**[0172]** Two nested primer pairs were designed to screen for Mlsn1 cDNA in the above libraries. The sequence of these primers is as follows:

Primer 1a (SEQ ID N°44) : TGAAGTAAAATCAATATCCAACCAGG

Primer 1b (SEQ ID N°45) : GCACTGCTCCTCGAACTCATGCAGCC

Primer 2a (SEQ ID N°46) : TCCAACCAGGTGTGGAAGTTCCAGCG

Primer 2b (SEQ ID N°47) : GGAAGTGCTCCTGCACGCACTGCTCC

Under the following PCR conditions:

**[0173]** All PCR reactions were carried out using Advantage cDNA PCR Kit (#K1905-1) in the suppliers buffer conditions. PCR primer concentration was 20 fmol/primer/100µl reaction. cDNA template concentration was 0.2µg. PCR reactions were carried out as two step cycles (denaturation 95°C for 30 sec, for annealing/elongation 68°C for 6min). PCR product was visualised by electrophoresis in an agarose gel and staining with ethidium bromide (0.5µg/ml).

**[0174]** Nested PCR using primer pair SEQ ID N°44 and 47 and nested pair SEQ ID N°45 and 46 indicated Mlsn1 expression in U937 cells. Similar experiments indicated the expression of Mlsn1 in Jurkat and KU-812 cells.

**[0175]** The presence of the expression of Mlsn1 in these libraries was further confirmed using nested PCR primers, SEQ ID NO: 44 and 46 together with CDS III/3 primer of SMART cDNA libraries (c.f. Advantage cDNA PCR Kit (#K1905-1)). A 2kb PCR product was obtained (figure 2). This corresponded to the predicted size of a Mlsn1 PCR product and indicated that the gene encoding Mlsn1 was being expressed in differentiated and undifferentiated U937 cell lines. However, the intensity of the band corresponding to the Mlsn1 PCR product was greater in the differentiated cell library than in the undifferentiated cell library. These results indicate that Mlsn1 cDNA was present in U937 cell lines and that this Mlsn1 expression was more pronounced in U937 cell lines differentiated to a macrophage phenotype.

## 2. MLSN1(TrpC8) expression is differentially increased in dbcAMP treated (macrophage-like) U937 cells

**[0176]** The relative abundance of nucleic acid encoding Mlsn1 in undifferentiated and dbcAMP differentiated U937 cell lines was determined. Briefly, undifferentiated and dbcAMP differentiated U937 cell lines were cultured and harvested as described above, and subtracted cDNA libraries prepared using Clontech PCR Select kit (K1804-1) and the protocols therein. This technology allows for selective enrichment of differentially expressed genes between the differentiated and undifferentiated cell lines.

**[0177]** Following PCR subtraction, Mlsn1 specific primer pair (SEQ ID N°44 and 47) were used to amplify cDNA-fragments from Mlsn1 from subtracted pools and controls using Advantage cDNA PCR, under suppliers conditions as two step cycles (95°C for 30 sec, 68°C for 1min annealing/elongation) (figure 3).

**[0178]** Forward subtraction of differentiated cells nucleic acid with non differentiated cell nucleic acid resulted in a strong Mlsn1 specific PCR product. Back subtraction of non-differentiated cells with differentiated cells resulted in undetectable amounts PCR product. This result indicates that there was greater Mlsn1 gene expression in the differentiated U937 cells than in the undifferentiated U937 cells. This result correlates with the observation that the Icrac phenotype increases during dbcAMP treatment of U937 cell line to a more macrophage like phenotype. The inventors conclude that the emergence of an Icrac phenotype in the dbcAMP differentiated U937 cells corresponds with an increase in expression of Mlsn1.

## 3. Down regulation of calcium entry in dbcAMP treated (macrophage-like) U937 cells using Mlsn1 anti-sense

**[0179]** U937 cells were cultured at 37°C in 5 % CO<sub>2</sub> in RPMI 1640, 10% heat inactivated FBS. Cells were incubated in absence or presence of 0, 1 or 10 µM Mlsn1 anti-sense nucleic acid (SEQ ID NO:26), 4 hours prior to and 48h during differentiation treatment by dbcAMP. All oligonucleotides used in this experiment contained phosphorothiorate modifications at their first two and last two linkages (terminal 5' and 3' linkage). A scrambled oligonucleotide (5'CTGGTGAAGAAGAGGACGTCCAT3', SEQ ID N°48) was used at 10 µM in the same conditions as negative con-

tol. Another oligonucleotide against FcγRII (CD 32) (5'TCTGGGACATACATTCTGAGACAT3', SEQ ID N° 49) was used at 10 μM in the same conditions as positive control for anti-sense uptake. DbcAMP induced differentiation was controlled by immunofluorescence using FACS quantitation of CD 32 and CD 64, as well as Ca<sup>2+</sup> signaling after Thapsigargin stimulation, before and after dbcAMP treatment using fura 2 and fluorimetry. Intra-cellular Ca<sup>2+</sup> after stimulation by ionomycin was measured during FACS analysis. Cells were loaded with 2 μg/ml indo-1 during 30 min at 37°C, 5 % CO<sub>2</sub> in RPMI, 2 mg/ml BSA. Cells were then activated by 2 μg/ml ionomycin. Ionomycin induced variations of intra-cellular calcium concentration were measured by flow cytometry (figure 4). CD 32 expression was quantitated by immunofluorescence with an FITC labeled anti CD32 antibody using flow cytometry.

Comparison of figures 4 a) and 4 b) shows apparition of a long lasting calcium current when differentiating U937 cells by dbcAMP.

Comparison of figures 4 b) and 4 c) shows a non specific inhibition of long lasting calcium current due to antisense treatment conditions.

Comparison of figures 4 c) and 4 d) show a specific inhibition of long lasting calcium current due to 1 μM Mln1 antisense.

Comparison of figures 4 c) and 4 e) show a specific inhibition of long lasting calcium current due to 10 μM Mln1 antisense.

Dibutyrate cAMP differentiation induced differences in calcium flux responses on U937 cells after Ionomycine 2 μg/ml stimulation (R2 were 3% in wild type Vs 29% after DBC differentiation. Particularly,

Even though anti-sense condition (absence of anti-sense itself) affected the level of differentiation (R2 were 29% Vs 23%), Mln1 anti-sense treatment seems to decrease the expression of R2 population in a dose-related manner, R2 moved from 23% to 17% and 14% according to anti-sense control, 1 μM and 10 μM Mln1 anti-sense treatment respectively.

These data show the existence of a long lasting calcium flux after DBC differentiation of U937 cells with 39% inhibition of this typical calcium flux induction after Mln1 anti-sense 10 μM treatment.

DbcAMP treatment was shown to increase intra-cellular Ca<sup>2+</sup> after stimulation according to Floto et al.. The results demonstrate that anti-sense oligonucleotide treated cultures have their calcium signal decreased by about 40%. This antisense experiment demonstrates that the down-regulation of Mln1 expression anti-sense oligonucleotides results in a diminished Icrac phenotype in U937 cell lines. This result also demonstrates that anti-sense nucleic acid specific to Mln1 is capable of down-regulating the Icrac phenotype.

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SEQUENCE LISTING

**[0181]**

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# EP 1 184 457 A1

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5	Tyr	Cys	Glu	Glu	Gly	Gly	Ile	Ile	Asn	Glu	Ser	Leu	Arg	Glu	Gln	Leu	
	225					230					235					240	
	Leu	Val	Thr	Ile	Gln	Lys	Thr	Phe	Asn	Tyr	Asn	Lys	Ala	Gln	Ser	His	
					245					250					255		
10	Gln	Leu	Phe	Ala	Ile	Ile	Met	Glu	Cys	Met	Lys	Lys	Lys	Glu	Leu	Val	
				260					265						270		
	Thr	Val	Phe	Arg	Met	Gly	Ser	Glu	Gly	Gln	Gln	Asp	Ile	Glu	Met	Ala	
			275					280					285				
15	Ile	Leu	Thr	Ala	Leu	Leu	Lys	Gly	Thr	Asn	Val	Ser	Ala	Pro	Asp	Gln	
	290						295					300					
	Leu	Ser	Leu	Ala	Leu	Ala	Trp	Asn	Arg	Val	Asp	Ile	Ala	Arg	Ser	Gln	
20	305					310					315					320	
	Ile	Phe	Val	Phe	Gly	Pro	His	Trp	Thr	Pro	Leu	Gly	Ser	Leu	Ala	Pro	
					325					330					335		
	Pro	Thr	Asp	Ser	Lys	Ala	Thr	Glu	Lys	Glu	Lys	Lys	Pro	Pro	Met	Ala	
25				340				345						350			
	Thr	Thr	Lys	Gly	Gly	Arg	Gly	Lys	Gly	Lys	Gly	Lys	Lys	Lys	Gly	Lys	
			355				360						365				
30	Val	Lys	Glu	Glu	Val	Glu	Glu	Glu	Thr	Asp	Pro	Arg	Lys	Ile	Glu	Leu	
	370						375					380					
	Leu	Asn	Trp	Val	Asn	Ala	Leu	Glu	Gln	Ala	Met	Leu	Asp	Ala	Leu	Val	
	385					390					395					400	
35	Leu	Asp	Arg	Val	Asp	Phe	Val	Lys	Leu	Leu	Ile	Glu	Asn	Gly	Val	Asn	
				405						410					415		
	Met	Gln	His	Phe	Leu	Thr	Ile	Pro	Arg	Leu	Glu	Glu	Leu	Tyr	Asn	Thr	
				420					425					430			
40	Arg	Leu	Gly	Pro	Pro	Asn	Thr	Leu	His	Leu	Leu	Val	Arg	Asp	Val	Lys	
			435				440						445				
	Lys	Ser	Asn	Leu	Pro	Pro	Asp	Tyr	His	Ile	Ser	Leu	Ile	Asp	Ile	Gly	
	450						455					460					
45	Leu	Val	Leu	Glu	Tyr	Leu	Met	Gly	Gly	Ala	Tyr	Arg	Cys	Asn	Tyr	Thr	
	465					470					475					480	
	Arg	Lys	Asn	Phe	Arg	Thr	Leu	Tyr	Asn	Asn	Leu	Phe	Gly	Pro	Lys	Arg	
				485					490					495			
50	Pro	Lys	Ala	Leu	Lys	Leu	Leu	Gly	Met	Glu	Asp	Asp	Glu	Pro	Pro	Ala	
				500					505					510			
	Lys	Gly	Lys	Lys	Lys	Lys	Lys	Lys	Lys	Lys	Glu	Glu	Glu	Ile	Asp	Ile	
55			515				520						525				

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	Asp Val Asp Asp Pro Ala Val Ser Arg Phe Gln Tyr Pro Phe His Glu	
	530	535 540
5	Leu Met Val Trp Ala Val Leu Met Lys Arg Gln Lys Met Ala Val Phe	
	545	550 555 560
	Leu Trp Gln Arg Gly Glu Glu Ser Met Ala Lys Ala Leu Val Ala Cys	
		565 570 575
10	Lys Leu Tyr Lys Ala Met Ala His Glu Ser Ser Glu Ser Asp Leu Val	
		580 585 590
	Asp Asp Ile Ser Gln Asp Leu Asp Asn Asn Ser Lys Asp Phe Gly Gln	
		595 600 605
15	Leu Ala Leu Glu Leu Leu Asp Gln Ser Tyr Lys His Asp Glu Gln Ile	
		610 615 620
	Ala Met Lys Leu Leu Thr Tyr Glu Leu Lys Asn Trp Ser Asn Ser Thr	
		625 630 635 640
20	Cys Leu Lys Leu Ala Val Ala Ala Lys His Arg Asp Phe Ile Ala His	
		645 650 655
	Thr Cys Ser Gln Met Leu Leu Thr Asp Met Trp Met Gly Arg Leu Arg	
		660 665 670
25	Met Arg Lys Asn Pro Gly Leu Lys Val Ile Met Gly Ile Leu Leu Pro	
		675 680 685
	Pro Thr Ile Leu Phe Leu Glu Phe Arg Thr Tyr Asp Asp Phe Ser Tyr	
		690 695 700
30	Gln Thr Ser Lys Glu Asn Glu Asp Gly Lys Glu Lys Glu Glu Glu Asn	
		705 710 715 720
35	Thr Asp Ala Asn Ala Asp Ala Gly Ser Arg Lys Gly Asp Glu Glu Asn	
		725 730 735
	Glu His Lys Lys Gln Arg Ser Ile Pro Ile Gly Thr Lys Ile Cys Glu	
		740 745 750
40	Phe Tyr Asn Ala Pro Ile Val Lys Phe Trp Phe Tyr Thr Ile Ser Tyr	
		755 760 765
	Leu Gly Tyr Leu Leu Leu Phe Asn Tyr Val Ile Leu Val Arg Met Asp	
		770 775 780
45	Gly Trp Pro Ser Leu Gln Glu Trp Ile Val Ile Ser Tyr Ile Val Ser	
		785 790 795 800
	Leu Ala Leu Glu Lys Ile Arg Glu Ile Leu Met Ser Glu Pro Gly Lys	
		805 810 815
50	Leu Ser Gln Lys Ile Lys Val Trp Leu Gln Glu Tyr Trp Asn Ile Thr	
		820 825 830
	Asp Leu Val Ala Ile Ser Thr Phe Met Ile Gly Ala Ile Leu Arg Leu	
		835 840 845
55		

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	Gln Asn Gln Pro Tyr Met	Gly Tyr Gly Arg Val	Ile Tyr Cys Val Asp
	850	855	860
5	Ile Ile Phe Trp Tyr Ile Arg Val Leu Asp	Ile Phe Gly Val Asn Lys	
	865	870	875 880
	Tyr Leu Gly Pro Tyr Val Met Met Ile Gly Lys Met Met Ile Asp Met		
		885	890 895
10	Leu Tyr Phe Val Val Ile Met Leu Val Val Leu Met Ser Phe Gly Val		
		900	905 910
	Ala Arg Gln Ala Ile Leu His Pro Glu Glu Lys Pro Ser Trp Lys Leu		
		915	920 925
15	Ala Arg Asn Ile Phe Tyr Met Pro Tyr Trp Met Ile Tyr Gly Glu Val		
		930	935 940
	Phe Ala Asp Gln Ile Asp Leu Tyr Ala Met Glu Ile Asn Pro Pro Cys		
		945	950 955 960
20	Gly Glu Asn Leu Tyr Asp Glu Glu Gly Lys Arg Leu Pro Pro Cys Ile		
		965	970 975
	Pro Gly Ala Trp Leu Thr Pro Ala Leu Met Ala Cys Tyr Leu Leu Val		
		980	985 990
25	Ala Asn Ile Leu Leu Val Asn Leu Leu Ile Ala Val Phe Asn Asn Thr		
		995	1000 1005
	Phe Phe Glu Val Lys Ser Ile Ser Asn Gln Val Trp Lys Phe Gln Arg		
		1010	1015 1020
30	Tyr Gln Leu Ile Met Thr Phe His Asp Arg Pro Val Leu Pro Pro Pro		
		1025	1030 1035 1040
	Met Ile Ile Leu Ser His Ile Tyr Ile Ile Ile Met Arg Leu Ser Gly		
		1045	1050 1055
35	Arg Cys Arg Lys Lys Arg Glu Gly Asp Gln Glu Glu Arg Asp Arg Gly		
		1060	1065 1070
40	Leu Lys Leu Phe Leu Ser Asp Glu Glu Leu Lys Arg Leu His Glu Phe		
		1075	1080 1085
	Glu Glu Gln Cys Val Gln Glu His Phe Arg Glu Lys Glu Asp Glu Gln		
		1090	1095 1100
45	Gln Ser Ser Ser Asp Glu Arg Ile Arg Val Thr Ser Glu Arg Val Glu		
		1105	1110 1115 1120
	Asn Met Ser Met Arg Leu Glu Glu Ile Asn Glu Arg Glu Thr Phe Met		
		1125	1130 1135
50	Lys Thr Ser Leu Gln Thr Val Asp Leu Arg Leu Ala Gln Leu Glu Glu		
		1140	1145 1150
	Leu Ser Asn Arg Met Val Asn Ala Leu Glu Asn Leu Ala Gly Ile Asp		
		1155	1160 1165
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Arg Ser Asp Leu Ile Gln Ala Arg Ser Arg Ala Ser Ser Glu Cys Glu  
 1170 1175 1180  
 5 Ala Thr Tyr Leu Leu Arg Gln Ser Ser Ile Asn Ser Ala Asp Gly Tyr  
 1185 1190 1195 1200  
 Ser Leu Tyr Arg Tyr His Phe Asn Gly Glu Glu Leu Leu Phe Glu Asp  
 1205 1210 1215  
 10 Thr Ser Leu Ser Thr Ser Pro Gly Thr Gly Val Arg Lys Lys Thr Cys  
 1220 1225 1230  
 Ser Phe Arg Ile Lys Glu Glu Lys Asp Val Lys Thr His Leu Val Pro  
 1235 1240 1245  
 15 Glu Cys Gln Asn Ser Leu His Leu Ser Leu Gly Thr Ser Thr Ser Ala  
 1250 1255 1260  
 Thr Pro Asp Gly Ser His Leu Ala Val Asp Asp Leu Lys Asn Ala Glu  
 1265 1270 1275 1280  
 20 Glu Ser Lys Leu Gly Pro Asp Ile Gly Ile Ser Lys Glu Asp Asp Glu  
 1285 1290 1295  
 Arg Gln Thr Asp Ser Lys Lys Glu Glu Thr Ile Ser Pro Ser Leu Asn  
 1300 1305 1310  
 25 Lys Thr Asp Val Ile His Gly Gln Asp Lys Ser Asp Val Gln Asn Thr  
 1315 1320 1325  
 Gln Leu Thr Val Glu Thr Thr Asn Ile Glu Gly Thr Ile Ser Tyr Pro  
 1330 1335 1340  
 Leu Glu Glu Thr Lys Ile Thr Arg Tyr Phe Pro Asp Glu Thr Ile Asn  
 1345 1350 1355 1360  
 35 Ala Cys Lys Thr Met Lys Ser Arg Ser Phe Val Tyr Ser Arg Gly Arg  
 1365 1370 1375  
 Lys Leu Val Gly Gly Val Asn Gln Asp Val Glu Tyr Ser Ser Ile Thr  
 1380 1385 1390  
 40 Asp Gln Gln Leu Thr Thr Glu Trp Gln Cys Gln Val Gln Lys Ile Thr  
 1395 1400 1405  
 Arg Ser His Ser Thr Asp Ile Pro Tyr Ile Val Ser Glu Ala Ala Val  
 1410 1415 1420  
 45 Gln Ala Glu Gln Lys Glu Gln Phe Ala Asp Met Gln Asp Glu His His  
 1425 1430 1435 1440  
 Val Ala Glu Ala Ile Pro Arg Ile Pro Arg Leu Ser Leu Thr Ile Thr  
 1445 1450 1455  
 50 Asp Arg Asn Gly Met Glu Asn Leu Leu Ser Val Lys Pro Asp Gln Thr  
 1460 1465 1470  
 Leu Gly Phe Pro Ser Leu Arg Ser Lys Ser Leu His Gly His Pro Arg  
 1475 1480 1485  
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Asn Val Lys Ser Ile Gln Gly Lys Leu Asp Arg Ser Gly His Ala Ser  
 1490 1495 1500  
 5 Ser Val Ser Ser Leu Val Ile Val Ser Gly Met Thr Ala Glu Glu Lys  
 1505 1510 1515 1520  
 Lys Val Lys Lys Glu Lys Ala Ser Thr Glu Thr Glu Cys  
 1525 1530  
 10  
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 15  
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 20 Leu Met Val Lys Asp Trp Gln Leu Glu Leu Pro Lys Leu Leu Ile Ser  
 20 25 30  
 Val His Gly Gly Leu Gln Asn Phe Glu Met Gln Pro Lys Leu Lys Gln  
 35 40 45  
 25 Val Phe Gly Lys Gly Leu Ile Lys Ala Ala Met Thr Thr Gly Ala Trp  
 50 55 60  
 Ile Phe Thr Gly Gly Val Ser Thr Gly Val Ile Ser His Val Gly Asp  
 65 70 75 80  
 30 Ala Leu Lys Asp His Ser Ser Lys Ser Arg Gly Arg Val Cys Ala Ile  
 85 90 95  
 Gly Ile Ala Pro Trp Gly Ile Val Glu Asn Lys Glu Asp Leu Val Gly  
 100 105 110  
 35 Lys Asp Val Thr Arg Val Tyr Gln Thr Met Ser Asn Pro Leu Ser Lys  
 115 120 125  
 Leu Ser Val Leu Asn Asn Ser His Thr His Phe Ile Leu Ala Asp Asn  
 130 135 140  
 40 Gly Thr Leu Gly Lys Tyr Gly Ala Glu Val Lys Leu Arg Arg Leu Leu  
 145 150 155 160  
 Glu Lys His Ile Ser Leu Gln Lys Ile Asn Thr Arg Leu Gly Gln Gly  
 165 170 175  
 45 Val Pro Leu Val Gly Leu Val Val Glu Gly Gly Pro Asn Val Val Ser  
 180 185 190  
 Ile Val Leu Glu Tyr Leu Gln Glu Glu Pro Pro Ile Pro Val Val Ile  
 195 200 205  
 50 Cys Asp Gly Ser Gly Arg Ala Ser Asp Ile Leu Ser Phe Ala His Lys  
 210 215 220  
 55 Tyr Cys Glu Glu Gly Gly Ile Ile Asn Glu Ser Leu Arg Glu Gln Leu

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	225		230		235		240
5	Leu Val Thr Ile Gln Lys Thr Phe Asn Tyr Asn Lys Ala Gln Ser His	245	250	255			
	Gln Leu Phe Ala Ile Ile Met Glu Cys Met Lys Lys Lys Glu Leu Val	260	265	270			
10	Thr Val Phe Arg Met Gly Ser Glu Gly Gln Gln Asp Ile Glu Met Ala	275	280	285			
	Ile Leu Thr Ala Leu Leu Lys Gly Thr Asn Val Ser Ala Pro Asp Gln	290	295	300			
15	Leu Ser Leu Ala Leu Ala Trp Asn Arg Val Asp Ile Ala Arg Ser Gln	305	310	315			320
	Ile Phe Val Phe Gly Pro His Trp Thr Pro Leu Gly Ser Leu Ala Pro	325	330	335			
20	Pro Thr Asp Ser Lys Ala Thr Glu Lys Glu Lys Lys Pro Pro Met Ala	340	345	350			
	Thr Thr Lys Gly Gly Arg Gly Lys Gly Lys Gly Lys Lys Lys Gly Lys	355	360	365			
25	Val Lys Glu Glu Val Glu Glu Glu Thr Asp Pro Arg Lys Ile Glu Leu	370	375	380			
	Leu Asn Trp Val Asn Ala Leu Glu Gln Ala Met Leu Asp Ala Leu Val	385	390	395			400
30	Leu Asp Arg Val Asp Phe Val Lys Leu Leu Ile Glu Asn Gly Val Asn	405	410	415			
	Met Gln His Phe Leu Thr Ile Pro Arg Leu Glu Glu Leu Tyr Asn Thr	420	425	430			
35	Arg Leu Gly Pro Pro Asn Thr Leu His Leu Leu Val Arg Asp Val Lys	435	440	445			
	Lys Ser Asn Leu Pro Pro Asp Tyr His Ile Ser Leu Ile Asp Ile Gly	450	455	460			
40	Leu Val Leu Glu Tyr Leu Met Gly Gly Ala Tyr Arg Cys Asn Tyr Thr	465	470	475			480
	Arg Lys Asn Phe Arg Thr Leu Tyr Asn Asn Leu Phe Gly Pro Lys Arg	485	490	495			
45	Pro Lys Ala Leu Lys Leu Leu Gly Met Glu Asp Asp Glu Pro Pro Ala	500	505	510			
50	Lys Gly Lys Lys Lys Lys Lys Lys Lys Lys Glu Glu Glu Ile Asp Ile	515	520	525			
	Asp Val Asp Asp Pro Ala Val Ser Arg Phe Gln Tyr Pro Phe His Glu	530	535	540			
55	Leu Met Val Trp Ala Val Leu Met Lys Arg Gln Lys Met Ala Val Phe						

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	545		550		555		560
	Leu Trp Gln Arg Gly Glu Glu Ser Met Ala Lys Ala Leu Val Ala Cys						
5		565		570		575	
	Lys Leu Tyr Lys Ala Met Ala His Glu Ser Ser Glu Ser Asp Leu Val						
		580		585		590	
10	Asp Asp Ile Ser Gln Asp Leu Asp Asn Asn Ser Lys Asp Phe Gly Gln						
		595		600		605	
	Leu Ala Leu Glu Leu Leu Asp Gln Ser Tyr Lys His Asp Glu Gln Ile						
		610		615		620	
15	Ala Met Lys Leu Leu Thr Tyr Glu Leu Lys Asn Trp Ser Asn Ser Thr						
		625		630		635	
	Cys Leu Lys Leu Ala Val Ala Ala Lys His Arg Asp Phe Ile Ala His						
		645		650		655	
20	Thr Cys Ser Gln Met Leu Leu Thr Asp Met Trp Met Gly Arg Leu Arg						
		660		665		670	
	Met Arg Lys Asn Pro Gly Leu Lys Val Ile Met Gly Ile Leu Leu Pro						
		675		680		685	
25	Pro Thr Ile Leu Phe Leu Glu Phe Arg Thr Tyr Asp Asp Phe Ser Tyr						
		690		695		700	
	Gln Thr Ser Lys Glu Asn Glu Asp Gly Lys Glu Lys Glu Glu Glu Asn						
		705		710		715	
30	Thr Asp Ala Asn Ala Asp Ala Gly Ser Arg Lys Gly Asp Glu Glu Asn						
		725		730		735	
	Glu His Lys Lys Gln Arg Ser Ile Pro Ile Gly Thr Lys Ile Cys Glu						
		740		745		750	
35	Phe Tyr Asn Ala						
		755					
40	<210> 3						
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45	<400> 3						
	Met Tyr Ile Arg Val Ser Tyr Asp Thr Lys Pro Asp Ser Leu Leu His						
	1 5 10 15						
	Leu Met Val Lys Asp Trp Gln Leu Glu Leu Pro Lys Leu Leu Ile Ser						
		20		25		30	
50	Val His Gly Gly Leu Gln Asn Phe Glu Met Gln Pro Lys Leu Lys Gln						
		35		40		45	
	Val Phe Gly Lys Gly Leu Ile Lys Ala Ala Met Thr Thr Gly Ala Trp						
		50		55		60	
55							

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	Ile	Phe	Thr	Gly	Gly	Val	Ser	Thr	Gly	Val	Ile	Ser	His	Val	Gly	Asp	65	70	75	80
5	Ala	Leu	Lys	Asp	His	Ser	Ser	Lys	Ser	Arg	Gly	Arg	Val	Cys	Ala	Ile		85	90	95
	Gly	Ile	Ala	Pro	Trp	Gly	Ile	Val	Glu	Asn	Lys	Glu	Asp	Leu	Val	Gly	100	105	110	
10	Lys	Asp	Val	Thr	Arg	Val	Tyr	Gln	Thr	Met	Ser	Asn	Pro	Leu	Ser	Lys	115	120	125	
	Leu	Ser	Val	Leu	Asn	Asn	Ser	His	Thr	His	Phe	Ile	Leu	Ala	Asp	Asn	130	135	140	
15	Gly	Thr	Leu	Gly	Lys	Tyr	Gly	Ala	Glu	Val	Lys	Leu	Arg	Arg	Leu	Leu	145	150	155	160
	Glu	Lys	His	Ile	Ser	Leu	Gln	Lys	Ile	Asn	Thr	Arg	Leu	Gly	Gln	Gly	165	170	175	
20	Val	Pro	Leu	Val	Gly	Leu	Val	Val	Glu	Gly	Gly	Pro	Asn	Val	Val	Ser	180	185	190	
	Ile	Val	Leu	Glu	Tyr	Leu	Gln	Glu	Glu	Pro	Pro	Ile	Pro	Val	Val	Ile	195	200	205	
25	Cys	Asp	Gly	Ser	Gly	Arg	Ala	Ser	Asp	Ile	Leu	Ser	Phe	Ala	His	Lys	210	215	220	
30	Tyr	Cys	Glu	Glu	Gly	Gly	Ile	Ile	Asn	Glu	Ser	Leu	Arg	Glu	Gln	Leu	225	230	235	240
	Leu	Val	Thr	Ile	Gln	Lys	Thr	Phe	Asn	Tyr	Asn	Lys	Ala	Gln	Ser	His	245	250	255	
35	Gln	Leu	Phe	Ala	Ile	Ile	Met	Glu	Cys	Met	Lys	Lys	Lys	Glu	Leu	Val	260	265	270	
	Thr	Val	Phe	Arg	Met	Gly	Ser	Glu	Gly	Gln	Gln	Asp	Ile	Glu	Met	Ala	275	280	285	
40	Ile	Leu	Thr	Ala	Leu	Leu	Lys	Gly	Thr	Asn	Val	Ser	Ala	Pro	Asp	Gln	290	295	300	
	Leu	Ser	Leu	Ala	Leu	Ala	Trp	Asn	Arg	Val	Asp	Ile	Ala	Arg	Ser	Gln	305	310	315	320
45	Ile	Phe	Val	Phe	Gly	Pro	His	Trp	Thr	Pro	Leu	Gly	Ser	Leu	Ala	Pro	325	330	335	
	Pro	Thr	Asp	Ser	Lys	Ala	Thr	Glu	Lys	Glu	Lys	Lys	Pro	Pro	Met	Ala	340	345	350	
50	Thr	Thr	Lys	Gly	Gly	Arg	Gly	Lys	Gly	Lys	Gly	Lys	Lys	Lys	Gly	Lys	355	360	365	
	Val	Lys	Glu	Glu	Val	Glu	Glu	Glu	Thr	Asp	Pro	Arg	Lys	Ile	Glu	Leu	370	375	380	

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	Leu Asn Trp Val Asn Ala Leu Glu Gln Ala Met Leu Asp Ala Leu Val	385	390	395	400
5	Leu Asp Arg Val Asp Phe Val Lys Leu Leu Ile Glu Asn Gly Val Asn	405	410	415	
	Met Gln His Phe Leu Thr Ile Pro Arg Leu Glu Glu Leu Tyr Asn Thr	420	425	430	
10	Arg Leu Gly Pro Pro Asn Thr Leu His Leu Leu Val Arg Asp Val Lys	435	440	445	
	Lys Ser Asn Leu Pro Pro Asp Tyr His Ile Ser Leu Ile Asp Ile Gly	450	455	460	
15	Leu Val Leu Glu Tyr Leu Met Gly Gly Ala Tyr Arg Cys Asn Tyr Thr	465	470	475	480
	Arg Lys Asn Phe Arg Thr Leu Tyr Asn Asn Leu Phe Gly Pro Lys Arg	485	490	495	
20	Pro Lys Ala Leu Lys Leu Leu Gly Met Glu Asp Asp Glu Pro Pro Ala	500	505	510	
	Lys Gly Lys Lys Lys Lys Lys Lys Lys Lys Glu Glu Glu Ile Asp Ile	515	520	525	
25	Asp Val Asp Asp Pro Ala Val Ser Arg Phe Gln Tyr Pro Phe His Glu	530	535	540	
	Leu Met Val Trp Ala Val Leu Met Lys Arg Gln Lys Met Ala Val Phe	545	550	555	560
30	Leu Trp Gln Arg Gly Glu Glu Ser Met Ala Lys Ala Leu Val Ala Cys	565	570	575	
	Lys Leu Tyr Lys Ala Met Ala His Glu Ser Ser Glu Ser Asp Leu Val	580	585	590	
35	Asp Asp Ile Ser Gln Asp Leu Asp Asn Asn Ser Lys Asp Phe Gly Gln	595	600	605	
	Leu Ala Leu Glu Leu Leu Asp Gln Ser Tyr Lys His Asp Glu Gln Ile	610	615	620	
40	Ala Met Lys Leu Leu Thr Tyr Glu Leu Lys Asn Trp Ser Asn Ser Thr	625	630	635	640
	Cys Leu Lys Leu Ala Val Ala Ala Lys His Arg Asp Phe Ile Ala His	645	650	655	
45	Thr Cys Ser Gln Met Leu Leu Thr Asp Met Trp Met Gly Arg Leu Arg	660	665	670	
50	Met Arg Lys Asn Pro Gly Leu Lys Val Ile Met Gly Ile Leu Leu Pro	675	680	685	
	Pro Thr Ile Leu Phe Leu Glu Phe Arg Thr Tyr Asp Asp Phe Ser Tyr	690	695	700	

55

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	Gln Thr Ser Lys Glu Asn Glu Asp Gly Lys Glu Lys Glu Glu Glu Asn	
	705	710 715 720
5	Thr Asp Ala Asn Ala Asp Ala Gly Ser Arg Lys Gly Asp Glu Glu Asn	
		725 730 735
	Glu His Lys Lys Gln Arg Ser Ile Pro Ile Gly Thr Lys Ile Cys Glu	
		740 745 750
10	Phe Tyr Asn Ala Pro Ile Val Lys Phe	
		755 760
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20	Ser Asn Gln Val Trp Lys Phe Gln Arg Tyr Gln Leu Ile Met Thr Phe	
	1	5 10 15
	His Asp Arg Pro Val Leu Pro Pro Pro Met Ile Ile Leu Ser His Ile	
		20 25 30
25	Tyr Ile Ile Ile Met Arg Leu Ser Gly Arg Cys Arg Lys Lys Arg Glu	
		35 40 45
	Gly Asp Gln Glu Glu Arg Asp Arg Gly Leu Lys Leu Phe Leu Ser Asp	
		50 55 60
30	Glu Glu Leu Lys Arg Leu His Glu Phe Glu Glu Gln Cys Val Gln Glu	
		65 70 75 80
	His Phe Arg Glu Lys Glu Asp Glu Gln Gln Ser Ser Ser Asp Glu Arg	
		85 90 95
35	Ile Arg Val Thr Ser Glu Arg Val Glu Asn Met Ser Met Arg Leu Glu	
		100 105 110
	Glu Ile Asn Glu Arg Glu Thr Phe Met Lys Thr Ser Leu Gln Thr Val	
		115 120 125
40	Asp Leu Arg Leu Ala Gln Leu Glu Glu Leu Ser Asn Arg Met Val Asn	
		130 135 140
	Ala Leu Glu Asn Leu Ala Gly Ile Asp Arg Ser Asp Leu Ile Gln Ala	
		145 150 155 160
45	Arg Ser Arg Ala Ser Ser Glu Cys Glu Ala Thr Tyr Leu Leu Arg Gln	
		165 170 175
	Ser Ser Ile Asn Ser Ala Asp Gly Tyr Ser Leu Tyr Arg Tyr His Phe	
50		180 185 190
	Asn Gly Glu Glu Leu Leu Phe Glu Asp Thr Ser Leu Ser Thr Ser Pro	
		195 200 205
55	Gly Thr Gly Val Arg Lys Lys Thr Cys Ser Phe Arg Ile Lys Glu Glu	
		210 215 220

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Lys Asp Val Lys Thr His Leu Val Pro Glu Cys Gln Asn Ser Leu His  
 225 230 235 240  
 5 Leu Ser Leu Gly Thr Ser Thr Ser Ala Thr Pro Asp Gly Ser His Leu  
 245 250 255  
 Ala Val Asp Asp Leu Lys Asn Ala Glu Glu Ser Lys Leu Gly Pro Asp  
 260 265 270  
 10 Ile Gly Ile Ser Lys Glu Asp Asp Glu Arg Gln Thr Asp Ser Lys Lys  
 275 280 285  
 Glu Glu Thr Ile Ser Pro Ser Leu Asn Lys Thr Asp Val Ile His Gly  
 290 295 300  
 15 Gln Asp Lys Ser Asp Val Gln Asn Thr Gln Leu Thr Val Glu Thr Thr  
 305 310 315 320  
 Asn Ile Glu Gly Thr Ile Ser Tyr Pro Leu Glu Glu Thr Lys Ile Thr  
 325 330 335  
 20 Arg Tyr Phe Pro Asp Glu Thr Ile Asn Ala Cys Lys Thr Met Lys Ser  
 340 345 350  
 Arg Ser Phe Val Tyr Ser Arg Gly Arg Lys Leu Val Gly Gly Val Asn  
 355 360 365  
 25 Gln Asp Val Glu Tyr Ser Ser Ile Thr Asp Gln Gln Leu Thr Thr Glu  
 370 375 380  
 Trp Gln Cys Gln Val Gln Lys Ile Thr Arg Ser His Ser Thr Asp Ile  
 385 390 395 400  
 Pro Tyr Ile Val Ser Glu Ala Ala Val Gln Ala Glu Gln Lys Glu Gln  
 405 410 415  
 35 Phe Ala Asp Met Gln Asp Glu His His Val Ala Glu Ala Ile Pro Arg  
 420 425 430  
 Ile Pro Arg Leu Ser Leu Thr Ile Thr Asp Arg Asn Gly Met Glu Asn  
 435 440 445  
 40 Leu Leu Ser Val Lys Pro Asp Gln Thr Leu Gly Phe Pro Ser Leu Arg  
 450 455 460  
 Ser Lys Ser Leu His Gly His Pro Arg Asn Val Lys Ser Ile Gln Gly  
 465 470 475 480  
 45 Lys Leu Asp Arg Ser Gly His Ala Ser Ser Val Ser Ser Leu Val Ile  
 485 490 495  
 Val Ser Gly Met Thr Ala Glu Glu Lys Lys Val Lys Lys Glu Lys Ala  
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 50 Ser Thr Glu Thr Glu Cys  
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	Ile Ile Leu Ser His Ile Tyr Ile Ile Ile Met Arg Leu Ser Gly Arg
	35 40 45
	Cys Arg Lys Lys Arg Glu Gly Asp Gln Glu Glu Arg Asp Arg Gly Leu
15	50 55 60
	Lys Leu Phe Leu Ser Asp Glu Glu Leu Lys Arg Leu His Glu Phe Glu
	65 70 75 80
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## Claims

1. The use of any purified or isolated nucleic acid encoding a human Mlsn1 or a sequence complementary thereto for the screening of Mlsn1 modulators, wherein the encoding nucleic acid encodes a polypeptide having the following characteristics:

- (1) the N-terminal region is encoded by a consecutive sequence of at least 75 nucleotides, located 5' with respect to nucleotides encoding the transmembrane and C- terminal regions of Mlsn1 protein,
- (2) a transmembrane region encoding six membrane spanning domains, encoded by a consecutive sequence of at least 150 nucleotides, located 3' with respect to the nucleotides encoding the N-terminal region and 5' with respect to nucleotides encoding the C-terminal region, and
- (3) a C-terminal region encoded by a consecutive sequence of at least 75 nucleotides, located 3' with respect to nucleotides encoding the N-terminal region and transmembrane region.

2. The use of any purified or isolated nucleic acid encoding human Mlsn1 according to claim 1, wherein said nucleic acid has at least 80% nucleotide identity with the nucleotide sequence of SEQ ID N°23 or a sequence complementary thereto.

3. The use of any purified or isolated human Mlsn1 polypeptide or a fragment thereof, for the screening of Mlsn1 modulators, wherein the polypeptide has the following characteristics:

- (1) the N-terminal region comprises a consecutive sequence of at least 25 amino acids, located N-terminal with respect to the transmembrane and C- terminal regions of Mlsn1 protein,
- (2) a transmembrane region forming six membrane spanning domains, comprising a consecutive sequence of at least 50 amino acids, located C-terminal with respect to the N-terminal region and N-terminal with respect to the C-terminal region, and
- (3) a C-terminal region comprising a consecutive sequence of at least 25 amino acids, located C-terminal with respect to the N-terminal and transmembrane regions.

4. The use of any purified or isolated polypeptide encoding human Mlsn1 according to claim 3, wherein said polypeptide has at least 90% amino acid identity with the sequence of SEQ ID N°1 or a fragment thereof.

5. The use of any purified or isolated nucleic acid encoding a human Mlsn1 or a sequence complementary thereto

for the diagnosis of disorders associated with aberrant Icrac function in immune cells, wherein the encoding nucleic acid encodes a polypeptide having the following characteristics:

- (1) the N-terminal region is encoded by a consecutive sequence of at least 75 nucleotides, located 5' with respect to nucleotides encoding the transmembrane and C- terminal regions of Mlsn1 protein,
- (2) a transmembrane region encoding six membrane spanning domains, encoded by a consecutive sequence of at least 150 nucleotides, located 3' with respect to the nucleotides encoding the N-terminal region and 5' with respect to nucleotides encoding the C-terminal region, and
- (3) a C-terminal region encoded by a consecutive sequence of at least 75 nucleotides, located 3' with respect to nucleotides encoding the N-terminal region and transmembrane region.

6. The use of any purified or isolated nucleic acid encoding human Mlsn1 according to claim 5, wherein said nucleic acid has at least 80% nucleotide identity with the nucleotide sequence of SEQ ID N°23 or a sequence complementary thereto.

7. The use of any purified or isolated polypeptide encoding a human Mlsn1 or fragment thereof for the diagnosis of disorders associated with aberrant Icrac function in immune cells, wherein the polypeptide has the following characteristics:

- (1) the N-terminal region comprises a consecutive sequence of at least 25 amino acids, located N-terminal with respect to the transmembrane and C- terminal regions of Mlsn1 protein,
- (2) a transmembrane region forming six membrane spanning domains, comprising a consecutive sequence of at least 50 amino acids, located C-terminal with respect to the N-terminal region and N-terminal with respect to the C-terminal region, and
- (3) a C-terminal region comprising a consecutive sequence of at least 25 amino acids, located C-terminal with respect to the N-terminal and transmembrane regions.

8. The use of any purified or isolated polypeptide encoding human Mlsn1 according to claim 7, wherein said polypeptide has at least 90% amino identity with the sequence of SEQ ID N° 1 or a fragment thereof.

9. A purified or isolated human Mlsn1 N-terminal polypeptide region, wherein the N-terminal region comprises a consecutive sequence of at least 25 amino acids, located N-terminal of the first transmembrane region of the Mlsn1 protein.

10. A purified or isolated human Mlsn1 N-terminal polypeptide region according to claim 9, wherein said N-terminal polypeptide has at least 90% amino acid identity with any of the amino acid sequences SEQ ID N°2 or 3, or fragments thereof.

11. A purified or isolated human Mlsn1 C-terminal polypeptide region, wherein the C-terminal region comprises a consecutive sequence of at least 25 amino acids, located C-terminal of the sixth transmembrane region of the Mlsn1 protein.

12. A purified or isolated human Mlsn1 C-terminal region according to claim 11, wherein said C-terminal polypeptide has at least 90% amino acid identity with any of the polypeptides of amino acid sequences SEQ ID N°4 or 5, or fragments thereof.

13. A purified or isolated human Mlsn1 Intra-cellular loop-1 polypeptide region, wherein the intra-cellular loop-1 region comprises a consecutive sequence of at least 7 amino acids, located C-terminal of the second transmembrane region and N-terminal to the third transmembrane region of the Mlsn1 protein.

14. A purified or isolated human Mlsn1 intracellular loop-1 region according to claim 13, wherein said intra-cellular loop-1 polypeptide has at least 90% amino acid identity with any of the polypeptides of amino acid sequences SEQ ID N°6 or 7, or fragments thereof.

15. A purified or isolated human Mlsn1 intra-cellular loop-2 polypeptide region, wherein the intra-cellular loop-2 polypeptide region comprises a consecutive sequence of at least 7 amino acids, located C-terminal of the fourth transmembrane region and N-terminal of the fifth transmembrane region of the Mlsn1 protein.

16. A purified or isolated human Mlsn1 intra-cellular loop-2 region according to claim 15, wherein said intra-cellular loop-2 region polypeptide has at least 90% amino acid identity with any of the polypeptides of amino acid sequences SEQ ID N°8 or 9, or fragments thereof.
- 5 17. A purified or isolated human Mlsn1 extra-cellular loop-1 polypeptide region, wherein the extra-cellular loop-1 region comprises a consecutive sequence of at least 5 amino acids, located C-terminal of the first transmembrane region and N-terminal of the second transmembrane region of the Mlsn1 protein.
- 10 18. A purified or isolated human Mlsn1 extra-cellular loop-1 region according to claim 17, wherein said extra-cellular loop-1 region polypeptide has at least 90% amino acid identity with any of the polypeptides of amino acid sequences SEQ ID N°10 or 11, or fragments thereof.
- 15 19. A purified or isolated human Mlsn1 extra-cellular loop-2 polypeptide region, wherein the extra-cellular loop-2 region comprises a consecutive sequence of at least 5 amino acids, located C-terminal of the third transmembrane region and N-terminal of the fourth transmembrane region of the Mlsn1 protein.
- 20 20. A purified or isolated human Mlsn1 extra-cellular loop-2 region according to claim 19, wherein said extra-cellular loop-2 region polypeptide has at least 90% amino acid identity with any of the polypeptides of amino acid sequences SEQ ID N°12 or 13, or fragments thereof.
- 25 21. A purified or isolated human Mlsn1 extra-cellular loop-3 polypeptide region, wherein the extra-cellular loop-3 region comprises a consecutive sequence of at least 7 amino acids C-terminal of the fifth transmembrane region and N-terminal of the sixth transmembrane region of the Mlsn1 protein.
- 30 22. A purified or isolated human Mlsn1 extra-cellular loop-3 region according to claim 21, wherein said extra-cellular loop-3 region polypeptide has at least 90% amino acid identity with any of the polypeptides of amino acid sequences SEQ ID N°14, 15, 16 or 17, or fragments thereof.
- 35 23. A purified or isolated human Mlsn1 polypeptide containing both intra- and extra-cellular loop regions, wherein said intra- and extra-cellular loop region comprises a consecutive sequence of at least 50 amino acids, located C-terminal to the N-terminal region and N-terminal to the C-terminal region of the Mlsn1 protein.
- 40 24. A purified or isolated human Mlsn1 peptide containing both intra- and extra-cellular loop regions according to claim 23, wherein said intra- and extra-cellular loop region polypeptide has at least 90% amino acid identity with any of the polypeptides of amino acid sequences SEQ ID N°18 or 19, or fragments thereof.
- 45 25. A purified or isolated nucleic acid encoding the human Mlsn1 N-terminal region, wherein the N-terminal region is encoded by a consecutive sequence of at least 75 nucleotides, located 5' with respect to nucleotides encoding the first transmembrane region of the Mlsn1 protein.
- 50 26. A purified or isolated nucleic acid encoding the human Mlsn1 N-terminal region according to claim 25, wherein said nucleic acid has at least 80% nucleic acid identity with any of the nucleotide sequences SEQ ID N°24 or 25, or a complementary sequence thereto.
- 55 27. A purified or isolated nucleic acid encoding the human Mlsn1 C-terminal region, wherein the C-terminal region is encoded by a consecutive sequence of at least 75 nucleotides, located 5' with respect to nucleotides encoding the sixth transmembrane region of the Mlsn1 protein.
28. A purified or isolated nucleic acid encoding the human Mlsn1 C-terminal region according to claim 27, wherein said nucleic acid has at least 80% nucleic acid identity with any of the nucleotide sequences of SEQ ID N°26 or 27, or a complementary sequence thereto.
29. A purified or isolated nucleic acid encoding the human Mlsn1 intra-cellular loop-1 region, wherein the intra-cellular loop-1 region is encoded by a consecutive sequence of at least 21 nucleotides, located 3' with respect to nucleotides encoding the second transmembrane region and 5' with respect to nucleotides encoding the third transmembrane region of the Mlsn1 protein.
30. A purified or isolated nucleic acid encoding the human Mlsn1 intra-cellular loop-1 region according to claim 29,

wherein said nucleic acid has at least 80% nucleic acid identity with any of the nucleotide sequences of SEQ ID N°28 or 29, or a complementary sequence thereto.

31. A purified or isolated nucleic acid encoding the human Mlsn1 intra-cellular loop-2 region, wherein the intra-cellular loops-2 region is encoded by a consecutive sequence of at least 21 nucleotides, located 3' with respect to nucleotides encoding the fourth transmembrane region and 5' with respect to nucleotides encoding the fifth transmembrane region of the Mlsn1 protein.

32. A purified or isolated nucleic acid encoding the human Mlsn1 intra-cellular loop-2 region according to claim 31, wherein said nucleic acid has at least 80% nucleic acid identity with any of the nucleotide sequences of SEQ ID N°30 or 31, or a complementary sequence thereto.

33. A purified or isolated nucleic acid encoding the human Mlsn1 extra-cellular loop-1 region, wherein the extra-cellular loop-1 region is encoded by a consecutive sequence of at least 15 nucleotides, located 3' with respect to nucleotides encoding the first transmembrane region and 5' with respect to nucleotides encoding the second transmembrane region of the Mlsn1 protein.

34. A purified or isolated nucleic acid encoding the human Mlsn1 extra-cellular loop-1 region according to claim 33, wherein said nucleic acid has at least 80% nucleic acid identity with any of the nucleotide sequences of SEQ ID N°32 or 33, or a complementary sequence thereto.

35. A purified or isolated nucleic acid encoding the human Mlsn1 extra-cellular loop-2 region, wherein the extra-cellular loop-2 region is encoded by a consecutive sequence of at least 15 nucleotides, located 3' with respect to nucleotides encoding the third transmembrane region and 5' with respect to nucleotides encoding the fourth transmembrane region of the Mlsn1 protein.

36. A purified or isolated nucleic acid encoding the human Mlsn1 extra-cellular loop-2 region according to claim 35, wherein said nucleic acid has at least 80% nucleic acid identity with any of the nucleotide sequences of SEQ ID N°34 or 35, or a complementary sequence thereto.

37. A purified or isolated nucleic acid encoding the human Mlsn1 extra-cellular loop-3 region, wherein the extra-cellular loop-3 region is encoded by a consecutive sequence of at least 21 nucleotides, located 3' with respect to nucleotides encoding the fifth transmembrane region and 5' with respect to nucleotides encoding the sixth transmembrane region of the Mlsn1 protein.

38. A purified or isolated nucleic acid encoding the human Mlsn1 extra-cellular loop-3 region according to claim 37, wherein said nucleic acid has at least 80% nucleic acid identity with any of the nucleotide sequences of SEQ ID N°36, 37 or 38, or a complementary sequence thereto.

39. A purified or isolated nucleic acid encoding a human Mlsn1 polypeptide containing both intra- and extra-cellular loop regions, wherein said intra- and extra-cellular loop region is encoded by a consecutive sequence of at least 150 nucleotides, located 3' with respect to nucleotides encoding the N-terminal region and 5' with respect to nucleotides encoding the C-terminal regions of the Mlsn1 protein.

40. A purified or isolated nucleic acid encoding the human Mlsn1 polypeptide containing both intra- and extra-cellular loop regions according to claim 39, wherein said nucleic acid has at least 80% nucleic acid identity with any of the nucleotide sequences of SEQ ID N°39 or 40, or a complementary sequence thereto.

41. A nucleic acid sequence encoding an Mlsn1 transcription promoter of SEQ ID N°41 or a complementary sequence thereto.

42. A nucleic acid sequence of claim 41, having at least 80% nucleotide identity with the nucleotide sequence of SEQ ID N°41 or a complementary sequence thereto.

43. A nucleic acid sequence encoding an Mlsn1 transcription promoter of SEQ ID N°42 or a complementary sequence thereto.

44. A nucleic acid sequence of claim 43, having at least 80% nucleotide identity with the nucleotide sequence of SEQ

ID N°42 or a complementary sequence thereto.

45. A nucleic acid sequence encoding an Mlsn1 transcription promoter of SEQ ID N°43 or a complementary sequence thereto.

46. A nucleic acid sequence of claim 45, having at least 80% nucleotide identity with the nucleotide sequence of SEQ ID N°43 or a complementary sequence thereto.

47. A recombinant vector comprising a nucleic acid according to any one of the claims 25 to 46.

48. A recombinant vector comprising a nucleic acid encoding an Mlsn1 polypeptide according to any one of the claims 9 to 24

49. A recombinant host cell comprising a nucleic acid according to any one of the claims 25 to 46 or a vector of claim 47 or 48.

50. A recombinant host cell comprising a nucleic acid encoding an Mlsn1 polypeptide according to any one of the claims 9 to 24.

51. A method for producing a Mlsn1 polypeptide, wherein the said method comprises the following steps of:

- a) culturing, in an appropriate culture medium, a host cell previously transformed or transfected with a polynucleotide according to any one of claims 25 to 48,
- b) harvesting the culture medium thus conditioned or lyse the host cell, for example by sonication or by osmotic shock ; and
- c) separating or purifying, from said culture medium, or from the pellet of the resulting cell lysate, the produced Mlsn1 polypeptide of interest.

52. A method for screening ligand substances or molecules that are able to bind to a Mlsn1, or fragment thereof, or Mlsn1 transcription promoter elements of any one of the claims 1 to 46, said method comprising ;

- a) contacting a test compound with Mlsn1 gene, an Mlsn1 gene product or Mlsn1 transcription promoter element and,
- b) measuring the ability of said test compound to interact with the Mlsn1 gene, Mlsn1 gene product or transcription promoter element (i.e., binding assay).

53. A method for screening ligand substances or molecules that are able to modulate the biological activity of Mlsn1, or fragment thereof, of any one of the claims 1 to 40, said method comprising ;

- a) contacting a test compound with an Mlsn1 expressing cell, and,
- b) determining the ability of said test compound to affect Mlsn1 gene or an Mlsn1 gene product activity within the cells.

54. A method for screening ligand substances or molecules that are able to modulate the biological activity of Mlsn1, or fragment thereof, of any one of the claims 1 to 40, said method comprising ;

- a) obtaining a recombinant cell expressing Mlsn1 gene or gene product,
- b) exposing said recombinant cell to a substance or molecule to be tested ; and
- c) Measuring the change in calcium flux or activation potential within the exposed recombinant cell.

55. A method for screening ligand substances or molecules that are able to modulate the biological activity of Mlsn1, or fragment thereof, of any one of the claims 41 to 46, said method comprising ;

- a) obtaining a recombinant cell expressing a reporter molecule under the control of an Mlsn1 transcription promoterMlsn1 gene,
- b) Exposing said recombinant cell to a substance or molecule to be tested ; and

c) Measuring the change in reporter expression or activity in said recombinant cell.

56. A method for screening ligand substances or molecules that are able to bind to a Mlsn1, or modulate the biological activity of Mlsn1, according to any one of claims 51 to 55, wherein a competitor Mlsn1 binding ligand, preferably selected from the class of pyrazol compounds such as SEW04225, KM02940, KM03000 or GK02421, is added and the binding of said competitor in the presence and/or absence of a test compound is determined.

57. A method for detecting any one of the nucleic acids of claims 6, 8, or 25 to 50, in a blood sample, said method comprising ;

- a) obtaining a blood sample,
- b) optionally separating immune cells from other blood components,
- c) reverse transcribing cellular RNA to cDNA, and
- d) amplifying Mlsn1 nucleic acid obtained in step c) by PCR with a plurality of Mlsn1 specific oligonucleotides capable of hybridizing, under stringent conditions to Mlsn1 nucleic acids and characterize the PCR product.

58. The use of a Mlsn1 gene, gene product or transcriptional element, to screen compounds that modulate the activity of an antigen presenting cell, particularly of macrophages, in particular macrophage maturation.

59. The use of a compound that modulates the activity of Mlsn1 gene, gene product or transcriptional element, for the manufacture of a composition to regulate the activity of an antigen presenting cell, particularly of macrophages, in particular macrophage maturation.

60. An antisense nucleic acid molecule comprising a sequence region which is complementary to at least a region of a Mlsn1 nucleic acid of claims 25 to 40.

61. The use of an active compound as identified, selected or **characterized by** the screening method described in claims 52 to 56 for the preparation of pharmaceutical compounds or compositions that bind and/or modulate the biological activity of Mlsn1.

[illegible]

### FIGURE 1

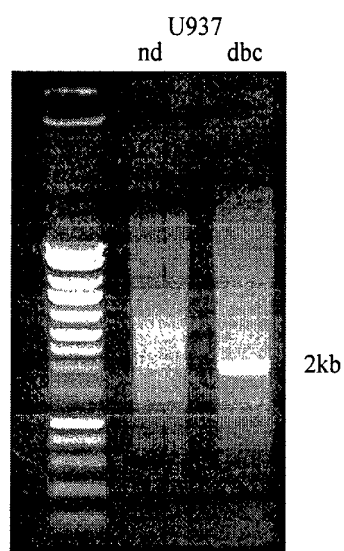


Figure 2

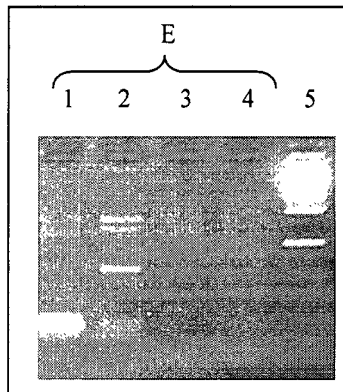
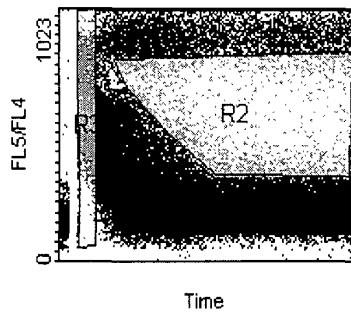
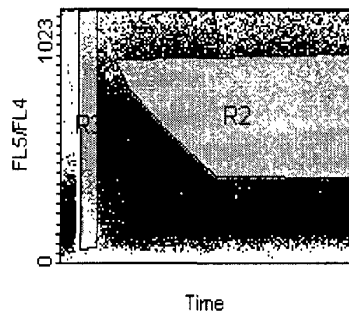


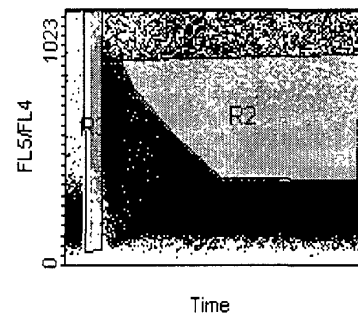
Figure 3

**Wild U937**

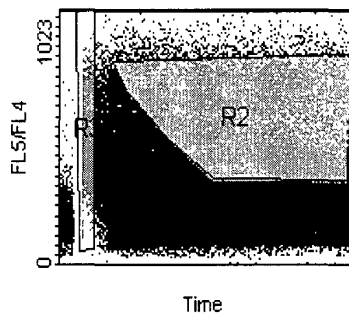
File: U937+IONO.002 Sample: U937+IONO  
 Date: 24-Jan-00  
 Total Events 106920 Gated Events 106920 100.00%  
 System: Log Parameter Means: Arithmetic  
 Time (8) vs FL5/FL4 (7)  
 Region X-Mean Y-Mean Events %Gated  
 R1 487.6 324.6 94171 88.08  
 R2 633.2 569.3 3203 3.00  
 R3 96.3 629.7 6979 6.53

**U937 DBC treated**

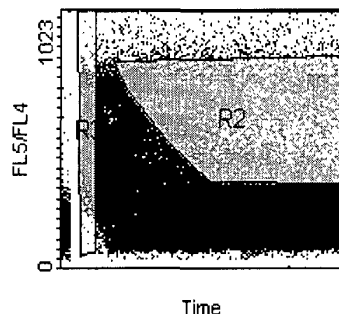
File: U937+dbc+IONO.003 Sample: U937+dbc+IONO  
 Date: 24-Jan-00  
 Total Events 128040 Gated Events 128040 100.00%  
 System: Log Parameter Means: Arithmetic  
 Time (8) vs FL5/FL4 (7)  
 Region X-Mean Y-Mean Events %Gated  
 R1 383.2 372.1 105000 82.01  
 R2 670.6 561.7 37006 28.90  
 R3 102.5 630.5 7369 5.76

**Anti-sens treatment conditions**

File: U937+dbc+a-sens Sample: U937+dbc+a-sens conditions  
 Date: 24-Jan-00  
 Total Events 130080 Gated Events 130080 100.00%  
 System: Log Parameter Means: Arithmetic  
 Time (8) vs FL5/FL4 (7)  
 Region X-Mean Y-Mean Events %Gated  
 R1 440.8 417.2 106547 81.91  
 R2 685.6 512.4 30202 23.22  
 R3 109.6 694.0 6435 4.95

**MLSN anti-sens 1μM**

File: U937+dbc+a-sens Sample: U937+dbc+a-sens  
 MLSN 1  
 Date: 24-Jan-00  
 Total Events 130665 Gated Events 130665 100.00%  
 System: Log Parameter Means: Arithmetic  
 Time (8) vs FL5/FL4 (7)  
 Region X-Mean Y-Mean Events %Gated  
 R1 420.5 312.3 100935 77.25  
 R2 634.6 567.9 22038 16.87  
 R3 106.3 599.4 5954 4.56

**MLSN anti-sens 10μM**

File: U937+dbc+a-sens Sample: U937+dbc+a-sens MLSN  
 10  
 Date: 24-Jan-00  
 Total Events 121995 Gated Events 121995 100.00%  
 System: Log Parameter Means: Arithmetic  
 Time (8) vs FL5/FL4 (7)  
 Region X-Mean Y-Mean Events %Gated  
 R1 478.4 358.6 91138 74.71  
 R2 595.9 602.9 17180 14.08  
 R3 99.7 605.8 7560 6.20



European Patent  
Office

# EUROPEAN SEARCH REPORT

Application Number  
EP 01 40 2283

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Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.Cl.7)
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X	HUNTER JOHN J ET AL: "Chromosomal localization and genomic characterization of the mouse melastatin gene (Mlnsl)." GENOMICS, vol. 54, no. 1, 15 November 1998 (1998-11-15), pages 116-123, XP000910696 ISSN: 0888-7543	9-40, 47-51	
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The present search report has been drawn up for all claims			
Place of search <b>MUNICH</b>		Date of completion of the search <b>10 January 2002</b>	Examiner <b>Morawetz, R</b>
CATEGORY OF CITED DOCUMENTS X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document			

EPO FORM 1503 03.82 (P04C01)



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## EUROPEAN SEARCH REPORT

Application Number  
EP 01 40 2283

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Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.Cl.7)
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The present search report has been drawn up for all claims			
Place of search <b>MUNICH</b>		Date of completion of the search <b>10 January 2002</b>	Examiner <b>Morawetz, R</b>
CATEGORY OF CITED DOCUMENTS X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document		T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document	

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European Patent  
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# EUROPEAN SEARCH REPORT

Application Number  
EP 01 40 2283

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.Cl.7)
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			TECHNICAL FIELDS SEARCHED (Int.Cl.7)
The present search report has been drawn up for all claims			
Place of search <b>MUNICH</b>		Date of completion of the search <b>10 January 2002</b>	Examiner <b>Morawetz, R</b>
<p><b>CATEGORY OF CITED DOCUMENTS</b></p> <p>X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document</p> <p>T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons &amp; : member of the same patent family, corresponding document</p>			

EPO FORM 1503 03.82 (P44C01)

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ON EUROPEAN PATENT APPLICATION NO.**

EP 01 40 2283

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The members are as contained in the European Patent Office EDP file on  
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10-01-2002

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EPO FORM P4459

For more details about this annex : see Official Journal of the European Patent Office, No. 12/82

专利名称(译)	鉴定抗原呈递细胞中的容量钙通道及其用途		
公开(公告)号	<a href="#">EP1184457A1</a>	公开(公告)日	2002-03-06
申请号	EP2001402283	申请日	2001-09-03
[标]申请(专利权)人(译)	华纳兰茂公司		
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当前申请(专利权)人(译)	华纳 - 兰伯特公司有限责任公司		
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IPC分类号	C07K14/705 C12N15/12 C12Q1/68 G01N33/48 G01N33/53 A61K38/00		
CPC分类号	C07K14/705		
代理机构(译)	贝克尔, PHILIPPE		
优先权	2000402436 2000-09-05 EP		
外部链接	<a href="#">Espacenet</a>		

#### 摘要(译)

本发明涉及用于免疫系统活化的电容性钙通道同源物的鉴定, 以及其用于报告和调节体外, 离体或体内调节免疫细胞活性的用途。本发明更具体地公开了Mlsn1基因产物, 其代表免疫细胞如巨噬细胞, 单核细胞, T细胞, B细胞和肥大细胞中的容量钙通道。本发明是证明在免疫细胞中表达电容性钙通道的基因的鉴定, 并且可以用于监测或调节受试者中免疫应答的各种组合物和方法中。本发明可用于开发用于免疫系统激活或炎症反应的生物标志物或筛选改变药物的特定免疫系统活性。

Primer 1a (SEQ ID N°4): TCAGTAAATCAATCTCAACAGG