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(54) Title: P2Y<sub>12</sub> RECEPTOR

(57) Abstract: The present invention includes a novel subtype of the P2-purinergic receptor, referred to as the P2Y<sub>12</sub> receptor. This receptor is expressed selectively in the platelets and brain, and couples to a pertussis toxin-sensitive G protein (Gi). Nucleic acids encoding the receptor and associated screening and therapeutic methods are also disclosed.

**P2Y<sub>12</sub> RECEPTOR**

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**CROSS-REFERENCES TO RELATED APPLICATIONS**

This application is related to Provisional Application 60/171,622, entitled Novel ADP Receptor, filed December 23, 1999, which is herein incorporated by reference in its entirety.

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**FIELD OF THE INVENTION**

The present invention relates to methods employing an ADP receptor, termed the P2Y<sub>12</sub> receptor, and its encoding nucleic acid molecules. The invention also relates to methods for the recombinant production of the receptor proteins and the proteins made by these methods, antibodies against the whole receptor or regions thereof, vectors, nucleotide probes, host cells transformed by genes encoding polypeptides having the receptor activity, methods for the discovery of novel modulators of receptor activity, along with diagnostic and therapeutic uses for these various agents.

15  
20**BACKGROUND OF THE INVENTION**

Thrombotic complications are a major cause of death in the industrialized world. Examples of these complications include acute myocardial infarction, unstable angina, chronic stable angina, transient ischemic attacks, strokes, peripheral vascular disease, preeclampsia, deep venous thrombosis, embolism, disseminated intravascular coagulation and thrombotic thrombocytopenic purpura. Thrombotic and restenotic complications also occur following invasive procedures, *e.g.*, angioplasty, carotid endarterectomy, post CABG (coronary artery bypass graft) surgery, vascular graft surgery, stent placements and insertion of endovascular devices and prostheses.

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It is generally thought that platelet aggregates play a critical role in these events. Blood platelets, which normally circulate freely in the vasculature, become activated and aggregate to form a thrombus with disturbed blood flow caused by ruptured atherosclerotic lesions or by invasive treatments such as angioplasty, resulting in vascular occlusion. Platelet activation can be initiated by a variety of agents, e.g., exposed subendothelial matrix molecules such as collagen, or by thrombin which is formed in the coagulation cascade.

An important mediator of platelet activation and aggregation is ADP (adenosine 5'-diphosphate) which is released from blood platelets in the vasculature upon activation by various agents, such as collagen and thrombin, and from damaged blood cells, endothelium or tissues. Activation of platelets by ADP results in the recruitment of more platelets and stabilization of existing platelet aggregates. Platelet ADP receptors mediating aggregation are activated by ADP and some of its derivatives and antagonized by ATP (adenosine 5'-triphosphate) and some of its derivatives (Mills, D.C.B. (1996) *Thromb. Hemost.* 76:835-56). Therefore, platelet ADP receptors are members of the family of P2 receptors activated by purine and/or pyrimidine nucleotides (King, B.F., Townsend-Nicholson, A. & Burnstock, G. (1998) *Trends Pharmacol. Sci.* 19:506-514). ADP receptors additionally have been characterized as belonging to the G protein-coupled receptor superfamily.

Studies of inherited disorders in humans and rats which result in a reduction of ADP release from platelets or reduced ADP receptor number and signaling confirm the critical role in platelet aggregation of ADP and the ADP receptors (Cattaneo M. & Gachet C., *Arterioscler. Thromb. Vasc. Biol.* (1999) 19:2281-2285). Inhibitors of ADP-induced platelet aggregation are efficacious antithrombotic drugs.

Recent pharmacological data using selective antagonists suggests that ADP-dependent platelet aggregation requires activation of at least two ADP receptors (Kunapuli, S.P. (1998) *TIPS* 19:391-394; Kunapuli S.P. et al., (1998) *Biochem J.* 336:513-523; Jantzen, H.M. et al., (1999) *Thromb. Haemost.* 81:111-117). One receptor appears to be identical to the cloned P2Y<sub>1</sub> receptor, is coupled to the G protein G<sub>q</sub> and mediates intracellular calcium mobilization. This receptor is also required for ADP-induced platelet shape change.

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The second platelet ADP receptor couples to inhibition of adenylyl cyclase. The gene or cDNAs for this receptor has not been identified previously and the receptor has been provisionally termed P2Y<sub>ADP</sub> (Fredholm B.B. et al., (1997) *TIPS* 18:79-82), P2T<sub>AC</sub>, (Kunapuli, S (1998) *TIPS* 19(10):391-394), and P2Y<sub>cyc</sub> based on  
5 its pharmacological and signaling properties. Based on the inhibition of adenylyl cyclase and the activation of the G protein G<sub>o</sub>2 by ADP in platelet membranes it was thought that this receptor may couple to G<sub>i</sub> *in vivo*, and may belong to the G-protein coupled receptor family. This receptor also appears to be the target of the orally active antithrombotic drugs ticlopidine and clopidogrel, which appear to act through  
10 unstable and irreversible acting liver metabolites (Quinn, M.J. & Fitzgerald, D.J. (1999) *Circulation* 100:1667-1672 ). Patients with mild bleeding disorders have been identified with functional defects in this receptor (Cattaneo M. & Gachet C., *Arterioscler. Thromb. Vasc. Biol.* (1999) 19:2281-2285). The molecular identity of the G<sub>i</sub>-linked receptor has remained elusive, even though it is the target of efficacious  
15 antithrombotic agents such as ticlopidine and clopidogrel (Gachet *et al.* (1990)). A sequence corresponding to this G<sub>i</sub>-linked receptor was previously published in WO 98/50549, but its function was not identified.

Despite the previous physiological identification of a second ADP receptor and the discovery of agents that may act at this receptor, there exists a need for  
20 platelet ADP receptor inhibitors with improved properties.

### SUMMARY OF THE INVENTION

The present inventors have cloned the G<sub>i</sub>-linked receptor, designated P2Y<sub>12</sub>, and show that a patient with a bleeding disorder (Nurden *et al.* (1995)) is defective in  
25 this gene. Cloning of the P2Y<sub>12</sub> receptor facilitates the development of better antiplatelet agents to treat a variety of cardiovascular diseases.

The invention includes an isolated nucleic acid molecule selected from the group consisting of: (a) an isolated nucleic acid molecule that encodes the amino acid sequence of SEQ ID NO: 2, an isolated nucleic acid molecule that encodes the amino  
30 acid sequence of SEQ ID NO: 12 (b) an isolated nucleic acid molecule which hybridizes to the complement of a nucleic acid molecule comprising SEQ ID NO: 1

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under conditions of sufficient stringency to produce a clear signal; and (c) an isolated nucleic acid molecule which hybridizes to a nucleic acid molecule that encodes the amino acid sequence of SEQ ID NO: 2 under conditions of sufficient stringency to produce a clear signal.

5           The present invention further includes isolated nucleic acid molecules wherein the nucleic acid molecule comprises the sequence of SEQ ID NO: 1, isolated nucleic acid molecules that comprise nucleotides 130-1158 of SEQ ID NO: 1 or nucleotides 130-1161 of SEQ ID NO: 1; isolated nucleic acid molecules that comprise the sequence of SEQ ID NO: 11, nucleic acid molecules that comprise nucleotides 73-873  
10 of SEQ ID NO: 11 or nucleotides 73-876 of SEQ ID NO: 11.

The present invention further includes the nucleic acids operably linked to one or more expression control elements, including vectors comprising the isolated nucleic acid molecules. The invention further includes host cells transformed to contain the nucleic acids of the invention, wherein said host is selected selected from  
15 the group consisting of prokaryotic hosts and eukaryotic hosts and methods for producing a protein comprising the step of culturing a host cell under conditions in which the protein encoded by said nucleic acid molecule is expressed.

The invention further provides a polypeptide, which may be expressed in a recombinant cell or may be purified, selected from the group consisting of an isolated  
20 receptor polypeptide comprising the amino acid sequence of SEQ ID NO: 2, an isolated receptor polypeptide comprising the amino acid sequence of SEQ ID NO: 12.

The invention further provides an isolated antibody that binds to a polypeptide of the invention, including monoclonal and polyclonal antibodies.

The invention further provides a method of identifying an agent which  
25 modulates the expression of a nucleic acid encoding a P2Y<sub>12</sub> receptor comprising the steps of exposing cells which express the nucleic acid to the agent and determining whether the agent modulates expression of said nucleic acid, thereby identifying an agent which modulates the expression of a nucleic acid encoding the protein having the sequence of P2Y<sub>12</sub> receptor.

30           The invention further provides a method of identifying an agent which modulates at least one activity of a P2Y<sub>12</sub> receptor comprising the steps of exposing

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cells which express the protein to the agent; determining whether the agent modulates at least one activity of said protein, thereby identifying an agent which modulates at least one activity of a P2Y<sub>12</sub> receptor.

Activities the agent may modify include but are not limited to the interaction  
5 between ADP, ATP or a derivative thereof, and a P2Y<sub>12</sub> receptor; potassium current or adenylyl cyclase activity, platelet activation or thrombotic activity; acute myocardial infarction, unstable angina, chronic stable angina, transient ischemic attacks, strokes, peripheral vascular disease, preeclampsia, deep venous thrombosis, embolism, disseminated intravascular coagulation, thrombotic thrombocytopenic  
10 purpura or a bleeding disorder; thrombotic and restenotic complications following angioplasty, carotid endarterectomy, post CABG (coronary artery bypass graft) surgery, vascular graft surgery, stent placements or insertion of endovascular devices and prostheses.

The invention further provides a method of identifying binding partners for a  
15 P2Y<sub>12</sub> receptor protein comprising the steps of exposing said protein to a potential binding partner and determining if the potential binding partner binds to said protein, thereby identifying binding partners for a P2Y<sub>12</sub> receptor protein.

The invention further provides a method of modulating the expression of a  
20 nucleic acid encoding a P2Y<sub>12</sub> receptor protein comprising the step of administering an effective amount of an agent which modulates the expression of a nucleic acid encoding encoding a P2Y<sub>12</sub> receptor protein.

The invention further provides a method of modulating at least one activity of  
a P2Y<sub>12</sub> receptor protein comprising the step of administering an effective amount of  
an agent which modulates at least one activity of a P2Y<sub>12</sub> receptor protein.

25 The invention further provides a non-human transgenic animal modified to contain a nucleic acid molecule of the invention including a nucleic acid molecule which encode a P2Y<sub>12</sub> receptor or a truncation mutant of P2Y<sub>12</sub> receptor.

The invention further provides a method of diagnosing a disease state in a  
subject, comprising the step of determining the level of expression of a nucleic acid  
30 molecule encoding a P2Y<sub>12</sub> receptor.

The invention further provides a method of diagnosing a disease state in a subject, comprising the step of determining the level of expression of a P2Y<sub>12</sub> receptor protein.

5 The invention further provides an isolated mutant P2Y<sub>12</sub> receptor protein which is a naturally occurring truncation mutant, wherein said truncation is caused by a frame-shift mutation in the region of the gene encoding transmembrane domain six of the protein.

10 The invention further provides an isolated mutant P2Y<sub>12</sub> receptor protein which is a naturally occurring truncation mutant, wherein said truncation is caused by a frame-shift mutation in the region of the gene encoding transmembrane domain six of the protein wherein said protein is associated with a bleeding disorder.

15 The invention further provides an isolated mutant P2Y<sub>12</sub> receptor protein which is a naturally occurring truncation mutant, wherein said truncation is caused by a frame-shift mutation in the region of the gene encoding transmembrane domain six of the protein wherein said frame shift mutation introduces a stop codon in the gene encoding the P2Y<sub>12</sub> protein and results in platelets with impaired ADP-dependent platelet aggregation activity, reduced ADP binding activity and reduced ability to inhibit cAMP levels in response to ADP.

20 The invention further provides an isolated nucleic acid encoding a mutant P2Y<sub>12</sub> receptor protein which is a naturally occurring truncation mutant, wherein said truncation is caused by a frame-shift mutation in the region of the gene encoding transmembrane domain six of the protein.

25 The invention further provides an isolated nucleic acid encoding a mutant P2Y<sub>12</sub> receptor protein which is a naturally occurring truncation mutant, wherein said truncation is caused by a frame-shift mutation in the region of the gene encoding transmembrane domain six of the protein wherein said nucleic acid encodes a protein associated with a bleeding disorder.

30 The invention further provides an isolated nucleic acid encoding a mutant P2Y<sub>12</sub> receptor protein which is a naturally occurring truncation mutant, wherein said truncation is caused by a frame-shift mutation in the region of the gene encoding transmembrane domain six of the protein wherein said frame shift mutation

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introduces a stop codon in the gene encoding the P2Y<sub>12</sub> protein and results in platelets with impaired ADP-dependent platelet aggregation activity, reduced ADP binding activity and reduced ability to inhibit cAMP levels in response to ADP.

5 The invention further provides a method of diagnosing a disease state in a subject, comprising the step of determining the level of expression of a nucleic acid molecule of the invention.

The invention further provides a method of diagnosing a disease state in a subject, comprising the step of determining the level of expression of a protein of the invention.

10 The invention further provides a method of diagnosing a disease state in a subject, comprising the step of identifying a nucleic acid molecule of the invention.

#### BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1A-1C P2Y<sub>12</sub> is a G protein-coupled receptor that responds to ADP.

15 Figure 1(A): Activation of potassium-dependent currents in *Xenopus* oocytes expressing P2Y<sub>12</sub> with Kir3.1 and 3.4. Figure 1(B): ADP-selective stimulation of potassium channel-dependent currents by P2Y<sub>12</sub> occurs via a pertussis toxin-sensitive pathway. Figure 1(C): The agonist profile of P2Y<sub>12</sub> recapitulates that observed for the G<sub>i</sub>-coupled platelet ADP receptor.

20

Figure 2A-2C Currents stimulated by ADP in oocytes expressing hP2Y<sub>12</sub> with Kir3.1 and 3.4 are inhibited by 2MeSAMP, C1330-7 and a thiol reagent. Figure 2(A): Current tracing showing reversible block of ADP (1 μM) responses by 2MeSAMP (10 μM) and C1330-7 (1 μM), but not A3P5P (300 μM). Figure 2(B): 2MeSAMP and C1330-7 inhibition curves. Figure 2(C): Selective ablation of P2Y<sub>12</sub> but not m2 muscarinic receptor signaling by the thiol reagent pCMBS.

25

Figure 3A-3C Activation of hP2Y<sub>12</sub> in CHO cells inhibits adenylyl cyclase. Figure 3(A): Receptor coupling to adenylyl cyclase. Figure 3(B): The effect of the

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specific antagonists 2MeSAMP (2-MES)(50  $\mu$ M) and C1330-7 (50  $\mu$ M). Figure 3(C): Effect of pertussis toxin (PTX) pretreatment.

Figure 4A-4E P2Y<sub>12</sub> receptor is selectively expressed in platelets and brain.

5 Figure 4(A) and (B): Northern analysis of hP2Y<sub>12</sub> transcripts. Figure 4(C): The rP2Y<sub>12</sub> protein is expressed in megakaryocytes and platelets in rat bone marrow. Figure 4(D): FACS analysis of rat platelets stained with rP2Y<sub>12</sub> antisera. Figure 4(E): FACS analysis of rat 2-9 fibroblasts transfected with the rP2Y<sub>12</sub> cDNA clone.

10 Figure 5A-5D A frame-shift mutation within the hP2Y<sub>12</sub> gene is associated with a bleeding disorder. Figure 5(A): Deduced amino acid sequence of the hP2Y<sub>12</sub> protein and alignment with other receptor sequences (SEQ ID NO: 6, SEQ ID NO: 13, SEQ ID NO: 14). Figure 5(B): A P2Y<sub>12</sub> allele from a patient (ML) with defective ADP-dependent aggregation contains a 2 base pair deletion, resulting in a frame-shift  
15 mutation and a premature truncation of the protein (SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 11, SEQ ID NO: 12). Figure 5(C): Mutant hP2Y<sub>12</sub> receptor from patient ML is non-functional and does not act in a dominant-negative capacity. Figure 5(D): Patient ML has abnormally low levels of RT-PCR product derived from P2Y<sub>12</sub> mRNA.

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Figure 6A-6B Evaluation of [<sup>3</sup>H]2MeSADP binding to the cloned hP2Y<sub>12</sub> Receptor. Figure 6(A): [<sup>3</sup>H]2MeSADP binding to membranes of COS7 cells transiently transfected with hP2Y<sub>12</sub>. Figure 6(B): Competition by 2MeSAMP and A3P5P of 1 nM [<sup>3</sup>H]2MeSADP binding to membranes from COS7 cells transfected  
25 with hP2Y<sub>12</sub>.

Figure 7 Representative potassium currents from voltage clamp assays on Xenopus oocytes. Oocytes were co-injected with cRNA for the P2Y<sub>12</sub> receptor and for GIRK1 and GIRK4 potassium channels, cultured for 48-72 hours, and  
30 challenged with different agonists and antagonists.

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Figure 8 Representative potassium currents from voltage clamp assays on Xenopus oocytes. Oocytes were co-injected with cRNA for the P2Y<sub>12</sub> receptor, for pertussis toxin, and for GIRK1 and GIRK4 potassium channels, cultured for 48-72 hours, and challenged with the agonist ADP.

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## DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENT

### I. General Description

The present inventors have characterized a novel cDNA from a platelet library  
5 that encodes the G<sub>i</sub>-linked platelet ADP receptor. Genetic (Leon *et al.* (1999); Fabre *et al.* (1999); Nurden *et al.* (1995); Cattaneo *et al.* (1999)) and pharmacological (Jarvis *et al.* (2000); Hechler *et al.* (1998)) studies demonstrate that the G<sub>i</sub>-linked receptor is critical for formation and stabilization of large platelet aggregates (Humbert *et al.* (1996)). Additionally, the G<sub>i</sub>-linked receptor is the target of the antithrombotic drugs  
10 clopidogrel and ticlopidine, which have been demonstrated to be efficacious in the treatment of a variety of thrombotic diseases (stroke, MI, peripheral vascular disease). However, these drugs work through a mechanism of covalent protein modification, which may underlie their recent association with the syndrome thrombotic  
thrombocytopenic purpura (TTP) (Bennett *et al.* (2000)), an immune-mediated  
15 response. The present studies demonstrate that the P2Y<sub>12</sub> receptor has a selective tissue distribution compared to other purinergic receptors (such as P2Y<sub>1</sub>), making this receptor an extremely attractive target for the development of novel antithrombotics.

The invention includes methods of using this receptor for identification of  
binding partners or for diagnostic applications. Additionally, the proteins of the  
20 invention provide targets for screening synthetic small molecules and combinatorial or naturally occurring compound libraries to discover therapeutics to regulate platelet aggregation, vascular injury or disease as well as schizophrenia, eating disorders, depression, migraine and other brain disorders.

### 25 II. Specific Embodiments

#### A. Protein Molecules

The present invention provides and employs isolated P2Y<sub>12</sub> receptor protein, allelic variants of the protein as well as conservative amino acid substitutions of the  
30 protein. As used herein, the "protein" or "polypeptide" refers in part to a protein that has the amino acid sequence depicted in SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO:

6 or SEQ ID NO: 12. The methods of the invention also employ naturally occurring allelic variants and proteins that have a slightly different amino acid sequence than that specifically recited above. Allelic variants, though possessing a slightly different amino acid sequence than those recited above, will still have the same or similar biological functions associated with the 267, 315, 342 or 343 amino acid protein.

As used herein, the “family of proteins” related to the amino acid sequence of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6 or SEQ ID NO: 12 refers to proteins that have been isolated from organisms in addition to rats and humans. The methods used to identify and isolate other members of the family of proteins related to the 267, 315, 342 or 343 amino acid protein are described below.

The proteins of the present invention may be in isolated form, may be recombinantly expressed so as to be present on the cell surface or may be in partially purified form, such as in cellular membrane preparations. For such preparations, the protein may be found at levels typically higher than normally found without recombinant expression of the protein. As used herein, a protein is said to be isolated when physical, mechanical or chemical methods are employed to remove the protein from cellular constituents that are normally associated with the protein. A skilled artisan can readily employ standard purification methods to obtain an isolated protein.

The proteins used in the methods of the invention further include insertion, deletion or conservative amino acid substitution variants of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6 or SEQ ID NO:12. As used herein, a conservative variant refers to at least one alteration in the amino acid sequence that does not adversely affect the biological functions of the protein. A substitution, insertion or deletion is said to adversely affect the protein when the altered sequence prevents or disrupts a biological function associated with the protein. For example, the overall charge, structure or hydrophobic/hydrophilic properties of the protein can be altered without adversely affecting a biological activity. Such biological activities of the P2Y<sub>12</sub> receptor include but are not limited to the activities described in the Examples discussed below. Accordingly, the amino acid sequence can often be altered, for example to render the peptide more hydrophobic or hydrophilic, without adversely affecting the biological activities of the protein.

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Ordinarily, the allelic variants, the conservative substitution variants, and the members of the protein family, will have an amino acid sequence having at least about 75% amino acid sequence identity with the sequence set forth in SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6 or SEQ ID NO: 12 more preferably at least about 80%,  
5 even more preferably at least about 90%, and most preferably at least about 95% sequence identity to said sequences. Identity or homology with respect to such sequences is defined herein as the percentage of amino acid residues in the candidate sequence that are identical with the known peptides, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent homology, and not  
10 considering any conservative substitutions as part of the sequence identity. A further discussion of the methods of determining sequence identity is provided below. Fusion proteins, or N-terminal, C-terminal or internal extensions, deletions, or insertions into the peptide sequence shall not be construed as affecting homology.

Contemplated variants further include those containing predetermined  
15 mutations by, *e.g.*, homologous recombination, site-directed or PCR mutagenesis, and the corresponding proteins of other animal species, including but not limited to canine, rabbit, mouse, porcine, bovine, ovine, equine and non-human primate species, and the alleles or other naturally occurring variants of the family of proteins; and derivatives wherein the protein has been covalently modified by substitution,  
20 chemical, enzymatic, or other appropriate means with a moiety other than a naturally occurring amino acid (for example a detectable moiety such as an enzyme or radioisotope).

The proteins of the present invention also include molecules having a portion of the amino acid sequence disclosed in SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO:  
25 6 or SEQ ID NO: 12 such as fragments having a consecutive sequence of at least about 3, 4, 5, 6, 10, 15, 20, 25, 30, 35 or more amino acid residues of the protein. Such fragments, also referred to as peptides or polypeptides, may contain antigenic regions, functional regions of the protein identified as regions of the amino acid sequence which correspond to known protein domains, as well as regions of  
30 pronounced hydrophilicity. The regions are all easily identifiable by using commonly

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available protein sequence analysis software such as MacVector™ (Oxford Molecular).

As described below, members of the family of proteins can be used: 1) to identify agents which modulate at least one activity of the P2Y<sub>12</sub> protein; 2) to identify binding partners for the protein, 3) as an antigen to raise polyclonal or monoclonal antibodies, and 4) as a therapeutic agent or target.

### B. Nucleic Acid Molecules

The present invention further provides and utilizes nucleic acid molecules that encode the proteins having SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6 or SEQ ID NO: 12 and the related proteins herein described, preferably in isolated form. As used herein, “nucleic acid” is defined as RNA or DNA that encodes a protein or peptide as defined above, is complementary to a nucleic acid sequence encoding such peptides, hybridizes to such a nucleic acid and remains stably bound to it under appropriate stringency conditions, or encodes a polypeptide sharing at least about 75% sequence identity, preferably at least about 80%, and more preferably at least about 85% sequence identity with the peptide sequences of the invention. Specifically contemplated are genomic DNA, cDNA, mRNA and antisense molecules, as well as nucleic acids based on alternative backbones or including alternative bases whether derived from natural sources or synthesized. Such hybridizing or complementary nucleic acids, however, are defined further as being novel and non-obvious over any prior art nucleic acid.

Homology or identity at the nucleotide or amino acid sequence level is determined by **BLAST** (Basic Local Alignment Search Tool) analysis using the algorithm employed by the programs **blastp**, **blastn**, **blastx**, **tblastn** and **tblastx** (Karlin *et al.* *Proceedings of the National Academy of Science USA*, Vol. 87 (1990) pp. 2264-2268 and Altschul, SF. *Journal of Molecular Evolution*, Vol. 36 (1993) pp. 290-300, herein incorporated by reference in their entirety) which are tailored for sequence similarity searching. The approach used by the **BLAST** program is to first consider similar segments between a query sequence and a database sequence, then to

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evaluate the statistical significance of all matches that are identified and finally to summarize only those matches which satisfy a preselected threshold of significance. For a discussion of basic issues in similarity searching of sequence databases, see Altschul *et al.* (*Nature Genetics*, Vol. 6 (1994) pp. 119-129) which is herein  
5 incorporated by reference in its entirety. The search parameters for **histogram**, **descriptions**, **alignments**, **expect** (*i.e.*, the statistical significance threshold for reporting matches against database sequences), **cutoff**, **matrix** and **filter** are at the default settings. The default scoring matrix used by **blastp**, **blastx**, **tblastn**, and **tblastx** is the **BLOSUM62** matrix (Henikoff *et al.* *Proceedings of the National*  
10 *Academy of Science USA*, Vol. 89 (1992) pp. 10915-10919, herein incorporated by reference in its entirety). For **blastn**, the scoring matrix is set by the ratios of **M** (*i.e.*, the reward score for a pair of matching residues) to **N** (*i.e.*, the penalty score for mismatching residues), wherein the default values for **M** and **N** are 5 and -4, respectively.

15 “Stringent conditions” are those that (1) employ low ionic strength and high temperature for washing, for example, 0.015 M NaCl/0.0015 M sodium titrate/0.1% SDS at 50 C, or (2) employ during hybridization a denaturing agent such as formamide, for example, 50% (vol/vol) formamide with 0.1% bovine serum albumin/0.1% Ficoll/0.1% polyvinylpyrrolidone/50 mM sodium phosphate buffer at  
20 pH 6.5 with 750 mM NaCl, 75 mM sodium citrate at 42 C. Another example is use of 50% formamide, 5x SSC (0.75M NaCl, 0.075 M sodium citrate), 50 mM sodium phosphate (pH 6.8), 0.1% sodium pyrophosphate, 5x Denhardt's solution, sonicated salmon sperm DNA (50 g/ml), 0.1% SDS, and 10% dextran sulfate at 42 C, with washes at 42 C. in 0.2x SSC and 0.1% SDS. As used herein, highly stringent  
25 conditions include the use of a wash at 65°C using 0.1xSSC and 0.1% SDS (see Sambrook *et al.*). A skilled artisan can readily determine and vary the stringency conditions appropriately to obtain a clear and detectable hybridization signal. Preferred molecules are those that hybridize under the above conditions to the complement of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5 or SEQ ID NO: 11 and  
30 which encode a functional protein. Preferred hybridizing molecules are those that hybridize under the above conditions to the complement strand of the open reading

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frame of SEQ ID NO:1 (nucleotides 130-1158), SEQ ID NO: 3 (nucleotides 163-1107), SEQ ID NO: 5 (nucleotides 73-1098) or SEQ ID NO: 11 (nucleotides 73-873).

As used herein, a clear signal is typically produced by hybridizing or annealing two nucleic acid molecules such that their complements exhibit at least about 80%,  
5 preferably at least about 85%, preferably at least about 90%, preferably at least about 95%, preferably at least about 96%, preferably at least about 97%, preferably at least about 98% or most preferably at least about 99% identity at the nucleotide level.

As used herein, a nucleic acid molecule is said to be "isolated" when the nucleic acid molecule is substantially separated from contaminant nucleic acid  
10 molecules encoding other polypeptides.

The present invention further provides fragments of the encoding nucleic acid molecule. As used herein, a fragment of an encoding nucleic acid molecule refers to a small portion of the entire protein coding sequence. The size of the fragment will be determined by the intended use. For example, if the fragment is chosen so as to  
15 encode an active portion of the protein, the fragment will need to be large enough to encode the functional region(s) of the protein. For instance, fragments which encode peptides corresponding to predicted antigenic regions may be prepared.

If the fragment is to be used as a nucleic acid probe or PCR primer, then the fragment length is chosen so as to obtain a relatively small number of false positives  
20 during probing/priming. Fragments of the encoding nucleic acid molecules of the present invention (*i.e.*, synthetic oligonucleotides) that are used as probes or specific primers for the polymerase chain reaction (PCR), or to synthesize gene sequences encoding proteins of the invention can easily be synthesized by chemical techniques, for example, the phosphotriester method of Matteucci *et al.* (*Journal of the American*  
25 *Chemical Society*, Vol. 103 (1981) pp. 3185-3191) or using automated synthesis methods. In addition, larger DNA segments can readily be prepared by well known methods, such as synthesis of a group of oligonucleotides that define various modular segments of the gene, followed by ligation of oligonucleotides to build the complete modified gene.

30 The encoding nucleic acid molecules of the present invention may further be modified so as to contain a detectable label for diagnostic and probe purposes. A

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variety of such labels are known in the art and can readily be employed with the encoding molecules herein described. Suitable labels include, but are not limited to, biotin, radiolabeled nucleotides and the like. A skilled artisan can readily employ any such labels to obtain labeled variants of the nucleic acid molecules of the invention.

5           Modifications to the primary structure itself by deletion, addition, or alteration of the amino acids incorporated into the protein sequence during translation can be made without destroying the activity of the protein. Such substitutions or other alterations result in proteins having an amino acid sequence encoded by a nucleic acid falling within the contemplated scope of the present invention.

10

### C.     **Isolation of Other Related Nucleic Acid Molecules**

As described above, the identification and characterization of the P2Y<sub>12</sub> nucleic acid molecule having SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5 or SEQ  
15 ID NO: 11 allows a skilled artisan to isolate nucleic acid molecules that encode other members of the protein family in addition to the sequences herein described.

Briefly, a skilled artisan can readily use the amino acid sequence of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6 or SEQ ID NO: 12 to generate antibody probes to screen expression libraries prepared from appropriate cells. Typically, polyclonal  
20 antiserum from mammals such as rabbits immunized with the purified protein (as described below) or monoclonal antibodies can be used to probe a mammalian cDNA or genomic expression library, such as lambda gtl1 library, to obtain the appropriate coding sequence for other members of the protein family. The cloned cDNA sequence can be expressed as a fusion protein, expressed directly using its own  
25 control sequences, or expressed by constructions using control sequences appropriate to the particular host used for expression of the enzyme.

Alternatively, a portion of the coding sequence herein described can be synthesized and used as a probe to retrieve DNA encoding a member of the protein family from any mammalian organism. Oligomers containing approximately 18-20  
30 nucleotides (encoding about a 6-7 amino acid stretch) are prepared and used to screen

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genomic DNA or cDNA libraries to obtain hybridization under stringent conditions or conditions of sufficient stringency to eliminate an undue level of false positives.

In PCR based methods, pairs of oligonucleotide primers can be prepared for use in a polymerase chain reaction (PCR) to selectively clone an encoding nucleic acid molecule. A PCR denature/anneal/extend cycle for using such PCR primers is well known in the art and can readily be adapted for use in isolating other encoding nucleic acid molecules.

Lastly, nucleic acid molecules encoding other members of the protein family may also be identified in existing genomic or other sequence information using any available computational method, including but not limited to: PSI-BLAST (Altschul, *et al.* (1997) *Nucleic Acids Res.* 25:3389-3402); PHI-BLAST (Zhang, *et al.* (1998), *Nucleic Acids Res.* 26:3986-3990), 3D-PSSM (Kelly *et al.* (2000) *J. Mol. Biol.* 299(2): 499-520); and other computational analysis methods (Shi *et al.* (1999) *Biochem. Biophys. Res. Commun.* 262(1):132-8 and Matsunami *et al.* (2000) *Nature* 404(6778):601-4.

#### **D. rDNA molecules Containing a Nucleic Acid Molecule**

The present invention further provides and utilizes recombinant DNA molecules (rDNAs) that contain a coding sequence. As used herein, a rDNA molecule is a DNA molecule that has been subjected to molecular manipulation *in situ*. Methods for generating rDNA molecules are well known in the art, for example, see Sambrook *et al.* *Molecular Cloning: A Laboratory Manual*. Cold Spring Harbor, NY. Cold Spring Harbor Laboratory Press, 1985. In the preferred rDNA molecules, a coding DNA sequence is operably linked to expression control sequences and/or vector sequences.

The choice of vector and/or expression control sequences to which one of the protein family encoding sequences of the present invention is operably linked depends directly, as is well known in the art, on the functional properties desired, *e.g.*, protein expression, and the host cell to be transformed. A vector contemplated by the present invention is at least capable of directing the replication or insertion into the host

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chromosome, and preferably also expression, of the structural gene included in the rDNA molecule.

Expression control elements that are used for regulating the expression of an operably linked protein encoding sequence are known in the art and include, but are not limited to, inducible promoters, constitutive promoters, secretion signals, and other regulatory elements. Preferably, the inducible promoter is readily controlled, such as being responsive to a nutrient in the host cell's medium.

In one embodiment, the vector containing a coding nucleic acid molecule will include a prokaryotic replicon, *i.e.*, a DNA sequence having the ability to direct autonomous replication and maintenance of the recombinant DNA molecule extra-chromosomally in a prokaryotic host cell, such as a bacterial host cell, transformed therewith. Such replicons are well known in the art. In addition, vectors that include a prokaryotic replicon may also include a gene whose expression confers a detectable marker such as a drug resistance. Typical bacterial drug resistance genes are those that confer resistance to ampicillin or tetracycline.

Vectors that include a prokaryotic replicon can further include a prokaryotic or bacteriophage promoter capable of directing the expression (transcription and translation) of the coding gene sequences in a bacterial host cell, such as *E. coli*. A promoter is an expression control element formed by a DNA sequence that permits binding of RNA polymerase and transcription to occur. Promoter sequences compatible with bacterial hosts are typically provided in plasmid vectors containing convenient restriction sites for insertion of a DNA segment of the present invention. Typical of such vector plasmids are pUC8, pUC9, pBR322 and pBR329 available from BioRad Laboratories, (Richmond, CA), pPL and pKK223 available from Pharmacia (Piscataway, NJ).

Expression vectors compatible with eukaryotic cells, preferably those compatible with vertebrate cells such as kidney cells, can also be used to form rDNA molecules that contain a coding sequence. Eukaryotic cell expression vectors are well known in the art and are available from several commercial sources. Typically, such vectors are provided containing convenient restriction sites for insertion of the desired DNA segment. Typical of such vectors are pSVL and pKSV-10 (Pharmacia),

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pBPV-1/pML2d (International Biotechnologies, Inc.), pTDT1 (ATCC, #31255), the vector pCDM8 described herein, and the like eukaryotic expression vectors.

Eukaryotic cell expression vectors used to construct the rDNA molecules of the present invention may further include a selectable marker that is effective in an eukaryotic cell, preferably a drug resistance selection marker. A preferred drug  
5 resistance marker is the gene whose expression results in neomycin resistance, *i.e.*, the neomycin phosphotransferase (*neo*) gene. (Southern *et al. Journal of Molecular and Applied Genetics*, Vol. 1, no. 4 (1982) pp. 327-341) Alternatively, the selectable marker can be present on a separate plasmid, and the two vectors are introduced by  
10 co-transfection of the host cell, and selected by culturing in the appropriate drug for the selectable marker.

**E. Host Cells Containing an Exogenously Supplied Coding Nucleic  
15 Acid Molecule**

The present invention further provides or utilizes host cells transformed with a nucleic acid molecule that encodes a protein of the present invention. The host cell can be either prokaryotic or eukaryotic. Eukaryotic cells useful for expression of a protein of the invention are not limited, so long as the cell line is compatible with cell  
20 culture methods and compatible with the propagation of the expression vector and expression of the gene product. Preferred eukaryotic host cells include, but are not limited to, yeast, insect and mammalian cells, preferably vertebrate cells such as those from a mouse, rat, monkey or human cell line. Preferred eukaryotic host cells include Chinese hamster ovary (CHO) cells available from the ATCC as CCL61, NIH Swiss  
25 mouse embryo cells (NIH3T3) available from the ATCC as CRL 1658, baby hamster kidney cells (BHK), COS and COS7 cells and like eukaryotic tissue culture cell lines.

Any prokaryotic host can be used to express a rDNA molecule encoding a protein of the invention, particularly peptides and fragments of the full-length receptor protein. The preferred prokaryotic host is *E. coli*.

30 Transformation of appropriate cell hosts with a rDNA molecule of the present invention is accomplished by well known methods that typically depend on the type

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of vector used and host system employed. With regard to transformation of prokaryotic host cells, electroporation and salt treatment methods are typically employed, see, for example, Cohen *et al. Proceedings of the National Academy of Science USA*, Vol. 69, no. 8 (1972) pp. 2110-2114; and Maniatis *et al. Molecular Cloning: A Laboratory Manual*. Cold Spring Harbor, NY. Cold Spring Harbor Laboratory Press, 1982). With regard to transformation of vertebrate cells with vectors containing rDNAs, electroporation, cationic lipid or salt treatment methods are typically employed, see, for example, Graham *et al. Virology*, Vol. 52, no. 2 (1973) pp. 456-467; and Wigler *et al. Proceedings of the National Academy of Science USA*, Vol. 76 (1979) pp. 1373-1376.

Successfully transformed cells, *i.e.*, cells that contain a rDNA molecule of the present invention, can be identified by well known techniques including the selection for a selectable marker. For example, cells resulting from the introduction of an rDNA of the present invention can be cloned to produce single colonies. Cells from those colonies can be harvested, lysed and their DNA content examined for the presence of the rDNA using a method such as that described by Southern, *Journal of Molecular Biology*, Vol. 98, no. 3 (1975) pp. 503-517; or Berent *et al. Biotechnic and Histochemistry*, Vol. 3 (1985) pp. 208; or the proteins produced from the cell assayed via an immunological method.

20

#### F. Production of Recombinant Proteins using a rDNA Molecule

The present invention further provides or utilizes methods for producing a protein of the invention using nucleic acid molecules herein described. In general terms, the production of a recombinant form of a protein typically involves the following steps:

First, a nucleic acid molecule is obtained that encodes a protein of the invention, such as the nucleic acid molecule depicted in SEQ ID NO: 1, nucleotides 130-1158 of SEQ ID NO: 1, nucleotides 130-1161 of SEQ ID NO: 1, SEQ ID NO: 3, nucleotides 163-1107 of SEQ ID NO:3, SEQ ID NO: 5, nucleotides 73-1098 of SEQ ID NO: 5, nucleotides 73-1101 of SEQ ID NO: 5, SEQ ID NO: 11, nucleotides 73-

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873 of SEQ ID NO: 11 or nucleotides 73-876 of SEQ ID NO: 11. If the encoding sequence is uninterrupted by introns as are SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5 or SEQ ID NO: 11, it is directly suitable for expression in any host.

The nucleic acid molecule is then preferably placed in operable linkage with suitable control sequences, as described above, to form an expression unit containing the protein open reading frame. The expression unit is used to transform a suitable host and the transformed host is cultured under conditions that allow the production of the recombinant protein. Optionally the recombinant protein is isolated from the medium or from the cells; recovery and purification of the protein may not be necessary in instances where some impurities may be tolerated, particularly when membrane fragments containing the receptor polypeptide are desired.

Each of the foregoing steps can be done in a variety of ways. For example, the desired coding sequences may be obtained from genomic fragments and used directly in appropriate hosts. The construction of expression vectors that are operable in a variety of hosts is accomplished using appropriate replicons and control sequences, as set forth above. The control sequences, expression vectors, and transformation methods are dependent on the type of host cell used to express the gene and were discussed in detail earlier. Suitable restriction sites can, if not normally available, be added to the ends of the coding sequence so as to provide an excisable gene to insert into these vectors. A skilled artisan can readily adapt any host/expression system known in the art for use with the nucleic acid molecules of the invention to produce recombinant protein.

#### 25           **G.       Methods to Identify Binding Partners**

Another embodiment of the present invention provides methods for isolating and identifying binding partners of proteins of the invention. In one method of the invention, a P2Y<sub>12</sub> protein, or fragment thereof, is mixed with a potential binding partner or an extract or fraction of a cell under conditions that allow the association of potential binding partners with the protein of the invention. After mixing, peptides, polypeptides, proteins or other molecules that have become associated with a protein

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of the invention are separated from the mixture. The binding partner that bound to the protein of the invention can then be removed and further analyzed. To identify and isolate a binding partner, the entire protein, for instance a protein comprising the entire amino acid sequence of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6 or SEQ ID NO: 12 can be used. Alternatively, a fragment of the protein can be used.

As used herein, a cellular extract refers to a preparation or fraction which is made from a lysed or disrupted cell, for instance, from platelets. The preferred source of cellular extracts will be cells that normally express the receptor polypeptide.

A variety of methods can be used to obtain an extract of a cell. Cells can be disrupted using either physical or chemical disruption methods. Examples of physical disruption methods include, but are not limited to, sonication and mechanical shearing. Examples of chemical lysis methods include, but are not limited to, detergent lysis and enzyme lysis. A skilled artisan can readily adapt methods for preparing cellular extracts in order to obtain extracts for use in the present methods.

Once an extract of a cell is prepared, the extract is mixed with the protein of the invention under conditions in which association of the protein with the binding partner can occur. A variety of conditions can be used, the most preferred being conditions that closely resemble conditions found in the cytoplasm of a human cell. Features such as osmolarity, pH, temperature, and the concentration of cellular extract used, can be varied to optimize the association of the protein with the binding partner.

After mixing under appropriate conditions, the bound complex is separated from the mixture. A variety of techniques can be utilized to separate the mixture. For example, antibodies specific to a protein of the invention can be used to immunoprecipitate the binding partner complex. Alternatively, standard chemical separation techniques such as chromatography and density/sediment centrifugation can be used.

After removal of non-associated cellular constituents found in the extract, the binding partner can be dissociated from the complex using conventional methods. For example, dissociation can be accomplished by altering the salt concentration or pH of the mixture.

To aid in separating associated binding partner pairs from the mixed extract, the protein of the invention can be immobilized on a solid support. For example, the protein can be attached to a nitrocellulose matrix or acrylic beads. Attachment of the protein to a solid support aids in separating peptide/binding partner pairs from other constituents found in the extract. The identified binding partners can be either a single protein or a complex made up of two or more proteins. Alternatively, binding partners may be identified using a Far-Western assay according to the procedures of Takayama *et al. Methods in Molecular Biology*, Vol. 69 (1997) pp. 171-184 or identified through the use of epitope tagged proteins or GST fusion proteins.

Alternatively, the nucleic acid molecules of the invention can be used in a yeast two-hybrid system. The yeast two-hybrid system has been used to identify other protein partner pairs and can readily be adapted to employ the nucleic acid molecules herein described.

#### **H. Radioligand Binding Assays**

Expression of the human P2Y<sub>12</sub> receptor in mammalian cells allows for preparation of cell membranes to be utilized in high throughput screening assays (*e.g.*, radioligand binding assays or other assays). Any standard procedure or assay format may be used such as those below described. The use of cell membranes eliminates the requirement of using platelets and/or blood products as a source of receptor for these assays, which improves the overall efficiency and convenience for the user.

#### **I. Methods to Identify Agents that Modulate the Expression of the Nucleic Acids**

Another embodiment of the present invention provides methods for identifying agents that modulate the expression of a nucleic acid encoding a protein of the invention such as a protein having the amino acid sequence of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6 or SEQ ID NO: 12. Such assays may utilize any available means of monitoring for changes in the expression level of the nucleic acids

of the invention. As used herein, an agent is said to modulate the expression of a nucleic acid of the invention, for instance a nucleic acid encoding the protein having the sequence of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6 or SEQ ID NO: 12, if it is capable of up- or down-regulating expression of the nucleic acid in a cell  
5 compared to a control.

In one assay format, cell lines that contain reporter gene fusions between the 3' and/or 5' regulatory sequences +/- the open reading frame defined by nucleotides 130-1158 of SEQ ID NO: 1 or nucleotides 130-1161 of SEQ ID NO: 1 or nucleotides 163-1107 of SEQ ID NO: 3 or nucleotides 73-1098 of SEQ ID NO: 5 or nucleotides  
10 73-1101 of SEQ ID NO: 5 or nucleotides 73-873 of SEQ ID NO: 11 or nucleotides 73-876 of SEQ ID NO: 11 and any assayable fusion partner may be prepared. Numerous assayable fusion partners are known and readily available including the firefly luciferase gene and the gene encoding chloramphenicol acetyltransferase (Alam *et al. Analytical Biochemistry*, Vol. 188 (1990) pp. 245-254). Cell lines  
15 containing the reporter gene fusions are then exposed to the agent to be tested under appropriate conditions and time. Differential expression of the reporter gene between samples exposed to the agent and control samples identifies agents which modulate the expression of a nucleic acid encoding the protein having the sequence of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6 or SEQ ID NO: 12.

20 Additional assay formats may be used to monitor the ability of the agent to modulate the expression of a nucleic acid encoding a protein of the invention such as the protein having SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6 or SEQ ID NO: 12. For instance, mRNA expression may be monitored directly by hybridization to the nucleic acids of the invention. Cell lines are exposed to the agent to be tested under  
25 appropriate conditions and time and total RNA or mRNA is isolated by standard procedures such as those disclosed in Sambrook *et al. Molecular Cloning: A Laboratory Manual*. Cold Spring Harbor, NY. Cold Spring Harbor Laboratory Press, 1985.

30 Probes to detect differences in RNA expression levels between cells exposed to the agent and control cells may be prepared from the nucleic acids of the invention. It is preferable, but not necessary, to design probes which hybridize only with target

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nucleic acids under conditions of high stringency. Only highly complementary nucleic acid hybrids form under conditions of high stringency. Accordingly, the stringency of the assay conditions determines the amount of complementarity which should exist between two nucleic acid strands in order to form a hybrid. Stringency  
5 should be chosen to maximize the difference in stability between the probe:target hybrid and potential probe:non-target hybrids.

Probes may be designed from the nucleic acids of the invention through methods known in the art. For instance, the G+C content of the probe and the probe length can affect probe binding to its target sequence. Methods to optimize probe  
10 specificity are commonly available in Sambrook *et al.* (*Molecular Cloning: A Laboratory Manual*. Cold Spring Harbor, NY. Cold Spring Harbor Laboratory Press, 1985); or Ausubel *et al.* (*Current Protocols in Molecular Biology*. NY, Greene Publishing Company, 1995).

Hybridization conditions are modified using known methods, such as those  
15 described by Sambrook *et al.* and Ausubel *et al.* as required for each probe. Hybridization of total cellular RNA or RNA enriched for polyA RNA can be accomplished in any available format. For instance, total cellular RNA or RNA enriched for polyA RNA can be affixed to a solid support and the solid support exposed to at least one probe comprising at least one, or part of one of the sequences  
20 of the invention under conditions in which the probe will specifically hybridize. Alternatively, nucleic acid fragments comprising at least one, or part of one of the sequences of the invention can be affixed to a solid support, such as a porous glass wafer. The glass wafer can then be exposed to total cellular RNA or polyA RNA from a sample under conditions in which the affixed sequences will specifically  
25 hybridize. Such glass wafers and hybridization methods are widely available, for example, those disclosed by Beattie (WO 95/11755). By examining for the ability of a given probe to specifically hybridize to an RNA sample from an untreated cell population and from a cell population exposed to the agent, agents which up or down regulate the expression of a nucleic acid encoding the protein having the sequence of  
30 SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6 or SEQ ID NO: 12 are identified.

Hybridization for qualitative and quantitative analysis of mRNAs may also be carried out by using a RNase Protection Assay (*i.e.*, RPA, see Ma *et al. Methods*, Vol. 10, no. 3 (1996) pp. 273-238). Briefly, an expression vehicle comprising cDNA encoding the gene product and a phage specific DNA dependent RNA polymerase promoter (*e.g.*, T7, T3 or SP6 RNA polymerase) is linearized at the 3' end of the cDNA molecule, downstream from the phage promoter, wherein such a linearized molecule is subsequently used as a template for synthesis of a labeled antisense transcript of the cDNA by *in vitro* transcription. The labeled transcript is then hybridized to a mixture of isolated RNA (*i.e.*, total or fractionated mRNA) by incubation at 45°C overnight in a buffer comprising 80% formamide, 40 mM Pipes, pH 6.4, 0.4 M NaCl and 1 mM EDTA. The resulting hybrids are then digested in a buffer comprising 40 g/ml ribonuclease A and 2 g/ml ribonuclease. After deactivation and extraction of extraneous proteins, the samples are loaded onto urea/polyacrylamide gels for analysis.

In another assay format, cells or cell lines are first be identified which express the gene products of the invention physiologically (*e.g.*, see for example, Figures 10 and 12 for tissue distribution via Northern blot). Cell and/or cell lines so identified would be expected to comprise the necessary cellular machinery such that the fidelity of modulation of the transcriptional apparatus is maintained with regard to exogenous contact of agent with appropriate surface transduction mechanisms and/or the cytosolic cascades. Further, such cells or cell lines would be transduced or transfected with an expression vehicle (*e.g.*, a plasmid or viral vector) construct comprising an operable non-translated 5'-promoter containing end of the structural gene encoding the instant gene products fused to one or more antigenic fragments, which are peculiar to the instant gene products, wherein said fragments are under the transcriptional control of said promoter and are expressed as polypeptides whose molecular weight can be distinguished from the naturally occurring polypeptides or may further comprise an immunologically distinct tag. Such a process is well known in the art (see Maniatis *et al. Molecular Cloning: A Laboratory Mammal*. Cold Spring Harbor, NY. Cold Spring Harbor Laboratory Press, 1982).

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Cells or cell lines transduced or transfected as outlined above would then be contacted with agents under appropriate conditions; for example, the agent comprises a pharmaceutically acceptable excipient and is contacted with cells comprised in an aqueous physiological buffer such as phosphate buffered saline (PBS) at physiological pH, Eagles balanced salt solution (BSS) at physiological pH, PBS or BSS comprising serum or conditioned media comprising PBS or BSS and/or serum incubated at 37°C . Said conditions may be modulated as deemed necessary by one of skill in the art. Subsequent to contacting the cells with the agent, said cells will be disrupted and the polypeptides from disrupted cells are fractionated such that a polypeptide fraction is pooled and contacted with an antibody to be further processed by immunological assay (*e.g.*, ELISA, immunoprecipitation or Western blot). The pool of proteins isolated from the "agent contacted" sample will be compared with a control sample where only the excipient is contacted with the cells and an increase or decrease in the immunologically generated signal from the "agent contacted" sample compared to the control will be used to distinguish the effectiveness of the agent.

**J. Methods to Identify Agents that Modulate at Least One Activity of the Proteins.**

Another embodiment of the present invention provides methods for identifying agents that modulate at least one activity of a protein of the invention such as the protein having the amino acid sequence of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6 or SEQ ID NO: 12. Such methods or assays may utilize any means of monitoring or detecting the desired activity.

In one format, the relative amounts of a protein of the invention between a cell population that has been exposed to the agent to be tested compared to an un-exposed control cell population may be assayed. In this format, probes such as specific antibodies are used to monitor the differential expression of the protein in the different cell populations. Cell lines or populations are exposed to the agent to be tested under appropriate conditions and time. Cellular lysates or membrane fractions may be prepared from the exposed cell line or population and a control, unexposed

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cell line or population. The cellular lysates or membrane fractions are then analyzed with the probe.

Antibody probes are prepared by immunizing suitable mammalian hosts in appropriate immunization protocols using the peptides, polypeptides or proteins of the invention if they are of sufficient length, or, if desired, or if required to enhance  
5 immunogenicity, conjugated to suitable carriers. Methods for preparing immunogenic conjugates with carriers such as BSA, KLH, or other carrier proteins are well known in the art. In some circumstances, direct conjugation using, for example, carbodiimide reagents may be effective; in other instances linking reagents such as  
10 those supplied by Pierce Chemical Co. (Rockford, IL), may be desirable to provide accessibility to the hapten. The hapten peptides can be extended at either the amino or carboxy terminus with a Cysteine residue or interspersed with cysteine residues, for example, to facilitate linking to a carrier. Administration of the immunogens is conducted generally by injection over a suitable time period and with use of suitable  
15 adjuvants, as is generally understood in the art. During the immunization schedule, titers of antibodies are taken to determine adequacy of antibody formation.

While the polyclonal antisera produced in this way may be satisfactory for some applications, for pharmaceutical compositions, use of monoclonal preparations is preferred. Immortalized cell lines which secrete the desired monoclonal antibodies  
20 may be prepared using the standard method of Kohler and Milstein (*Nature*, Vol. 256, no. 5517 (Aug. 1975) pp. 495-497) or modifications which effect immortalization of lymphocytes or spleen cells, as is generally known. The immortalized cell lines secreting the desired antibodies are screened by immunoassay in which the antigen is the peptide hapten, polypeptide or protein. When the appropriate immortalized cell  
25 culture secreting the desired antibody is identified, the cells can be cultured either *in vitro* or by production in ascites fluid.

The desired monoclonal antibodies are then recovered from the culture supernatant or from the ascites supernatant. Fragments of the monoclonals or the polyclonal antisera which contain the immunologically significant portion can be used  
30 as antagonists, as well as the intact antibodies. Use of immunologically reactive fragments, such as the Fab, Fab', of F(ab')<sub>2</sub> fragments is often preferable, especially in

a therapeutic context, as these fragments are generally less immunogenic than the whole immunoglobulin.

The antibodies or fragments may also be produced, using current technology, by recombinant means. Antibody regions that bind specifically to the desired regions  
5 of the protein can also be produced in the context of chimeras with multiple species origin, particularly humanized antibodies.

As mentioned above, isolated cells providing a P2Y<sub>12</sub> receptor polypeptide on their surface and the availability of the recombinant DNA encoding a receptor polypeptide which permits display and expression of the receptor on host cell surfaces  
10 are all valuable tools for evaluating the ability of candidate agonists or antagonists to bind to the receptor and thus contribute to the receptor's activation or deactivation. In this manner, the invention includes cell assay systems which utilize an isolated or a recombinantly produced receptor polypeptide to screen for agonist and antagonist activity of candidate drugs. These assays are especially useful in assuring that these  
15 candidate therapeutic agents have the desired effect of either activating or inhibiting the receptor polypeptide. Determination of these properties is essential in evaluating the specificity of drugs intended for binding other related receptors.

The most useful host cells are typically animal cells, including mammalian  
20 cells. In order to be useful in the assays, the cells must have intracellular mechanisms which permit the receptor to be displayed on the cell surface. Particularly useful cells for use in the method of the invention are *Xenopus laevis* frog oocytes, which typically utilize cRNA rather than standard recombinant expression systems proceeding from the DNA encoding the desired protein. Capped RNA (at the 5' end)  
25 is typically produced from linearized vectors containing DNA sequences encoding the receptor. The reaction is conducted using RNA polymerase and standard reagents. cRNA is recovered, typically using phenol/chloroform precipitation with ethanol and injected into the oocytes.

The animal host cells expressing the DNA encoding the receptor or the  
30 cRNA-injected oocytes are then cultured to effect the expression of the encoding nucleic acids so as to produce the receptor display on the cell surface. These cells

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then are used directly in assays for assessment of a candidate drug to bind, antagonize, or activate the receptor.

One method of evaluating candidates as potential therapeutic agents typically involves a binding assay in which the candidate (such as a peptide or a small organic molecule) would be tested to measure if, or to what extent, it binds the receptor. Preferably, a mammalian or insect cell line that expresses the receptor or plasma membrane preparations thereof, will be used in a binding assay. For example, a candidate antagonist competes for binding to the receptor with either a labeled nucleotide agonist or antagonist. Varying concentrations of the candidate are supplied, along with a constant concentration of the labeled agonist or antagonist. The inhibition of binding of the labeled material can then be measured using established techniques. This measurement is then correlated to determine the amount and potency of the candidate that is bound to the receptor.

Another method of evaluating candidates for potential therapeutic applications typically involves a functional assay in which the candidate's effect upon cells expressing the recombinant receptor is measured, rather than simply determining its ability to bind the receptor (see Jantzen *et al.* (1999) *Thromb. Haemost.* 81:111-117). Suitable functional assays include those that measure calcium mobilization,  $^{45}\text{Ca}$  efflux or measurements of intracellular  $\text{Ca}^{+2}$  concentration with fluorescent dyes such as fura-2 and voltage clamp, described below.

For example, agonist-induced increases in  $^{45}\text{Ca}$  release by oocytes expressing cRNA encoding the receptor or other mammalian recombinant cells producing the receptor can be measured by the techniques described by Williams, et al., (1988) *Proc. Natl. Acad. Sci. USA* 85:4939-4943. Intracellular calcium pools are labeled by incubating groups of 30 oocytes in 300  $\mu\text{l}$  calcium-free modified Barth's solution (MBSH) containing 50  $\mu\text{Ci}$   $^{45}\text{CaCl}_2$  (10-40  $\text{mCi/mg Ca}$ ; Amersham) for 4 hours at room temperature. The labeled oocytes or cells are washed, then incubated in MBSH II without antibiotics for 90 minutes. Groups of 5 oocytes are selected and placed in individual wells in a 24-well tissue culture plate containing 0.5 ml/well MBSH II without antibiotics. This medium is removed and replaced with fresh medium every 10 minutes; the harvested medium is analyzed by scintillation counting to determine

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<sup>45</sup>Ca released by the oocytes during each 10-minute incubation. The 10-minute incubations are continued until a stable baseline of <sup>45</sup>Ca release per unit time is achieved. Two additional 10-minute collections are obtained, then test medium including agonist is added and <sup>45</sup>Ca release determined.

5           Using the above assay, the ability of a candidate drug to activate the receptor can be tested directly. In this case, ADP or the agonists of the invention are used as controls. In addition, by using the agonists of the invention to activate the recombinant receptor, the effect of the candidate drug on this activation can be tested directly. Cells expressing the nucleic acids encoding the receptor are incubated in the  
10 assay in the presence of agonist with and without the candidate compound. A diminution in activation in the presence of the candidate will indicate an antagonist effect. Conversely, the ability of a candidate drug to reverse the antagonist effects of an antagonist of the invention may also be tested.

As indicated above, receptor activation can also be measured by means of the  
15 two-electrode voltage clamp assay. In this assay, agonist-induced inward chloride currents are measured in voltage-clamped oocytes that express the receptor coinjected with a chimeric G alpha subunit which converts Gi-responses to Gq responses. The technique suitable for use in the instant invention is described by Julius, *et al.*, (1988) *Science* 241:558-563.

20           Agents that are assayed in the above method can be randomly selected or rationally selected or designed. As used herein, an agent is said to be randomly selected when the agent is chosen randomly without considering the specific sequences involved in the association of a protein of the invention alone or with its associated substrates, binding partners, etc. An example of randomly selected agents  
25 is the use a chemical library or a peptide combinatorial library, or a growth broth of an organism.

As used herein, an agent is said to be rationally selected or designed when the agent is chosen on a nonrandom basis which takes into account the sequence of the target site and/or its conformation in connection with the agent's action.

30           The agents of the present invention can be, as examples, peptides, small molecules, vitamin derivatives, as well as carbohydrates. A skilled artisan can readily

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recognize that there is no limit as to the structural nature of the agents of the present invention. Dominant negative proteins, DNAs encoding these proteins, antibodies to these proteins, peptide fragments of these proteins or mimics of these proteins may be introduced into cells to affect function. "Mimic" used herein refers to the modification  
5 of a region or several regions of a peptide molecule to provide a structure chemically different from the parent peptide but topographically and functionally similar to the parent peptide (see Grant GA. in: Meyers (ed.) *Molecular Biology and Biotechnology* (New York, VCH Publishers, 1995), pp. 659-664).

The peptide agents of the invention can be prepared using standard solid phase  
10 (or solution phase) peptide synthesis methods, as is known in the art. In addition, the DNA encoding these peptides may be synthesized using commercially available oligonucleotide synthesis instrumentation and produced recombinantly using standard recombinant production systems. The production using solid phase peptide synthesis is necessitated if non-gene-encoded amino acids are to be included.

15 Another class of agents of the present invention are antibodies immunoreactive with critical positions of proteins of the invention. Antibody agents are obtained by immunization of suitable mammalian subjects with peptides, containing as antigenic regions, those portions of the protein intended to be targeted by the antibodies.

20

#### **K. cAMP assays**

Expression of the P2Y<sub>12</sub> receptor in mammalian cells allows for measurement  
25 of ADP-dependent cAMP responses in transfected cells. This functional read-out is useful for high throughput screening using standard screening assay techniques. In addition, the demonstrated coupling to the cAMP pathway is useful for screening assays using other G-protein dependent read-outs, such as changes in intracellular calcium mediated by G $\alpha$ 16 or Gi-Gq-chimeric subunits. (see Figure 3 A, B, C).

30

**L. Uses for Agents that Modulate at Least One Activity of the Proteins.**

As provided in the Examples, the proteins and nucleic acids of the invention, such as the proteins having the amino acid sequences of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6 or SEQ ID NO: 12, are expressed in various tissues, including but not limited to the brain, and blood cells such as platelets. Agents that modulate, up-or-down-regulate the expression of the protein or agents such as agonists or antagonists of at least one activity of the protein may be used to modulate biological and pathologic processes associated with the protein's function and activity.

Agonists or antagonists of the invention have therapeutic utility (1) in treating diseases caused by aberrant activation of this receptor in tissues where it is customarily found, for example in the vascular system or the brain and (2) in treating diseases whose symptoms can be ameliorated by stimulating or inhibiting the activity of the receptor, for example diseases of the vascular system or injured vascular tissue.

Vascular disease may refer to any disease of the cardiovascular system, including but not limited to acute myocardial infarction, unstable angina, chronic stable angina, transient ischemic attacks, strokes, peripheral vascular disease, preeclampsia, deep venous thrombosis, embolism, disseminated intravascular coagulation or thrombotic cytopenic purpura. Vascular injury may refer to an injury arising by any means, including but not limited to procedures such as angioplasty, carotid endarterectomy, post CABG (coronary artery bypass graft) surgery, vascular graft surgery, stent placements or insertion of endovascular devices and prostheses.

As used herein, a subject can be any mammal, so long as the mammal is in need of modulation of a pathological or biological process mediated by a protein of the invention. The term mammal is defined as an individual belonging to the class Mammalia. The invention is particularly useful in the treatment of human subjects.

Pathological processes refer to a category of biological processes which produce a deleterious effect. For example, expression of a protein of the invention may be associated with improved ADP-induced platelet activation and aggregation.

As used herein, an agent is said to modulate a pathological process when the agent reduces the degree or severity of the process. For instance, bleeding disorders or

thrombosis may be prevented or disease progression modulated by the administration of agents which modulates in some way the expression or at least one activity of a protein of the invention.

The agents of the present invention can be provided alone, or in combination  
5 with other agents that modulate a particular pathological process. For example, an agent of the present invention can be administered in combination with other known drugs or may be combined with surgery or with known blood-thinning drugs. As used herein, two agents are said to be administered in combination when the two agents are administered simultaneously or are administered independently in a fashion  
10 such that the agents will act at the same time.

The agents of the present invention can be administered via parenteral, subcutaneous, intravenous, intramuscular, intraperitoneal, transdermal, or buccal routes. Alternatively, or concurrently, administration may be by the oral route. The dosage administered will be dependent upon the age, health, and weight of the  
15 recipient, kind of concurrent treatment, if any, frequency of treatment, and the nature of the effect desired.

The present invention further provides compositions containing one or more agents which modulate expression or at least one activity of a protein of the invention. While individual needs vary, determination of optimal ranges of effective amounts of  
20 each component is within the skill of the art. Typical dosages comprise 0.1 to 100  $\mu\text{g}/\text{kg}$  body wt. The preferred dosages comprise 0.1 to 10  $\mu\text{g}/\text{kg}$  body weight. The most preferred dosages comprise 0.1 to 1  $\mu\text{g}/\text{kg}$  body weight.

In addition to the pharmacologically active agent, the compositions of the present invention may contain suitable pharmaceutically acceptable carriers  
25 comprising excipients and auxiliaries which facilitate processing of the active compounds into preparations which can be used pharmaceutically for delivery to the site of action. Suitable formulations for parenteral administration include aqueous solutions of the active compounds in water-soluble form, for example, water-soluble salts. In addition, suspensions of the active compounds as appropriate oily injection  
30 suspensions may be administered. Suitable lipophilic solvents or vehicles include

fatty oils, for example, sesame oil, or synthetic fatty acid esters, for example, ethyl oleate or triglycerides. Aqueous injection suspensions may contain substances which increase the viscosity of the suspension include, for example, sodium carboxymethyl cellulose, sorbitol, and/or dextran. Optionally, the suspension may also contain stabilizers. Liposomes can also be used to encapsulate the agent for delivery into the cell.

The pharmaceutical formulation for systemic administration according to the invention may be formulated for enteral, parenteral or topical administration. Indeed, all three types of formulations may be used simultaneously to achieve systemic administration of the active ingredient.

Suitable formulations for oral administration include hard or soft gelatin capsules, pills, tablets, including coated tablets, elixirs, suspensions, syrups or inhalations and controlled release forms thereof.

In practicing the methods of this invention, the compounds of this invention may be used alone or in combination, or in combination with other therapeutic or diagnostic agents. In certain preferred embodiments, the compounds of this invention may be coadministered along with other compounds typically prescribed for these conditions according to generally accepted medical practice. The compounds of this invention can be utilized *in vivo*, ordinarily in mammals, such as humans, sheep, horses, cattle, pigs, dogs, cats, rats and mice, or *in vitro*.

#### **M. Transgenic Animals**

Transgenic animals containing, unmodified, mutant, knock-out or modified genes corresponding to the nucleic acid molecules of the invention, particularly the cDNA sequence of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5 or SEQ ID NO: 11, are also included in the invention. Transgenic animals are genetically modified animals into which recombinant, exogenous or cloned genetic material has been experimentally transferred. Such genetic material is often referred to as a "transgene". The nucleic acid sequence of the transgene, in this case a form of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5 or SEQ ID NO: 11 may be integrated either at a locus

of a genome where that particular nucleic acid sequence is not otherwise normally found or at the normal locus for the transgene. The transgene may consist of nucleic acid sequences derived from the genome of the same species or of a different species than the species of the target animal.

5           The term "germ cell line transgenic animal" refers to a transgenic animal in which the genetic alteration or genetic information was introduced into a germ line cell, thereby conferring the ability of the transgenic animal to transfer the genetic information to offspring. If such offspring in fact possess some or all of that alteration or genetic information, then they too are transgenic animals.

10           The alteration or genetic information may be foreign to the species of animal to which the recipient belongs, foreign only to the particular individual recipient, or may be genetic information already possessed by the recipient. In the last case, the altered or introduced gene may be expressed differently than the native gene.

          Transgenic animals can be produced by a variety of different methods  
15 including transfection, electroporation, microinjection, gene targeting in embryonic stem cells and recombinant viral and retroviral infection (see, *e.g.*, U.S. Patent No. 4,736,866; U.S. Patent No. 5,602,307; Mullins *et al.* *Hypertension* (1993) 22(4):630-633; Brenin *et al.* *Surgical Oncology*, Vol. 6, no. 2 (1997) pp. 99-110; "Recombinant Gene Expression Protocols" in: Tuan (ed.), *Methods in Molecular*  
20 *Biology*, No. 62 (Humana Press, 1997)).

          A number of recombinant or transgenic mice have been produced, including those which express an activated oncogene sequence (U.S. Patent No. 4,736,866); express simian SV40 T-antigen (U.S. Patent No. 5,728,915); lack the expression of interferon regulatory factor 1 (IRF-1) (U.S. Patent No. 5,731,490); exhibit  
25 dopaminergic dysfunction (U.S. Patent No. 5,723,719); express at least one human gene which participates in blood pressure control (U.S. Patent No. 5,731,489); display greater similarity to the conditions existing in naturally occurring Alzheimer's disease (U.S. Patent No. 5,720,936); have a reduced capacity to mediate cellular adhesion (U.S. Patent No. 5,602,307); possess a bovine growth hormone gene (Clutter *et al.*  
30 *Genetics*, Vol. 143, no. 4 (1996) pp. 1753-1760); or, are capable of generating a fully human antibody response (McCarthy. *The Lancet*, Vol. 349, no. 9049 (1997) pp. 405).

While mice and rats remain the animals of choice for most transgenic experimentation, in some instances it is preferable or even necessary to use alternative animal species. Transgenic procedures have been successfully utilized in a variety of non-murine animals, including sheep, goats, pigs, dogs, cats, monkeys, chimpanzees, hamsters, rabbits, cows and guinea pigs (see, e.g., Kim *et al. Molecular Reproduction and Development*, Vol. 46, no. 4 (1997) pp. 515-526; Houdebine. *Reproduction, Nutrition, Development*, Vol. 35, no. 6 (1995) pp. 609-617; Petters *Reproduction, Fertility and Development*, Vol. 6, no. 5 (1994) pp. 643-645; Schnieke *et al. Science* Vol. 278, no. 5346 (1997) pp. 2130-2133; and Amoah, *Journal of Animal Science*, Vol. 75, no. 2 (1997) pp. 578-585.

The method of introduction of nucleic acid fragments into recombination competent mammalian cells can be by any method which favors co-transformation of multiple nucleic acid molecules. Detailed procedures for producing transgenic animals are readily available to one skilled in the art, including the disclosures in U.S. Patent No. 5,489,743 and U.S. Patent No. 5,602,307.

#### N. Diagnostic Methods

One means of diagnosing a vascular disease or disorder using the nucleic acid molecules or proteins of the invention involves obtaining a tissue or blood sample from living subjects.

The use of molecular biological tools has become routine in forensic technology. For example, nucleic acid probes may be used to determine the expression of a nucleic acid molecule comprising all or at least part of the sequence of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5 or SEQ ID NO: 11 in forensic/pathology specimens. Further, nucleic acid assays may be carried out by any means of conducting a transcriptional profiling analysis. In addition to nucleic acid analysis, forensic methods of the invention may target the protein encoded by SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5 or SEQ ID NO: 11 to determine up or down regulation of the genes (Shiverick *et al., Biochim Biophys Acta* (1975) 393(1):124-33).

Assays to detect nucleic acid or protein molecules of the invention may be in any available format. Typical assays for nucleic acid molecules include hybridization or PCR based formats. Typical assays for the detection of proteins, polypeptides or peptides of the invention include the use of antibody probes in any available format  
5 such as *in situ* binding assays, etc. See Harlow *et al.*, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988. In preferred embodiments, assays are carried-out with appropriate controls. Methods of the invention may involve treatment of tissues with collagenases or other proteases to make the tissue amenable to cell lysis (Semenov *et al.*, *Biull Eksp Biol Med* (1987) 104(7):113-6).

10 The above methods may also be used in other diagnostic protocols including for forensic purposes, and in diagnostic protocols and methods to detect disease states in other tissues or organs, for example the brain.

Without further description, it is believed that one of ordinary skill in the art can, using the preceding description and the following illustrative examples, make  
15 and utilize the compounds of the present invention and practice the claimed methods. The following working examples therefore, specifically point out preferred embodiments of the present invention, and are not to be construed as limiting in any way the remainder of the disclosure.

**EXAMPLES**Example 1Platelet cDNA library

In the parent provisional application 60/171,622, the P2Y<sub>12</sub> receptor protein  
5 was referred to as the H11 receptor. Poly-A<sup>+</sup> mRNA from rat platelets was used to  
generate a directional oligo-dT primed cDNA library in the pcDNA3.1<sup>+</sup> vector.  
Approximately 320,000 clones were divided into 48 individual pools. Linearized  
cDNA templates from these pools were transcribed in vitro using T7 RNA  
polymerase (Ambion). Sib selection of a positive pool was performed to  
10 subfractionate the signal to the level of 96 clones. All were sequenced and a novel  
GPCR was further characterized. Rat P2Y<sub>12</sub> cDNA was used to isolate a human  
orthologue from a platelet λ ZAP cDNA library. A full length hP2Y<sub>12</sub> cDNA  
expression construct was obtained by ligation of a λ clone and a fragment derived by  
3' RACE into the pcIneo expression vector (Promega). GenBank accession number  
15 for human P2Y<sub>12</sub> is XXXXX. SEQ ID NO:1 corresponds to a full length rat P2Y<sub>12</sub>  
sequence, SEQ ID NO:3 corresponds to a partial human P2Y<sub>12</sub> sequence, SEQ ID  
NO:5 corresponds to a full length human P2Y<sub>12</sub> sequence, SEQ ID NO:11  
corresponds to a truncated allelic variant human P2Y<sub>12</sub> sequence.

20 Platelet RT-PCR

Whole blood (30 ml) was lysed and total RNA isolated using TriReagent BD  
(Molecular Research Center). First-strand cDNA was generated (Superscript 2, Life  
Technologies) and PCR (35 cycles) performed using the following mRNA-specific  
primers: The P2Y<sub>12</sub> 5' (5'-CCAGAATCAACAGTTATCAGGTAACC-3') (SEQ ID  
25 NO: 7); and 3' (5'-GTCAGTTAATATTTTACTTAGCGCTTTGC-3') (SEQ ID NO:  
8) primers were annealed at 57°C, while the GPIIb  
5' (5'-GTCAACGGGGATGGGAGGCATGA-3') (SEQ ID NO: 9) and  
3' (5'-GTCTGCCTCATCTCGAAGGAAGG-3') (SEQ ID NO: 10) primers were  
annealed at 60°C. PCR products were analyzed by electrophoresis in 1% agarose and  
30 bands of the correct size were isolated for direct sequencing.

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### Electrophysiology

Defolliculated *Xenopus laevis* oocytes were injected with a positive 500 clone pool (10 ng), rP2Y<sub>12</sub> (10 pg), hP2Y<sub>12</sub> (50 pg), Kir3.1, Kir3.4, PTX and hm2 (1 ng each) cRNAs as indicated. Three to seven days after injection, two-electrode voltage-clamp recordings were performed using a Geneclamp 500 amplifier (Axon Instruments) and a Maclab A/D converter (Maclab). Membrane potentials were clamped at -70 mV while the recording chamber was perfused at a rate of 2 ml/min with a solution containing (in mM) 70 KCl, 20 NaCl, 3 MgCl<sub>2</sub>, 5 HEPES, pH 7.4, at room temperature. The KCl was replaced with NaCl to examine responses in zero potassium. Agonists and antagonists (Roche Molecular Biochemistry or Sigma) were diluted in the recording solution. Experiments using C1330-7 included 0.1 % dimethylsulfoxide to enhance its solubility in the perfusate.

### 15 Generation of stable mammalian cell lines and cAMP assays

Chinese Hamster Ovary (CHO) cells or rat 2-9 fibroblasts, which are null for G<sub>i</sub>-linked purinergic receptors, were transfected with hP2Y<sub>12</sub> or rP2Y<sub>12</sub> cDNA's, respectively using FuGene reagent (Roche), and cells were cultured in the presence of G418 for 2 weeks to select for stable transfectants. For cAMP assays, stably transfected CHO cells expressing the hP2Y<sub>12</sub> plasmid were plated in 12-well dishes. Forty-eight hours later media was removed from the cells and replaced with serum-free media containing IBMX (0.25 mM final) and incubated at 37°C for 5 minutes. Cells were incubated for an additional 5 min with 10 μM forskolin, as well as the indicated agonists and antagonists. Pertussis toxin treatment (30 ng/ml) occurred for 20 hrs at 37°C prior to assay. Cyclic AMP levels were determined from aliquots of cell extracts in a radioimmunoassay (Amersham Biotrak cAMP <sup>125</sup>I assay system).

### Northern and in situ hybridizations

Northern blots of poly-A<sup>+</sup> RNA from human tissues (Clontech) or total human platelet RNA was hybridized with radiolabeled hP2Y<sub>12</sub> cDNA fragments under standard conditions. Digoxigenin-labeled *in situ* hybridization was performed

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on coronal rat brain sections using an RNA probe corresponding to the antisense sequence of rP2Y<sub>12</sub> (Caterina *et al.* (1997).

#### Flow cytometry

5           Adult male Sprague-Dawley rats were anesthetized and whole blood isolated using citrate as anticoagulant. Platelet-rich plasma (PRP) was isolated by centrifugation and used for flow-cytometry analysis. A rabbit anti-sera (SynPep Corporation) was produced to the amino-terminal 23 residues of rP2Y<sub>12</sub>. IgG was purified using protein-G sepharose. Rat PRP ( $2 \times 10^6$  cells) and cultured rat 2-9  
10 fibroblasts transfected with rP2Y<sub>12</sub> cDNA ( $1 \times 10^5$  cells) were incubated with purified IgG (10-50  $\mu$ g/ml) in FACS buffer (phosphate-buffered saline containing 0.1% BSA and 2% heat-inactivated fetal bovine serum) in a total volume of 100  $\mu$ l for 1 hr at 4°C. Cells and platelets were then washed with cold FACS buffer and incubated with 2.5  $\mu$ g/ml of FITC-conjugated goat anti-rabbit antibody for 30 min at 4°C. Cells and  
15 platelets were washed, resuspended in cold FACS buffer, and fluorescence of cell-bound secondary antibody was determined with a FACSort flow cytometer (Becton-Dickinson). Control samples contained cells without antibodies (for determination of autofluorescence), cells with control rabbit IgG, or secondary antibodies alone.

#### 20 Figure 1: P2Y<sub>12</sub> is a G protein-coupled receptor that responds to ADP

Figure 1(A). Activation of potassium-dependent currents in *Xenopus* oocytes expressing P2Y<sub>12</sub> with Kir3.1 and 3.4. ADP (10  $\mu$ M) was applied (short bars) in the presence or absence (long bar) of extracellular potassium (70 mM) while recording membrane currents in the whole-cell voltage clamp configuration. Oocytes injected  
25 with mRNA for Kir3.1 and 3.4 alone (top trace) do not exhibit significant currents in response to ADP application unless messages from a positive cDNA pool, the isolated rat P2Y<sub>12</sub> cRNA or the human P2Y<sub>12</sub> homologue are included (subsequent traces). (B) ADP-selective stimulation of potassium channel-dependent currents by P2Y<sub>12</sub> occurs via a pertussis toxin-sensitive pathway. UDP, adenosine (A), ATP- $\alpha$ -S or ADP (10  
30  $\mu$ M each) were sequentially applied to oocytes expressing the rat or human receptor

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with or without Kir3.1, 3.4 and pertussis toxin (PTX). (C) The agonist profile of P2Y<sub>12</sub> recapitulates that observed for the G<sub>i</sub>-coupled platelet ADP receptor. Concentration-response curves for ADP and 2MeSADP are presented. Membrane currents were normalized in each oocyte to a response obtained with 10 μM ADP.

- 5 Each point represents the mean values (+/- s.d.) from five independent oocytes. The Hill equation was used to fit the response data.

Figure 2: Currents stimulated by ADP in oocytes expressing hP2Y<sub>12</sub> with Kir3.1 and 3.4 are inhibited by 2MeSAMP, C1330-7 and a thiol reagent

- 10 Figure 2(A). Current tracing showing reversible block of ADP (1 μM) responses by 2MeSAMP (10 μM) and C1330-7 (1 μM), but not A3P5P (300 μM). The dot (.) indicates the start of a 15 sec application of ADP, while bars denote co-application with the antagonist. (B) 2MeSAMP and C1330-7 inhibition curves. Current responses were normalized to that elicited by ADP (500 nM) alone in each
- 15 oocyte and plotted as the mean +/- s.d. Curves were fitted to the data using the Hill equation (n=5 independent oocytes for each point). (C) Selective ablation of P2Y<sub>12</sub> but not m2 muscarinic receptor signaling by the thiol reagent pCMBS. ADP (10 μM), carbachol (1 μM), and pCMBS (1 mM) were applied sequentially to an oocyte
- 20 expressing both receptors concurrently with Kir3.1 and 3.4. Bars indicate periods of drug application (10 sec).

Figure 3: Activation of hP2Y<sub>12</sub> in CHO cells inhibits adenylyl cyclase

- Figure 3(A). Receptor coupling to adenylyl cyclase was assessed as ADP-mediated (0.1-10 μM) inhibition of forskolin-stimulated (10 μM) cAMP accumulation
- 25 (CON=control, normalized to 100%). (B) The effect of the specific antagonists 2MeSAMP (2-MES)(50 μM) and C1330-7 (50 μM) on repression of ADP-mediated (1 μM) forskolin-stimulated cAMP levels. (C) Effect of pertussis toxin (PTX) pretreatment on the inhibition by 10 μM ADP of forskolin-stimulated cAMP levels. Results are the mean +/- s.d. of three representative experiments performed in
- 30 triplicate.

Figure 4: P2Y<sub>12</sub> receptor is selectively expressed in platelets and brain

Figure 4(A, B). Northern analysis of hP2Y<sub>12</sub> transcripts. All lanes contain 2 µg poly-A<sup>+</sup> mRNA except samples from platelet and Jurkat cells (20 µg each). (C) rP2Y<sub>12</sub> transcripts are distributed throughout the brain in presumptive glia. Staining was equally abundant in fiber tracts (corpus callosum, cc) and regions enriched for neuronal cell bodies (dentate gyrus, dg; arcuate nucleus of the hypothalamus, an), but absent from vasculature (pericallosal artery, pa). Control (sense) riboprobes did not stain these regions. Ventricular structures are also indicated (dorsal third ventricle, d3v; third ventricle, 3v). (D) FACS analysis of rat platelets stained with rP2Y<sub>12</sub> antisera (filled peak) or a control IgG (unfilled peak). (E) FACS analysis of rat 2-9 fibroblasts transfected with the rP2Y<sub>12</sub> cDNA clone (filled peak) or untransfected rat 2-9 fibroblasts (unfilled peak).

Figure 5: A frame-shift mutation within the hP2Y<sub>12</sub> gene is associated with a bleeding disorder

Figure 5(A). Deduced amino acid sequence of the hP2Y<sub>12</sub> protein and alignment with other homologous receptor sequences. The putative membrane-spanning domains are designated with bars above the sequence. hP2Y<sub>12</sub> sequence is aligned with the sequences of hP2Y<sub>1</sub> receptor (also expressed in platelets and activated by ADP), as well as with the human UDP-glucose receptor, with which it shares greatest homology. Shading denotes amino acid identity (black) or similarity (gray); asterisks (\*) denote extracellular cysteine residues. (B) A P2Y<sub>12</sub> allele from a patient (ML) with defective ADP-dependent aggregation contains a 2 base pair deletion, resulting in a frame-shift mutation and a premature truncation of the protein. No such mutation was observed in PCR products amplified and sequenced from genomic DNA of one hundred randomly chosen individuals. (C) Mutant hP2Y<sub>12</sub> receptor from patient ML is non-functional and does not act in a dominant-negative capacity. Representative ADP-evoked membrane currents from an oocyte injected with 50 pg of wildtype (WT) hP2Y<sub>12</sub> cRNA (upper left panel), 50 pg of mutant (MT) hP2Y<sub>12</sub> cRNA (upper right panel) or with 50 pg WT and increasing amounts of MT

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hP2Y<sub>12</sub> cRNAs (bottom panels). Oocytes were also injected with 1 ng Kir3.1 and 1 ng Kir3.4 cRNAs. Dot indicates onset of ADP application (10 μM for 5 s). (D) Patient ML has abnormally low levels of RT-PCR product derived from P2Y<sub>12</sub> mRNA. RT-PCR using either P2Y<sub>12</sub>- (lanes 1-4) or GPIIb-(lanes 5-8) specific primers was performed using whole blood RNA from patient ML (lanes 1,2,5,6) or a control (CON) sample (lanes 3,4,7,8). PCR reactions performed on RNA samples without reverse transcriptase control for genomic DNA contamination (lanes 1,3,5,7). A 1.1 kb product encoding the P2Y<sub>12</sub> ORF was amplified from the control sample, but virtually absent from ML (a faint product can be observed upon longer exposure). In contrast, the amount of product (0.77 kb) amplified from GPIIb mRNA was equivalent between ML and control. Sequence analysis reveals that ML's P2Y<sub>12</sub> RT-PCR product derived solely from the mutant allele.

15 Figure 6: Evaluation of [<sup>3</sup>H]2MeSADP Binding to the Cloned hP2Y<sub>12</sub> Receptor; High Throughput Screening (HTS) Assay

hP2Y<sub>12</sub> cDNA was transiently transfected into cos7 cells and crude membranes were prepared using standard procedures. Binding of 1 nM [<sup>3</sup>H]2MeSADP at various membrane protein concentrations and in the absence and presence of various inhibitors was determined at 4C for 30 minutes in a rapid filtration assay (triplicates). Non-specific binding was determined with 10 μM 2MeSADP and subtracted from all data. Each experiment included membranes from untransfected cells.

At 1 nM [<sup>3</sup>H]2MeSADP (approx. K<sub>D</sub> in whole platelet binding assays), specific binding was increased up to 5-fold in membranes from transfected vs. untransfected cells. This ratio was optimal around 25 ug membrane protein, an amount used in many GPCR binding assays. Nonspecific binding (incl. counter background) was <10% of total binding and identical in transfected and untransfected cells. Similar results were obtained with membranes from TSA-201 cells (data not shown). This result indicates that a HTS assay using the cloned P2Y<sub>12</sub> receptor and available radioligands such as [<sup>3</sup>H]2MeSADP might be feasible, especially when the receptor expression can be increased another 2-fold and assay conditions will be

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further optimized. SPA technology should be evaluated to facilitate automation. The amount of membranes required could be further reduced using [ $^{33}\text{P}$ ]2MeSADP with higher specific activity.

The selectivity of the expressed receptor was assessed using competition binding with the P2Y<sub>12</sub> antagonist 2MeSAMP and the P2Y<sub>1</sub> antagonist A3P5P. 2MeSAMP at 50  $\mu\text{M}$  reduced [ $^3\text{H}$ ]2MeSADP binding to membranes from transfected cells essentially to the level of untransfected cells with a potency similar to whole platelet binding assays. Binding to membranes from untransfected cells was only marginally affected. A3P5P had only little effect on binding even at 300  $\mu\text{M}$ , and the degree of inhibition can be explained by inhibiting endogenous receptors in the untransfected cells. This preliminary experiment suggests that the cloned P2Y<sub>12</sub> receptor displays a pharmacological profile similar to the high-affinity binding site for [ $^3\text{H}$ ]2MeSADP on whole platelets further validating its use for HTS.

To identify the G<sub>i</sub>-linked platelet ADP receptor, the present inventors engineered *Xenopus* oocytes to allow detection of G<sub>i</sub>-linked responses through a sensitive electrophysiological assay. This strategy is based on the fact that several G<sub>i</sub>-coupled receptors, such as the m2 muscarinic receptor, release G $\beta\gamma$  subunits from heterotrimeric G proteins, thereby activating inwardly rectifying potassium (K<sup>+</sup>) channels (Kir3.1-4) (Krapivinsky *et al.* (1995)). A cDNA library from rat platelets was screened in oocytes expressing Kir3.1 and 3.4 and three positive pools that responded to 10  $\mu\text{M}$  ADP (as determined by an increase in K<sup>+</sup> current) were identified. Subfractionation of one of these pools led to the identification of a single clone tentatively designated as P2Y<sub>12</sub> (SEQ ID NO:1). The current induced by ADP was K<sup>+</sup>-dependent since replacement of K<sup>+</sup> in the bath solution resulted in a complete loss of current (Figure 1A). Additionally, injection of Kir or P2Y<sub>12</sub> cRNAs alone gave no ADP-dependent currents, indicating that the observed signal was not due to activation of an endogenous purinergic receptor and was Kir-dependent (Figure 1B). Moreover, when cRNA encoding pertussin toxin was injected together with the rat P2Y<sub>12</sub> clone, the response to ADP was abolished (Figure 1B), as predicted for the G<sub>i</sub>-linked platelet ADP receptor (Ohlmann *et al.* (1995)). The human P2Y<sub>12</sub> homolog

(SEQ ID NO: 5) was isolated from a human platelet library and similar results were obtained when this cRNA was expressed in *Xenopus* oocytes (Figure 1A, B).

One hallmark of the G<sub>i</sub>-linked platelet ADP receptor is that substitution of alkylthio groups at the 2-position of the adenine ring increases potency at the receptor (Mills (1996); MacFarlane *et al.* (1983); Hourani *et al.* (1994)). Consistent with this, 2MeSADP displayed 2 orders of magnitude greater potency compared to ADP (with an EC<sub>50</sub> of 0.9 nM and 300 nM, respectively) (Figure 1C). In contrast, other nucleoside or nucleotide derivatives were without effect (Figure 1B). The present inventors also examined the actions of several antagonists specific for the platelet G<sub>i</sub>-linked ADP receptor. Treatment of *Xenopus* oocytes expressing the rat or human P2Y<sub>12</sub> receptor with the nucleotide derivative 2MeSAMP (Jantzen *et al.* (1999)) or a non-nucleotide inhibitor C1330-7 (Jantzen *et al.* (1998)), blocked ADP-induced K<sup>+</sup> currents with IC<sub>50</sub>'s of 1.4 μM and 40 nM, respectively (Figure 2B). In contrast, the P2Y<sub>1</sub>-selective antagonist A3P5P (Boyer *et al.* (1996)) had no inhibitory effect on the signal evoked by ADP at the rat or human P2Y<sub>12</sub> (Figure 2A). Thus, when expressed in *Xenopus* oocytes, the P2Y<sub>12</sub> receptor recapitulates the pharmacological profile previously described for the platelet G<sub>i</sub>-linked ADP receptor. The only anomaly that we observed relates to the action of ATP-αS, which behaved as a weak agonist, rather than an antagonist at the cloned receptor. This finding is somewhat unexpected since ATP derivatives reportedly antagonize the platelet G<sub>i</sub>-linked receptor. However, this discrepancy may reflect partial degradation or impurities in commercially available preparations of ATP-αS, or differences between the platelet and oocyte environments, such as the degree of ectonucleotidase activity. Indeed, recombinant P2Y<sub>1</sub> receptors respond differentially to ATP, depending on the expression system utilized (Palmer *et al.* (1998); Filippov *et al.* (2000)).

Chinese Hamster Ovary (CHO) cells expressing the hP2Y<sub>12</sub> receptor displayed ADP-mediated repression of forskolin-stimulated cAMP levels in a dose-dependent manner, reaching a maximum of 47% reduction at 10 μM ADP (Figure 3A). The repression of cAMP levels by 1 μM ADP was reversed by the selective antagonists 2MeSAMP and C1330-7 (Figure 3B), in agreement with the pharmacological profile

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observed in *Xenopus* oocytes, and as described for the G<sub>i</sub>-coupled receptor on platelets. Neither of these antagonists had effects on forskolin-stimulated cAMP levels in the absence of agonist. Similar responses to ADP were observed in rat 2-9 fibroblasts stably expressing rP2Y<sub>12</sub>. Pretreatment of transfected cells with pertussis toxin abolished ADP effects on forskolin-stimulated cAMP (Figure 3C), suggesting that the response is G<sub>i</sub>-mediated.

Northern blot analysis demonstrated that P2Y<sub>12</sub> is abundantly expressed in human platelets, and to a lesser extent in brain (Figure 4A, B). The predominant transcript of 2.4 kb was absent from all other tissues examined, including peripheral blood leukocytes. A fainter species of ~4.5 kb was also detected in platelet and brain, while a prominent band of ~1.0 kb (Figure 4B) was observed only in platelet RNA. Among rat tissues, selective expression in platelets and brain was also seen. Thus, the mRNA for this novel GPCR has a restricted expression pattern and is abundantly present in platelets, consistent with this cDNA encoding the platelet G<sub>i</sub>-linked receptor. Within the brain, the 2.4 kb species was observed in numerous subregions, including the amygdala, caudate nucleus, corpus collosum, hippocampus, substantia nigra, and thalamus. Cellular resolution of rP2Y<sub>12</sub> expression was obtained by *in situ* hybridization histochemistry of brain sections where punctate staining was noted throughout white and gray matter (Figure 4C). Principal cells of the hippocampus did not stain, nor was a laminar pattern of expression observed in the neocortex. These observations are consistent with a glial expression pattern. Interestingly, the only cell line previously described to express a P2Y purinergic receptor that is negatively coupled to adenylyl cyclase is the rat C6 glioma cell line (Boyer *et al.* (1993)). Indeed, a 2.4 kb mRNA species was detected in these cells by Northern analysis with a rP2Y<sub>12</sub> probe.

Using a rabbit polyclonal antisera directed to the predicted amino-terminus of rP2Y<sub>12</sub>, we assessed surface expression of receptor protein on stably-transfected rat 2-9 fibroblasts or rat platelets using flow cytometry. At an antibody concentration of 25 µg/ml, a 9-fold (Figure 4E) and 4-fold (Figure 4D) increase in mean fluorescence intensity (compared to a control antibody) was observed with transfected cells and

platelets, respectively, demonstrating that P2Y<sub>12</sub> protein is, indeed, expressed on the platelet surface.

When the chromosomal localization of the P2Y<sub>12</sub> gene was determined using the Stanford G3 panel (Stewart *et al.* (1997)) (Research Genetics), P2Y<sub>12</sub>-specific primers mapped closest to STS-D13626, corresponding to the KIAA0001 gene recently identified as a UDP-glucose GPCR (Chambers *et al.* (2000)). Both of these genes reside on chromosome 3q24-25, interval D3S1279-1280, a region that also includes the human P2Y<sub>1</sub> gene, (GeneMap 99, [www.ncbi.nlm.nih.gov](http://www.ncbi.nlm.nih.gov)). Thus, this interval contains genes encoding at least three receptors, two of which (P2Y<sub>1</sub> and P2Y<sub>12</sub>) mediate ADP-dependent platelet aggregation. Among GPCR's, P2Y<sub>12</sub> is most closely related to the UDP-glucose receptor (Chambers *et al.* (2000)) (44% identical) but much less so to P2Y<sub>1</sub> (19% identical), suggesting that the UDP-glucose and P2Y<sub>12</sub> receptors are the product of a relatively recent gene duplication on chromosome 3.

The predicted hP2Y<sub>12</sub> protein encodes four extracellular cysteines (see Figure 5). A critical role of cysteine residues in the function of the platelet ADP receptor has been suggested by the ability of thiol reagents to ablate ADP responses in platelets (Mills (1996)). Indeed, the antithrombotic agent clopidogrel is proposed to inactivate the G<sub>i</sub>-linked platelet ADP receptor through a mechanism in which it is metabolized to a sulphhydryl species that modifies a cysteine residue on the receptor (Savi *et al.* (1999)). The present inventors found that brief exposure of oocytes expressing Kir3.1, 3.4 and hP2Y<sub>12</sub> to the thiol reagent p-chloromercuriphenylsulfonic acid (pCMBS) eliminated ADP-evoked current responses (Figure 2C). Inhibition was selective for the P2Y<sub>12</sub> receptor since activation of this signaling pathway by m2 muscarinic receptors expressed in the same oocytes was unaffected by treatment with pCMBS.

Nurden *et al.* (1995) have previously described a patient (ML) with a mild bleeding disorder. Platelets from ML exhibit impaired ADP-dependent platelet aggregation, greatly reduced ADP binding activity and lack the ability to inhibit cAMP levels in response to ADP. However, the P2Y<sub>1</sub>-receptor mediated responses, such as intracellular calcium mobilization and shape change, are not affected, suggesting that this patient has a selective defect in the G<sub>i</sub>-linked receptor. Analysis of PCR products from the P2Y<sub>12</sub> coding region from ML's genomic DNA revealed the

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presence of one mutant allele at this locus, as confirmed by direct sequencing of at least three independent PCR reactions. The mutation found in the P2Y<sub>12</sub> gene consists of a deletion of two nucleotides (TTCATT) within the coding region, at amino acid 240 (near the amino-terminal end of TM6), thus shifting the reading frame for 28  
5 residues before introducing a premature stop codon (Figure 5B). Biochemical studies suggest that platelets from ML lack G<sub>i</sub>-linked ADP receptors, yet sequence analysis indicates that this individual has one mutant and one wild-type P2Y<sub>12</sub> allele, at least so far as the protein coding region is concerned. This suggests one of two possibilities: the P2Y<sub>12</sub> mutation identified exerts a dominant-negative effect, or ML  
10 harbors an additional mutation that eliminates expression of the allele containing a wild-type coding region. We evaluated the former possibility using an electrophysiological assay (Figure 5C). First, no significant activity was observed when oocytes were injected with cRNA transcripts corresponding to the frame-shifted allele, demonstrating that this mutant is indeed non-functional. Moreover, when  
15 mutant and wild-type cRNA's were co-injected into oocytes at different ratios, no inhibition of the signal from the wild-type allele was observed, demonstrating that the mutant allele does not act in a dominant negative manner. Further support for this conclusion comes from sequence analysis of the P2Y<sub>12</sub> coding region from ML's daughter, who has previously been shown to have an intermediate number of ADP  
20 binding sites and impaired ADP-dependent aggregation (Nurden *et al.* (1995)). Like her father, she has one wt and one frame-shifted allele, and is therefore likely to be a true heterozygote, both genotypically and phenotypically. If so, then the truncated receptor does not act as a dominant negative *in vivo*. Finally, it was asked whether ML's alleles are both expressed by carrying out RT-PCR analysis with RNA from his  
25 platelets. Extremely low levels of P2Y<sub>12</sub>-derived product were obtained compared to levels amplified from an unaffected individual or compared to a control transcript encoding platelet GPIIB (Figure 5D). In addition, sequence analysis of P2Y<sub>12</sub> RT-PCR products demonstrated that ML's P2Y<sub>12</sub> transcripts derive only from the mutant allele (*i.e.*, no wild-type product was detected). We therefore conclude that ML's lack  
30 of functional G<sub>i</sub>-coupled platelet ADP receptor activity is due to the fact that he expresses only the frame-shifted allele.

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Although the present invention has been described in detail with reference to examples above, it is understood that various modifications can be made without departing from the spirit of the invention. Accordingly, the invention is limited only by the following claims. All cited patents, patent applications and publications  
5 referred to in this application are herein incorporated by reference in their entirety.

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## WHAT IS CLAIMED:

1. An isolated nucleic acid molecule selected from the group consisting of: (a) an isolated nucleic acid molecule that encodes the amino acid sequence of SEQ ID NO: 2; (b) an isolated nucleic acid molecule which hybridizes to the complement of a nucleic acid molecule comprising SEQ ID NO: 1 under conditions of sufficient stringency to produce a clear signal; and (c) an isolated nucleic acid molecule which hybridizes to a nucleic acid molecule that encodes the amino acid sequence of SEQ ID NO: 2 under conditions of sufficient stringency to produce a clear signal.
2. The isolated nucleic acid molecule of claim 1, wherein the nucleic acid molecule comprises the sequence of SEQ ID NO: 1.
3. The isolated nucleic acid molecule of claim 1, wherein the nucleic acid molecule consists of the sequence of SEQ ID NO: 1.
4. The isolated nucleic acid molecule of claim 1, wherein the nucleic acid molecule comprises nucleotides 130-1158 of SEQ ID NO: 1 or nucleotides 130-1161 of SEQ ID NO: 1.
5. The isolated nucleic acid molecule of claim 1, wherein the nucleic acid molecule consists of nucleotides 130-1158 of SEQ ID NO: 1 or nucleotides 130-1161 of SEQ ID NO: 1.
6. The isolated nucleic acid molecule of any one of claims 1-5, wherein said nucleic acid molecule is operably linked to one or more expression control elements.
7. A vector comprising an isolated nucleic acid molecule of any one of claims 1-5.

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8. A host cell transformed to contain the nucleic acid molecule of any one of claims 1-5.

9. A host cell comprising a vector of claim 7.

5

10. A host cell of claim 9, wherein said host is selected from the group consisting of prokaryotic hosts and eukaryotic hosts.

11. A method for producing a polypeptide comprising the step of culturing a host cell transformed with the nucleic acid molecule of any one of claims 1-5 under conditions in which the protein encoded by said nucleic acid molecule is expressed.

10

12. The method of claim 11, wherein said host cell is selected from the group consisting of prokaryotic hosts and eukaryotic hosts.

15

13. An isolated receptor polypeptide produced by the method of claim 11.

14. An isolated receptor polypeptide comprising the amino acid sequence of SEQ ID NO: 2.

20

15. An isolated antibody that binds to a polypeptide of either claim 13 or 14.

16. An isolated antibody that binds to the polypeptide of claim 14.

25

17. The antibody of claim 16 wherein said antibody is a monoclonal or polyclonal antibody.

18. An isolated nucleic acid molecule that encodes the amino acid sequence of SEQ ID NO: 12.

30

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19. The isolated nucleic acid molecule of claim 18, wherein the nucleic acid molecule comprises the sequence of SEQ ID NO: 11.

20. The isolated nucleic acid molecule of claim 18, wherein the nucleic acid molecule comprises nucleotides 73-873 of SEQ ID NO: 11 or nucleotides 73-876 of SEQ ID NO: 11.

21. The isolated nucleic acid molecule of any one of claims 18-20, wherein said nucleic acid molecule is operably linked to one or more expression control elements.

22. A vector comprising an isolated nucleic acid molecule of any one of claims 18-20.

23. A host cell transformed to contain the nucleic acid molecule of any one of claims 18-20.

24. A host cell comprising a vector of claim 22.

25. A host cell of claim 24, wherein said host is selected from the group consisting of prokaryotic hosts and eukaryotic hosts.

26. A method for producing a polypeptide comprising the step of culturing a host cell transformed with the nucleic acid molecule of any one of claims 18-20 under conditions in which the protein encoded by said nucleic acid molecule is expressed.

27. The method of claim 26, wherein said host cell is selected from the group consisting of prokaryotic hosts and eukaryotic hosts.

28. An isolated receptor polypeptide produced by the method of claim 27.

29. An isolated receptor polypeptide comprising the amino acid sequence of SEQ ID NO: 12.

5 30. An isolated antibody that binds to a polypeptide of either of claims 28 or 29.

31. An isolated antibody that binds to the polypeptide of claim 29.

10 32. The antibody of claim 31 wherein said antibody is a monoclonal or polyclonal antibody.

33. A method of identifying an agent which modulates the expression of a nucleic acid encoding a P2Y<sub>12</sub> receptor comprising the steps of:

- 15 (a) exposing cells which express the nucleic acid to the agent; and  
(b) determining whether the agent modulates expression of said nucleic acid, thereby identifying an agent which modulates the expression of a nucleic acid encoding the protein having the sequence of P2Y<sub>12</sub> receptor.

20 34. A method of identifying an agent which modulates at least one activity of a P2Y<sub>12</sub> receptor comprising the steps of:

- (a) exposing cells which express the protein to the agent;  
(b) determining whether the agent modulates at least one activity of said protein, thereby identifying an agent which modulates at least one activity of a P2Y<sub>12</sub> receptor.  
25

35. The method of claim 34, wherein the agent modulates the interaction between ADP, or a derivative thereof, and a P2Y<sub>12</sub> receptor.

30 36. The method of claim 34 wherein the agent modulates potassium current.

37. The method of claim 34 wherein the agent modulates an activity selected from the group consisting of adenylyl cyclase activity, platelet activation and thrombotic activity.

5

38. A method of identifying binding partners for a P2Y<sub>12</sub> receptor protein comprising the steps of:

- (a) exposing said protein to a potential binding partner; and
- (b) determining if the potential binding partner binds to said protein, thereby identifying binding partners for a P2Y<sub>12</sub> receptor protein.

10

39. A method of modulating the expression of a nucleic acid encoding a P2Y<sub>12</sub> receptor protein comprising the step of:

- (a) administering an effective amount of an agent which modulates the expression of a nucleic acid encoding encoding a P2Y<sub>12</sub> receptor protein.

15

40. A method of modulating at least one activity of a P2Y<sub>12</sub> receptor protein comprising the step of:

- (a) administering an effective amount of an agent which modulates at least one activity of a P2Y<sub>12</sub> receptor protein.

20

41. The method of claim 40, wherein the agent modulates the interaction between ADP or ATP and the P2Y<sub>12</sub> receptor protein.

25

42. The method of claim 40 wherein the agent modulates potassium current.

43. The method of claim 40 wherein the agent modulates an activity selected from the group consisting of adenylyl cyclase activity, platelet activation and thrombotic activity.

30

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44. The method of claim 40 wherein the agent modulates acute myocardial infarction, unstable angina, chronic stable angina, transient ischemic attacks, strokes, peripheral vascular disease, preeclampsia, deep venous thrombosis, embolism, disseminated intravascular coagulation, thrombotic thrombocytopenic purpura or a  
5 bleeding disorder.

45. The method of claim 40 wherein the agent modulates thrombotic and restenotic complications following angioplasty, carotid endarterectomy, post CABG (coronary artery bypass graft) surgery, vascular graft surgery, stent placements or  
10 insertion of endovascular devices and prostheses.

46. A non-human transgenic animal modified to contain a nucleic acid molecule which encodes a P2Y<sub>12</sub> receptor or a truncation mutant of P2Y<sub>12</sub> receptor.

15 47. A method of diagnosing a disease state in a subject, comprising the step of determining the level of expression of a nucleic acid molecule encoding a P2Y<sub>12</sub> receptor.

20 48. A method of diagnosing a disease state in a subject, comprising the step of determining the level of expression of a P2Y<sub>12</sub> receptor protein.

25 49. An isolated mutant P2Y<sub>12</sub> receptor protein which is a naturally occurring truncation mutant, wherein said truncation is caused by a frame-shift mutation in the region of the gene encoding transmembrane domain six of the protein.

50. The receptor protein of claim 49 wherein said protein is associated with a bleeding disorder.

30 51. The receptor protein of claim 49 wherein said frame shift mutation introduces a stop codon in the gene encoding the P2Y<sub>12</sub> protein and results in platelets

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with impaired ADP-dependent platelet aggregation activity, reduced ADP binding activity and reduced ability to inhibit cAMP levels in response to ADP.

52. An isolated nucleic acid encoding a mutant P2Y<sub>12</sub> receptor protein  
5 which is a naturally occurring truncation mutant, wherein said truncation is caused by a frame-shift mutation in the region of the gene encoding transmembrane domain six of the protein.

53. An isolated nucleic acid of claim 52 wherein said nucleic acid encodes  
10 a protein associated with a bleeding disorder.

54. An isolated nucleic acid of claim 52 wherein said frame shift mutation  
introduces a stop codon in the gene encoding the P2Y<sub>12</sub> protein and results in platelets  
with impaired ADP-dependent platelet aggregation activity, reduced ADP binding  
15 activity and reduced ability to inhibit cAMP levels in response to ADP.

55. A method of diagnosing a disease state in a subject, comprising the  
step of determining the level of expression of a nucleic acid molecule of any one of  
claims 52-54.

20

56. A method of diagnosing a disease state in a subject, comprising the  
step of determining the level of expression of a protein of any one of claims 49-51.

57. A method of diagnosing a disease state in a subject, comprising the  
25 step of identifying a nucleic acid molecule of any of claims 52-54.

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10

Figure 1A

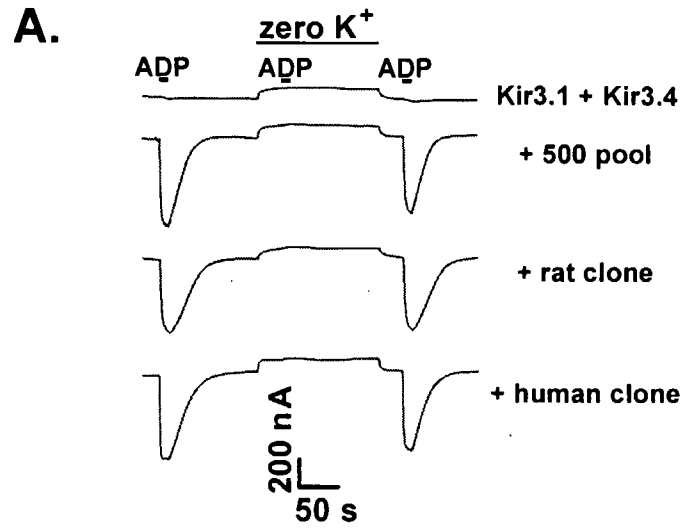


Figure 1B

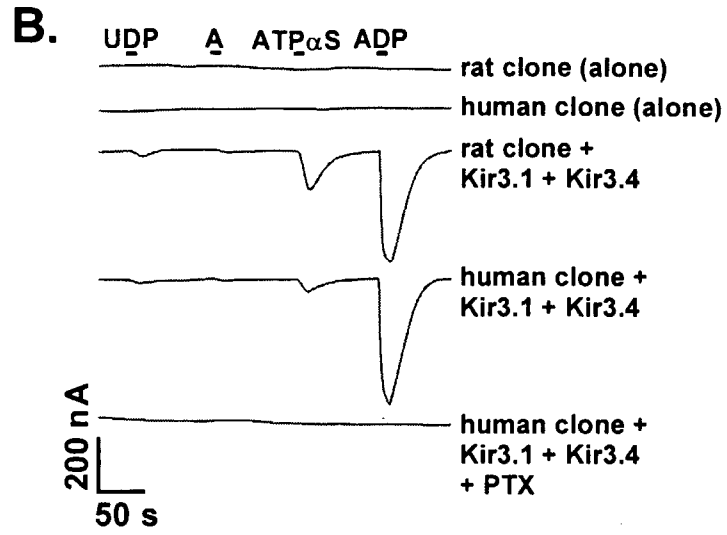


Figure 1C

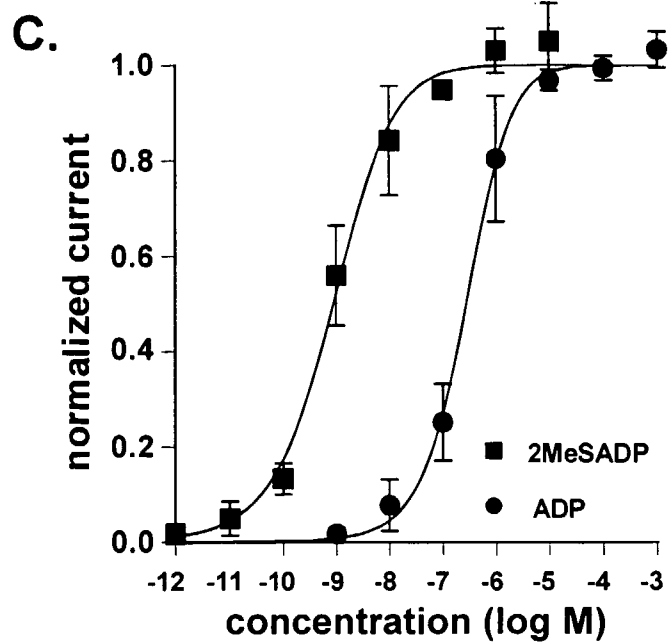


Figure 2A

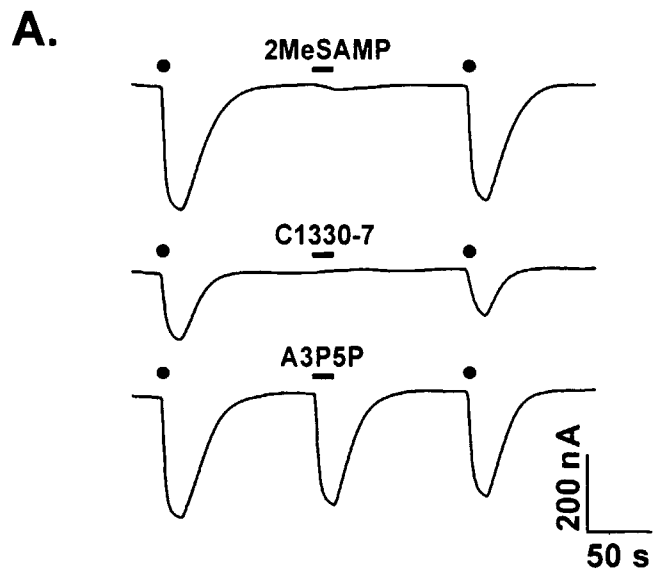


Figure 2B

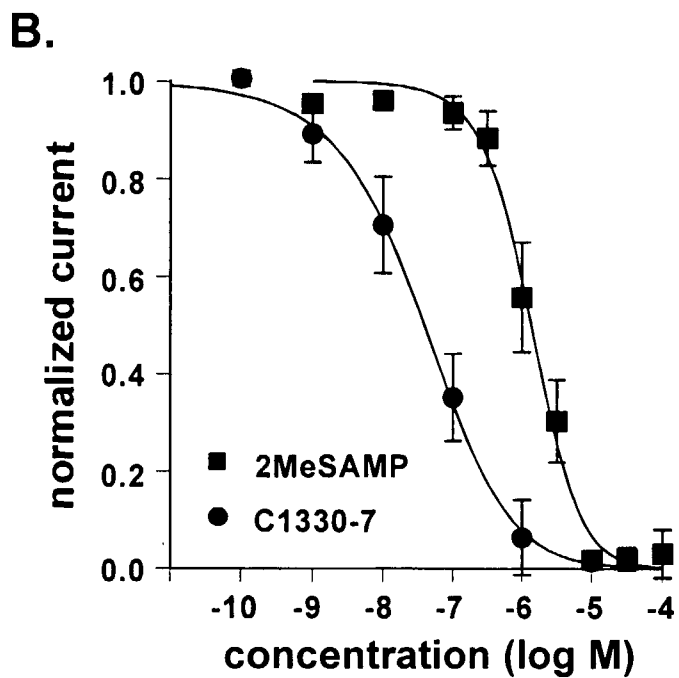
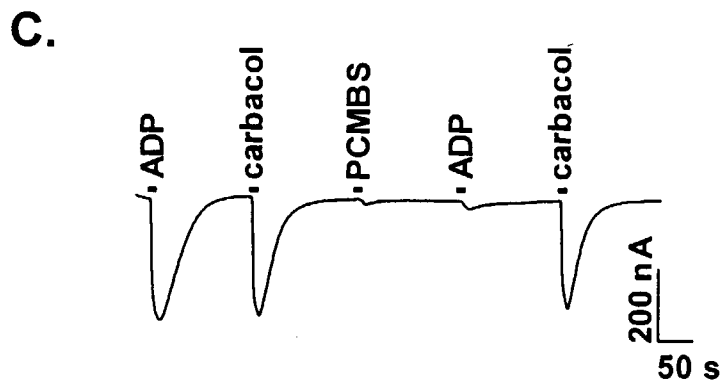
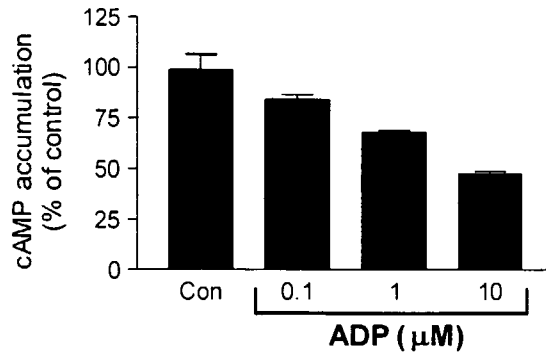


Figure 2C

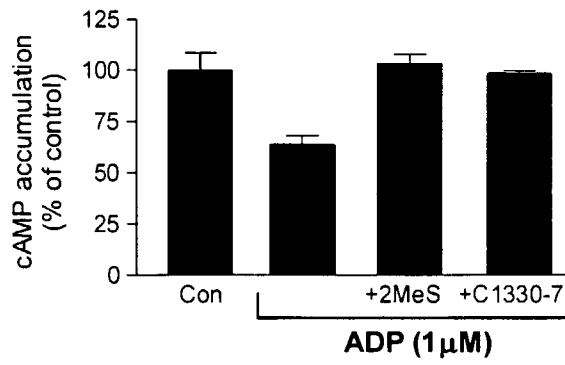


**Figure 3**

**A.**



**B.**



**C.**

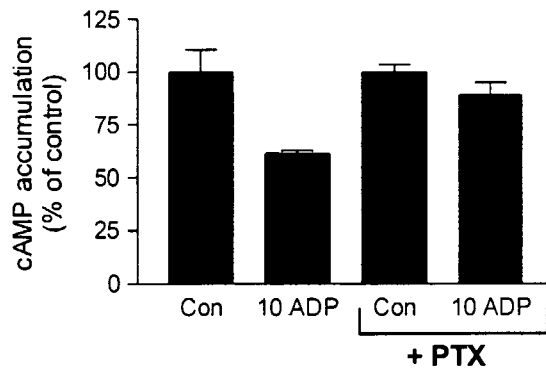
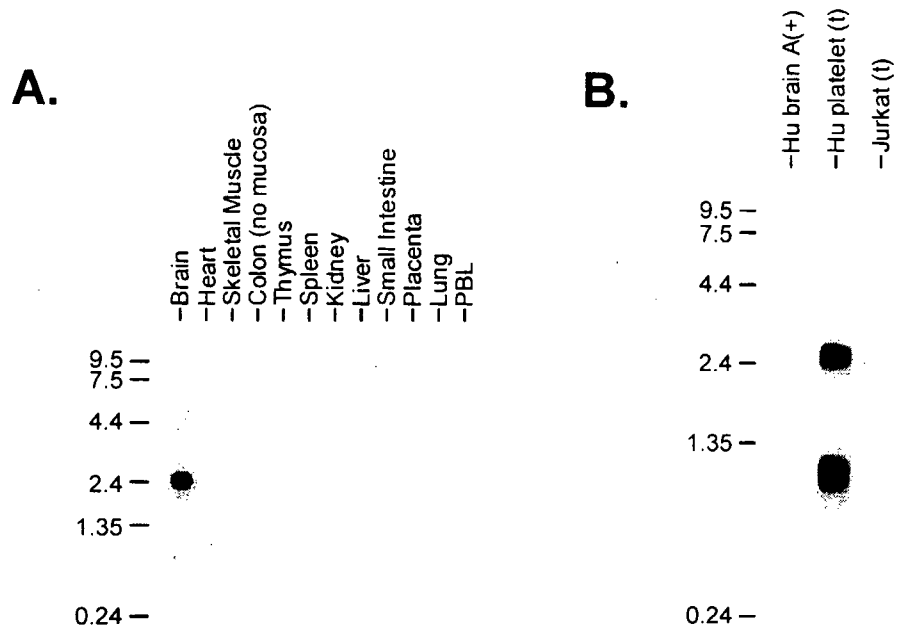
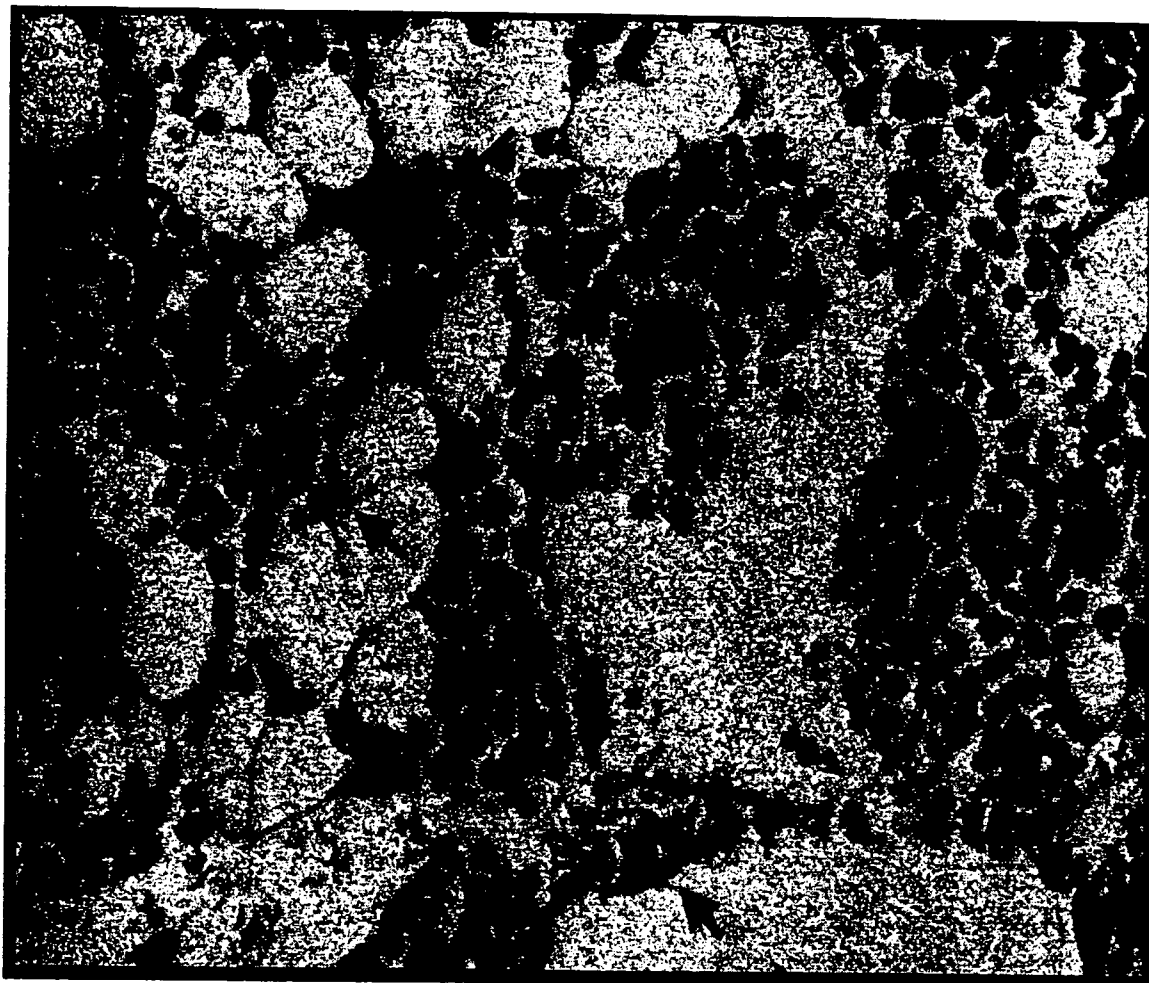


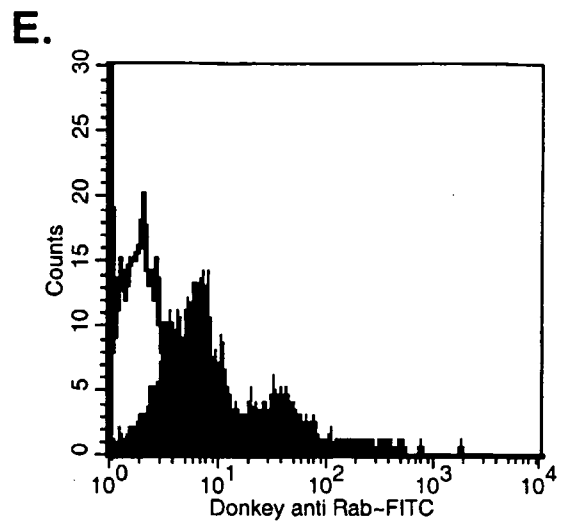
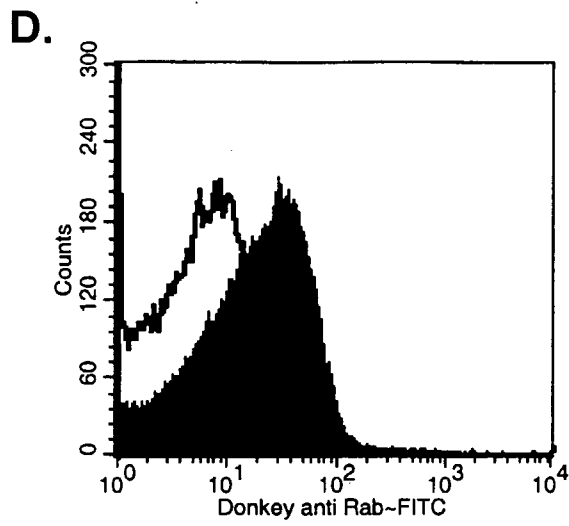
Figure 4A and 4B





**FIG. 4C**

Figure 4D and 4E





**Figure 5B**

Wild type aa	236	V	K	V	F	I	I	I	A	V	F	F																					
Wild type nucleotide		G	T	C	A	A	G	T	T	T	T	C	A	T	T	A	T	C	A	T	T	G	C	T	G	T	A	T	T	C	T	T	T
Mutant nucleotide		G	T	C	A	A	G	T	T	T	T	-	-	T	T	A	T	C	A	T	T	G	C	T	G	T	A	T	T	C	T	T	.
Mutant aa	236	V	K	V	F			Y	H	C	C	I	L																				

Figure 5C

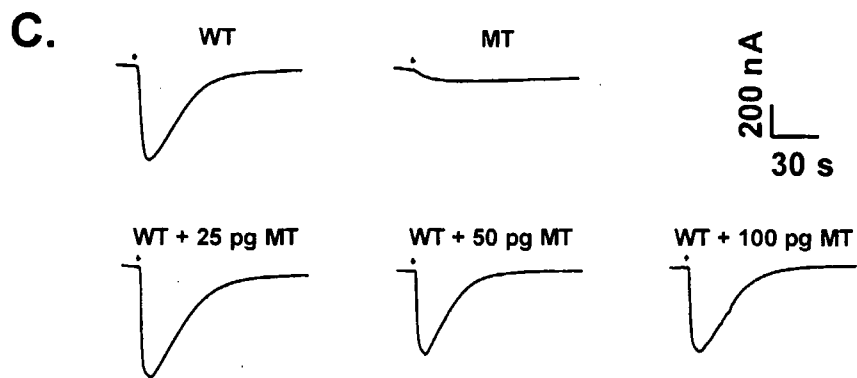
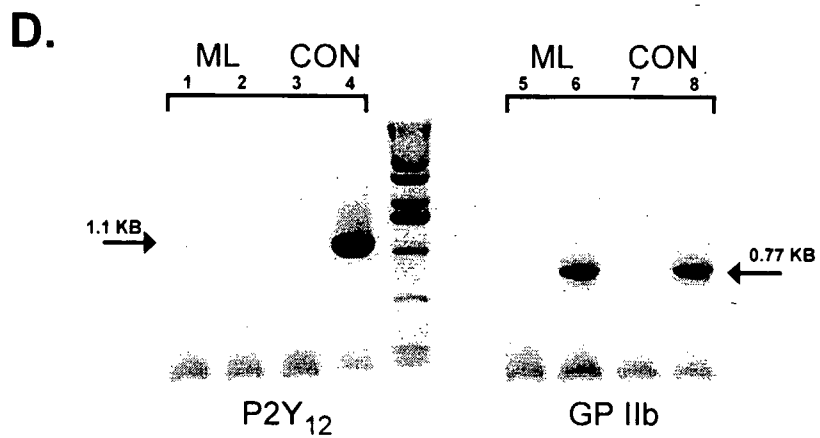
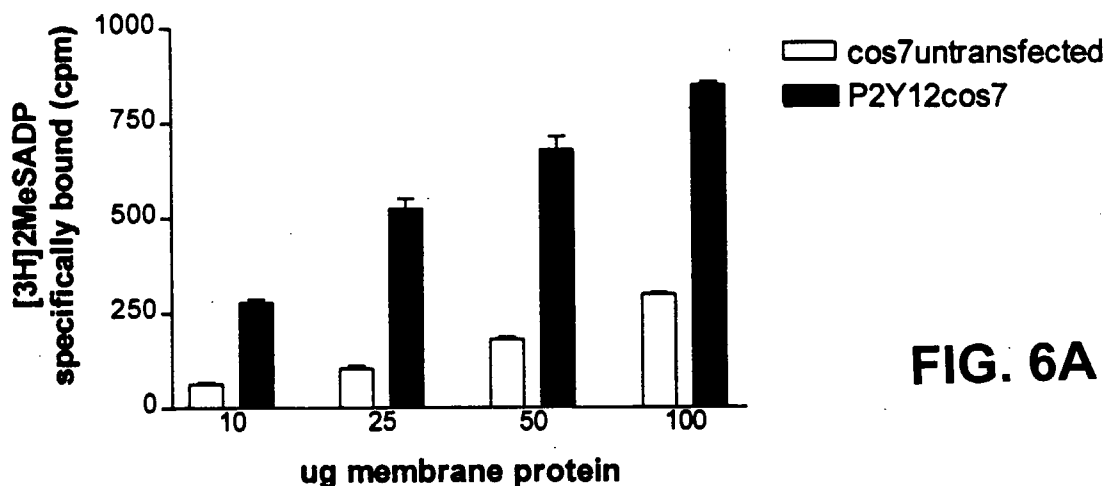


Figure 5D

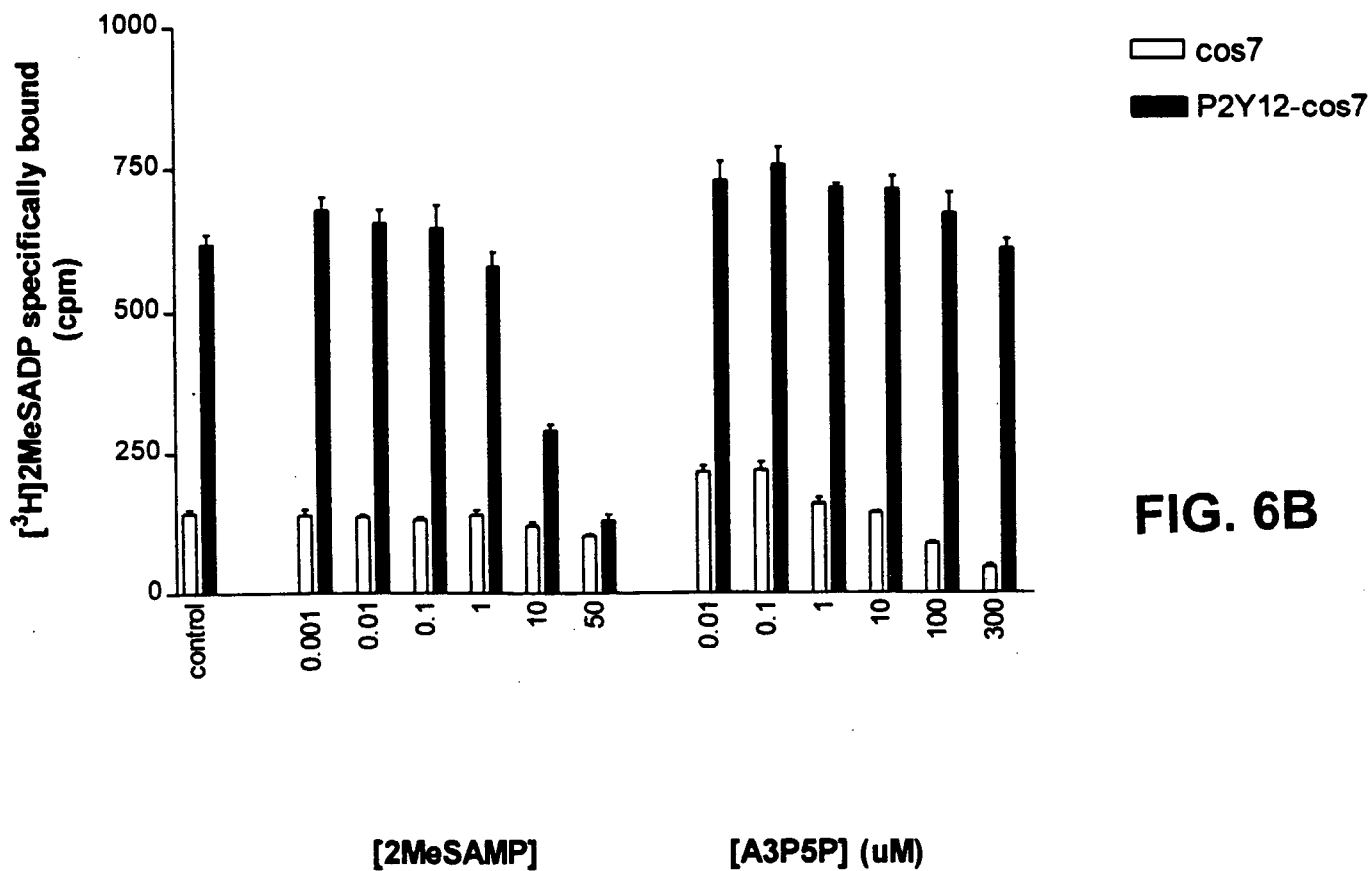


**[<sup>3</sup>H]2MeSADP Binding to Membranes of COS7 cells transiently transfected with hP2Y12**



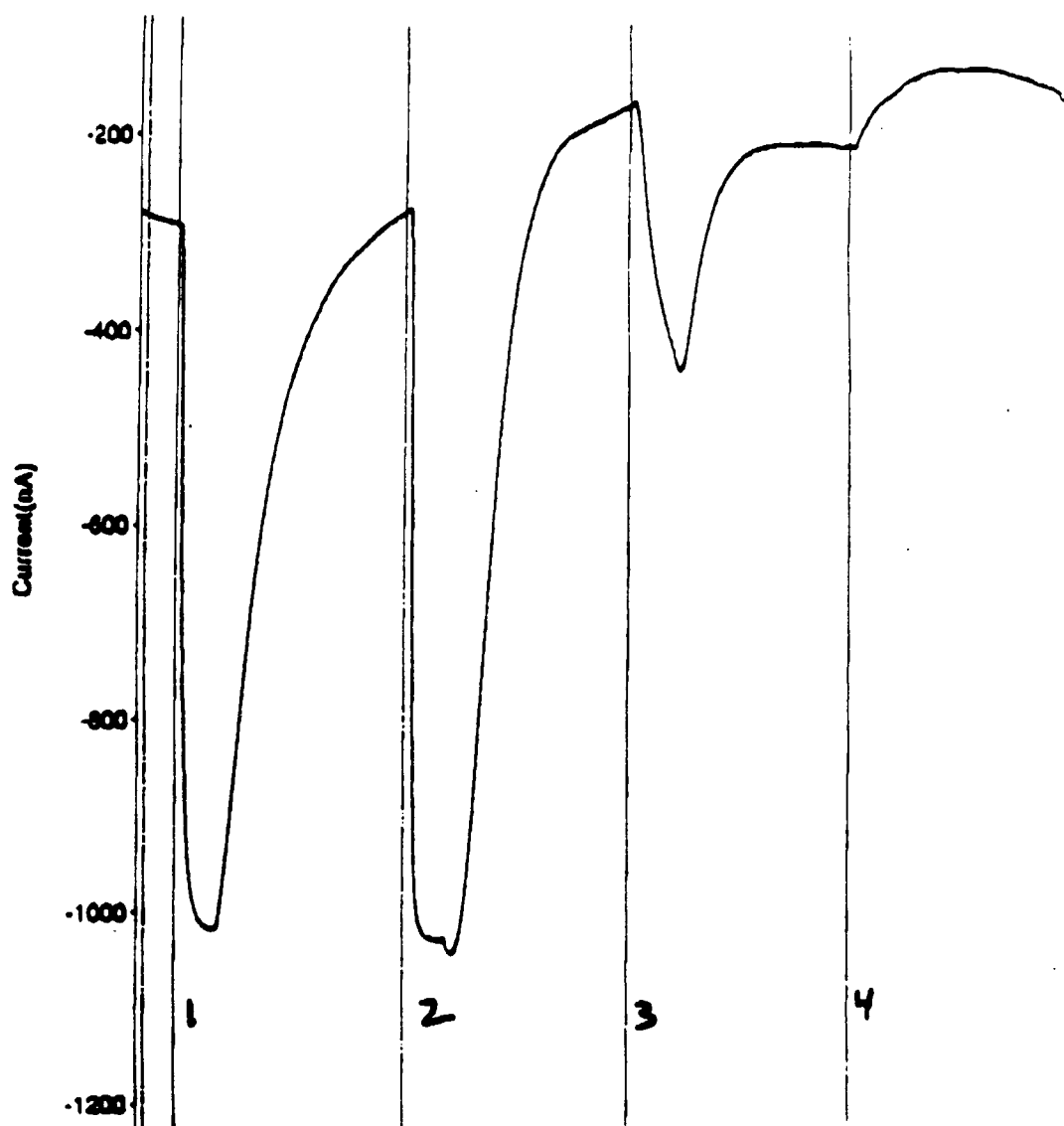
**FIG. 6A**

**Competition by 2MeSAMP and A3P5P of 1 nM [<sup>3</sup>H]2MeSADP binding to membranes from cos7 cells transfected with hP2Y12**



**FIG. 6B**

Response of oocytes injected with cRNA for H11 GPCR and GIRK1 & 4 cRNA's



drug applications:

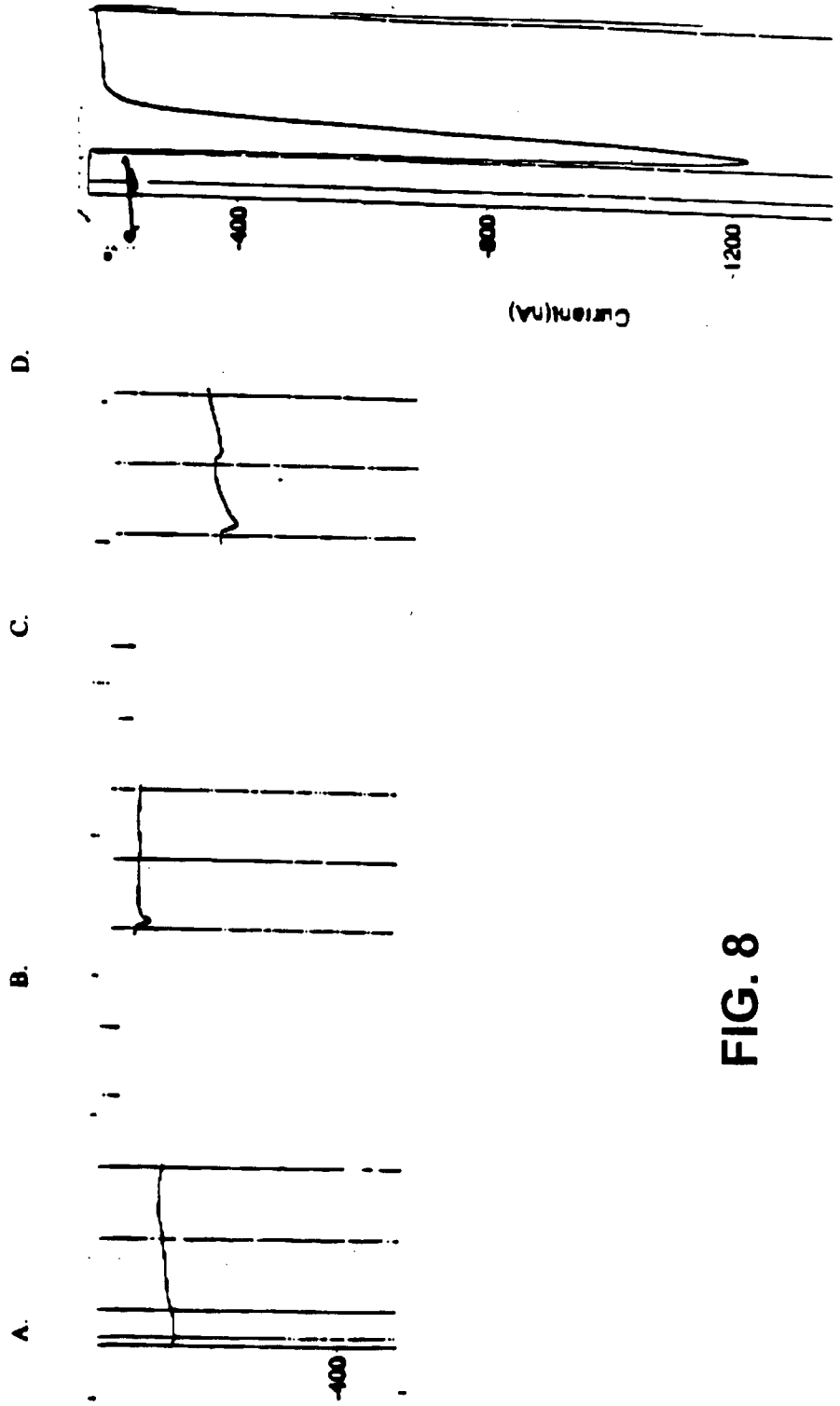
- 1)  $1\mu\text{M}$  ADP 30s
- 2)  $1\mu\text{M}$  ADP +  $10\mu\text{M}$  A3P5P 30s
- 3)  $1\mu\text{M}$  ADP +  $10\mu\text{M}$  2MeSAMP 30s
- 4)  $1\mu\text{M}$  ADP +  $10\mu\text{M}$  CT compound 30s

FIG. 7

**Effect of pertussis toxin on response of oocytes injected with H11, GIRK 1 & 4 cRNA's  
on measured GIRK current (nanoAmps)**

- A. Water injected oocyte
- B. Oocyte injected with H11 cRNA alone
- C. Oocyte injected with H11, GIRK 1&4 cRNA's plus 25 ng of pertussis toxin cRNA
- D. Oocyte injected with H11, GIRK 1&4 cRNA's

(vertical lines indicate addition of ADP as agonist)



**FIG. 8**

SEQUENCE LISTING

<110> Conley, Pamela B.  
 Jantzen, Hans-Michael  
 Ramakrishnan-DuBridge, Vanitha  
 Julius, David  
 Hollopeter, Gunter  
 COR Therapeutics, Inc.

<120> P2Y12 Receptor

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 <151> 1999-12-23

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 accaggacc atg gag gtg cct ggt gcc aac gcc acc tca gcc aac acc acc 171  
 Met Glu Val Pro Gly Ala Asn Ala Thr Ser Ala Asn Thr Thr  
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 Ser Ile Pro Gly Thr Ser Thr Leu Cys Ser Arg Asp Tyr Lys Ile Thr  
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 cag gtt ctc ttc oca ttg ctc tac act gtc ctg ttt ttt gct ggg ctc 267  
 Gln Val Leu Phe Pro Leu Leu Tyr Thr Val Leu Phe Phe Ala Gly Leu  
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Ser Asn Phe Ile Ile Phe Leu Lys Asn Thr Val Ile Ser Asp Leu Leu	
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atg atc cta act ttt cct ttc aaa att ctc agt gat gcc aaa ctg gga	411
Met Ile Leu Thr Phe Pro Phe Lys Ile Leu Ser Asp Ala Lys Leu Gly	
80	85 90
gct ggg cac ctg aga acc ctg gtg tgc caa gtc act tca gtc acg ttt	459
Ala Gly His Leu Arg Thr Leu Val Cys Gln Val Thr Ser Val Thr Phe	
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tac ttc aca atg tac atc agt atc tcg ttc ctc gga ttg ata acc att	507
Tyr Phe Thr Met Tyr Ile Ser Ile Ser Phe Leu Gly Leu Ile Thr Ile	
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gac cga tac ctg aag acc acc aga cca ttt aaa act tcc agc ccc agc	555
Asp Arg Tyr Leu Lys Thr Thr Arg Pro Phe Lys Thr Ser Ser Pro Ser	
130	135 140
aat ctt ttg ggt gcg aag att ctt tct gtt gcc atc tgg gcc ttc atg	603
Asn Leu Leu Gly Ala Lys Ile Leu Ser Val Ala Ile Trp Ala Phe Met	
145	150 155
ttc ctg ctg tca ctg cct aac atg att ctc acc aac agg agg cca aaa	651
Phe Leu Leu Ser Leu Pro Asn Met Ile Leu Thr Asn Arg Arg Pro Lys	
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Asp Lys Asp Ile Thr Lys Cys Ser Phe Leu Lys Ser Glu Phe Gly Leu	
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gtc tgg cac gag ata gtc aat tac atc tgc caa gtc att ttc tgg att	747
Val Trp His Glu Ile Val Asn Tyr Ile Cys Gln Val Ile Phe Trp Ile	
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Tyr Arg Ser Tyr Val Arg Thr Arg Gly Ser Ala Lys Ala Pro Lys Lys	
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Arg Val Asn Ile Lys Val Phe Ile Ile Ile Ala Val Phe Phe Ile Cys	
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ttc gtt ccc ttc cac ttt gca cgg att ccc tac acc ctg agc cag aca	939
Phe Val Pro Phe His Phe Ala Arg Ile Pro Tyr Thr Leu Ser Gln Thr	
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cgg gcc gtc ttt gac tgc aat gcc gag aac act ctg ttc tac gtg aag	987

Arg Ala Val Phe Asp Cys Asn Ala Glu Asn Thr Leu Phe Tyr Val Lys  
 275 280 285

gag agc acc ctg tgg ctg acg tcc ttg aac gcc tgc ctt gat cca ttc 1035  
 Glu Ser Thr Leu Trp Leu Thr Ser Leu Asn Ala Cys Leu Asp Pro Phe  
 290 295 300

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 Gly Asp Pro Ser Glu Glu Thr Pro Met  
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Phe Ile Ile Phe Leu Lys Asn Thr Val Ile Ser Asp Leu Leu Met Ile  
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Leu Thr Phe Pro Phe Lys Ile Leu Ser Asp Ala Lys Leu Gly Ala Gly  
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His Leu Arg Thr Leu Val Cys Gln Val Thr Ser Val Thr Phe Tyr Phe  
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Thr Met Tyr Ile Ser Ile Ser Phe Leu Gly Leu Ile Thr Ile Asp Arg  
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Tyr Leu Lys Thr Thr Arg Pro Phe Lys Thr Ser Ser Pro Ser Asn Leu  
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Leu Gly Ala Lys Ile Leu Ser Val Ala Ile Trp Ala Phe Met Phe Leu  
 145 150 155 160

Leu Ser Leu Pro Asn Met Ile Leu Thr Asn Arg Arg Pro Lys Asp Lys  
 165 170 175

Asp Ile Thr Lys Cys Ser Phe Leu Lys Ser Glu Phe Gly Leu Val Trp  
 180 185 190

His Glu Ile Val Asn Tyr Ile Cys Gln Val Ile Phe Trp Ile Asn Phe  
 195 200 205

Leu Ile Val Ile Val Cys Tyr Ser Leu Ile Thr Lys Glu Leu Tyr Arg  
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Ser Tyr Val Arg Thr Arg Gly Ser Ala Lys Ala Pro Lys Lys Arg Val  
 225 230 235 240

Asn Ile Lys Val Phe Ile Ile Ile Ala Val Phe Phe Ile Cys Phe Val  
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Pro Phe His Phe Ala Arg Ile Pro Tyr Thr Leu Ser Gln Thr Arg Ala  
 260 265 270

Val Phe Asp Cys Asn Ala Glu Asn Thr Leu Phe Tyr Val Lys Glu Ser  
 275 280 285

Thr Leu Trp Leu Thr Ser Leu Asn Ala Cys Leu Asp Pro Phe Ile Tyr  
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Phe Phe Leu Cys Lys Ser Phe Arg Asn Ser Leu Met Ser Met Leu Arg  
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Ser Phe Leu Gly Leu Ile Thr Ile Asp Arg Tyr Gln Lys Thr Thr Arg  
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Pro Phe Lys Thr Ser Asn Pro Lys Asn Leu Leu Gly Ala Lys Ile Leu  
 130 135 140

Ser Val Val Ile Trp Ala Phe Met Phe Leu Leu Ser Leu Pro Asn Met  
 145 150 155 160

Ile Leu Thr Asn Arg Gln Pro Arg Asp Lys Asn Val Lys Lys Cys Ser  
 165 170 175

Phe Leu Lys Ser Glu Phe Gly Leu Val Trp His Glu Ile Val Asn Tyr  
 180 185 190

Ile Cys Gln Val Ile Phe Trp Ile Asn Phe Leu Ile Val Ile Val Cys  
 195 200 205

Tyr Thr Leu Ile Thr Lys Glu Leu Tyr Arg Ser Tyr Val Arg Thr Arg  
 210 215 220

Gly Val Gly Lys Val Pro Arg Lys Lys Val Asn Val Lys Val Phe Ile  
 225 230 235 240

Ile Ile Ala Val Phe Phe Ile Cys Phe Val Pro Phe His Phe Ala Arg  
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Ile Pro Tyr Thr Leu Ser Gln Thr Arg Asp Val Phe Asp Cys Thr Ala  
 260 265 270

Glu Asn Thr Leu Phe Tyr Val Lys Glu Ser Thr Leu Trp Leu Thr Ser  
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Thr	Ser	Leu	Cys	Thr	Arg	Asp	Tyr	Lys	Ile	Thr	Gln	Val	Leu	Phe	Pro		
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ctg	ctc	tac	act	gtc	ctg	ttt	ttt	gtt	gga	ctt	atc	aca	aat	ggc	ctg		207
Leu	Leu	Tyr	Thr	Val	Leu	Phe	Phe	Val	Gly	Leu	Ile	Thr	Asn	Gly	Leu		
	30				35				40					45			
gcg	atg	agg	att	ttc	ttt	caa	atc	cgg	agt	aaa	tca	aac	ttt	att	att		255
Ala	Met	Arg	Ile	Phe	Phe	Gln	Ile	Arg	Ser	Lys	Ser	Asn	Phe	Ile	Ile		
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Phe	Leu	Lys	Asn	Thr	Val	Ile	Ser	Asp	Leu	Leu	Met	Ile	Leu	Thr	Phe		
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cca	ttc	aaa	att	ctt	agt	gat	gcc	aaa	ctg	gga	aca	gga	cca	ctg	aga		351
Pro	Phe	Lys	Ile	Leu	Ser	Asp	Ala	Lys	Leu	Gly	Thr	Gly	Pro	Leu	Arg		
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act	ttt	gtg	tgt	caa	gtt	acc	tcc	gtc	ata	ttt	tat	ttc	aca	atg	tat		399
Thr	Phe	Val	Cys	Gln	Val	Thr	Ser	Val	Ile	Phe	Tyr	Phe	Thr	Met	Tyr		
	95					100					105						
atc	agt	att	tca	ttc	ctg	gga	ctg	ata	act	atc	gat	cgc	tac	cag	aag		447
Ile	Ser	Ile	Ser	Phe	Leu	Gly	Leu	Ile	Thr	Ile	Asp	Arg	Tyr	Gln	Lys		
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Thr	Thr	Arg	Pro	Phe	Lys	Thr	Ser	Asn	Pro	Lys	Asn	Leu	Leu	Gly	Ala		
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Lys	Ile	Leu	Ser	Val	Val	Ile	Trp	Ala	Phe	Met	Phe	Leu	Leu	Ser	Leu		
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cct	aac	atg	att	ctg	acc	aac	agg	cag	ccg	aga	gac	aag	aat	gtg	aag		591
Pro	Asn	Met	Ile	Leu	Thr	Asn	Arg	Gln	Pro	Arg	Asp	Lys	Asn	Val	Lys		
		160					165					170					
aaa	tgc	tct	ttc	ctt	aaa	tca	gag	ttc	ggt	cta	gtc	tgg	cat	gaa	ata		639
Lys	Cys	Ser	Phe	Leu	Lys	Ser	Glu	Phe	Gly	Leu	Val	Trp	His	Glu	Ile		
	175					180					185						
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Val	Asn	Tyr	Ile	Cys	Gln	Val	Ile	Phe	Trp	Ile	Asn	Phe	Leu	Ile	Val		
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Ile	Val	Cys	Tyr	Thr	Leu	Ile	Thr	Lys	Glu	Leu	Tyr	Arg	Ser	Tyr	Val		
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Phe Ala Arg Ile Pro Tyr Thr Leu Ser Gln Thr Arg Asp Val Phe Asp			
255	260	265	
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Cys Thr Ala Glu Asn Thr Leu Phe Tyr Val Lys Glu Ser Thr Leu Trp			
270	275	280	285
tta act tcc tta aat gca tgc ctg gat ccg ttc atc tat ttt ttc ctt			975
Leu Thr Ser Leu Asn Ala Cys Leu Asp Pro Phe Ile Tyr Phe Phe Leu			
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Cys Lys Ser Phe Arg Asn Ser Leu Ile Ser Met Leu Lys Cys Pro Asn			
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Ser Ala Thr Ser Leu Ser Gln Asp Asn Arg Lys Lys Glu Gln Asp Gly			
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Gly Asp Pro Asn Glu Glu Thr Pro Met			
335	340		
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acc agt ctg tgc acc aga gac tac aaa atc acc cag gtc ctc ttc cca 159
Thr Ser Leu Cys Thr Arg Asp Tyr Lys Ile Thr Gln Val Leu Phe Pro
      15             20             25

ctg ctc tac act gtc ctg ttt ttt gtt gga ctt atc aca aat ggc ctg 207
Leu Leu Tyr Thr Val Leu Phe Phe Val Gly Leu Ile Thr Asn Gly Leu
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gcg atg agg att ttc ttt caa atc cgg agt aaa tca aac ttt att att 255
Ala Met Arg Ile Phe Phe Gln Ile Arg Ser Lys Ser Asn Phe Ile Ile
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Phe Leu Lys Asn Thr Val Ile Ser Asp Leu Leu Met Ile Leu Thr Phe
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Pro Phe Lys Ile Leu Ser Asp Ala Lys Leu Gly Thr Gly Pro Leu Arg
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act ttt gtg tgt caa gtt acc tcc gtc ata ttt tat ttc aca atg tat 399
Thr Phe Val Cys Gln Val Thr Ser Val Ile Phe Tyr Phe Thr Met Tyr
      95             100            105

atc agt att tca ttc ctg gga ctg ata act atc gat cgc tac cag aag 447
Ile Ser Ile Ser Phe Leu Gly Leu Ile Thr Ile Asp Arg Tyr Gln Lys
      110            115            120            125

acc acc agg cca ttt aaa aca tcc aac ccc aaa aat ctc ttg ggg gct 495
Thr Thr Arg Pro Phe Lys Thr Ser Asn Pro Lys Asn Leu Leu Gly Ala
      130            135            140

aag att ctc tct gtt gtc atc tgg gca ttc atg ttc tta ctc tct ttg 543
Lys Ile Leu Ser Val Val Ile Trp Ala Phe Met Phe Leu Leu Ser Leu
      145            150            155

cct aac atg att ctg acc aac agg cag ccg aga gac aag aat gtg aag 591
Pro Asn Met Ile Leu Thr Asn Arg Gln Pro Arg Asp Lys Asn Val Lys
      160            165            170

aaa tgc tct ttc ctt aaa tca gag ttc ggt cta gtc tgg cat gaa ata 639
Lys Cys Ser Phe Leu Lys Ser Glu Phe Gly Leu Val Trp His Glu Ile
      175            180            185

gta aat tac atc tgt caa gtc att ttc tgg att aat ttc tta att gtt 687
Val Asn Tyr Ile Cys Gln Val Ile Phe Trp Ile Asn Phe Leu Ile Val
      190            195            200            205
    
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att gta tgt tat aca ctc att aca aaa gaa ctg tac cgg tca tac gta 735  
 Ile Val Cys Tyr Thr Leu Ile Thr Lys Glu Leu Tyr Arg Ser Tyr Val  
 210 215 220

aga acg agg ggt gta ggt aaa gtc ccc agg aaa aag gtg aac gtc aaa 783  
 Arg Thr Arg Gly Val Gly Lys Val Pro Arg Lys Lys Val Asn Val Lys  
 225 230 235

gtt ttt tat cat tgc tgt att ctt tat ttg ttt tgt tcc ttt cca ttt 831  
 Val Phe Tyr His Cys Cys Ile Leu Tyr Leu Phe Cys Ser Phe Pro Phe  
 240 245 250

tgc ccg aat tcc tta cac cct gag cca aac ccg gga tgt ctt 873  
 Cys Pro Asn Ser Leu His Pro Glu Pro Asn Pro Gly Cys Leu  
 255 260 265

tgactgcact gctgaaaata ctctgttcta tgtgaaagag agcactctgt ggtaacttc 933

cttaaagtca tgccctggatc cgttcatcta ttttttcctt tgcaagtcct tcagaaattc 993

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aaaagaacag gatggtggtg acccaaatga agagactcca atgtaaacaa attaactaag 1113

gaaatatttc aatctctttg tgttcagaac tcgttaaagc aaagcgctaa gtaaaaatat 1173

taactgacga agaagcaact aagttaataa taatgactct aaagaaacag aagattacaa 1233

aagcaatfff catttacctt tccagtatga aaagctatct taaaatatag aaaactaatc 1293

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Cys Thr Arg Asp Tyr Lys Ile Thr Gln Val Leu Phe Pro Leu Leu Tyr  
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Thr Val Leu Phe Phe Val Gly Leu Ile Thr Asn Gly Leu Ala Met Arg  
 35 40 45

Ile Phe Phe Gln Ile Arg Ser Lys Ser Asn Phe Ile Ile Phe Leu Lys  
 50 55 60

Asn Thr Val Ile Ser Asp Leu Leu Met Ile Leu Thr Phe Pro Phe Lys  
 65 70 75 80

Ile Leu Ser Asp Ala Lys Leu Gly Thr Gly Pro Leu Arg Thr Phe Val  
 85 90 95

Cys Gln Val Thr Ser Val Ile Phe Tyr Phe Thr Met Tyr Ile Ser Ile  
 100 105 110

Ser Phe Leu Gly Leu Ile Thr Ile Asp Arg Tyr Gln Lys Thr Thr Arg  
 115 120 125

Pro Phe Lys Thr Ser Asn Pro Lys Asn Leu Leu Gly Ala Lys Ile Leu  
 130 135 140

Ser Val Val Ile Trp Ala Phe Met Phe Leu Leu Ser Leu Pro Asn Met  
 145 150 155 160

Ile Leu Thr Asn Arg Gln Pro Arg Asp Lys Asn Val Lys Lys Cys Ser  
 165 170 175

Phe Leu Lys Ser Glu Phe Gly Leu Val Trp His Glu Ile Val Asn Tyr  
 180 185 190

Ile Cys Gln Val Ile Phe Trp Ile Asn Phe Leu Ile Val Ile Val Cys  
 195 200 205

Tyr Thr Leu Ile Thr Lys Glu Leu Tyr Arg Ser Tyr Val Arg Thr Arg  
 210 215 220

Gly Val Gly Lys Val Pro Arg Lys Lys Val Asn Val Lys Val Phe Tyr  
 225 230 235 240

His Cys Cys Ile Leu Tyr Leu Phe Cys Ser Phe Pro Phe Cys Pro Asn  
 245 250 255

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 260 265

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<220>  
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Asn Leu Leu Ile Thr Gln Gln Ile Ile Pro Val Leu Tyr Cys Met Val  
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Phe Ile Ala Gly Ile Leu Leu Asn Gly Val Ser Gly Trp Ile Phe Phe  
 35 40 45

Tyr Val Pro Ser Ser Lys Ser Phe Ile Ile Tyr Leu Lys Asn Ile Val  
 50 55 60

Ile Ala Asp Phe Val Met Ser Leu Thr Phe Pro Phe Lys Ile Leu Gly  
 65 70 75 80

Asp Ser Gly Leu Gly Pro Trp Gln Leu Asn Val Phe Val Cys Arg Val  
 85 90 95

Ser Ala Val Leu Phe Tyr Val Asn Met Tyr Val Ser Ile Val Phe Phe  
 100 105 110

Gly Leu Ile Ser Phe Asp Arg Tyr Tyr Lys Ile Val Lys Pro Leu Trp  
 115 120 125

Thr Ser Phe Ile Gln Ser Val Ser Tyr Ser Lys Leu Leu Ser Val Ile  
 130 135 140

Val Trp Met Leu Met Leu Leu Leu Ala Val Pro Asn Ile Ile Leu Thr  
 145 150 155 160

Asn Gln Ser Val Arg Glu Val Thr Gln Ile Lys Cys Ile Glu Leu Lys  
 165 170 175

Ser Glu Leu Gly Arg Lys Trp His Lys Ala Ser Asn Tyr Ile Phe Val  
 180 185 190

Ala Ile Phe Trp Ile Val Phe Leu Leu Leu Ile Val Phe Tyr Thr Ala  
 195 200 205

Ile Thr Lys Lys Ile Phe Lys Ser His Leu Lys Ser Ser Arg Asn Ser  
 210 215 220

Thr Ser Val Lys Lys Lys Ser Ser Arg Asn Ile Phe Ser Ile Val Phe  
 225 230 235 240

Val Phe Phe Val Cys Phe Val Pro Tyr His Ile Ala Arg Ile Pro Tyr  
 245 250 255

Thr Lys Ser Gln Thr Glu Ala His Tyr Ser Cys Gln Ser Lys Glu Ile  
 260 265 270

Leu Arg Tyr Met Lys Glu Phe Thr Leu Leu Leu Ser Ala Ala Asn Val  
 275 280 285

Cys Leu Asp Pro Ile Ile Tyr Phe Phe Leu Cys Gln Pro Phe Arg Glu  
 290 295 300

Ile Leu Cys Lys Lys Leu His Ile Pro Leu Lys Ala Gln Asn Asp Leu  
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Thr Leu

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<212> PRT

<213> Homo sapiens

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 Phe Gln Phe Tyr Tyr Leu Pro Ala Val Tyr Ile Leu Val Phe Ile Ile  
 50 55 60  
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 Lys Pro Trp Ser Gly Ile Ser Val Tyr Met Phe Asn Leu Ala Leu Ala  
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 Asp Phe Leu Tyr Val Leu Thr Leu Pro Ala Leu Ile Phe Tyr Tyr Phe  
 100 105 110  
 Asn Lys Thr Asp Trp Ile Phe Gly Asp Ala Met Cys Lys Leu Gln Arg  
 115 120 125  
 Phe Ile Phe His Val Asn Leu Tyr Gly Ser Ile Leu Phe Leu Thr Cys  
 130 135 140  
 Ile Ser Ala His Arg Tyr Ser Gly Val Val Tyr Pro Leu Lys Ser Leu  
 145 150 155 160  
 Gly Arg Leu Lys Lys Lys Asn Ala Ile Cys Ile Ser Val Leu Val Trp  
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 Leu Ile Val Val Val Ala Ile Ser Pro Ile Leu Phe Tyr Ser Gly Thr  
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 Gly Val Arg Lys Asn Lys Thr Ile Thr Cys Tyr Asp Thr Thr Ser Asp  
 195 200 205  
 Glu Tyr Leu Arg Ser Tyr Phe Ile Tyr Ser Met Cys Thr Thr Val Ala  
 210 215 220  
 Met Phe Cys Val Pro Leu Val Leu Ile Leu Gly Cys Tyr Gly Leu Ile  
 225 230 235 240  
 Val Arg Ala Leu Ile Tyr Lys Asp Leu Asp Asn Ser Pro Leu Arg Arg  
 245 250 255

Lys Ser Ile Tyr Leu Val Ile Ile Val Leu Thr Val Phe Ala Val Ser  
260 265 270

Tyr Ile Pro Phe His Val Met Lys Thr Met Asn Leu Arg Ala Arg Leu  
275 280 285

Asp Phe Gln Thr Pro Ala Met Cys Ala Phe Asn Asp Arg Val Tyr Ala  
290 295 300

Thr Tyr Gln Val Thr Arg Gly Leu Ala Ser Leu Asn Ser Cys Val Asp  
305 310 315 320

Pro Ile Leu Tyr Phe Leu Ala Gly Asp Thr Phe Arg Arg Arg Leu Ser  
325 330 335

Arg Ala Thr Arg Lys Ala Ser Arg Arg Ser Glu Ala Asn Leu Gln Ser  
340 345 350

Lys Ser Glu Asp Met Thr Leu Asn Ile Leu Pro Glu Phe Lys Gln Asn  
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Gly Asp Thr Ser Leu  
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<210> 15

<211> 374

<212> PRT

<213> Meleagris gallopavo

<220>

<223> Turkey P2Y nucleotide receptor; tp2ynovel

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Pro Thr Pro Trp Leu Gly Gly Asn Thr Thr Ala Ala Ala Glu Ala Lys  
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Cys Val Phe Asn Glu Glu Phe Lys Phe Ile Leu Leu Pro Ile Ser Tyr  
35 40 45

Gly Ile Val Phe Val Val Gly Leu Pro Leu Asn Ser Trp Ala Met Trp  
50 55 60

Ile Phe Val Ser Arg Met Arg Pro Trp Asn Ala Thr Thr Thr Tyr Met  
65 70 75 80

Phe Asn Leu Ala Ile Ser Asp Thr Leu Tyr Val Phe Ser Leu Pro Thr  
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Leu Val Tyr Tyr Tyr Ala Asp Arg Asn Asn Trp Pro Phe Gly Lys Val  
100 105 110

Phe Cys Lys Ile Val Arg Phe Leu Phe Tyr Ala Asn Leu Tyr Ser Ser



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<223> P2Y4 pyrimidinergic receptor

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 Phe Lys Phe Ile Leu Leu Pro Val Ser Tyr Ala Val Val Phe Val Leu  
 35 40 45  
 Gly Leu Gly Leu Asn Ala Pro Thr Leu Trp Leu Phe Ile Phe Arg Leu  
 50 55 60  
 Arg Pro Trp Asp Ala Thr Ala Thr Tyr Met Phe His Leu Ala Leu Ser  
 65 70 75 80  
 Asp Thr Leu Tyr Val Leu Ser Leu Pro Thr Leu Ile Tyr Tyr Tyr Ala  
 85 90 95  
 Ala His Asn His Trp Pro Phe Gly Thr Glu Ile Cys Lys Phe Val Arg  
 100 105 110  
 Phe Leu Phe Tyr Trp Asn Leu Tyr Cys Ser Val Leu Phe Leu Thr Cys  
 115 120 125  
 Ile Ser Val His Arg Tyr Leu Gly Ile Cys His Pro Leu Arg Ala Leu  
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 Arg Trp Gly Arg Pro Arg Leu Ala Gly Leu Leu Cys Leu Ala Val Trp  
 145 150 155 160  
 Leu Val Val Ala Gly Cys Leu Val Pro Asn Leu Phe Phe Val Thr Thr  
 165 170 175  
 Ser Asn Lys Gly Thr Thr Val Leu Cys His Asp Thr Thr Arg Pro Glu  
 180 185 190  
 Glu Phe Asp His Tyr Val His Phe Ser Ser Ala Val Met Gly Leu Leu  
 195 200 205  
 Phe Gly Val Pro Cys Leu Val Thr Leu Val Cys Tyr Gly Leu Met Ala  
 210 215 220  
 Arg Arg Leu Tyr Gln Pro Leu Pro Gly Ser Ala Gln Ser Ser Ser Arg  
 225 230 235 240  
 Leu Arg Ser Leu Arg Thr Ile Ala Val Val Leu Thr Val Phe Ala Val  
 245 250 255  
 Cys Phe Val Pro Phe His Ile Thr Arg Thr Ile Tyr Tyr Leu Ala Arg  
 260 265 270  
 Leu Leu Glu Ala Asp Cys Arg Val Leu Asn Ile Val Asn Val Val Tyr



Val Leu Ala Cys Gln Ala Pro Val Leu Tyr Phe Val Thr Thr Ser Ala  
 165 170 175

Arg Gly Gly Arg Val Thr Cys His Asp Thr Ser Ala Pro Glu Leu Phe  
 180 185 190

Ser Arg Phe Val Ala Tyr Ser Ser Val Met Leu Gly Leu Leu Phe Ala  
 195 200 205

Val Pro Phe Ala Val Ile Leu Val Cys Tyr Val Leu Met Ala Arg Arg  
 210 215 220

Leu Leu Lys Pro Ala Tyr Gly Thr Ser Gly Gly Leu Pro Arg Ala Lys  
 225 230 235 240

Arg Lys Ser Val Arg Thr Ile Ala Val Val Leu Ala Val Phe Ala Leu  
 245 250 255

Cys Phe Leu Pro Phe His Val Thr Arg Thr Leu Tyr Tyr Ser Phe Arg  
 260 265 270

Ser Leu Asp Leu Ser Cys His Thr Leu Asn Ala Ile Asn Met Ala Tyr  
 275 280 285

Lys Val Thr Arg Pro Leu Ala Ser Ala Asn Ser Cys Leu Asp Pro Val  
 290 295 300

Leu Tyr Phe Leu Ala Gly Gln Arg Leu Val Arg Phe Ala Arg Asp Ala  
 305 310 315 320

Lys Pro Pro Thr Gly Pro Ser Pro Ala Thr Pro Ala Arg Arg Arg Leu  
 325 330 335

Gly Leu Arg Arg Ser Asp Arg Thr Asp Met Gln Arg Ile Gly Asp Val  
 340 345 350

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Ser Glu Asn Thr Lys Asp Ile Arg Leu  
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<220>  
 <223> P2Y6 receptor

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Thr	Leu	Asn	Leu	Ala	Leu	Ala	Asp	Leu	Leu	Tyr	Ala	Cys	Ser	Leu	Pro		
65					70					75					80		
Leu	Leu	Ile	Tyr	Asn	Tyr	Ala	Gln	Gly	Asp	His	Trp	Pro	Phe	Gly	Asp		
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Phe	Ala	Cys	Arg	Leu	Val	Arg	Phe	Leu	Phe	Tyr	Ala	Asn	Leu	His	Gly		
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Ser	Ile	Leu	Phe	Leu	Thr	Cys	Ile	Ser	Phe	Gln	Arg	Tyr	Leu	Gly	Ile		
	115						120					125					
Cys	His	Pro	Leu	Ala	Pro	Trp	His	Lys	Arg	Gly	Gly	Arg	Arg	Ala	Ala		
130						135					140						
Trp	Leu	Val	Cys	Val	Ala	Val	Trp	Leu	Ala	Val	Thr	Thr	Gln	Cys	Leu		
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				165					170					175			
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Gly	Met	Ala	Leu	Thr	Val	Ile	Gly	Phe	Leu	Leu	Pro	Phe	Ala	Ala	Leu		
	195						200					205					
Leu	Ala	Cys	Tyr	Cys	Leu	Leu	Ala	Cys	Arg	Leu	Cys	Arg	Gln	Asp	Gly		
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Pro	Ala	Glu	Pro	Val	Ala	Gln	Glu	Arg	Arg	Gly	Lys	Ala	Ala	Arg	Met		
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Ala	Val	Val	Val	Ala	Ala	Ala	Phe	Ala	Ile	Ser	Phe	Leu	Pro	Phe	His		
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Ile	Thr	Lys	Thr	Ala	Tyr	Leu	Ala	Val	Arg	Ser	Thr	Pro	Gly	Val	Pro		
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Gln	Lys	Lys	Phe	Arg	Arg	Arg	Pro	His	Glu	Leu	Leu	Gln	Lys	Leu	Thr		
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325

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<212> PRT  
<213> Homo sapiens

<220>  
<223> P2Y11 purinergic receptor

<400> 19

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Val Val Glu Phe Leu Val Ala Val Ala Ser Asn Gly Leu Ala Leu Tyr  
35 40 45  
Arg Phe Ser Ile Arg Lys Gln Arg Pro Trp His Pro Ala Val Val Phe  
50 55 60  
Ser Val Gln Leu Ala Val Ser Asp Leu Leu Cys Ala Leu Thr Leu Pro  
65 70 75 80  
Pro Leu Ala Ala Tyr Leu Tyr Pro Pro Lys His Trp Arg Tyr Gly Glu  
85 90 95  
Ala Ala Cys Arg Leu Glu Arg Phe Leu Phe Thr Cys Asn Leu Leu Gly  
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Ser Val Ile Phe Ile Thr Cys Ile Ser Leu Asn Arg Tyr Leu Gly Ile  
115 120 125  
Val His Pro Phe Phe Ala Arg Ser His Leu Arg Pro Lys His Ala Trp  
130 135 140  
Ala Val Ser Ala Ala Gly Trp Val Leu Ala Ala Leu Leu Ala Met Pro  
145 150 155 160  
Thr Leu Ser Phe Ser His Leu Lys Arg Pro Gln Gln Gly Ala Gly Asn  
165 170 175  
Cys Ser Val Ala Arg Pro Glu Ala Cys Ile Lys Cys Leu Gly Thr Ala  
180 185 190  
Asp His Gly Leu Ala Ala Tyr Arg Ala Tyr Ser Leu Val Leu Ala Gly  
195 200 205  
Leu Gly Cys Gly Leu Pro Leu Leu Leu Thr Leu Ala Ala Tyr Gly Ala  
210 215 220  
Leu Gly Arg Ala Val Leu Arg Ser Pro Gly Met Thr Val Ala Glu Lys



Asn Met Tyr Leu Ser Ile Ile Phe Leu Ala Phe Val Ser Ile Asp Arg  
 100 105 110

Cys Leu Gln Leu Thr His Ser Cys Lys Ile Tyr Arg Ile Gln Glu Pro  
 115 120 125

Gly Phe Ala Lys Met Ile Ser Thr Val Val Trp Leu Met Val Leu Leu  
 130 135 140

Ile Met Val Pro Asn Met Met Ile Pro Ile Lys Asp Ile Lys Glu Lys  
 145 150 155 160

Ser Asn Val Gly Cys Met Glu Phe Lys Lys Glu Phe Gly Arg Asn Trp  
 165 170 175

His Leu Leu Thr Asn Phe Ile Cys Val Ala Ile Phe Leu Asn Phe Ser  
 180 185 190

Ala Ile Ile Leu Ile Ser Asn Cys Leu Val Ile Arg Gln Leu Tyr Arg  
 195 200 205

Asn Lys Asp Asn Glu Asn Tyr Pro Asn Val Lys Lys Ala Leu Ile Asn  
 210 215 220

Ile Leu Leu Val Thr Thr Gly Tyr Ile Ile Cys Phe Val Pro Tyr His  
 225 230 235 240

Ile Val Arg Ile Pro Tyr Thr Leu Ser Gln Thr Glu Val Ile Thr Asp  
 245 250 255

Cys Ser Thr Arg Ile Ser Leu Phe Lys Ala Lys Glu Ala Thr Leu Leu  
 260 265 270

Leu Ala Val Ser Asn Leu Cys Phe Asp Pro Ile Leu Tyr Tyr His Leu  
 275 280 285

Ser Lys Ala Phe Arg Ser Lys Val Thr Glu Thr Phe Ala Ser Pro Lys  
 290 295 300

Glu Thr Lys Ala Gln Lys Glu Lys Leu Arg Cys Glu Asn Asn Ala  
 305 310 315

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 <223> G protein-coupled receptor 34; hugpr34

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	50					55					60				
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Gly	Ile	His	Arg	Lys	Arg	Asn	Ser	Ile	Gln	Ile	Tyr	Leu	Leu	Asn	Val
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Ala	Ile	Ala	Asp	Leu	Leu	Leu	Ile	Phe	Cys	Leu	Pro	Phe	Arg	Ile	Met
			100					105					110		
Tyr	His	Ile	Asn	Gln	Asn	Lys	Trp	Thr	Leu	Gly	Val	Ile	Leu	Cys	Lys
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Val	Val	Gly	Thr	Leu	Phe	Tyr	Met	Asn	Met	Tyr	Ile	Ser	Ile	Ile	Leu
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Leu	Gly	Phe	Ile	Ser	Leu	Asp	Arg	Tyr	Ile	Lys	Ile	Asn	Arg	Ser	Ile
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Gln	Gln	Arg	Lys	Ala	Ile	Thr	Thr	Lys	Gln	Ser	Ile	Tyr	Val	Cys	Cys
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Ile	Val	Trp	Met	Leu	Ala	Leu	Gly	Gly	Phe	Leu	Thr	Met	Ile	Ile	Leu
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Thr	Leu	Lys	Lys	Gly	Gly	His	Asn	Ser	Thr	Met	Cys	Phe	His	Tyr	Arg
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Asp	Lys	His	Asn	Ala	Lys	Gly	Glu	Ala	Ile	Phe	Asn	Phe	Ile	Leu	Val
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Val	Met	Phe	Trp	Leu	Ile	Phe	Leu	Leu	Ile	Ile	Leu	Ser	Tyr	Ile	Lys
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Ile	Gly	Lys	Asn	Leu	Leu	Arg	Ile	Ser	Lys	Arg	Arg	Ser	Lys	Phe	Pro
				245					250					255	
Asn	Ser	Gly	Lys	Tyr	Ala	Thr	Thr	Ala	Arg	Asn	Ser	Phe	Ile	Val	Leu
			260					265					270		
Ile	Ile	Phe	Thr	Ile	Cys	Phe	Val	Pro	Tyr	His	Ala	Phe	Arg	Phe	Ile
		275					280					285			
Tyr	Ile	Ser	Ser	Gln	Leu	Asn	Val	Ser	Ser	Cys	Tyr	Trp	Lys	Glu	Ile
	290					295					300				
Val	His	Lys	Thr	Asn	Glu	Ile	Met	Leu	Val	Leu	Ser	Ser	Phe	Asn	Ser
305					310					315					320

Cys Leu Asp Pro Val Met Tyr Phe Leu Met Ser Ser Asn Ile Arg Lys  
 325 330 335  
 Ile Met Cys Gln Leu Leu Phe Arg Arg Phe Gln Gly Glu Pro Ser Arg  
 340 345 350  
 Ser Glu Ser Thr Ser Glu Phe Lys Pro Gly Tyr Ser Leu His Asp Thr  
 355 360 365  
 Ser Val Ala Val Lys Ile Gln Ser Ser Ser Lys Ser Thr  
 370 375 380

**INTERNATIONAL SEARCH REPORT**

International application No.

PCT/US00/34998

**A. CLASSIFICATION OF SUBJECT MATTER**

IPC(7) : C12P 21/06, G01N 33/53, C12N 5/06, 5/16, C07K 1/00  
 US CL : 435/69.1, 7.1, 334, 530/350, 387.7

According to International Patent Classification (IPC) or to both national classification and IPC

**B. FIELDS SEARCHED**

Minimum documentation searched (classification system followed by classification symbols)  
 U.S. : 435/69.1, 7.1, 334, 530/350, 387.7

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)  
 Please See Continuation Sheet

**C. DOCUMENTS CONSIDERED TO BE RELEVANT**

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 98/50549 A2 (HUMAN GENOME SCIENCES, INC.) 12 November 1998, SEQ ID No: 2 pages 42-43	1 and 6-13
Y	BOYER et al. Identification of Competitive Antagonists of the P2Y1 Receptor. Molecular Pharmacology. November 1996, Vol. 50, pages 1323-1329, especially pages 1324-1327.	33-45
Y	FABRE et al. Decreased platelet aggregation, increased bleeding time and resistance to thromboembolism in P2Y1-deficient mice. Nature Medicine. October 1999, Vol. 5, No. 10, pages 1199-1202, especially pages 1199-1201.	49-51
T,E	HOLLOPETER et al. Identification of the platelet ADP receptor targeted by antithrombotic drugs. Nature. January 2001, Vol. 409, pages 202-207.	1-14, 18-29, 33-45 and 49-51

Further documents are listed in the continuation of Box C.

See patent family annex.

\* Special categories of cited documents:

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier application or patent published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

- "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
- "&" document member of the same patent family

Date of the actual completion of the international search

21 February 2001 (21.02.2001)

Date of mailing of the international search report

**APR 30 2001**

Name and mailing address of the ISA/US  
 Commissioner of Patents and Trademarks  
 Box PCT  
 Washington, D.C. 20231  
 Facsimile No. (703)305-3230

Authorized officer  
 Jegatheesan Seharaseyon

Telephone No. 703-308-0196

# INTERNATIONAL SEARCH REPORT

International application No.

PCT/US00/34998

## Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1.  Claim Nos.:  
because they relate to subject matter not required to be searched by this Authority, namely:
  
2.  Claim Nos.:  
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
  
3.  Claim Nos.: 15 and 30  
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

## Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:  
Please See Continuation Sheet

1.  As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
  2.  As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
  3.  As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.: 1-14, 18-29,33-45 and 49-51
  
  4.  No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
- Remark on Protest**  The additional search fees were accompanied by the applicant's protest.  
 No protest accompanied the payment of additional search fees.

# INTERNATIONAL SEARCH REPORT

International application No.

PCT/US00/34998

**BOX II. OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING** This application contains the following inventions or groups of inventions which are not so linked as to form a single general inventive concept under PCT Rule 13.1. In order for all inventions to be examined, the appropriate additional examination fees must be paid.

Group I, claim(s) 1-12 and 18-27, drawn to a nucleic acid, the polypeptide encoded by the nucleic acid, an expression vector containing the nucleic acid, a host cell expressing the polypeptide, a cell line transformed with the vector and a method of making the polypeptide..

Group II, claim(s) 13, 14, 28, 29 and 49-51 drawn to isolated receptor polypeptide.

Group III, claim(s) 16, 17, 31 and 32 drawn to an antibody.

Group IV, claim(s) 33-45, drawn to a method of identifying an agent, which modulates expression.

Group V, claim(s) 46, drawn to a method of generating a transgenic animal

Group VI, claim(s) 47,52-55 and 57, drawn to a method of diagnosing using the nucleic acid expression

Group VII, claim(s) 48 and 56, drawn to a method of diagnosing using the protein expression

Inventions I, II and III are compositions and are different the methods IV-VII.

The compositions of Inventions I,II and III are different from each other as they are directed to nonequivalent types of compounds with different chemical characters. Invention I is nucleic acid and a cell containing it. Invention II is a polypeptide. Invention III is an antibody. Inventions IV-VII are different, from each other as they are directed to nonequivalent methods. Invention IV is a method to identify an agent to modulate expression. Invention V uses polynucleotide to generate transgenic animals. Invention VI uses polynucleotide and Invention VII uses polypeptide for diagnosis.

The claims of these groups are directed to different inventions, which are not linked to form a single general inventive concept under PCT Rule 13.1. The claims in the different groups lack the same or corresponding special technical features. In particular, each group is directed to different compounds and /or methods. Accordingly, the claims are not so linked by a special technical feature within the meaning of PCT Rule 13.2 so as to form a single inventive concept and lack of unity is deemed proper

**Continuation of B. FIELDS SEARCHED Item 3:** STN search on CAPLUS, MEDLIN, BIOSIS, EUROPATFULL, PCTFULL,USPATFULL

WEST search on JPO and USPAT

Search terms: P2Y12 Receptor and G protein coupled receptor

专利名称(译)	P2y12受体		
公开(公告)号	<a href="#">EP1240350A1</a>	公开(公告)日	2002-09-18
申请号	EP2000986695	申请日	2000-12-26
[标]申请(专利权)人(译)	加利福尼亚大学董事会		
申请(专利权)人(译)	COR THERAPEUTICS , INC. 加利福尼亚大学董事会		
当前申请(专利权)人(译)	加利福尼亚大学董事会 PORTOLA制药公司.		
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IPC分类号	C07K14/705 A61K38/00 C07K16/28 G01N33/50 G01N33/74 C12P21/06 C07K1/00 C12N5/06 C12N5/16 G01N33/53		
CPC分类号	G01N33/5008 A61K38/00 C07K14/705 C07K16/28 G01N33/502 G01N33/5091 G01N33/74 G01N2333/726 G01N2500/02		
优先权	60/171622 1999-12-23 US		
其他公开文献	EP1240350A4		
外部链接	<a href="#">Espacenet</a>		

#### 摘要(译)

本发明包括P2-嘌呤能受体的新亚型，称为P2Y12受体。该受体在血小板和脑中选择性表达，并与百日咳毒素敏感的G蛋白 (Gi) 偶联。还公开了编码受体的核酸和相关的筛选和治疗方法。