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(54) **INSULIN-LIKE GROWTH FACTOR 1 RECEPTOR -SPECIFIC ANTIBODIES AND USES THEREOF**

FÜR INSULINÄHNLICHEN WACHSTUMSFAKTOR-1-REZEPTOR SPEZIFISCHE ANTIKÖRPER UND VERWENDUNGEN DAVON

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(74) Representative: **Fleuchaus, Andrea et al**
Fleuchaus & Gallo Partnerschaft mbB
Patent- und Rechtsanwälte
Steinerstrasse 15/A
81369 München (DE)

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(73) Proprietor: **National Research Council of Canada**
Ottawa, Ontario K1A 0R6 (CA)

- (72) Inventors:
- **STANIMIROVIC Danica**
Ottawa, Ontario K4P 1R7 (CA)
 - **KEMMERICH Kristin**
Ottawa, Ontario K1Y 0A5 (CA)
 - **HAQQANI, Arsalan S.**
Kanata, Ontario K2T 0A8 (CA)
 - **SULEA, Traian**
Kirkland, Quebec H9J 3A2 (CA)
 - **ARBABI-GHAHROUDI, Mehdi**
Ottawa, Ontario K1W 1K1 (CA)
 - **MASSIE Bernard**
Laval, Québec H7H 1R2 (CA)
 - **GILBERT Rénald**
Montreal, Québec H4V 2B4 (CA)

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- **STANIMIROVIC DANICA ET AL: "Engineering and pharmacology of blood-brain barrier-permeable bispecific antibodies", ADVANCES IN PHARMACO, ELSEVIER, USA, vol. 71, 1 January 2014 (2014-01-01), pages 301-335, XP009188677, ISSN: 1557-8925, DOI: 10.1016/BS.APHA.2014.06.005**
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- **GABATHULER R: 'Approaches to transport therapeutic drugs across the blood-brain barrier to treat brain diseases.' NEUROBIOL DIS. vol. 37, no. 1, January 2010, ISSN 0969-9961 pages 48 - 57, XP026765785**

DescriptionFIELD OF THE INVENTION

5 **[0001]** The present invention relates to Insulin-Like Growth Factor 1 Receptor-specific antibody fragments and uses thereof. More specifically, the present invention relates to Insulin-Like Growth Factor 1 Receptor-specific antibody fragments that transigrate the blood-brain barrier, and uses thereof.

BACKGROUND OF THE INVENTION

10 **[0002]** Neurodegenerative diseases, such as Alzheimer's and Parkinson's disease, are an increasing burden on our ageing society because there are currently no effective treatments for these disabling conditions. Treatment as well as early diagnosis of these and other diseases that originate in the brain remain challenging because the majority of suitable therapeutic molecules and diagnostics cannot penetrate the tight and highly restrictive blood-brain barrier (BBB) (Abbott, 2013). The BBB constitutes a physical barricade that is formed by brain endothelial cells (BECs) that line the blood vessels and connect with each other through tight junctions (Abbott, 2013). The tight junctions formed between the BECs are essential for the integrity of the BBB and prevent the paracellular transport of molecules larger than 500 daltons (Da). Because brain endothelial cells exhibit very low pinocytosis rates (Abbott, 2013), transcellular transport of larger molecules is limited to the highly specific receptor mediated transcytosis (RMT) pathway, and the passive, charge-based adsorption mediated transcytosis (Abbott, 2013; Pardridge, 2002). Additionally, the high density of efflux pumps, such as P-glycoprotein or the multi-drug resistance protein -1 (MDR-1), contribute to the removal of unwanted substances from the brain (Abbott, 2013).

20 **[0003]** While all these characteristics protect the brain from pathogens and toxins, they equally prevent the entry of most therapeutics. In fact, less than 5% of small molecule therapeutics and virtually none of the larger therapeutics can cross the BBB in pharmacologically relevant concentrations (i.e., sufficient to engage a central nervous system (CNS) target and elicit pharmacologic/therapeutic response) unless they are specifically 'ferried', that is, coupled to a transporter molecule. Due to the lack of effective 'carriers' to transport molecules across the BBB, numerous drugs against neurodegenerative diseases have been 'shelved' or eliminated from further development as they cannot be delivered to the brain in sufficient amount.

25 **[0004]** Different approaches to deliver larger molecules into the brain have been explored. For example, the integrity of the BBB may be disrupted, resulting in a leaky BBB, which in turn allows for unrestricted, paracellular entry of larger molecules into the brain. Tight junctions can be successfully loosened or disrupted by various approaches. For example, injection of substances that induce osmotic shock (for example, mannitol, hypertonic solutions) into the blood stream causes cell shrinkage and results in the disruption of tight junctions, therefore severely compromising the BBB (Guillaume, 2010). Other modulators of tight junctions include alkylglycerols, bradykinin and several analogues thereof, as well as viruses that modulate expression of proteins involved in maintaining the tight junctions (Erdlenbruch et al., 2003; Preston et al., 2008; Gan et al., 2013). A more localized disruption of the BBB is possible through application of ultrasound (Nhan et al., 2013). However, the periods during which the BBB is disrupted are sufficient to alter brain homeostasis and allow harmful chemicals, toxins and pathogens to enter the brain; this can result in serious side-effects, e.g., seizures and brain swelling, infection and possibly permanent neuropathological changes. As would be evident to those of skill in the art, repeated treatments with these techniques for chronic and diffuse brain diseases affecting multiple brain regions are not practical. Most of these treatments are costly, necessitate hospitalisation, and some approaches require anesthesia.

35 **[0005]** Another approach for circumventing the BBB is direct injection of therapeutic molecules into the cerebrospinal fluid (CSF), the parenchymal space, or other parts of the brain. Several delivery methods have been developed, including: intracerebral (intra-parenchymal), intraventricular, and intrathecal delivery via infusion or convection-enhanced diffusion (CED) pumps. However, any type of direct injection into the brain or intracerebral implant is an invasive and costly procedure, as it requires hospitalization, anesthesia, and often surgery. Moreover, the poor diffusion rates of the therapeutics, particularly large biologics, within brain parenchyma limit the penetration of therapeutics to only small areas surrounding the site of injection/implantation. The correct placement of injections, catheters, and implants is challenging yet crucial to achieve diffusion of the drug to the targeted region of the brain. Additionally, catheters and implants provide a site for infection and/or immune response against the foreign material.

40 **[0006]** In another attempt to increase delivery across the BBB, CNS drugs have been modified to increase their brain uptake. Such modifications can include a change of their surface charge, a reduction in molecule size, and change to the lipophilicity of the drugs. However, any modifications to increase brain penetration are also likely to alter the overall pharmacology of the drug, including its desired activity and/or specificity. In addition, lipophilic molecules are more prone to being exported from the brain through the P-glycoprotein efflux pump.

45 **[0007]** Finally, endogenous transport mechanisms across the BBB have been exploited. Physiological mechanisms

that allow transport of large molecules across the BBB can be divided into the highly specific receptor mediated transcytosis (RMT) and the non-specific charge based adsorptive mediated endocytosis pathways. Endocytosis is triggered upon binding of the specific ligand to its receptor, or upon electrostatic interaction between the cationic ligand or drug and the anionic functional groups on the brain endothelial cell surface (luminal side), respectively. Subsequently, the newly formed endosome is transcytosed across the cell to the abluminal side, to release its cargo.

[0008] Because adsorptive mediated transcytosis is non-specific, charge-mediated interaction, it occurs in all vascular beds and organs, limiting the availability of drug for brain delivery. Therefore, exploiting the RMT pathway remains the only physiological, non-invasive yet highly receptor-specific brain delivery method.

[0009] Only a few receptors are presently known to undergo RMT at the BBB and 'ferry' across their natural ligands: the well-studied transferrin receptor (TfR), the insulin receptor (IR), low-density lipoprotein receptor related proteins 1 and 2 (LRP-1 and -2), diphtheria toxin receptor, and TMEM30A. Peptides, natural ligands, and antibodies or antibody fragments have been developed that bind to these receptors (Pardridge et al., 1991; Yu et al., 2011; Muruganandam et al., 2001; Abulrob et al., 2005; Demeule, 2008; Sumbria et al., 2013), functioning as drug-to-brain transporters that utilize endogenous RMT pathways. However, to date only a single peptide (Angiopep ANG1005, targeting LRP-1) has been analyzed in phase I clinical studies, while other candidates are being studied in laboratory settings. The RMT pathway appears to be the most promising pathway for drug transport to the brain, but current approaches have limitations, including: non-selective expression of the target receptor at the BBB, competition between the carrier and the natural ligands to the receptor, ineffective transcytosis of a receptor as well as lysosomal degradation of endocytosed carriers (Xiao and Gun, 2013). IGF1R-binding peptides have been described as part of fusion proteins to mediate crossing the blood-brain barrier in WO2011/15061, whilst WO2011/044542 describes fusions of antibodies to a payload to cross the blood-brain barrier, whereby the antibody can amongst others be directed to IGF1R. Antibodies specifically binding IGF1R in general are known from e.g. WO2011/066721.

[0010] The lack of high-capacity and high-selectivity BBB carriers delays the development of new therapeutics and diagnostics for diseases originating in the brain, including brain tumors and neurodegenerative diseases. There is clearly a need for a non-invasive method to deliver small and large therapeutic and diagnostic molecules in pharmacologically efficacious doses into the brain without disrupting the physiology and homeostasis of the BBB.

SUMMARY OF THE INVENTION

[0011] The present invention relates to Insulin-Like Growth Factor 1 Receptor (IGF1R) -specific antibody fragments and uses thereof. More specifically, the present invention relates to Insulin-Like Growth Factor 1 Receptor-specific antibody fragments that transmigrate the blood-brain barrier, and uses thereof.

[0012] The present invention provides isolated or purified antibody fragments specifically binding to an Insulin-Like Growth Factor 1 Receptor (IGF1 R) epitope, wherein the antibody fragment transmigrates the blood-brain barrier, and wherein the epitope is specifically bound by the antibody of SEQ ID NO:5. The IGF1R epitope may be in the extracellular domain of IGF1R, and may comprise the sequence FENFLHNSIFVPR (SEQ ID NO:11), or fragments thereof.

[0013] The present invention further provides an isolated or purified antibody fragment, comprising

a complementarity determining region (CDR) 1 sequence of GRTIDNYA (SEQ ID NO:1);
 a CDR2 sequence of IDWGDGGX (SEQ ID NO:2), where X is A or T; and
 a CDR3 sequence of AMARQSRVNLDVARYDY (SEQ ID NO:3),

wherein the antibody fragment specifically binds the Insulin-Like Growth Factor 1 Receptor (IGF1R). In one embodiment, the isolated or purified antibody fragment comprises a CDR2 sequence of IDWGDGGA.

[0014] For example, and without wishing to be limiting in any manner, the isolated or purified antibody fragment specific for IGF1R may be

X₁VX₂LX₃ESGGGLVQX₄GGSLRLSCAASGRTIDNYAMAWX₅RQAPGKX₆X₇EX₈VX₉TID
 WGDGGX₁₀RYANSVKGRFTISRDNX₁₁KX₁₂TX₁₃YLQMNX₁₄LX₁₅X₁₆EDTAVYX₁₇CAMAR
 QSRVNLDVARYDYWGQGTX₁₈VTVSS (SEQ ID NO:4), where X₁ is E or Q; X₂ is K or Q; X₃ is V or E; X₄ is A or P; X₅
 is V or S; X₆ is D or G; X₇ is L or R; X₈ is F or W; X₉ is A or S; X₁₀ is A or T; X₁₁ is A or S; X₁₂ is G or N; X₁₃ is M or L;
 X₁₄ is N or R; X₁₅ is E or R; X₁₆ is P or A; X₁₇ is S or Y; and X₁₈ is Q or L,
 or a sequence substantially identical thereto. In more specific, non-limiting examples, the isolated or purified antibody
 fragment may comprise a sequence selected from the group consisting of:

QVKLEESGGGLVQAGGSLRLSCAASGRTIDNYAMAWSRQAPGKDFVATIDWGDG
 GARYANSVKGRFTISRDNKGTMYLQMNLEPEDTAVYSCAMARQSRVNLDVARYD YWQGTQVTVSS (SEQ
 ID NO:5), referred to herein as IGF1R-5;
 EVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWVRQAPGKGLEWVSTIDWGD

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GGTRYANSVKGRFTISRDN SKNTLYLQMNSLRAEDTAVYYCAMARQSRVNL DVARYD YWGQGLTVTVSS (SEQ ID NO:6), referred to herein as IGF1R-5_H1;

QVQLVESGGGLVQP GGGSLRLS CAASGRTIDNYAMAWVRQAPGK GLEWVATIDWGD

5 GGTRYANSVKGRFTISRDN SKNTMYLQMNSLRAEDTAVYYCAMARQSRVNL DVARYDYWGQGLTVTVSS (SEQ ID NO:7), referred to herein as IGF1R-5_H2;

QVQLVESGGGLVQP GGGSLRLS CAASGRTIDNYAMAWSRQAPGK GLEFVATIDWGDG

GTRYANSVKGRFTISRDN SKNTMYLQMNSLRAEDTAVYYCAMARQSRVNL DVARYD YWGQGLTVTVSS (SEQ ID NO:8), referred to herein as IGF1R-5_H3;

QVQLVESGGGLVQP GGGSLRLS CAASGRTIDNYAMAWSRQAPGK DREFVATIDWGDG

10 GTRYANSVKGRFTISRDN SKGTMYLQMNSLRAEDTAVYSCAMARQSRVNL DVARYD YWGQGLTVTVSS (SEQ ID NO:9), referred to herein as IGF1R-5_H5; and

EVQLVESGGGLVQP GGGSLRLS CAASGRTIDNYAMAWSRQAPGK DREFVSTIDWGDG

GTRYANSVKGRFTISRDN SKNTLYLQMNSLRAEDTAVYYCAMARQSRVNL DVARYDY WGQGLTVTVSS (SEQ ID NO:10), referred to herein as IGF1R-5_H6,

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or a sequence substantially identical thereto.

[0015] The isolated or purified antibody fragment as described above may be a single-domain antibody (sdAb); the sdAb may be of camelid origin. The antibody fragment may bind to an epitope comprising the sequence FENFLHNSIFVPR (SEQ ID NO:11).

20 **[0016]** The isolated or purified antibody fragment of the present invention may be presented in a multivalent display format. In a multivalent display format, the antibody fragment may be linked to a Fc fragment; the Fc fragment may be the mouse Fc2b or human Fc1. For example, and without wishing to be limiting in any manner, the isolated or purified antibody fragment in multivalent display may comprise the sequence of SEQ ID NO:42 (referred to herein as IGF1R-5-mFc consensus), or 43 (referred to herein as mFc-IGF1R-5 consensus), SEQ ID NO:12 (referred to herein as IGF1R-5-mFc) or 13 (referred to herein as mFc-IGF1R-5).

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[0017] The isolated or purified antibody fragment as described herein, in any format, may transmigrate the blood-brain barrier.

[0018] The present invention also provides a nucleic acid molecule encoding the isolated or purified antibody fragment as described herein. A vector comprising the nucleic acid molecule as just described is also provided.

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[0019] The isolated or purified antibody fragment as described herein may be immobilized onto a surface.

[0020] The present invention further provides the isolated or purified antibody fragment as described herein linked to a cargo molecule; the cargo molecule may have a molecular weight in the range of about 1kD to about 200 kDa. The cargo molecule linked to the antibody fragment may be a detectable agent, a therapeutic, a drug, a peptide, a growth factor, a cytokine, a receptor trap, a chemical compound, a carbohydrate moiety, an enzyme, an antibody or fragments thereof, a DNA-based molecule, a viral vector, or a cytotoxic agent; one or more liposomes or nanocarriers loaded with a detectable agent, a therapeutic, a drug, a peptide, an enzyme, and antibody or fragments thereof, a DNA-based molecule, a viral vector, or a cytotoxic agent; or one or more nanoparticle, nanowire, nanotube, or quantum dots. Additionally, the present invention provides a composition comprising one or more than one isolated or purified antibody fragment as described herein and a pharmaceutically-acceptable carrier, diluent, or excipient.

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[0021] An *in vitro* method of detecting IGF1R is also provided, the method comprising

a) contacting a tissue sample with one or more than one isolated or purified antibody fragment of as described herein linked to a detectable agent; and

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b) detecting the detectable agent linked to the antibody fragment bound to IGF1R in the tissue sample.

[0022] In the method described above, the sample may be a serum sample, a vascular tissue sample, tumour tissue sample, or a brain tissue sample from a human or animal subject. In the method as described, the step of detecting (step b)) may be performed using optical imaging, immunohistochemistry, molecular diagnostic imaging, ELISA, imaging mass spectrometry, or other suitable method.

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[0023] Further provided is an *in vivo* method of detecting IGF1R expression in a subject, the method comprising:

a) administering one or more than one isolated or purified antibody fragment as described herein linked to a detectable agent to the subject; and

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b) detecting the detectable agent linked to the antibody fragment bound to IGF1 R.

[0024] In the method described above, the step of detecting (step b)) may be performed using PET (positron emission tomography), SPECT (single-photon emission computed tomography), fluorescence imaging, or any other suitable

method.

[0025] Presently provided is a method of transporting a molecule of interest across the blood-brain barrier (BBB), the method comprising:

- 5 a) administering one or more than one isolated or purified antibody fragment as described herein linked to the molecule of interest to a subject, the isolated or purified antibody fragment transmigration the blood-brain barrier,

wherein the one or more than one antibody fragment ferries the molecule of interest across the BBB. In the method as just described, the molecule of interest may have a molecular weight in the range of about 1kD to about 200 kDa; the molecule of interest may be a detectable agent, a therapeutic, a drug, a peptide, a growth factor, a cytokine, a receptor trap, a chemical compound, a carbohydrate moiety, an enzyme, an antibody or fragment thereof, a DNA-based molecule, a viral vector, or a cytotoxic agent; one or more liposomes loaded with a detectable agent, a therapeutic, a drug, a peptide, an enzyme, an antibody or fragment thereof, a DNA-based molecule, a viral vector, or a cytotoxic agent; or one or more nanoparticle, nanowire, nanotube, or quantum dots. In the method as described, the administration may be intravenous (iv), subcutaneous (sc), or intramuscular (im).

[0026] The invention also encompasses a method of quantifying an amount of a cargo molecule delivered across the BBB of a subject, wherein the cargo molecule is linked to one or more than one isolated or purified antibody fragment as described herein, the method comprising

- 20 a) collecting cerebrospinal fluid (CSF) from the subject; and
b) using targeted proteomics methods to quantify the amount of the cargo molecule linked to one or more than one isolated or purified antibody fragment in the CSF.

[0027] The cargo molecule may be any desired molecule, including the cargo molecules previously described; the antibody fragment transmigrates the BBB; and the molecule may be "linked" to the antibody fragment as previously described. In the above method, the CSF is collected from a subject using any suitable method known in the art. The amount of CSF required for targeted proteomics method in step b) may be between about 1 to 10 μ l. The targeted proteomics methods used to quantify the amount of the one or more than antibody fragment linked to the cargo molecule may be any suitable method known in the art. For example and without wishing to be limiting, the targeted proteomics method may be a mass spectrometry method, such as multiple reaction monitoring - isotope labeled internal standards (MRM - ILIS).

[0028] The poor delivery of diagnostics or drugs across the tight and highly selective BBB compromises the development of treatments for brain diseases, such as, but not limiting to, brain tumors and neurodegenerative diseases. The lack of carriers to transport molecules across the BBB delays the development of new therapeutics and diagnostics for such diseases. As described herein, an IGF1R-binding V_HH has been produced that provides an effective transport platform for delivery of drugs conjugated to the antibody across the BBB to their targets in the brain. The presently described antibody fragments exploit the natural RMT pathway of the IGF1R from the luminal to abluminal side of the BBB-forming brain endothelial cells.

[0029] Following binding of the antibody fragment to IGF1R, RMT is initiated and the antibody fragment, together with a conjugated molecule (cargo), is transcytosed through the cell to the abluminal side where they are both released into the brain microenvironment. This is the first identification and demonstration of an anti-IGF1R binding domain antibody fragment that transmigrates the BBB by RMT.

[0030] The IGF1R V_HH was confirmed to bind to IGF1R (Fig. 2B), internalize into BBB cells (Fig. 4), and cross to abluminal side of the *in vitro* BBB model (Fig. 5B). Drug-to-brain delivery studies *in vivo* also showed that the IGF1R V_HH 'carried' a conjugated peptide (Galanin; about 3kDa) as well as a large protein fusion (about 80kDa) across the BBB (Fig. 7A and B; Fig. 9).

[0031] The results also show that the anti-IGF1R V_HH can be expressed in fusion with Fc (fragment crystallisable) fragment to prolong circulation half-life by about 75 fold (about 25h compared to about 20 min for V_HH alone). This high molecular weight fusion construct (about 80kDa) is also efficiently transported across the BBB. The long plasma half-life increases CSF exposure of the IGF1R V_HH-mFc (mFc = mouse Fc) conjugate significantly compared to the V_HH alone and is useful as a BBB delivery carrier for the treatment of chronic diseases with targets in the CNS. The conjugate is readily detected in brain parenchyma using immunofluorescence detection. The results demonstrate that the IGF1R V_HH carrier can "ferry" large molecules (similar in size to: antibodies, enzymes, growth factors, peptides, cytokines, receptor traps) across the BBB.

[0032] Thus, the antibody-delivery may not only be useful for short term treatment (e.g. epileptic seizure), but may also be useful for medium-term (e.g. cancer) and long-term (e.g. Alzheimer's or Parkinson's disease) treatments.

BRIEF DESCRIPTION OF THE DRAWINGS

[0033] These and other features of the invention will now be described by way of example, with reference to the appended drawings, wherein:

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 FIGURE 1 is a schematic diagram of the insulin-like growth factor 1 receptor (IGF1R). The IGF1R is found on the cell surface, and comprises two subunits: the alpha subunit and the beta subunit. The alpha subunit (comprising an extracellular part with the insulin-like growth factor 1 binding site) is connected by a disulphide bond to the beta subunit (comprising a small extracellular domain, a transmembrane region, and an intracellular portion). The IGF1 receptor can form a dimer. A 933 amino acid long fragment comprising the alpha subunit and the extracellular portion of the beta subunit, as indicated within the grey box, (M1 - F933, SwissProt Accession No. P08069; see FIGURE 2) was recombinantly produced and used for immunization of a llama.

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 FIGURE 2 shows the sequence of IGF1R (SwissProt Accession No. P08069). The 933 amino acid long protein fragment used for immunization and panning is shown in bold; the full ectodomain is 2 amino acids longer. The furin cleavage site, separating alpha and beta subunits, is shown in italicized lowercase letters. The signal peptide is shown in bold italics. The putative epitope to which IGF1R-5 V_HH might bind is underlined.

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 FIGURE 3A shows a size exclusion chromatogram of the IGF1R-binding V_HH IGF1R-5, run through a Superdex 75 column. The profile suggests that the V_HH is monomeric and non-aggregating. FIGURE 3B shows the melting temperature (T_m) as determined by circular dichroism (CD) for IGF1R-5-V_HH, and its humanized variants (labelled 5H1, 5H2, 5H3, 5H5, 5H6). The proteins were heated to above 90°C and measurements were taken in the CD instrument to determine the melting curve (black or filled circles) and the T_m. Subsequently, the proteins were cooled to room temperature, heated once more and analysed by CD (grey curve or squares). This allowed the determination of the fraction of refolded protein. FIGURE 3C shows a surface plasmon resonance (SPR) sensogram overlay for binding of 0.1 - 10nM IGF1R-5 V_HH and its humanized variants (labelled 5H1, 5H2, 5H3, 5H5, 5H6) to the recombinant extracellular portion of the human IGF1R fragment (h-IGF1R). The data obtained with this specific method and experimental set up fit well to a 1:1 model, giving K_D values in the low nanomolar (nM) range (0.6 - 16.4nM), as detailed in Table 2. FIGURE 3D shows SPR sensograms of IGF1R-5 V_HH binding to the recombinant extracellular portion of the human IGF1R fragment and subsequent addition of IGF1, which can simultaneously bind to the receptor. The experiment was repeated twice, as shown in the graph.

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 FIGURE 4 shows imaging results of cell uptake of Cy-5.5-labeled IGF1R-5 V_HH and control V_HH. Cy5.5-labeled IGF1R-5 V_HH (or FC5 V_HH as a positive control; Muruganandam et al., 2002; Haqqani et al., 2012) were incubated with SV40 immortalized rat brain endothelial cells (svARBEC) at 4°C (top panels) or at 37°C (bottom panels) to assess whether IGF1R-5 is internalized passively (4°C) or through active mechanisms (37°C) such as receptor mediated endocytosis. Co-staining with wheat germ agglutinin and DAPI was carried out to visualize the cell surface and the nucleus, respectively. Top panel: When incubated at 4°C, IGF1R-5 and FC5 V_HH were found outside the cells (arrowheads), bound to cell membrane (co-localized with the wheat germ agglutinin). Bottom panel: At 37°C both FC5 and IGF1R-5 V_HH accumulated inside the cells in vesicle-like structures (arrowheads), likely endosomes, suggesting internalization through an active transport mechanism.

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 FIGURE 5A shows the sequence for the C-terminal fusion of IGF1R-5 V_HH with the murine Fc fragment (IGF1R-5-mFc) while FIGURE 5B shows the N-terminal fusion sequence (mFc-IGF1R-5-Gal). A schematic representation of the assembled fusion protein is shown in FIGURE 5C. The IGF1R-5 V_HH are shown in black and the murine Fc (CH2 and CH3) are shown in grey in Figure 5 A-C.

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 FIGURE 6A is a flowchart summarizing the use of the *in vitro* BBB model to assess the ability of various V_HHs to cross the BBB. Equimolar amounts (5.6 μM or 1.25 μM) of positive (FC5) and negative control (A20.1, a *Clostridium difficile* toxin A-binding V_HH; and EG2, an EGFR binding V_HH) V_HHs and IGF1R-5 were tested simultaneously for their ability to cross a rat *in vitro* BBB model. SV40-immortalized brain endothelial cells from adult rat (svARBECs) are grown in a monolayer on the membrane of an insert in the presence of rat astrocyte-conditioned medium in the bottom chamber and standard medium in the top chamber. Following co-addition of equimolar amounts of the various V_HH to the luminal side of the BBB model, samples were taken from the bottom chamber after 15, 30 and 60min. The concentrations of each V_HH were then quantified by mass spectrometry (multiple reaction monitoring - isotope labeled internal standards; MRM-ILIS) in these samples. The P_{app} value [Qr/dt = cumulative amount in the receiver compartment versus time; A = area of the cell monolayer; C₀ = initial concentration of the dosing solution] is commonly used to determine the ability of a molecule to cross the BBB. FIGURE 6B shows the P_{app} values of the four co-

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administered V_{HH} . IGF1R-5 has a similar P_{app} value to FC5, while the negative controls have both very low P_{app} values, indicating facilitated transport of FC5 and IGF1R5 V_{HH} compared to very low non-specific transport or paracellular transport of control V_{HH} s. The results are average P_{app} values obtained in 5-6 independent experiments. FIGURE 6C shows the P_{app} values of the humanized IGF1R-5 single-domain antibodies (H1, H2, H3, H5, H6) (black bars), compared to A20.1 V_{HH} (white bars), which was tested as a control in the same well; the average FC5 and A20.1 value is indicated by a grey dotted line. FIGURE 6D shows the P_{app} values of the N-terminal and C-terminal fusion of IGF1R-5 to mouse Fc (light grey), compared to the A20.1-mouse Fc (A20.1mFc) (dark grey), which was tested in the same well as control and has a very low P_{app} value. Average P_{app} values of FC5-Fc (FC5 fused to an Fc), FC5 (V_{HH}), IGF1R-5 (V_{HH}) and A20.1 (V_{HH}) are indicated by dotted lines for comparison.

FIGURE 7 shows the scheme for chemical synthesis of IGF1R-5 V_{HH} -Galanin conjugate. IGF1R-5 was first conjugated to the NHS group of a Sulfo-SMCC cross-linker (1); then maleimide-activated IGF1R-5-sulfo-SMCC was conjugated to the reduced cysteine of Galanin (2).

FIGURE 8 shows IGF1R-5-mediated brain delivery of the chemically conjugated peptide Galanin. FIGURE 8A is a graph showing the ability of IGF1R-5 to deliver pharmacologically efficacious doses of the conjugated analgesic peptide Galanin (3.2 kD) into the brain using Hargreaves pain model. In this model, localised chronic pain is induced in male Wistar rats (4-6 weeks age), by injecting 100 μ l of complete Freund's adjuvant (CFA) into the left plantar surface, causing a local inflammation within a few hours. Following tail vein injection of the BBB carrier V_{HH} -drug conjugate or Galanin alone, the rats are placed into Plexiglas enclosures set on a glass surface. A thermal stimulus is focused on the inflamed or contralateral paw via an angled mirror. The latency between stimulus application and paw withdrawal (lick or flick of paw) is interpreted as a measure of the analgesic effect (inhibition of thermal hyperalgesia). The peptide Galanin alone cannot penetrate the BBB, as demonstrated by the lack of analgesic effect after systemic (tail-vein) injection of 3 mg/kg Galanin (grey solid circles). Systemic injection of IGF1R-5-Galanin conjugate (2.93 mg/kg or 5.85 mg/kg) induced a dose-dependent analgesic effect over a 3 hour period, more pronounced than that observed with the 6mg/kg of FC5-Galanin conjugate. FIGURE 8B shows these results as area under the curve (AUC) of pharmacodynamics response compared to the maximal possible effect (MPE; control paw). 2.93 mg/kg IGF1R-5-Galanin induced 28% of MPE, while 5.85 mg/kg IGF1R-5-Galanin induced 55% of MPE over a 3 hour period, demonstrating a significant brain penetration of the conjugate compared to the Galanin alone after systemic injection (<5% of MPE). FIGURE 8C is a graph showing the ability of the mFc-IGF1R-5-Gal to suppress thermal hyperalgesia in Hargreaves pain model after intravenous injection different concentrations. FIGURE 8D shows a dose-dependent response (percent of reversal at peak response) of thermal hyperalgesia in Hargreaves model after iv injection of either IGF1R-5-mFc-Gal or mFc-IGF1 R-5-Gal conjugate.

FIGURE 9 shows plasma and CSF pharmacokinetics of IGF1R-5-mFc and control V_{HH} , A20.1 mouse Fc fusion (A20.1-mFc) after systemic (tail vein) administration of 5 mg/kg. The cisterna magna was cannulated for serial CSF collections (1 - 48 h). Plasma and CSF levels of IGF1R-5-mFc and A20.1-mFc were determined using the MRM-ILIS method that 'tracks' and quantifies specific protein peptide signatures. Albumin levels in the CSF were concurrently determined by MRM. All CSF samples having a plasma/CSF ratio lower than 1500 were excluded as potentially blood-contaminated. The plasma half-life of IGF1R-5-mFc and A20.1-mFc was similar (23 h and 25 h, respectively). The plasma/CSF ratio was 2% and 0.04% for IGF1R-5-mFc and A20.1-mFc, respectively, 24 h after systemic injection of the fusion protein.

FIGURE 10 shows immuno-detection of IGF1R-5-mFc in brain sections 48 h after tail-vein administration of a 5 mg/kg dose. Sacrifice perfusion with PBS was carried out on the rats and brain sections (35 μ m) were obtained using the vibratome. IGF1R-5-mFc was immuno-detected using an anti-mouse Fc antibody. Blood vessels in the cortex section were detected using lectin RCA1 (top-left). Astrocytes were detected in the same section by an antibody against glial fibrillary acidic protein (GFAP) (top-right). IGF1R-5-mFc could be detected in both vessels and outside the vessels (i.e. in the brain parenchyma, transmigrated across the BBB) (bottom-left) as indicated by arrowheads. A composite micrograph of all stainings is shown in the bottom-right, indicating the transmigration of IGF1R-5-mFc from the luminal side of the blood vessel, across the BBB into the brain parenchyma.

FIGURE 11 shows that IGF1R-5 does not interfere with insulin or IGF-1 signaling through either the insulin receptor or IGF1R. FIGURE 11A is a representative Western blot showing that neither IGF1R-5, nor any of the other anti-IGF1R V_{HH} tested (IGF1R-1, -3, -4, or -6) induce downstream Akt phosphorylation alone at a concentration of 100nM. Neither does the presence of 100nM of IGF1R-5, or any of the other anti-IGF1R V_{HH} s inhibit Akt phosphorylation as induced by 10 μ g/ml of insulin. The quantitation of Western blot band densities from 3 independent experiments is shown in the bar graph (average \pm SD) below the gel image. FIGURE 11B is a representative

Western blot showing that neither IGF1R-5, nor any of the other anti-IGF1R V_HH tested (IGF1R, -3, -4, or -6) at 100nM induce phosphorylation of Akt on their own and neither inhibit IGF-1 induced Akt phosphorylation (i.e. signaling) induced upon stimulation with 200ng/ml of IGF-1. The quantitation of Western blot band densities from 3 independent experiments is shown in the bar graph (average +/-SD) below the gel image. FIGURE 11C shows Western blots probed for phosphorylated IGF1R. Cells were incubated with either 100nM or 500nM IGF1R-5, or any of the other anti-IGF1R V_HHs (IGF1R-1, -3, or -4) fused at their C-terminus to a murine Fc (e.g. IGF1R-5-mFc) alone or stimulated with 200ng/ml IGF-1 in presence of the respective IGF1R-V_HH-mFc fusion proteins. The Western blots indicate that none of the fusion constructs inhibited IGF-1 induced phosphorylation of IGF1R and neither induced receptor phosphorylation on their own.

DETAILED DESCRIPTION OF THE INVENTION

[0034] The present invention relates to Insulin-Like Growth Factor 1 Receptor-specific antibody fragments, and uses thereof. More specifically, the present invention relates to Insulin-Like Growth Factor 1 Receptor-specific antibody fragments that transigrate the blood-brain barrier, and uses thereof.

[0035] The present invention provides isolated or purified antibody fragments specifically binding to an Insulin-Like Growth Factor 1 Receptor (IGF1R) epitope, wherein the antibody fragment transigrates the blood-brain barrier, and wherein the epitope is specifically bound by the antibody of SEQ ID NO:5. The IGF1R epitope may be in the extracellular domain of IGF1R, and may comprise the sequence FENFLHNSIFVPR (SEQ ID NO:11), or fragments thereof.

[0036] The present invention further provides an isolated or purified antibody fragment, comprising

- a complementarity determining region (CDR) 1 sequence of GRTIDNYA (SEQ ID NO:1);
- a CDR2 sequence of IDWGDGGX (SEQ ID NO:2), where X is A or T; and
- a CDR3 sequence of AMARQSRVNLDVARYDY (SEQ ID NO:3),

wherein the antibody fragment specifically binds to the Insulin-Like Growth Factor 1 Receptor (IGF1R). In one embodiment, the isolated or purified antibody fragment comprises a CDR2 sequence of IDWGDGGA.

[0037] The term "antibody", also referred to in the art as "immunoglobulin" (Ig), as used herein refers to a protein constructed from paired heavy and light polypeptide chains; various Ig isotypes exist, including IgA, IgD, IgE, IgG, and IgM. When an antibody is correctly folded, each chain folds into a number of distinct globular domains joined by more linear polypeptide sequences. For example, the immunoglobulin light chain folds into a variable (V_L) and a constant (C_L) domain, while the heavy chain folds into a variable (V_H) and three constant (C_{H1}, C_{H2}, C_{H3}) domains. Interaction of the heavy and light chain variable domains (V_H and V_L) results in the formation of an antigen binding region (Fv). Each domain has a well-established structure familiar to those of skill in the art.

[0038] The light and heavy chain variable regions are responsible for binding the target antigen and can therefore show significant sequence diversity between antibodies. The constant regions show less sequence diversity, and are responsible for binding a number of natural proteins to elicit important biochemical events. The variable region of an antibody contains the antigen-binding determinants of the molecule, and thus determines the specificity of an antibody for its target antigen. The majority of sequence variability occurs in six hypervariable regions, three each per variable heavy (V_H) and light (V_L) chain; the hypervariable regions combine to form the antigen-binding site, and contribute to binding and recognition of an antigenic determinant. The specificity and affinity of an antibody for its antigen is determined by the structure of the hypervariable regions, as well as their size, shape, and chemistry of the surface they present to the antigen. Various schemes exist for identification of the regions of hypervariability, the two most common being those of Kabat and of Chothia and Lesk. Kabat et al (1991) define the "complementarity-determining regions" (CDR) based on sequence variability at the antigen-binding regions of the V_H and V_L domains. Chothia and Lesk (1987) define the "hypervariable loops" (H or L) based on the location of the structural loop regions in the V_H and V_L domains. These individual schemes define CDR and hypervariable loop regions that are adjacent or overlapping, those of skill in the antibody art often utilize the terms "CDR" and "hypervariable loop" interchangeably, and they may be so used herein. The CDR/loops are referred to herein according to the more recent IMGT numbering system (Lefranc, M.-P. et al., 2003), which was developed to facilitate comparison of variable domains. In this system, conserved amino acids (such as Cys23, Trp41, Cys104, Phe/Trp118, and a hydrophobic residue at position 89) always have the same position. Additionally, a standardized delimitation of the framework regions (FR1: positions 1 to 26; FR2: 39 to 55; FR3: 66 to 104; and FR4: 118 to 129) and of the CDR (CDR1: 27 to 38, CDR2: 56 to 65; and CDR3: 105 to 117) is provided.

[0039] An "antibody fragment" as referred to herein may include any suitable antigen-binding antibody fragment known in the art. The antibody fragment may be a naturally-occurring antibody fragment, or may be obtained by manipulation of a naturally-occurring antibody or by using recombinant methods. For example, an antibody fragment may include, but is not limited to a Fv, single-chain Fv (scFv; a molecule consisting of V_L and V_H connected with a peptide linker), Fab, F(ab')₂, single-domain antibody (sdAb; a fragment composed of a single V_L or V_H), and multivalent presentations

of any of these. Antibody fragments such as those just described may require linker sequences, disulfide bonds, or other type of covalent bond to link different portions of the fragments; those of skill in the art will be familiar with the requirements of the different types of fragments and various approaches and various approaches for their construction.

5 **[0040]** In a non-limiting example, the antibody fragment may be an sdAb derived from naturally-occurring sources. Heavy chain antibodies of camelid origin (Hamers-Casterman et al, 1993) lack light chains and thus their antigen binding sites consist of one domain, termed V_HH . sdAb have also been observed in shark and are termed V_{NAR} (Nuttall et al, 2003). Other sdAb may be engineered based on human Ig heavy and light chain sequences (Jespersen et al, 2004; To et al, 2005). As used herein, the term "sdAb" includes those sdAb directly isolated from V_H , V_HH , V_L , or V_{NAR} reservoir of any origin through phage display or other technologies, sdAb derived from the aforementioned sdAb, recombinantly produced sdAb, as well as those sdAb generated through further modification of such sdAb by humanization, affinity maturation, stabilization, solubilization, camelization, or other methods of antibody engineering. Also encompassed by the present invention are homologues, derivatives, or fragments that retain the antigen-binding function and specificity of the sdAb.

10 **[0041]** SdAb possess desirable properties for antibody molecules, such as high thermostability, high detergent resistance, relatively high resistance to proteases (Dumoulin et al, 2002) and high production yield (Arbabi-Ghahroudi et al, 1997); they can also be engineered to have very high affinity by isolation from an immune library (Li et al, 2009) or by *in vitro* affinity maturation (Davies & Riechmann, 1996). Further modifications to increase stability, such as the introduction of non-canonical disulfide bonds (Hussack et al, 2011; Kim et al, 2012), may also be brought to the sdAb.

15 **[0042]** A person of skill in the art would be well-acquainted with the structure of a single-domain antibody (see, for example, 3DWT, 2P42 in Protein Data Bank). An sdAb comprises a single immunoglobulin domain that retains the immunoglobulin fold; most notably, only three CDR/hypervariable loops form the antigen-binding site. However, and as would be understood by those of skill in the art, not all CDR may be required for binding the antigen. For example, and without wishing to be limiting, one, two, or three of the CDR may contribute to binding and recognition of the antigen by the sdAb of the present invention. The CDR of the sdAb or variable domain are referred to herein as CDR1, CDR2, and CDR3, and numbered as defined by Lefranc, M.-P. et al. (2003).

20 **[0043]** The antibody fragment of the present invention is specific for Insulin-Like Growth Factor 1 Receptor (IGF1R), a receptor found on cell surfaces. The IGF1R comprises an alpha subunit, which comprises an extracellular part having the insulin-like growth factor 1 binding site, connected by a disulphide bond to the beta subunit, which comprises a small extracellular domain, a transmembrane region, and an intracellular portion. The IGF1 receptor assembles into a homodimer or may form a heterodimer with the insulin receptor. The sequence of IGF1R may be, but is not limited to that shown in Figure 2 (SwissProt Accession No. P08069; SEQ ID NO:14), or a sequence substantially identical thereto.

25 **[0044]** The antibody fragment as described herein should not interfere with signaling through the Insulin Receptor (IR) or IGF1R. Specifically, the antibody fragments as described herein should not inhibit AKT phosphorylation induced by insulin, nor should they induce phosphorylation of the IR on their own or inhibit insulin-induced signaling; additionally, the antibody fragments described herein should not inhibit IGF-1-induced phosphorylation of IGF1R. Moreover, they should not bind to the Insulin Receptor.

30 **[0045]** As previously stated, the antibody fragment may be an sdAb. The sdAb may be of camelid origin or derived from a camelid V_HH , and thus may be based on camelid framework regions; alternatively, the CDR described above may be grafted onto V_{NAR} , V_HH , V_H or V_L framework regions. In yet another alternative, the hypervariable loops described above may be grafted onto the framework regions of other types of antibody fragments (Fv, scFv, Fab) of any source (for example, mouse) or proteins of similar size and nature onto which CDR can be grafted (for example, see Nicaise et al, 2004).

35 **[0046]** The present invention further encompasses an antibody fragment that is "humanized" using any suitable method known in the art, for example, but not limited to CDR grafting and veneering. Humanization of an antibody fragment comprises replacing an amino acid in the sequence with its human counterpart, as found in the human consensus sequence, without loss of antigen-binding ability or specificity; this approach reduces immunogenicity of the antibody fragment when introduced into human subjects. In the process of CDR grafting, one or more than one of the CDR defined herein may be fused or grafted to a human variable region (V_H , or V_L), to other human antibody (IgA, IgD, IgE, IgG, and IgM), to antibody fragment framework regions (Fv, scFv, Fab), or to proteins of similar size and nature onto which CDR can be grafted (Nicaise et al, 2004). In such a case, the conformation of said one or more than one hypervariable loop is likely preserved, and the affinity and specificity of the sdAb for its target (i.e., IGF1R) is likely minimally affected. CDR grafting is known in the art and is described in at least the following: US Patent No. 6180370, US Patent No. 5693761, US Patent No. 6054297, US Patent No. 5859205, and European Patent No. 626390. Veneering, also referred to in the art as "variable region resurfacing", involves humanizing solvent-exposed positions of the antibody fragment; thus, buried non-humanized residues, which may be important for CDR conformation, are preserved while the potential for immunological reaction against solvent-exposed regions is minimized. Veneering is known in the art and is described in at least the following: US Patent No. 5869619, US Patent No. 5766886, US Patent No. 5821123, and European Patent No. 519596. Persons of skill in the art would also be amply familiar with methods of preparing such humanized antibody

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fragments and humanizing amino acid positions.

[0047] For example, and without wishing to be limiting in any manner, the isolated or purified antibody fragment specific for IGF1R may be

X₁VX₂LX₃ESGGGLVQX₄GGSLRLSCAASGRTIDNYAMAWX₅RQAPGKX₆X₇EX₈VX₉TID
5 WGDGGX₁₀RYANSVKGRFTISRDNX₁₁KX₁₂TX₁₃YLQMNX₁₄LX₁₅X₁₆EDTAVYX₁₇CAMAR
QSRVNLDVARYDYWGQGT₁₈VTVSS (SEQ ID NO:4), where X₁ is E or Q; X₂ is K or Q; X₃ is V or E; X₄ is A or P; X₅
is V or S; X₆ is D or G; X₇ is L or R; X₈ is F or W; X₉ is A or S; X₁₀ is A or T; X₁₁ is A or S; X₁₂ is G or N; X₁₃ is M or L;
X₁₄ is N or R; X₁₅ is E or R; X₁₆ is P or A; X₁₇ is S or Y; and X₁₈ is Q or L,

10 or a sequence substantially identical thereto. Alternatively, the isolated or purified antibody fragment may comprise a
sequence selected from the group consisting of:

QVKLEESGGGLVQAGGSLRLSCAASGRTIDNYAMAWSRQAPGKDFVATIDWGDG
GARYANSVKGRFTISRDNKGTMYLQMNNLEPEDTAVYSCAMARQSRVNLDVARYD YWGQGTQVTVSS (SEQ
ID NO:5), referred to herein as IGF1R-5;

15 EVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWVRQAPGKGLEWVSTIDWGD
GGTRYANSVKGRFTISRDNKNTLYLQMNSLRAEDTAVYYCAMARQSRVNLDVARYD YWGQGTQVTVSS (SEQ
ID NO:6), referred to herein as IGF1R-5_H1;

QVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWVRQAPGKGLEWVATIDWGD
GGTRYANSVKGRFTISRDNKNTMYLQMNSLRAEDTAVYYCAMARQSRVNLDVARYDYWGQGTQVTVSS (SEQ
20 ID NO:7), referred to herein as IGF1R-5_H2;

QVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWSRQAPGKGLEFVATIDWGDG
GTRYANSVKGRFTISRDNKNTMYLQMNSLRAEDTAVYYCAMARQSRVNLDVARYD YWGQGTQVTVSS (SEQ
ID NO:8), referred to herein as IGF1R-5_H3;

25 QVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWSRQAPGKDFVATIDWGDG
GTRYANSVKGRFTISRDNKGTMYLQMNSLRAEDTAVYSCAMARQSRVNLDVARYD YWGQGTQVTVSS (SEQ
ID NO:9), referred to herein as IGF1R-5_H5; and

EVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWSRQAPGKDFVSTIDWGDG
GTRYANSVKGRFTISRDNKNTLYLQMNSLRAEDTAVYYCAMARQSRVNLDVARYDY WGQGTQVTVSS (SEQ
ID NO:10), referred to herein as IGF1 R-5_H6,

30 or a sequence substantially identical thereto.

[0048] A substantially identical sequence may comprise one or more conservative amino acid mutations. It is known
in the art that one or more conservative amino acid mutations to a reference sequence may yield a mutant peptide with
no substantial change in physiological, chemical, physico-chemical or functional properties compared to the reference
35 sequence; in such a case, the reference and mutant sequences would be considered "substantially identical" polypep-
tides. A conservative amino acid substitution is defined herein as the substitution of an amino acid residue for another
amino acid residue with similar chemical properties (e.g. size, charge, or polarity). These conservative amino acid
mutations may be made to the framework regions of the sdAb while maintaining the CDR sequences listed above and
the overall structure of the CDR of the antibody fragment; thus the specificity and binding of the antibody fragment are
40 maintained.

[0049] In a non-limiting example, a conservative mutation may be an amino acid substitution. Such a conservative
amino acid substitution may substitute a basic, neutral, hydrophobic, or acidic amino acid for another of the same group.
By the term "basic amino acid" it is meant hydrophilic amino acids having a side chain pK value of greater than 7, which
are typically positively charged at physiological pH. Basic amino acids include histidine (His or H), arginine (Arg or R),
45 and lysine (Lys or K). By the term "neutral amino acid" (also "polar amino acid"), it is meant hydrophilic amino acids
having a side chain that is uncharged at physiological pH, but which has at least one bond in which the pair of electrons
shared in common by two atoms is held more closely by one of the atoms. Polar amino acids include serine (Ser or S),
threonine (Thr or T), cysteine (Cys or C), tyrosine (Tyr or Y), asparagine (Asn or N), and glutamine (Gln or Q). The term
"hydrophobic amino acid" (also "non-polar amino acid") is meant to include amino acids exhibiting a hydrophobicity of
50 greater than zero according to the normalized consensus hydrophobicity scale of Eisenberg (1984). Hydrophobic amino
acids include proline (Pro or P), isoleucine (Ile or I), phenylalanine (Phe or F), valine (Val or V), leucine (Leu or L),
tryptophan (Trp or W), methionine (Met or M), alanine (Ala or A), and glycine (Gly or G). "Acidic amino acid" refers to
hydrophilic amino acids having a side chain pK value of less than 7, which are typically negatively charged at physiological
pH. Acidic amino acids include glutamate (Glu or E), and aspartate (Asp or D).

55 **[0050]** Sequence identity is used to evaluate the similarity of two sequences; it is determined by calculating the percent
of residues that are the same when the two sequences are aligned for maximum correspondence between residue
positions. Any known method may be used to calculate sequence identity; for example, computer software is available
to calculate sequence identity. Without wishing to be limiting, sequence identity can be calculated by software such as

NCBI BLAST2 service maintained by the Swiss Institute of Bioinformatics (and as found at ca.expasy.org/tools/blast/), BLAST-P, Blast-N, or FASTA-N, or any other appropriate software that is known in the art.

5 [0051] The substantially identical sequences of the present invention may be at least 90% identical; in another example, the substantially identical sequences may be at least 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, or 100% identical, or any percentage therebetween, at the amino acid level to sequences described herein. Importantly, the substantially identical sequences retain the activity and specificity of the reference sequence. In a non-limiting embodiment, the difference in sequence identity may be due to conservative amino acid mutation(s). In a non-limiting example, the present invention may be directed to an antibody fragment comprising a sequence at least 95%, 98%, or 99% identical to that of the antibody fragments described herein.

10 [0052] The antibody fragment of the present invention may also comprise additional sequences to aid in expression, detection or purification of a recombinant antibody fragment. Any such sequences or tags known to those of skill in the art may be used. For example, and without wishing to be limiting, the antibody fragment may comprise a targeting or signal sequence (for example, but not limited to ompA), a detection/purification tag (for example, but not limited to c-Myc, His₅, or His₆), or a combination thereof. In another example, the additional sequence may be a biotin recognition site such as that described by Cronan et al in WO 95/04069 or Voges et al in WO/2004/076670. As is also known to those of skill in the art, linker sequences may be used in conjunction with the additional sequences or tags, or may serve as a detection/purification tag.

15 [0053] The antibody fragment of the present invention may also be in a multivalent display format, also referred to herein as multivalent presentation. Multimerization may be achieved by any suitable method of known in the art. For example, and without wishing to be limiting in any manner, multimerization may be achieved using self-assembly molecules such as those described in Zhang et al (2004a; 2004b) and WO2003/046560, where pentabodies are produced by expressing a fusion protein comprising the antibody fragment of the present invention and the pentamerization domain of the B-subunit of an AB₅ toxin family (Merritt & Hol, 1995). A multimer may also be formed using the multimerization domains described by Zhu et al. (2010); this form, referred to herein as a "combody" form, is a fusion of the antibody fragment of the present invention with a coiled-coil peptide resulting in a multimeric molecule. Other forms of multivalent display are also encompassed by the present invention. For example, and without wishing to be limiting, the antibody fragment may be presented as a dimer, a trimer, or any other suitable oligomer. This may be achieved by methods known in the art, for example direct linking connection (Nielson et al, 2000), c-jun/Fos interaction (de Kruijff & Logtenberg, 1996), "Knob into holes" interaction (Ridgway et al, 1996).

20 [0054] Another method known in the art for multimerization is to dimerize the antibody fragment using an Fc domain, for example, but not limited to human Fc domains. The Fc domains may be selected from various classes including, but not limited to, IgG, IgM, or various subclasses including, but not limited to IgG1, IgG2, etc. In this approach, the Fc gene is inserted into a vector along with the sdAb gene to generate a sdAb-Fc fusion protein (Bell et al, 2010; Iqbal et al, 2010); the fusion protein is recombinantly expressed then purified. For example, and without wishing to be limiting in any manner, multivalent display formats may encompass chimeric formats of anti-IGF1R-5 V_HH linked to an Fc domain, or bi- or tri-specific antibody fusions with two or three anti-IGF1R-5 V_HH recognizing unique epitopes. Such antibody fragments are easy to engineer and to produce, can greatly extend the serum half-life of sdAb, and may be excellent tumor imaging reagents (Bell et al., 2010).

25 [0055] The Fc domain in the multimeric complex as just described may be any suitable Fc fragment known in the art. The Fc fragment may be from any suitable source; for example, the Fc may be of mouse or human origin. In a specific, non-limiting example, the Fc may be the mouse Fc2b fragment or human Fc1 fragment (Bell et al, 2010; Iqbal et al, 2010). In a specific, non-limiting example, the multimerized isolated or purified antibody fragment as just described may comprise the sequence of SEQ ID NO:42, 43, 12, or 13.

30 [0056] Each subunit of the multimers described above may comprise the same or different antibody fragments of the present invention, which may have the same or different specificity. Additionally, the multimerization domains may be linked to the antibody fragment using a linker, as required; such a linker should be of sufficient length and appropriate composition to provide flexible attachment of the two molecules, but should not hamper the antigen-binding properties of the antibody fragment.

35 [0057] The antibody fragment as described herein may transigrate across the blood-brain barrier. The brain is separated from the rest of the body by a specialized endothelial tissue known as the blood-brain barrier (BBB). The endothelial cells of the BBB are connected by tight junctions and efficiently prevent many therapeutic compounds from entering the brain. In addition to low rates of vesicular transport, one specific feature of the BBB is the existence of enzymatic barrier(s) and high level(s) of expression of ATP-dependent transporters on the abluminal (brain) side of the BBB, including P-glycoprotein (Gottesman et al., 1993; Watanabe, 1995), which actively transport various molecules from the brain into the blood stream (Samuels, 1993). Only small (<500 Daltons) and hydrophobic (Pardridge, 1995) molecules can more readily cross the BBB. Thus, the ability of the antibody fragment as described above to specifically bind the surface receptor, internalize into brain endothelial cells, and undergo transcytosis across the BBB by evading lysosomal degradation is useful in the neurological field.

[0058] The present invention also encompasses nucleic acid sequences encoding the molecules as described herein. Given the degeneracy of the genetic code, a number of nucleotide sequences would have the effect of encoding the polypeptide, as would be readily understood by a skilled artisan. The nucleic acid sequence may be codon-optimized for expression in various micro-organisms. The present invention also encompasses vectors comprising the nucleic acids as just described. Furthermore, the invention encompasses cells comprising the nucleic acid and/or vector as described.

[0059] The present invention further encompasses the isolated or purified antibody fragments immobilized onto a surface using various methodologies; for example, and without wishing to be limiting, the antibody fragment may be linked or coupled to the surface via His-tag coupling, biotin binding, covalent binding, adsorption, and the like. Immobilization of the antibody fragment of the present invention may be useful in various applications for capturing, purifying or isolating proteins. The solid surface may be any suitable surface, for example, but not limited to the well surface of a microtiter plate, channels of surface plasmon resonance (SPR) sensorchips, membranes, beads (such as magnetic-based or sepharose-based beads or other chromatography resin), glass, plastic, stainless steel, a film, or any other useful surface such as nanoparticles, nanowires and cantilever surfaces.

[0060] The invention also encompasses the antibody fragment as described above linked to a cargo molecule. The cargo molecule may be any suitable molecule, which is delivered across the BBB by the antibody fragment. The cargo molecule may have a molecular weight in the range of about 1kD to about 200 kDa; for example, the cargo molecule may have a molecular weight of about 1, 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, 150, 155, 160, 165, 170, 175, 180, 185, 190, 195, or 200 kDa, or any weight therebetween, or any range of weights defined by any two aforementioned weights. In specific, non-limiting examples, the cargo molecule may have a molecular weight of 1 kDa (for example, but not limited to a small molecule such as Cy5.5), 1-10 kDa (for example, but not limited to a peptide such as galanin, 3kDa), about 80 kDa (for example, but not limited to a Fc fragment, enzyme, protein, antibody etc), or about 200kDa (for example, but not limited to a monoclonal antibody).

[0061] For example, and without wishing to be limiting in any manner, the cargo molecule may be a detectable agent, a therapeutic agent, a drug, a peptide, an enzyme, a growth factor, a cytokine, a receptor trap, an antibody or fragment thereof (e.g., IgG, scFv, Fab, V_HH, V_H, V_L, etc) a chemical compound, a carbohydrate moiety, DNA-based molecules (anti-sense oligonucleotide, microRNA, siRNA, plasmid), a cytotoxic agent, viral vector (adeno-, lenti-, retro), one or more liposomes or nanocarriers loaded with any of the previously recited types of cargo molecules, or one or more nanoparticle, nanowire, nanotube, or quantum dots. The cargo molecule as described above may be a detectable agent. For example, the IGF1R-specific antibody fragment may be linked to a radioisotope, a paramagnetic label, a fluorophore, a fluorescent agent, Near Infra-Red (NIR; for example Cy5.5) fluorochrome or dye, an echogenic microbubble, an affinity label, a detectable protein-based molecule, nucleotide, quantum dot, nanoparticle, nanowire, or nanotube or any other suitable agent that may be detected by imaging methods. The antibody fragment may be linked to the cargo molecule using any method known in the art (recombinant technology, chemical conjugation, etc.).

[0062] The cargo molecule as described herein may be linked, also referred to herein as "conjugated", to the antibody fragment by any suitable method known in the art. For example, and without wishing to be limiting, the cargo molecule may be linked to the peptide by a covalent bond or ionic interaction. The linkage may be achieved through a chemical cross-linking reaction, or through fusion using recombinant DNA methodology combined with any peptide expression system, such as bacteria, yeast or mammalian cell-based systems. When conjugating the cargo molecule to the antibody fragment, a suitable linker may be used. Methods for linking an antibody fragment to a cargo molecule such as a therapeutic or detectable agent would be well-known to a person of skill in the art.

[0063] In one non-limiting example, the cargo molecule may be a detectable label, a radioisotope, a paramagnetic label such as gadolinium or iron oxide, a fluorophore, Near Infra-Red (NIR) fluorochrome or dye, an echogenic microbubble, an affinity label (for example biotin, avidin, etc), enzymes, or any other suitable agent that may be detected by diagnostic imaging methods. In a specific, non-limiting example, the anti-IGF1R-5 or fragment thereof may be linked to a near infrared fluorescence (NIRF) imaging dye, for example and not wishing to be limiting Cy5.5, Alexa680, Dylight680, or Dylight800.

[0064] Thus, the present invention further provides an *in vitro* method of detecting IGF1R, comprising contacting a tissue sample with one or more than one isolated or purified antibody fragment of the present invention linked to a detectable agent. The IGF1 R-antibody complex can then be detected using detection and/or imaging technologies known in the art. The tissue sample in the method as just described may be any suitable tissue sample, for example but not limited to a serum sample, a vascular tissue sample, a tumour tissue sample, or a brain tissue sample; the tissue sample may be from a human or animal subject. The step of contacting is done under suitable conditions, known to those skilled in the art, for formation of a complex between the antibody or fragment thereof and IGF1R. The step of detecting may be accomplished by any suitable method known in the art, for example, but not limited to optical imaging, immunohistochemistry, molecular diagnostic imaging, ELISA, imaging mass spectrometry, or other suitable method. For example, and without wishing to be limiting in any manner, the isolated or purified antibody fragment linked to a

detectable agent may be used in immunoassays (IA) including, but not limited to enzyme IA (EIA), ELISA, "rapid antigen capture", "rapid chromatographic IA", and "rapid EIA". (For example, see Planche et al, 2008; Sloan et al, 2008; Rüssmann et al, 2007; Musher et al, 2007; Turgeon et al, 2003; Fenner et al, 2008).

[0065] The present invention also provides an *in vivo* method of detecting IGF1R expression in a subject. The method comprises administering one or more than one isolated or purified antibody or fragment thereof as described herein linked to a detectable agent to the subject, then detecting the labelled antibody fragment bound to IGF1R. The step of detecting may comprise any suitable method known in the art, for example, but not limited to PET, SPECT, or fluorescence imaging, or any other suitable method. The method as just described may be useful in detecting the expression of IGF1R in blood vessels or tissues, for example but not limited to tumor tissues.

[0066] The *in vivo* detection step in the methods described above may be whole body imaging for diagnostic purposes or local imaging at specific sites, such as but not limited to brain vessels or brain tumor vessels, in a quantitative manner to assess the progression of disease or host response to a treatment regimen. The detection step in the methods as described above may be immunohistochemistry, or a non-invasive (molecular) diagnostic imaging technology including, but not limited to:

- Optical imaging;
- Positron emission tomography (PET), wherein the detectable agent is an isotopes such as ^{11}C , ^{13}N , ^{15}O , ^{18}F , ^{64}Cu , ^{62}Cu , ^{124}I , ^{76}Br , ^{82}Rb and ^{68}Ga , with ^{18}F being the most clinically utilized;
- Single photon emission computed tomography (SPECT), wherein the detectable agent is a radiotracer such as $^{99\text{m}}\text{Tc}$, ^{111}In , ^{123}I , ^{201}Tl , ^{133}Xe , depending on the specific application;
- Magnetic resonance imaging (MRI), wherein the detectable agent may be, for example and not limited to gadolinium, iron oxide nanoparticles and carbon-coated iron-cobalt nanoparticles thereby increasing the sensitivity of MRI for the detection of plaques.
- Contrast-Enhanced Ultrasonography (CEUS) or ultrasound, wherein the detectable agent is at least one acoustically active and gas-filled microbubble. Ultrasound is a widespread technology for the screening and early detection of human diseases. It is less expensive than MRI or scintigraphy and safer than molecular imaging modalities such as radionuclide imaging because it does not involve radiation.

[0067] The present invention further provides a method of transporting a molecule of interest across the blood-brain barrier. The method comprises administering the molecule linked to an antibody fragment as described herein to a subject; the antibody fragment transmigrates the blood-brain barrier. The molecule may be any desired molecule, including the cargo molecules as previously described; the molecule may be "linked" to the antibody fragment using any suitable method, including, but not limited to conjugation or expression in a fusion protein. The administration may be by any suitable method, for example parenteral administration, including but not limited to intravenous (iv), subcutaneous (sc), and intramuscular (im) administration. In this method, the antibody fragment of the present invention 'ferries' the molecule of interest across the BBB to its brain target.

[0068] The present invention also encompasses a composition comprising one or more than one isolated or purified antibody fragment as described herein. The composition may comprise a single antibody fragment as described above, or may be a mixture of antibody fragments. Furthermore, in a composition comprising a mixture of antibodies or fragments of the present invention, the antibodies may have the same specificity, or may differ in their specificities; for example, and without wishing to be limiting in any manner, the composition may comprise antibody fragments thereof specific to IGF1R (same or different epitope).

[0069] The composition may also comprise a pharmaceutically acceptable diluent, excipient, or carrier. The diluent, excipient, or carrier may be any suitable diluent, excipient, or carrier known in the art, and must be compatible with other ingredients in the composition, with the method of delivery of the composition, and is not deleterious to the recipient of the composition. The composition may be in any suitable form; for example, the composition may be provided in suspension form, powder form (for example, but limited to lyophilised or encapsulated), capsule or tablet form. For example, and without wishing to be limiting, when the composition is provided in suspension form, the carrier may comprise water, saline, a suitable buffer, or additives to improve solubility and/or stability; reconstitution to produce the suspension is effected in a buffer at a suitable pH to ensure the viability of the antibody or fragment thereof. Dry powders may also include additives to improve stability and/or carriers to increase bulk/volume; for example, and without wishing to be limiting, the dry powder composition may comprise sucrose or trehalose. In a specific, non-limiting example, the composition may be so formulated as to deliver the antibody fragment to the gastrointestinal tract of the subject. Thus, the composition may comprise encapsulation, time-release, or other suitable technologies for delivery of the antibody fragment. It would be within the competency of a person of skill in the art to prepare suitable compositions comprising the present compounds.

[0070] The invention also encompasses a method of quantifying an amount of a cargo molecule delivered across the BBB of a subject, wherein the cargo molecule is linked to one or more than one isolated or purified antibody fragment

as described herein, the method comprising

- a) collecting cerebrospinal fluid (CSF) from the subject; and
- b) using targeted proteomics methods to quantify the amount of the cargo molecule linked to one or more than one antibody fragment in the CSF.

[0071] The cargo molecule may be any desired molecule, including the cargo molecules, as previously described; the isolated or purified antibody fragment transmigrates the blood-brain barrier; and the molecule may be "linked" to the antibody fragment using any suitable method, including, but not limited to conjugation or expression in a fusion protein, as previously described. In the above method, the CSF is collected from a subject using any suitable method known in the art. The amount of CSF required for targeted proteomics method in step b) may be between about 1 to 10 μ l; for example, the amount of CSF required may be about 1.0, 1.5, 2.0, 2.5, 3.0, 3.5, 4.0, 4.5, 5.0, 5.5, 6.0, 6.5, 7.0, 7.5, 8.0, 8.5, 9.0, 9.5, or 10 μ l, or any amount therebetween, or any range defined by the amount just described. The antibody fragment linked to the cargo molecule may have been administered to the subject prior to collection of the CSF. A suitable delay between administration and delivery of the antibody fragment linked to the cargo molecule across the BBB may be required. The delay may be at least 30 minutes following administration of the antibody fragment linked to the cargo molecule; for example and without wishing to be limiting in any manner, the delay may be at least 30 minutes, 1 hour, 1.5 hour, 2 hours, 2.5 hours, 3 hours, 3.5 hours, 4 hours, 4.5 hours, or 5 hours. The targeted proteomics methods used to quantify the amount of the one or more than antibody or fragment thereof linked to the cargo molecule may be any suitable method known in the art. For example and without wishing to be limiting, the targeted proteomics method may be a mass spectrometry method, such as but not limited to multiple reaction monitoring using an isotopically labeled internal standard (MRM-ILIS; see for example Haqqani et al., 2013). MRM is advantageous in that it allows for rapid, sensitive, and specific quantification of unlabelled targeted analytes (for example, an antibody or fragment thereof as described herein) in a biological sample. The multiplexing capability of the assay may allow for quantification of both the antibody fragment and the cargo molecule.

[0072] The present invention will be further illustrated in the following examples. However, it is to be understood that these examples are for illustrative purposes only and should not be used to limit the scope of the present invention in any manner.

Example 1: Purification of IGF1R recombinant fragment

[0073] A 933 amino acid long recombinant fragment of the extracellular domain of IGF1R (shown by the grey box in Figure 1; see also amino acids 1-933 of SEQ ID NO:14) was prepared. The fragment comprised an N-terminal 30 amino acid signal peptide, the full alpha subunit, a furin cleavage site (RKRR, SEQ ID NO:15; separating alpha and beta subunits), as well as the majority of the extracellular portion of the beta subunit (Figures 1 and 2).

[0074] *Cloning.* The sequence of the IGF1R ectodomain of interest was amplified by PCR using the following primers:

5'-CGGGATCCGCCACCATGAAGTCTGGCTCCGGAG-3' (forward; SEQ ID NO:16)

5'-GCTCTAGATCAGAAGTTTTTCATATCCTGTTTTGG-3' (reverse; SEQ ID NO:17)

and subcloned into the SmaI site of Puc19. The IGF1R⁹³³ sequence was then sub-cloned into pCDN4/myc-His (Invitrogen) to generate pIGF1R⁹³³-His, which allows for expression of a His-tagged ectodomain as described previously (Samani et al. 2004).

[0075] *Transient transfection.* Lentivirus particles expressing IGF1R⁹³³-His were generated in the packaging cell line 293SF-PacLV, as detailed previously (Broussau et al., 2008). Briefly, the cells were transfected with the vector using polyethylenimine. Fresh medium (LC-SFM), containing 1 μ g/ml doxycycline and 10 μ g/ml cumate, was added to the cells 5 hours post-transfection and the supernatants containing LV particles were collected after 48-72 hours, concentrated by ultracentrifugation at 100,000xg for 2h at 4°C on a 20% sucrose cushion (Gaillet B et al. 2007), re-suspended in LC-SFM medium supplemented with 1% FBS and stored at -70°C until used.

[0076] *Stable expression.* A stable cell line, 293SF-cum2-CR5-IGF1R-His, was generated by transduction of 293SF-Cum2 cell lines with the respective lentivirus particles using a protocol described previously (Gaillet B. et al. 2010). Briefly, 0.5-1.0 X 10⁵ 293SF-Cum2 cells were seeded in 24 wells plates into 200 μ l of LC-SFM medium without dextran sulfate. The LV suspension was prepared by mixing 200-500 μ L of LV with 8 μ g/mL of polybrene and incubating it for 30min at 37°C. The freshly made LV suspension was added to the cells 4 hours after seeding. After 24h, 500 μ L medium supplemented with dextran sulfate was added to the cells. To increase the level of expression the cells were re-transduced up to 6 times using the same protocol after 3-4 days of cell recovery.. Finally the cells were expanded in 6-well plates and shaker flasks.

[0077] *Large scale protein production and purification.* The clone identified as the highest producer was expanded in

shaker or spinner flasks. Protein production was initiated by the addition of 1 $\mu\text{g/ml}$ cumate in fresh medium, followed by a 24 h incubation at 37°C and a 4-8 day incubation at 30°C. Cells were removed by centrifugation and the supernatants filtered and concentrated (10x) using the Tangential Flow Filtration Systems (Pellicon ultrafiltration cassettes, EMD Millipore).

[0078] The IGF1R⁹³³-His was purified using a HisPrep column (GE Healthcare) according to the manufacturer's instructions. Briefly, the concentrated sample was applied to a His-prep FF (16/10) column (GE Healthcare), equilibrated and washed with 50 mM sodium phosphate, 300mM NaCl, 5mM imidazole pH 7.5 and eluted with the 50 mM sodium phosphate, 300mM NaCl, 500mM imidazole pH 7.5. A step elution with 0.1 M sodium citrate pH 4.5 to pH 2.5 was used to elute the protein and peak fractions were pooled. Buffer exchange was performed by ultrafiltration using a 50 kDa cut-off membrane or a desalting column with a buffer containing 50 mM sodium phosphate, 150 mM NaCl and 0.01% Tween-80, pH 7.2. Purity of both proteins was verified by SDS-PAGE and they were stored at -80°C until used (see subsequent examples).

Example 2: Llama immunization and serum response

[0079] To isolate V_HHs that targets the extracellular domain of IGF1R, a llama was immunized with the recombinant IGF1R⁹³³-His fragment obtained in Example 1.

[0080] One male llama (*Lama glama*) was immunized by sub-cutaneous, lower-back injection of IGF1R⁹³³-His recombinant antigen (Example 1). On day 1, 200 μg of antigen diluted in PBS to 1 ml was injected together with 1 ml of Freund's Complete Adjuvant (Sigma, St. Louis, MO). Three further injections of 100 μg of IGF1R⁹³³-His antigen plus Freund's Incomplete Adjuvant (Sigma) were performed on days 22, 36, and 50. A final injection of 100 μg of antigen with no adjuvant was performed on day 77. Pre-immune blood was drawn before the first injection on day 1 and served as a negative control. Blood (10-15 ml) was collected on days 29, 43, 57 and 84. The blood from day 84 was processed immediately to isolate peripheral blood mononuclear cells (PBMC). The blood was diluted 1:1 with phosphate buffered saline (PBS) and PBMCs were purified from the blood using the Lymphoprep Tube (Axis Shield). The cells were counted and stored as aliquots of about 1×10^7 cells at -80°C for future use.

[0081] Pre-immune and post-immune total serum was analyzed for a specific response to IGF1R⁹³³-His antigen by ELISA on day 57. Llama sera from day 84 were fractionated as previously described (Doyle et al, 2008). The resulting fractions, A1 (HCAb), A2 (HCAb), G1 (HCAb) and G2 (cIgG) were analyzed for specific binding to the IGF1R⁹³³-His antigen by ELISA. Briefly, 5 μg of IGF1R⁹³³-His recombinant antigen diluted in PBS was incubated overnight (100 $\mu\text{l/well}$, 18 h, 4°C) in 96 well Maxisorp plates (Nalgene, Nunc) to coat the plates. Plates were blocked with bovine serum albumin (BSA), washed with PBS-T (PBS + 0.05% (v/v) Tween-20), and serial dilutions of pre-immune total serum, post-immune total serum (day 57), and fractionated serum (day 84) were applied. After incubation at room temperature for 1.5 h the plates were washed with PBS-T, before goat anti-llama IgG (1:1,000 in PBS) was added and plates were incubated for 1 h at 37°C. After washing with PBS-T, pig anti-goat IgG-HRP conjugate (1:3,000 in PBS) was added and plates were incubated for 1 h at 37°C. A final PBS-T wash was carried out prior to the addition of 100 $\mu\text{l/well}$ TMB substrate (KPL, Gaithersburg, MD); the substrate was incubated for 10 min. The reaction was stopped with 100 $\mu\text{l/well}$ 1 M H₃PO₄. Absorbance was read at 450 nm.

Example 3: Library construction and selection of IGF1R-binding V_HHs

[0082] A hyperimmunized llama V_HH library was constructed based on RNA isolated from the PBMCs in Example 2.

[0083] Library construction and panning was performed essentially as previously described (Arbabi-Ghahroudi et al, 2009a, 2009b; Tanha et al, 2003). Total RNA was isolated from approximately 10^7 PBMCs collected on day 84 post-immunization (Example 2) using the QIAamp RNA Blood Mini Kit (Qiagen). About 5 μg of total RNA was used as template for first strand cDNA synthesis with oligo dT primers using the First-Strand cDNA Synthesis Kit (GE Healthcare). The cDNA was amplified by an equimolar mix of three variable region-specific sense primers:

MJ1: 5'-GCCAGCCGGCCATGGCCSMKGTGCAGCTGGTGGAKTCTGGGGGA-3' (SEQ ID NO:18)

MJ2: 5'-GCCAGCCGGCCATGGCCAGGTAAAGCTGGAGGAGTCTGGGGGA-3' (SEQ ID NO:19)

MJ3: 5'-GCCAGCCGGCCATGGCCAGGCTCAGGTACAGCTGGTGGAGTCT-3' (SEQ ID NO:20),

and two antisense CH₂-specific primers:

CH₂: 5'-CGCCATCAAGGTACCAGTTGA-3' (SEQ ID NO:21)

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CH₂b₃: 5'-GGGGTACCTGTCATCCACGGACCAGCTGA-3' (SEQ ID NO:22).

[0084] Briefly, the PCR reaction mixture was set up in a total volume of 50 μ l with the following components: 1-3 μ l cDNA, 5 pmol of MJ1-3 primer mixture, 5 pmol of CH₂ or CH₂b₃ primers, 5 μ l of 10x reaction buffer, 1 μ l of 10 mM dNTP, 2.5 unit of Taq DNA polymerase (Hoffmann-La Roche). The PCR protocol consisted of an (i) initial step at 94°C for 3 min, (ii) followed by 30 cycles of 94°C for 1 min, 55°C for 30 s, 72°C for 30 s and (iii) a final extension step at 72°C for 7 min. The amplified PCR products were run on a 2% agarose gel and two major bands were observed: a band of about 850 bp, corresponding to conventional IgG, and a second band of around 600 bp, corresponding to the V_HH-CH2 region of camelid heavy chain antibodies. The smaller bands were cut and purified using the QIAquick Gel Extraction Kit (Qiagen) and re-amplified in a second PCR in a total volume of 50 μ l using 1 μ l (30ng) of DNA template, 5 pmol of each of MJ7 primer (5'-CATGTGTAGACTCGCGGCCAGCCGGCCATGGCC-3' SEQ ID NO:23) and MJ8 primer (5'-CATGTGTAGATTCTGCGCGGCCTGGCCTGAGGAGACGGTGACCTGG-3' SEQ ID NO:24), 5 μ l of 10x reaction buffer, 1 μ l of 10 mM dNTP, 2.5 unit of Taq DNA polymerase. The PCR protocol consisted of (i) an initial step at 94°C for 3 min, (ii) followed by 30 cycles of 94°C for 30 s, 57°C for 30 s and 72°C for 1 min and (iii) a final extension step at 72°C for 7 min. The amplified PCR products, ranging between 340 bp and 420 bp and corresponding to V_HH fragments of heavy chain antibodies, were purified using the QIAquick PCR Purification Kit (Qiagen), digested with *Sfi*I restriction enzyme (New England BioLabs) and re-purified using the same kit.

[0085] 80 μ g of pMED1 phagemid vector (Arbabi-Ghahroudi et al, 2009b) were digested with *Sfi*I overnight at 50°C. To minimize self-ligation, 20 units of *Xho*I and *Pst*I restriction enzymes were added to cut the excised fragment and the digestion reaction was incubated for an additional 2 h at 37°C. 60 μ g of digested phagemid DNA was ligated with 6 μ g of digested (*Sfi*I for 5h at 50°C) V_HH fragments (molar ratio 1:1) for 3 h at room temperature using LigaFast Rapid DNA Ligation System (Promega) according to the manufacturer's instructions. The ligated plasmids were purified using the QIAquick PCR Purification Kit (Qiagen), eluted in a final volume of 100 μ l, and transformed into electrocompetent TG1 *E. coli* (Stratagene) using 5 μ l of ligated DNA aliquot per transformation reaction, as described (Arbabi-Ghahroudi et al, 2009b). The size of the library was determined to be 5 x 10⁷ as described in (Arbabi-Ghahroudi et al, 2009b). 20 clones were sequenced and contained all unique V_HH sequences. The *E. coli*, containing the library was grown for 2-3 h at 37°C, 250 rpm in the presence of 2% (w/v) glucose. The bacteria were then pelleted, resuspended in 2xYT/Amp/Glu (2xYT medium with 100 μ g/ml ampicillin and 2% (w/v) glucose) with 35% (v/v) glycerol and stored at -80°C in small aliquots.

[0086] Panning experiments were essentially performed as described in (Arbabi et al, 1997). Two milliliters of the library (2.0 x 10¹⁰ bacteria) were thawed on ice and grown in 2xYT/Amp/Glu for about 2 h at 37°C (A₆₀₀ = 0.4 - 0.5). The *E. coli* were subsequently infected with 20x excess M13KO7 helper phage (New England Biolabs) for 1 h at 37°C. The culture was then centrifuged at 4°C and infected bacterial pellets were re-suspended in 200 ml of 2xYT/Amp with 50 μ g/ml kanamycin and incubated at 37°C and 250 rpm. The phage particles in the culture supernatant were incubated with 1/5 volume of 20% PEG 8000/2.5M NaCl on ice for 1 h and centrifuged at 10,000 rpm for 15 min. The phage pellets were re-suspended in 1.5 ml of sterile PBS, titrated and used as input phage for panning. For panning round 1, 96-well Maxisorp™ plates were coated with 10 μ g of recombinant IGF1R⁹³³-His per well in 100 μ l PBS overnight at 4°C. The wells were rinsed with PBS and blocked with PBS plus 1% (w/v) casein for 2 h at 37°C. Approximately 10¹² phages were added to the blocked wells and incubated for 2 h at 37°C. After 10x washing with PBS/0.1% (v/v) Tween 20, the bound phages were eluted with 0.1 M triethylamine, neutralized (50 μ l of 1M Tris-HCl, pH 7.4) and mixed with exponentially growing TG1 *E. coli*. Titration of eluted phage was performed and infected bacteria were superinfected with M13KO7 and grown overnight at 37°C. The purified phage from the overnight culture was used as the input for the next round of panning. The panning was continued for three further rounds. The same protocol as described above was used, except that the amount of recombinant antigen used to coat the plates was reduced to 7 μ g, 5 μ g and 5 μ g for the second, third and fourth rounds of panning, respectively.

[0087] Individual TG1 colonies obtained after round four of panning were subjected to phage ELISA screening, essentially as described elsewhere (Doyle et al, 2008), with the exception that 5 μ g/ml of IGF1R⁹³³-His recombinant antigen were used to coat the microtiter plates. All positive clones were sent for DNA sequencing. Unique clones that gave high phage ELISA signals were selected for large-scale expression and purification using known methods (see Example 4). A clone dubbed IGF1R-5 was identified for further study; its sequence is shown below.

QVKLEESGGGLVQAGGSLRLSCAASGRTIDNYAMAWSRQAPGKDREVFATIDW
GDGGARYANSVKGRFTISRDNAGKTMYLQMNLEPEDTAVYSCAMARQSRVNL
DVARYDYWGQGTQVTVSS (SEQ ID NO:5)

Example 4: Humanization of IGF1R-5

[0088] To avoid potential immunogenicity of llama-derived IGF1R-5 when applied as BBB carrier for therapeutics, to the camelid-derived sdAb was "humanized" by mutation of "camelid" residues in the V_HH. It should be noted that, for the purpose of humanization, Kabat numbering (Kabat et al, 1991) was used for identification of CDR residues.

[0089] *3D-structure modeling of camelid V_HHs.* Template structures similar to IGF1R-5 V_HH were identified using BLAST searches against the Protein Data Bank (PDB). The 3D structure of the IGF1R-5 was approximated using homology modeling based on 4KRP|B (PDB code | Chain ID) as the main template, with additional information from 4FHB|D. The IGF1R-5 V_HH structure was then built by mutating the main template structure to the IGF1R-5 sequence; this included 35 mutations at various positions. The IGF1R-5 V_HH model was then refined by energy minimization with the AMBER force-field and a stepwise release of constraints, ranging from the CDR loops, which were relaxed first, to the backbone heavy atoms of the framework region, which were fully relaxed only in the last stage. The CDR-H3 loop of the V_HH model was then refined by Monte-Carlo-minimization (MCM) conformational sampling, in which dihedral angles in the CDR-H3 region were sampled followed by energy minimization.

[0090] *Selection of the human heavy-chain framework for the camelid CDR.* Human heavy-chain framework was selected by standard sequence homology comparison against the human germline databases (VBASE), against other sequence databases (Genbank and SwissProt), and against the human framework consensus sequences. BLAST searches were conducted to retrieve sequence matches with highest homology in the framework region only (i.e., excluding CDR) while matching the length of the CDR. The closest human frameworks identified for IGF1R-5 V_HH corresponded to the human VH-3 subgroup. Several human germline VH-3 framework sequences that were most similar to IGF1R-5 V_HH were also retained in addition to the human VH-3 consensus sequence. The IGF1R-5 V_HH framework sequences required 18 mutations in order to arrive at the consensus human VH-3 sequence for 100% framework humanization.

[0091] *Identification of framework residues for back-mutations.* The IGF1R-5 V_HH model and its fully-humanized counterpart were characterized to estimate the humanness index, antigen contact propensity index, to delineate the CDR, canonical residues, unusual framework residues, potential glycosylation sites, buried residues, Vernier zone residues, and proximity to CDR. The analysis of these data suggested the design of several humanized variants for the anti-IGF1R V_HH, each variant having varying numbers of back-mutations to the parent camelid residues at various positions. 5 humanized variants were designed for IGF1R-5 V_HH (IGF1R-5 H1, H2, H3, H5, H6), where variants contained up to 10 back-mutations. Some of these camelid back-mutations residues were buried inside the V_HH domain core and hence are not expected to induce an immune response.

Example 5: Expression and purification of selected V_HH constructs

[0092] IGF1R-5 identified in Example 3 and the humanized versions constructed in Example 4 (collectively referred to herein as "V_HH constructs") were sub-cloned into expression plasmids for protein expression and purification.

[0093] A phagemid vector containing the DNA of the IGF1R-5 clone was purified using the MiniPrep Kit (Qiagen). The IGF1R-binding V_HH IGF1R-5 was PCR amplified from the pMED1 phagemid vector, adding an N-terminal BbsI cleavage site and a BamHI cleavage site at the C-terminus, using primers:

5'-TATGAAGACACCAGGCCAGGTAAGCTGGAGGAGTCT-3' (forward; SEQ ID NO:25)

5'-TTGTTTCGGATCCTGAGGAGACGGTGACCTG-3' (reverse; SEQ ID NO:26)

[0094] The PCR fragment and the pSJF2H expression vector was digested with BbsI and BamHI restriction enzymes (NEB) according to the manufacturer's instructions. The digested IGF1R-5 V_HH fragment was ligated into the digested pSJF2H expression vector, using methods similar to those described in Arbabi-Ghahroudi et al. (2009b); the ligation products were then transformed into electro-competent TG1 *E. coli*. Clones were selected on LB agar plates + 100 µg/ml ampicillin and verified by DNA sequencing.

[0095] The humanized clones were synthesized and directly cloned into pSJF2H, using methods similar to those described above, and subsequently transformed into *E. coli* TG1; clones were selected as described above.

[0096] *Protein Expression.* All IGF1R-5 V_HH constructs were expressed in TG1 *E. coli*. An overnight culture in LB/amp/glu medium (LB medium supplemented with 100 µg/ml ampicillin and 1% glucose) was subcultured at 1:100 dilution in 1L LB/amp/glu. Protein expression was induced at an OD₆₀₀ of 0.8 - 0.9 by the addition of IPTG to a final concentration of 0.2 mM. The culture was grown at 220 rpm overnight at 37°C. The bacteria were pelleted by centrifuging at 6000 rpm for 12 min; the pellet was re-suspended in 35 ml of cold TES buffer (0.2M Tris-Cl pH 8.0, 20% sucrose, 0.5mM EDTA). The suspension was incubated on ice and vortexed every 10 min for 1 hour. Then 45 ml of cold TES (1/8 volume of total volume) was added and immediately vortexed for 1 minute and for 15 seconds every 10 min thereafter

for 1 hour to extract the protein from the periplasm. The resulting supernatant containing V_HH was filtered through a 0.22 μm membrane and dialysed overnight into immobilized metal-affinity chromatography (IMAC) buffer A (10 mM HEPES pH 7.0, 500 mM NaCl). The protein was purified using HiTrap Chelating HP columns (GE Healthcare) as described previously (Arbabi-Ghahroudi 2009b). Eluted protein fractions were analyzed by SDS-PAGE and Western blotting before being dialysed against PBS as described previously (Arbabi-Ghahroudi 2009b). The purified protein fractions were pooled and dialyzed against PBS + 3 mM EDTA, and the protein concentration was determined.

Example 6: Biophysical characterization of anti-IGF1R V_HH IGF1R-5

[0097] The anti-IGF1R V_HH IGF1R-5 constructs expressed and purified in Example 5 were characterized using size exclusion chromatography, surface plasmon resonance analysis, and melting temperature analysis. Epitope mapping of the IGF1R-5 V_HH was also performed.

[0098] *Size exclusion chromatography:* Size exclusion chromatography employing Superdex™ 75 (GE Healthcare) was used to eliminate any possible aggregates prior to Surface Plasmon Resonance (SPR) analysis. The running buffer used was 10mM HEPES, pH7.4 containing 150mM NaCl, 3mM EDTA and 0.005% P20. Concentrations of fractions used for SPR analysis were determined by measuring absorbance at 280nm wavelength. The SEC analysis suggested that the antibodies were monomeric, based on the elution volume compared to standards (Figure 3A).

[0099] *Melting temperature:* The thermal stability of the IGF1R-5 V_HH constructs was evaluated using melting temperature (T_m) measurement by CD spectroscopy. A Jasco J-815 spectropolarimeter equipped with a Peltier thermoelectric type temperature control system (Jasco, Easton, MD, USA) was used to carry out experiments. A CD cuvette with a path length of 1 mm was used. The spectra were recorded over a wavelength range of 180 - 260 nm with scanning speed of 50 nm/min, digital integration time (DIT) of 4 s, a band width of 1 nm, data pitch of 1 nm, and an integration time of 1 s. To measure melting temperature or T_m (Greenfield, 2006a; 2006b), CD spectra were recorded over a temperature range of 30°C to 96°C. All CD spectra were subtracted from the blank corresponding to buffer spectra. Measurements were performed with 50 μg/mL V_HH in 100 mM sodium phosphate buffer, pH 7.4. Heat-induced protein denaturation was monitored at 210 nm for all variants. The fraction folded (ff) was obtained by a formula as described (Greenfield, 2006a; 2006b):

$$ff = ([\theta]_T - [\theta]_U) / ([\theta]_F - [\theta]_U) \quad \text{formula 1}$$

where [θ]_T is the molar ellipticity at any temperature, [θ]_F is the molar ellipticity of the fully folded protein at 30°C and [θ]_U is the molar ellipticity of the unfolded protein at 90°C. Melting temperature (T_m) was obtained as a midpoint of the unfolding curve (fraction folded (ff) versus temperature) by a nonlinear regression curve fit (Boltzman sigmoidal equation) using the graphing software GraphPad Prism (version 4.02 for Windows). The melting temperatures (T_m) of V_HH were determined based on ellipticity data assuming a two-state system, which is in agreement with the observed denaturation curves corresponding to a sharp transition into denaturation (Fig. 2D). The T_m values were taken at midpoint of the sigmoidal denaturation curves of fraction folded (ff) versus temperature. Results are shown in Figure 3B. The melting temperatures of all humanized V_HH were improved (higher) in comparison to the IGF1R-5 V_HH, suggesting improved biophysical properties

[0100] *Surface Plasmon Resonance (SPR):* The binding of monomeric IGF1R-5 V_HH constructs to immobilized recombinant human IGF1R (Example 1) was determined by SPR using BIACORE 3000 (GE Healthcare). Approximately 3000 Resonance Units (RU) of recombinant human IGF1R (R&D Systems) were immobilized on a Sensor chip CM5. Immobilization was carried out at a concentration of 10 μg/ml in 10mM acetate at pH4.0 using the amine coupling kit supplied by the manufacturer. The remaining binding sites were blocked with 1 M ethanolamine pH 8.5. An ethanolamine blocked surface was used as a reference surface. For the binding studies, analyses were carried out at 25°C in 10mM HEPES, pH7.4 containing 150mM NaCl, 3mM EDTA and 0.005% surfactant P20 (Polyoxyethylenesorbitan; GE Healthcare). Various concentrations of the IGF1R-5 V_HH were injected over the immobilized human IGF1R and reference surfaces at a flow rate of 20 μl/min. Surfaces were regenerated with 10mM glycine pH 2.0 with a contact time of 24 seconds. Data were analyzed with BIAevaluation 4.1 software (GE Healthcare). The results (see Figure 3C) show that the data fit well to a 1:1 model, giving K_D shown in Table 1. This shows that IGF1R-5 and the humanized variants are high-affinity single-domain antibody binders to the extracellular domain of human IGF1R.

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Table 1. Affinity of IGF1R-5 V_HH constructs for human (h) IGF1R as determined by surface plasmon resonance.

		h-IGF1R surface		
		K _D (nM)	k _d (s ⁻¹)	k _a (M ⁻¹ s ⁻¹)
5	IGF1R-5	0.6	3.4 x 10 ⁻⁴	5.3 x 10 ⁵
	IGF1R-5 H1	11.0	4.3 x 10 ⁻³	3.9 x 10 ⁵
	IGF1R-5 H2	7.5	2.9 x 10 ⁻¹³	3.8 x 10 ⁵
10	IGF1R-5 H3	0.8	4.1 x 10 ⁻⁴	5.2 x 10 ⁵
	IGF1R-5 H5	1.6	8.0 x 10 ⁻⁴	5.0 x 10 ⁵
	IGF1R-5 H6	1.7	7.2 x 10 ⁻⁴	4.3 x 10 ⁵

15 **[0101]** The SPR analyses were further used to demonstrate that IGF1R-5 V_HH does not bind to the same epitope on the receptor as the natural ligand IGF-1 (Figure 3D). The experiment was set up, carried out and analysed as described above. Binding to freshly immobilized human IGF1R surface was studied by injection of IGF1R-5 V_HH at a concentration of 25xK_D followed by co-injection of IGF1R-5 and human IGF-1 ligand, both at concentrations 25xK_D, with flow rates of 20μl/min and injection times of 5 minutes. Surfaces were regenerated by washing with running buffer. The IGF1R-5 V_HH bound the receptor, with a saturation reached at about 38RU on the non-regenerated surface. The IGF-1 ligand was able to bind to the IGF1R receptor in presence of the IGF1R-5 V_HH with the expected ~70RU (relative units). The simultaneous binding of both IGF1R-5 V_HH and IGF-1 to the receptor demonstrates that both bind to different epitopes.. SPR analysis of IGF1R-5 V_HH and Insulin receptor suggested that IGF1R-5 V_HH cannot bind the Insulin receptor (data not shown). Despite different relative units (RU) of IGF1R-5 binding to a regenerated versus a fresh, non-regenerated surface, the binding affinity determined on both surfaces was highly similar and could be reproduced in different experiments (data not shown).

20 **[0102]** *Epitope mapping:* Epitope mapping services were provided by Pepscan (Lelystad, The Netherlands; pepscan.com). The full-length sequence of the IGF1R is shown in Figure 2. Based on the epitope mapping results and the three-dimensional rendering of the related insulin receptor structure (as published in Protein Data Bank accession no. 2DTG, not shown), the epitope to which IGF1R-5 V_HH binds was suggested to be FENFLHNSIFVPR (SEQ ID NO:11).

Example 7: Internalization of the IGF1R-5 by brain endothelial cells

35 **[0103]** To determine whether IGF1R-5 is internalized into cells, svARBEc cells were incubated with Cy5-5-labelled IGF1R-5.

[0104] IGF1R-5 V_HH was labeled with NHS-Cy5.5. The labeling was done through a stable amine bond between primary amines (at the N-terminus and/or on lysine side chains of the protein) and the NHS ester. Typically, 10% v/v of 1M carbonate/bicarbonate buffer (759mM bicarbonate, 258mM carbonate) pH 9.3 was added to 4mg of V_HH prepared in PBS (1.06mM KH₂PO₄, 154mM NaCl, 5.6mM Na₂HPO₄) pH7.4 and adjusted to a final concentration of 4mg/mL. The NHS-Cy5.5, dissolved in DMSO at 10mg/mL, was added at a 2X molar ratio of dye to protein. The mixture was incubated at room temperature for 2 h with several inversions in a 1.5mL microcentrifuge tube. Following the incubation, unbound dye and reaction bi-products were filtered using Zeba Spin Desalting Columns, 7K MWCO (Pierce) and measured using a Beckman DU530 spectrophotometer (Beckman Coulter). Cy5.5-labeled IGF1R-5 or FC5 as a positive control (1mg/ml) were incubated with SV40 immortalized rat brain endothelial cells (svARBEc) at 4°C (Fig. 4, top panels), thus allowing only for passive, non-specific transport mechanism to occur, or at 37°C (Fig. 4, bottom panels) to allow for active transport such as receptor mediated endocytosis to take place. Co-staining with wheat germ agglutinin and DAPI was carried out to visualize the cell surface and the nucleus, respectively. Cells were observed under fluorescent microscope and images were captured.

50 **[0105]** If incubated at 4°C, IGF1R-5 was found outside the cells co-localizing with the cell membrane stained with wheat germ agglutinin. In contrast, when incubated at 37°C, IGF1R-5 accumulated in vesicles inside the cells, likely endosomes, suggesting that the antibody internalized into cells through an active transport mechanism. Similar behaviour was observed for FC5, previously shown to enter the cells by energy-dependent endocytosis via clathrin-coated vesicles (Abulrob et al. 2005)

Example 8: Production of IGF1R-5-Fc constructs

55 **[0106]** Constructs comprising IGF1R-5 V_HH fused to a murine antibody fragment crystallisable (Fc; mFc2b) were prepared, expressed, and isolated. Both a C-terminal construct (IGF1R-5-mFc) and an N-terminal IGF1R-5 mouse Fc

fusion construct (mFc-IGF1R-5) were made, the sequences of which are shown in Figures 5A-B, with a schematic of the molecules shown in Figure 5C. The fusion proteins (~80kDa) also comprised a N-terminal signal peptide (MEF-GLSWVFLVAILKGVQC; SEQ ID NO:40) that is not shown in the sequence of Figures 5A-B since it is cleaved following secretion.

5 **[0107]** The IGF1R-5 cDNA was cloned into mammalian expression vector pTT5 (Durocher 2002) containing the mouse Fc2b fragment. Polyplexes of the resulting vector were pre-formed by mixing 25 ml of plasmid DNA solution containing 187.5 µg pTT5-IR5mFc2b, 56.25 µg pTT-AKTdd (activated mutant of Protein Kinase B), 18.75 µg pTTo-GFP (to monitor transfection efficiency), and 112.5 µg of salmon testis DNA (Sigma-Aldrich); and 25 ml of PEI solution containing 1.125 mg of PEIproTM (PolyPlus Transfection), both made in F17 medium. The mixture was incubated for 10 minutes prior to addition to the cell culture. A 450 ml culture of CHO cells stably expressing a truncated EBNA1 protein (CHO-3E7) and grown in F17 medium (Invitrogen) was transfected with 50 ml of polyplexes. Twenty four hours post-transfection, the culture was fed with 12.5 ml of 40% (w/v) tryptone N1 (Organotechnie) solution and 1.25 ml of 200 mM valproic acid solution. The culture was harvested 8 days post-transfection and clarified by centrifugation. Clarified medium was filtered through a 0.22 µm membrane prior to its application on a column packed with 5 ml of protein-A MabSelect SuRe resin (GE Healthcare). After loading, the column was washed with 5 volumes of phosphate-buffered saline pH 7.1 (PBS) and the antibody was eluted with 100 mM sodium citrate buffer pH 3.0. Fractions containing the eluted antibody were pooled and a buffer exchange was performed by loading on a desalting Econo-Pac column (BioRad) equilibrated in PBS. Desalted antibody was then sterile-filtered by passing through a Millex GP (Millipore) filter unit (0.22 µm) and aliquoted.

20 Example 9: Pharmacokinetics of IGF1R-5-mFc in CSF and plasma

[0108] An *in vivo* assay was carried out to determine whether IGF1R-5-mFc (Example 8) is able to cross into the brain, and specifically into the cerebrospinal fluid (CSF), as well as to quantify its presence in CSF and serum.

25 **[0109]** The technique used for multiple sampling of cisterna magna CSF was developed at NRC by modification of previously described methods (Huang et al., 1995; Kornhuber et al., 1986; Yaksh and Rudy, 1976). Animals were housed singly in polypropylene cages, and were allowed free access to food and water. Experiments were done in a 12 h light/dark cycle at a temperature of 24°C and a relative humidity of 50 ± 5%. All animal procedures were approved by the NRC's Animal Care Committee and were in compliance with the Canadian Council of Animal Care guidelines.

30 **[0110]** Male Wistar rats, 8-10 weeks of age (weight range, 230-250 g) were anaesthetized with a ketamine/xylazine mixture (45/12 mg/kg, i.p.) and mounted in a stereotaxic instrument (any other device which will hold the animal's head firmly could be used). An incision beginning at a line joining the ears and extending about 3 cm in a caudal direction was made in the midline: the fascia was retracted from the skull exposing a superficial layer of neck muscles which were then separated by a midline incision beginning at the occipital crest and extending caudally about 2 cm. Separation of the superficial musculature exposed an underlying layer of muscles which was easily separated along the midline by blunt dissection. A scraping tool was used to free the muscles from their point of origin on the occipital bone for about 35 0.5 cm on either side of the midline; gentle retraction of the neck musculature exposed the atlanto-occipital membrane; the rat's head was then rotated downward to about a 45° angle. A small hole was made using the tip of a 27G disposable needle; when the incision is performed correctly, clear CSF should immediately well up through the slit; polyethylene tubing (PE-20) was then inserted 0.5 cm into the slit. The cannula was then fixed to the atlanto-occipital membrane with the aid of Histoacryl glue or Vetbond glue, allowing sample collection for long time periods (more than one week) and also preventing infections or other contaminants from entering the CSF; the cannula was then flushed thoroughly with artificial CSF. The collecting portion of the cannula was firmly attached to the occipital bone with dental cement. Once the cannula was fixed, the free part (4-5 cm) was threaded through the skin with a 20G needle, the muscle and the skin were then closed, the exposed portion was cut to desired length, and the tip was slightly heated to prevent backflow of CSF. Finally, the animal was placed in a recovery room for at least one week to allow the closing of the cisterna magna. This technique does not involve drilling through the skull, as described by other methods, avoiding damage of the meninges and the adjacent nervous tissue.

40 **[0111]** One week post-cisternal cannulation, 5 mg/kg of IGF1 R-5mFc or control A20.1mFc fusion was injected into the tail vein. Rats were anesthetized with 3% isoflurane; the first 10 µl of the CSF accumulated in the cannula were discarded. 10 µl of CSF was collected at 0.5, 1, 2, 24 and 48 h) post-injection. Blood samples (50 µl) were taken from the tail vein (Fluttert et al., 2000) using polymer gel tubes (Becton, Dickinson and Company Franklin Lakes, NJ USA) at the same time points; samples were centrifuged (15 min at 1600 rcf; room temperature). Samples were stored at -80°C until analysed. At the end of the experiments the rats were sacrificed by decapitation under anaesthetization.

45 **[0112]** Serum and CSF samples were analyzed by mass spectrometry and nanoLC-SRM based quantification as described in Example 7.

55 **[0113]** CSF collection is a delicate procedure during which CSF can be easily contaminated with blood. Since the amounts of V_HH s are expected to be much smaller in the CSF (<0.1%) than blood, even a slight contamination with blood could seriously compromise the value of an individual CSF sample. It was therefore necessary to develop stringent

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exclusion criteria for blood-contaminated CSF samples. To evaluate blood-CSF albumin ratio, a nanoLC-SRM method was developed for quantifying albumin levels in plasma and CSF. An albumin peptide APQVSTPTLVEAAR (SEQ ID NO:38) was selected based on its unique retention time and m/z value (Mol Pharm) in order to have minimum interference with other peptide peaks in the multiplex assay. The intensity of the peptide was quantified in both CSF and plasma samples using SRM as described above. The albumin ratio was calculated as follows for each rat:

$$\text{Albumin Ratio} = \text{Intensity per nL of plasma analyzed} / \text{Intensity per nL of CSF analyzed}$$

A ratio of 1500 and below was considered as blood contaminated.

[0114] Results are shown in Figure 9 and Table 2. The figure shows a 'typical' serum pharmacokinetic profile of Fc-containing molecule. CSF levels of IGF1R-5Fc increase over-time, peaking 24 h after injection, when they reach 2% of serum levels. While serum levels/decay is similar for control A20.1 Fc fusion molecules, its CSF levels are minimal, reaching 0.04% of serum levels at 24h. Table 2 shows pharmacokinetic parameters of IGF1R5-Fc and control A20.1-Fc in CSF and in serum after systemic intravenous administration of equimolar doses (5mg/kg V_HH--Fc fusions). Serum half-life of IGF1 R5-Fc and control A20.1-Fc were 23 h and 25 h, respectively.

[0115] Negative control A20.1 Fc (75kD) shows serum/CSF ratio at 24h similar to those described in the literature for molecules of the similar size. Typical serum/CSF (lumbar) ratio for albumin (60kD) at steady state is 0.1% whereas serum/CSF ratio for IgG is 0.07 (Shen et al., 2004; Lin, 2008). IGF1 R5-Fc serum/CSF ratio at 24h is 50-fold higher than that for A20.1mFc.

[0116] CSF/serum AUC ratio for each molecule is indicative of CNS 'exposure' of the construct over a 48h period. IGF1R-5-Fc shows 33-fold higher CNS exposure compared to A20.1Fc. The results indicate that IGF1R-5Fc gains facilitated access to CSF compared to control V_HH-Fc fusion molecule; this could occur via a specific 'secretion' by choroid plexus, or via 'washout' (diffusion through ependymal layer lining brain ventricular system or via a bulk flow of interstitial fluid) from brain parenchyma after crossing the BBB, or both (Abbott 2004; DeLange, 2002; Groothuis, 2007).

[0117] Additional constructs of ~110kDa and 180kDa have also been shown to be ferried across the BBB by IGF1R-5 and a humanized version (data not shown).

Table 2. Pharmacokinetic parameters of systemically injected IGF1R-5Fc and A20.1Fc in serum and cisterna magna CSF of rats.

		IGF1R-5Fc	A20.1Fc
CSF	AUC (h*ng/ml)	15750.5	774
Serum	AUC (h*ng/ml)	1369563	2247094
	Half-life (distribution phase) (h)	22.89	25.46
	Clearance (ml/h)	1.003	0.66
CSF/serum ratio (24h) (%)		1.988	0.04
CSF/serum AUC Ratio (%)		1.150038	0.034
Fold increase (CSF exposure)			33.82

Example 10: Transport of the IGF1R-5 across in vitro blood brain barrier model

[0118] To evaluate whether the IGF1R-5 V_HH and the constructs of Example 8 transmute the blood-brain barrier, an *in vitro* assay was used as described below. A flow chart summarizing the experiment is shown at Figure 6A.

[0119] SV40-immortalized Adult Rat Brain Endothelial Cells (Sv-ARBE) were used to generate an *in vitro* blood-brain barrier (BBB) model as described (Garberg et al., 2005; Haqqani et al., 2012). Sv-ARBE (80,000 cells/membrane) were seeded on a 0.1mg/mL rat tail collagen type I-coated tissue culture inserts (pore size-1 μm; surface area 0.9 cm², Falcon) in 1 ml of growth medium. The bottom chamber of the insert assembly contained 2 ml of growth medium supplemented with the immortalized neonatal rat astrocytes-conditioned medium in a 1:1 (v/v) ratio.

[0120] Equimolar amounts (5.6 μM) of positive (FC5) or negative controls (A20.1, a *Clostridium difficile* toxin A binding V_HH; and EG2, an EGFR binding V_HH), IGF1R-5 V_HH, and the C- and N-terminal mFc-IFG1R-5 constructs of Example

8 were tested for their ability to cross this rat *in vitro* BBB model.

[0121] Following exposure of equimolar amounts of the sdAb to the luminal side of the BBB, samples were taken after 15, 30 and 60min from the abluminal side. The sdAb content of each sample was then quantified by mass spectrometry (multiple reaction monitoring - isotope labeled internal standards; MRM - ILIS) as described by Haqqani et al.(2012) (see method description below).

[0122] *Determination of the apparent permeability coefficient:* Quantified values can be directly plotted or the P_{app} (apparent permeability coefficient) values can be determined with the given formula (Fig 5A) and plotted. The P_{app} value is commonly used to determine the ability of a molecule to cross the BBB. [Qr/dt = cumulative amount in the receiver compartment versus time; A = area of the cell monolayer; C0 = initial concentration of the dosing solution]. P_{app} values are a measure of the specific permeability of the compound across brain endothelial monolayer.

[0123] Results are shown in Figures 6B-D. The results given are average P_{app} values obtained from several independent experiments. Both negative controls have a very low P_{app} value, indicating that non-specific transport or paracellular transport of these V_{HH} s across the BBB model is minimal. IGF1R-5 has a high P_{app} value, indicating high rate of transport across the *in vitro* BBB model. The P_{app} for IGF1R-5 V_{HH} (Figure 6B) is similar to that of a positive control - BBB-permeable V_{HH} FC5 (WO 02/057445). Humanized IGF1R-5 variants showed similar or higher P_{app} values to wild-type IGF1R-5 (Figure 6C). Notably, IGF1R-5 H2 showed a significant 35% increase in P_{app} value compared to wild-type IGF1R5 (Figure 6C). Two humanized variants, IGF1R-5 H1 and IGF1R H2, that showed improved *in vitro* BBB crossing (P_{app} values), also showed faster 'off-rates' in the receptor binding studies by SPR carried out here (Figure 3C), suggesting that faster 'off-rates' are desirable to improve transcytosis of antibodies binding to BBB receptor-mediated transcytosis receptor IGF1R. IGF1R-5 fused to either N- or C-terminus of murine Fc (80kD) had similar P_{app} values to single-domain antibody IGF1R-5 (15kD), showing that molecules of 80kD can efficiently cross the BBB *in vitro* when fused with IGF1R. Notably, fusion to N-terminus of Fc provided some advantage (15% improvement in P_{app}). These data show that IGF1R-5 can be used as a platform to engineer bi-specific antibodies in particular to be fused to the C-terminus of a full IgG with therapeutic function that requires transport into the brain. These results provide strong indication that IGF1R-5 undergoes a facilitated trans-cellular transport across brain endothelial cells *in vitro* and could have similar properties *in vivo*. It is worth noting that constructs comprising IGF1R-5 or a humanized version linked to a cargo molecule (MW ~110kDa or 180kDa) have also been shown to be ferried across the BBB (data not shown).

[0124] *Absolute quantitation of V_{HH} using MRM-ILIS method.* The methods are all as described in Haqqani et al. (2012). Briefly, to develop the SRM (selected reaction monitoring also known as multiple reaction monitoring (MRM) assay for VHHs, each V_{HH} was first analyzed by nanoLC-MS/MS using data-dependent acquisition to identify all ionizable peptides. For each peptide, the 3 to 5 most intense fragment ions were chosen. An initial SRM assay was developed to monitor these fragments at attomole amounts of the digest (about 100-300 amol). Fragments that showed reproducible intensity ratios at low amounts (i.e., had Pearson $r^2 \geq 0.95$ compared to higher amounts) were considered stable and were chosen for the final SRM assay. To further optimize the assay, elution times for each peptide were also included, with care taken to not choose peptides that have close m/z (mass-to-charge ratio) and elution times.

[0125] A typical multiplexed SRM analysis of V_{HH} in cell media or body fluids (serum or cerebrospinal fluid (CSF)) involved spiking known amount of ILIS (0.1-10 nM) followed by injecting 100-400 ng of CSF or cultured media proteins (0.3-1 μ L) or about 50-100 ng of serum proteins (1-3 nanoliters) into the nanoLC-MS system. The precursor m/z of each target peptide ion was selected in the ion trap (and the remaining unrelated ions were discarded) at the specified elution time for the target, followed by collision induced dissociation (CID) fragmentation, and selection of only the desired fragment ions in the ion trap for monitoring by the detector. For quantification analysis, raw files generated by the LTQ (ThermoFisher) were converted to the standard mass spectrometry data format mzXML and intensities were extracted using an in-house software called Q-MRM (Quantitative-MRM; see Haqqani et al. 2012), which is a modified version of MatchRx software. For each V_{HH} , extracted-ion chromatograms were generated for each of its fragment ion that consisted of combined intensities within 0.25 Da of the fragment m/z over the entire elution time. To obtain a final intensity value for each fragment, all intensities within 0.5 min of the expected retention times were summed. A V_{HH} was defined as detectable in a sample if the fragments of at least one of its peptides showed the expected intensity ratios, i.e., the final intensity values showed a strong Pearson correlation $r \geq 0.95$ and $p < 0.05$ compared with the final intensities values of its corresponding pure V_{HH} .

[0126] Samples containing mixtures of V_{HH} (media, serum, CSF) were reduced, alkylated and trypsin-digested as previously described (Haqqani et al., 2012; Gergov et al., 2003). The digests (tryptic peptides) were acidified with acetic acid (5% final concentration) and analyzed on a reversed-phase nanoAcquity UPLC (Waters, Milford, MA) coupled to LTQ XL ETD or LTQ Orbitrap ETD mass spectrometer (ThermoFisher, Waltham, MA). The desired aliquot of the sample was injected and loaded onto a 300 μ m I.D. x 0.5 mm 3 μ m PepMaps C18 trap (ThermoFisher) then eluted onto a 100 μ m I.D. x 10 cm 1.7 μ m BEH130C18 nanoLC column (Waters) using a gradient from 0% - 20% acetonitrile (in 0.1% formic) in 1 minute, 20% - 46% in 16 min, and 46% - 95% in 1 min at a flow rate of 400 nL/min. The eluted peptides were ionized into the mass spectrometer by electrospray ionization (ESI) for MS/MS and SRM analysis using CID for fragmentation of the peptide ions. The CID was performed with helium as collision gas at normalized collision energy

of 35% and 30 ms of activation time. Ion injection times into linear ion trap were adjusted by the instrument using an automatic gain control (AGC) target value of 6×10^3 and a maximum accumulation time of 200 ms

[0127] The V_{HH} -specific peptides used for detection and quantification of each V_{HH} in multiplexed assay are shown in Table 3.

Table 3. Peptides used in nanoLC-SRM detection of FC5, FC5-ILIS, EG2, A20.1, IGF-1R-5 and albumin. (a) In various studies described, assays were multiplexed in different combinations for simultaneous monitoring in the same sample; (b) Heavy-labeled peptide; (c) Limits of detection and quantification of the SRM assay for each peptide ranged from 1.5-2.5 ng/ml. 1 ng/mL corresponds to about 60-70 pM of V_{HH} . A20-1 as described in Hussack et al, 2011b; EG2 as described in Iqbal et al, 2010.

Protein	Signatures	SEQ ID NO:	Unique
IGF1R-5	EFVATIDWGDGGAR	27	Yes
	TIDNYAMAWSR	28	Yes
	LEESGGGLVQAGGSLR	29	
FC5	ITWGGDNTFYNSVK	30	Yes
FC5-ILIS	ITWGGDNTFYNSVK ^(b)	30	Yes
A20.1	TTYADSVK	31	Yes
	EFVAAGSSTGR	32	Yes
	TFSMDPMAWFR	33	Yes
	DEYAYWGQGTQVTVSSGQAGQGSEK	34	Yes
EG2	DFSDYVMGWFR	35	Yes
	LEESGGGLVQAGDSLK	36	Yes
	NMYYLQMNSLKPEDTAVYYCAVNSAGTYVSPR	37	Yes
Albumin	APQVSTPTLVEAAR	38	Yes

Example 11: Conjugation of IGF1R-5 to Galanin

[0128] To determine whether IGF1R-5 can cross the blood-brain barrier (BBB) *in vivo* and 'ferry' across the BBB a molecule that cannot cross the BBB on its own, the neuropeptide Galanin was chemically conjugated to IGF1R-5 V_{HH} or IGF1R-5 fused to either C- or N-terminal of murine Fc and administered systemically. Galanin is a neuroactive peptide that produces analgesia by binding GalR1 and GalR2 expressed in brain tissue. When given peripherally, Galanin has no analgesic effects because it cannot cross the BBB on its own (Robertson et al., 2011).

[0129] The IGF1R-5 V_{HH} and Fc-fusion constructs were conjugated to a rat Galanin (Gal) fragment with cysteamide modified C-terminus (Biomatic) (GWTLNSAGYLLGPHAIIDNHRFSFSDKHGLT-cysteamide, SEQ ID NO:39). The scheme for conjugation is shown in Figure 7.

[0130] Briefly, 5mg of IGF1R-5 V_{HH} (Example 5) in 0.5X PBS, 2.5mM EDTA at [2mg/ml] were mixed with 436.4 μ l of 2.5mg/ml sulfosuccinimidyl-4-(N-maleimidomethyl)cyclohexane-1-carboxylate (Sulfo-SMCC) (7.5x excess molar ratio). The mixture was then flushed with nitrogen gas and incubated for 30 minutes at room temperature (RT) to allow the NHS ester arm of the Sulfo-SMCC to react with amines on the V_{HH} . Subsequently, the unreacted Sulfo-SMCC was removed from the maleimide-activated IGF1R-5 V_{HH} using a 10ml 7K Zeba column (Pierce). Prior to sample loading, the column was washed 3 times with 5ml PBS and spun at 1000xg for 2 min. After sample loading, the column was topped-up with 200 μ l of PBS and was spun for 2 min at 1000xg. For the IGF1 R-Fc constructs, 5 mg were reacted with -68 μ l of Sulfo-SMCC (6.5x excess molar ratio) as described above.

[0131] Separately and concurrently, cysteamide modified C-TERM galanin (Gal-cya) was prepared by dissolving 10 mg of lyophilized powder in 10 ml of endotoxin free water to make a 1mg/ml stock (the galanin-cya powder has a small amount of DTT to prevent disulfide bridge formation during purification). Finally, 100 μ l of 0.5M EDTA was added (5mM final concentration).

[0132] The purified maleimide-activated IGF1R-5 V_{HH} and IGF1R-Fc constructs (2.6 ml) were diluted to 5ml with 0.5X PBS, 2.5mM EDTA and then 5ml or 1ml Gal-cya respectively was added while vortexing. The samples were flushed with nitrogen, sealed and incubated overnight at 4°C. The next day, the unreacted Gal-cya was removed using Amicon-15 10K and 30K column (Millipore), respectively. The samples were added to the column and spun at 4000xg for 7 minutes until the volume was reduced to 5ml. 5ml of 0.5X PBS, 2.5mM EDTA was added to the remaining 5ml sample in the column's insert and was spun again until the sample was reduced to 4ml. The conjugated samples were then

added to a 10ml 7K Zeba column (Pierce), prepared as described above, and then spun for 2 min at 1000xg.

[0133] Conjugated IGF1R-5-Gal and IGF1R-Fc-Gal samples were then ran on a 16% or 10% SDS-PAGE non-reducing gel and silver stained to confirm shift in molecular weight size after conjugation. The reaction was titrated to achieve ~1 to 2 galanin molecules per V_HH or IGF1R-Fc constructs.

[0134] *Endotoxin removal and determination of endotoxin levels:* Endotoxins were removed using Amicon Ultra - molecular weight cut-off (MWCO) cellulose membrane spin columns (Millipore). First, 15ml of the V_HH preparation was passed through an Amicon-15-50K MWCO column by centrifugation at 4000xg for 10minutes; the elution was collect. This elution was then added to an Amicon-15-10K MWCO column and spun at 4000xg for 7-10minutes resulting in a reduction of the supernatant volume from 15 ml to 7.5ml. The supernatant volume in the column was adjusted back to 15 ml by adding PBS. The column was spun again as described above. The supernatant was collect and the endotoxin levels were measured with EndoSafe-PTS system using cartridges with a sensitivity range of 10-0.1EU/ml (Charles River Laboratories International). 25ul of sample was loaded in each of the 4 wells on the cartridge and diluted if necessary. Only samples with EU<1 per 1 mg were used for animal studies.

[0135] The Fc-fusion constructs were produced in mammalian cells under sterile conditions. Endotoxin levels were measured as described above in the samples purified from cells and only samples with <0.5 EU per 1 mg were used for animal studies.

Example 12: Transport of the IGF1R-5-Gal using the Hargreaves model

[0136] To evaluate whether the IGF1R-5-Gal, mFc-IGF1R-5-Gal, and IGF1R-5-mFc-Gal (Example 11) transmigrate the blood-brain barrier, an *in vivo* assay, was utilized, previously described in International Patent Publication No. WO 2011/127580.

[0137] A rat model of inflammatory hyperalgesia, similar to that described by Hargreaves et al. (1988), was used. Animals were housed in groups of three (Hargreaves model) per polypropylene cage, and were allowed free access to food and water. Experiments were done in a 12 h light/dark cycle at a temperature of 24°C and a relative humidity of 50 ± 5%. All animal procedures were approved by the NRC's Animal Care Committee and were in compliance with the Canadian Council of Animal Care guidelines.

[0138] In this model, male Wistar rats, 6-8 weeks (weight range 230-250 g) old were injected with low volume (100 µl with a 30-gauge needle) of complete Freund's adjuvant (CFA; heat-killed *M. tuberculosis* (Sigma) suspended in oil:saline 1:1 emulsion) into the right hind paw under brief isoflurane anesthesia (3%). CFA induces the release of pro-inflammatory substances that activate nociceptors and create a chronic pain state and hyperalgesia (a heightened sensitivity to noxious heat). The paw withdrawal latency was measured by the application of a radiant stimulus in the plantar surface in both hind paws (inflamed and non-inflamed control paw) using the plantar Analgesia Meter equipment for paw stimulation (IITC Model # 336TG Life Science, Inc.). The time taken by the animal to respond by licking or flicking its paw was interpreted as positive response (paw withdrawal latency). The light-intensity lamp was adjusted to elicit baseline paw withdrawal latencies between 17 to 20 s in both hind paws before CFA administration. If a withdrawal response does not occur within 20 s, the light beam was automatically turned off to avoid tissue damage and the paw was assigned the maximum score.

[0139] Two days post-CFA injection and prior to the administration of the compounds, the animals were manipulated and acclimatized in the analgesia meter equipment for at least 60 min with the aim to reduce stress and prevent false positive responses. The baseline was measured in both paws to verify the developed pain (thermal hyperalgesia); the non-inflamed paw was used as a control against the injected paw. Animals with paw withdrawal latency of more than 6 s for the "inflamed paw" and less than 17s for the "normal paw" were excluded from the experiment.

[0140] To determine whether IGF1R-5-Gal, mFc-IGF1R-5-Gal, and IGF1R-5-mFc-Gal are delivered across the blood brain barrier and can engage target receptors (GalR1 and 2) in brain parenchyma, the rats received one tail vein injection of IGF1R-5-Galanin (2.93mg/kg or 5.85mg/kg; endotoxin EU=<1/mg), or mFc-IGF1R-5-Gal Gal (1.98 mg/kg or 4.96 mg/kg; <0.5EU/mg), or IGF1R-5-mFc-Gal (2.0 mg/kg or 5.0 mg/kg; <0.5EU/mg) or control compounds three days post-CFA injection. The paw withdrawal latency (PWL) was tested for each hind paw (inflamed and non-inflamed) every 15 min for 3 hours. An increased latency of paw withdrawal indicates successfully induction of analgesia, which can only be obtained through successful delivery of Galanin into the brain parenchyma by IGF1R-5. Galanin can only induce analgesia when present in the brain parenchyma and on its own cannot cross an intact BBB (Robertson et al., 2011).

[0141] Results were analyzed as temporal courses of Paw Withdrawal Latencies (PWL, sec) versus time (min or hrs) (Fig 8A-D).

[0142] The results show that intravenously administered galanin does not reduce pain compared to PBS. In contrast, a single injection of FC5-Gal, IGF1R5-Gal, mFc-IGF1R-5-Gal, or IGF1R-5-mFc-Gal all produced dose-dependent suppression of thermal hyperalgesia, suggesting that IGF1R-5 V_HH is the entity responsible for 'ferrying' Galanin across the BBB to produce a pharmacological effect by binding to GalR1 and/or 2 in the brain parenchyma. The IGF1R-5-Gal effect is dose-dependent and significantly more pronounced than that induced by FC5-Gal, suggesting that the IGF1R

receptor has a higher rate of BBB transport than the putative FC5 receptor. These results demonstrate that IGF1R-5 V_HH can 'ferry' large molecules (3kDa - 80 kDa) across the BBB using receptor-mediated transcytosis pathway. Active, receptor mediated transport is required since the BBB is known to prevent the passage of all hydrophilic molecules greater than 0.5 kDa.

Example 13: Immunodetection of IGF1R-5-mFc

[0143] To ascertain that high levels of IGF1R-5mFc detected in the CSF after peripheral administration originate at least in part from parenchymal extracellular space, in other words, that the intact construct had crossed the BBB, immunodetection of IGF1R-5-mFc in rat brains was performed.

[0144] Briefly, brains of animals from the experiment of Example 11 were harvested immediately following animal perfusion with PBS (i.e. 48h after 5mg/kg tail-vein injection of IGF1R-5-mFc). ; The brains were frozen and sectioned on cryotome into 12 μm sections. Sections were fixed for 10 min RT in 100%Methanol, washed 3x in PBS and incubated for 1 h in 10% normal goat serum (NGS) containing 0.3% TritonX-100 in 1x PBS. Goat anti-m-IgG Fcy-cy3 (Cat#115-165-071, Jackson Immuno Research, lot#106360) 1:200 in 5%NGS containing 0.3% TritonX-100 in 1XPBS was applied overnight at 4°C Sections were washed three times in 1x PBS and incubated in rabbit-anti-GFAP (Cat# Z0334, Dako) 1:500 in 5%NGS/1XPBS for 1hr at room temperature. Section were washed gain three times in 1xPBS, and then incubated in 1:300 goat anti-mouse Alexa 647 (A21235, Invitrogen) or 1:500 goat anti-rabbit Alexa 647(A21244, Invitrogen) in 1xPBS. Vasculature-staining lectin RCAI (Cat# FL-1081, Vector) 1:500 in 1xPBS was then added for 10 min. After washing three times with 1xPBS, sections were covered with a cover slip in Dako fluorescent mounting medium (Cat#S3023, Dako) and spiked with 2μg/mL Hoechst (Cat#H3570, Invitrogen) to stain nuclei. Images were captured with Olympus 1X81 Fluorescent Microscope using 10X and 60X Objectives and channels as shown in Table 4.

Table 4. Objectives and channels used for fluorescent microscopy.

	Fluorescent molecule	Excitation (nm)	Emmision (nm)
RCAI-vessels	FITC	495	518
Hoechst33342- nuclei	Hoechst	350	461
IGF1R-5-m-Fc	Cy3	531	593
GFAP	Alexa 647	628	692

[0145] Results are shown in Figure 10. Immunodetection of mouse Fc showed strong staining of brain vessels throughout different brain regions, as well as staining of perivascular brain parenchyma, indicating that the IGF1R-5Fc is accumulated in brain vessels and also transmigrated the BBB into surrounding brain parenchyma. The results support the assertion that increased CSF levels of IGF1 R-Fc are indicative of the construct transmigration across the BBB. This is further strongly supported by the observation that galanin linked to IGF1R-5 induced pharmacological response (analgesia) on parenchymal GalR1 and GalR2 receptors. Collectively, in vitro BBB transmigration results, in vivo pharmacokinetic (serum/CSF levels) and pharmacodynamic (Hargreaves model) results demonstrate that IGF1R-5 V_HH transmigrates the intact BBB at significantly higher rates than other V_HHs via active receptor-mediated transcytosis triggered by its binding to IGF1R epitopes and that it can 'ferry' a range (1-80kD) of otherwise non-permeable molecules across the blood-brain barrier.

Example 14: IGF1R-5 effect on 'physiological' function of IGF1R

[0146] From a safety perspective, it is important to show that the antibody of the invention does not interfere with the physiological function of the receptor- i.e., signaling induced by its natural ligand, IGF-1 - when engaging its receptor for drug delivery via receptor-mediated transcytosis. In view of this, it is important to demonstrate that IGF1R-5 V_HH or IGF1R-5-mFc do not interfere with physiological signaling through IGF1R or the related insulin receptor (IR) induced by their natural ligands.

[0147] To determine whether IGF1R-5 induces signaling through IGF1R or IR alone, or interferes with signalling as stimulated by the receptor's natural ligands, IGF-1 or insulin, their effect on phosphorylation of the receptors themselves or receptor-stimulated downstream kinase, Akt, was determined in SV-ARBEC cells.

[0148] SV-ARBEC were grown to confluence in M199 base medium supplemented with peptone, D-glucose, BME amino acids, BME vitamins, antibiotic/antimycotic solution and fetal bovine serum, according to art-known methods. The cells were switched into serum free medium 18 h prior to treatment. IGF1R-5 V_HH or IGF1R-5-Fc fusion (100nM or 500nM) was added to the cells 1 h prior to the addition of either 200ng/ml IGF-1, 10μg/ml of insulin or vehicle. The cells were incubated with ligands or vehicle for 20 minutes and then washed twice in Hank's balanced salt solution. Cells

were subsequently lysed using 1x RIPA buffer (Cell Signaling Technology) supplemented with 1% Triton-x 100 and protease inhibitor cocktail (Sigma). The cells were given 2 x 20 second bursts in a water bath sonicator and lysates were clarified by centrifugation at 14,000rpm for 10 minutes. Protein concentration was determined using DC protein assay system (BIO-RAD laboratories). Equal μg of protein samples were resolved on a 4-20% gradient SDS polyacrylamide gel at 125V and transferred to PVDF membrane. Phospho-Akt (Ser 473) was detected by overnight incubation in 1:1000 dilution of the primary antibody against this target (Cell Signaling Technology) followed by a 1h incubation with goat anti-rabbit IgG-HRP secondary antibody then reacted with ECL Plus reagent and visualized on autoradiography film. Densitometry values were determined using Un-Scan-It software (Silk Scientific Inc.).

[0149] The results are shown in Figure 11. Western blot analyses of Akt phosphorylation showed that IGF1R-5 did not inhibit Akt phosphorylation induced by 10 $\mu\text{g}/\text{ml}$ of insulin or by 200ng/ml of IGF-1 when co-applied with at 100 nM of IGF1R-5 or IGF1R-5-mFc or 500nM IGF1 R-5-mFc. Neither did either of the V_HH or Fc fusions induce Akt signalling on its own (Fig 11A, 11B and 11C, labelled "-5"). The results demonstrate that even in bivalent display in the Fc fusion format IGF1R-5 does not trigger receptor dimerization and down-stream signaling, and therefore does not interfere with the receptor function in the presence of the natural ligand. This feature of IGF1R-5 ('silent binder') is important for its application as a BBB carrier for therapeutics, since it confers a favourable safety profile.

[0150] The embodiments and examples described herein are illustrative and are not meant to limit the scope of the invention as claimed. Variations of the foregoing embodiments, including alternatives, modifications and equivalents, are intended by the inventors. Furthermore, the discussed combination of features might not be necessary for the inventive solution.

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European Patent No. 626390

US Patent No. 5693761

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US Patent No. 5766886

US Patent No. 5821123

5 US Patent No. 5859205

US Patent No. 5869619

10 US Patent No. 6054297

US Patent No. 6180370

WO 02/057445

15 WO 2011/127580

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20 WO/2004/076670

WO2003/046560

SEQUENCE LISTING

25 [0152]

<110> National Research Council of Canada Stanimirovic, Danica Kemmerich, Kristin Sulea, Traian Arbabi-Ghahroudi, Mehdi Haqqani, Arsalan Gilbert, Renald Massie, Bernard

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<150> 61/948,831

35 <151> 2014-03-06

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 35 40 45

 Xaa Thr Ile Asp Trp Gly Asp Gly Gly Xaa Arg Tyr Ala Asn Ser Val
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 Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn Xaa Lys Xaa Thr Xaa Tyr
 65 70 75 80

 Leu Gln Met Asn Xaa Leu Xaa Xaa Glu Asp Thr Ala Val Tyr Xaa Cys
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 15 Ala Thr Ile Asp Trp Gly Asp Gly Gly Ala Arg Tyr Ala Asn Ser Val
 20 Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn Ala Lys Gly Thr Met Tyr
 25 Leu Gln Met Asn Asn Leu Glu Pro Glu Asp Thr Ala Val Tyr Ser Cys
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 15 Ser Thr Ile Asp Trp Gly Asp Gly Gly Thr Arg Tyr Ala Asn Ser Val
 20 Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn Ser Lys Asn Thr Leu Tyr
 25 Leu Gln Met Asn Ser Leu Arg Ala Glu Asp Thr Ala Val Tyr Tyr Cys
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 10 Ala Met Ala Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val
 15 Ala Thr Ile Asp Trp Gly Asp Gly Gly Thr Arg Tyr Ala Asn Ser Val

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15	Ala	Met	Ala	Arg	Gln	Ser	Arg	Val	Asn	Leu	Asp	Val	Ala	Arg	Tyr	Asp
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50	Lys	Gly	Arg	Phe	Thr	Ile	Ser	Arg	Asp	Asn	Ser	Lys	Asn	Thr	Met	Tyr
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60	Ala	Met	Ala	Arg	Gln	Ser	Arg	Val	Asn	Leu	Asp	Val	Ala	Arg	Tyr	Asp
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Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly Arg Thr Ile Asp Asn Tyr
 20 25 30

20

Ala Met Ala Trp Ser Arg Gln Ala Pro Gly Lys Asp Arg Glu Phe Val
 35 40 45

25

Ala Thr Ile Asp Trp Gly Asp Gly Gly Thr Arg Tyr Ala Asn Ser Val
 50 55 60

30

Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn Ser Lys Gly Thr Met Tyr
 65 70 75 80

Leu Gln Met Asn Ser Leu Arg Ala Glu Asp Thr Ala Val Tyr Ser Cys
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Ala Met Ala Arg Gln Ser Arg Val Asn Leu Asp Val Ala Arg Tyr Asp
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Tyr Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ser
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 10 Ala Met Ala Trp Ser Arg Gln Ala Pro Gly Lys Asp Arg Glu Phe Val
 15 Ser Thr Ile Asp Trp Gly Asp Gly Gly Thr Arg Tyr Ala Asn Ser Val
 20 Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn Ser Lys Asn Thr Leu Tyr
 25 Leu Gln Met Asn Ser Leu Arg Ala Glu Asp Thr Ala Val Tyr Tyr Cys
 30 Ala Met Ala Arg Gln Ser Arg Val Asn Leu Asp Val Ala Arg Tyr Asp
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5 Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly Arg Thr Ile Asp Asn Tyr
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10 Ala Met Ala Trp Ser Arg Gln Ala Pro Gly Lys Asp Arg Glu Phe Val
 35 40 45

15 Ala Thr Ile Asp Trp Gly Asp Gly Gly Ala Arg Tyr Ala Asn Ser Val
 50 55 60

Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn Ala Lys Gly Thr Met Tyr
 65 70 75 80

20

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Leu Gln Met Asn Asn Leu Glu Pro Glu Asp Thr Ala Val Tyr Ser Cys
 85 90 95
 5 Ala Met Ala Arg Gln Ser Arg Val Asn Leu Asp Val Ala Arg Tyr Asp
 100 105 110
 Tyr Trp Gly Gln Gly Thr Gln Val Thr Val Ser Ser Met Thr Val Asp
 115 120 125
 10 Lys Lys Leu Glu Pro Ser Gly Pro Ile Ser Thr Ile Asn Pro Cys Pro
 130 135 140
 15 Pro Cys Lys Glu Cys His Lys Cys Pro Ala Pro Asn Leu Glu Gly Gly
 145 150 155 160
 20 Pro Ser Val Phe Ile Phe Pro Pro Asn Ile Lys Asp Val Leu Met Ile
 165 170 175
 Ser Leu Thr Pro Lys Val Thr Cys Val Val Val Asp Val Ser Glu Asp
 180 185 190
 25 Asp Pro Asp Val Gln Ile Ser Trp Phe Val Asn Asn Val Glu Val His
 195 200 205
 30 Thr Ala Gln Thr Gln Thr His Arg Glu Asp Tyr Asn Ser Thr Ile Arg
 210 215 220
 35 Val Val Ser Thr Leu Pro Ile Gln His Gln Asp Trp Met Ser Gly Lys
 225 230 235 240
 Glu Phe Lys Cys Lys Val Asn Asn Lys Asp Leu Pro Ser Pro Ile Glu
 245 250 255
 40 Arg Thr Ile Ser Lys Ile Lys Gly Leu Val Arg Ala Pro Gln Val Tyr
 260 265 270
 45 Ile Leu Pro Pro Pro Ala Glu Gln Leu Ser Arg Lys Asp Val Ser Leu
 275 280 285
 Thr Cys Leu Val Val Gly Phe Asn Pro Gly Asp Ile Ser Val Glu Trp
 290 295 300
 50 Thr Ser Asn Gly His Thr Glu Glu Asn Tyr Lys Asp Thr Ala Pro Val
 305 310 315 320
 55 Leu Asp Ser Asp Gly Ser Tyr Phe Ile Tyr Ser Lys Leu Asn Met Lys
 325 330 335

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Thr Ser Lys Trp Glu Lys Thr Asp Ser Phe Ser Cys Asn Val Arg His
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 Gly Lys
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 20 25 30

30
 Phe Ile Phe Pro Pro Asn Ile Lys Asp Val Leu Met Ile Ser Leu Thr
 35 40 45

35
 Pro Lys Val Thr Cys Val Val Val Asp Val Ser Glu Asp Asp Pro Asp
 50 55 60

40
 Val Gln Ile Ser Trp Phe Val Asn Asn Val Glu Val His Thr Ala Gln
 65 70 75 80

45
 Thr Gln Thr His Arg Glu Asp Tyr Asn Ser Thr Ile Arg Val Val Ser
 85 90 95

50
 Thr Leu Pro Ile Gln His Gln Asp Trp Met Ser Gly Lys Glu Phe Lys
 100 105 110

55
 Cys Lys Val Asn Asn Lys Asp Leu Pro Ser Pro Ile Glu Arg Thr Ile
 115 120 125

Ser Lys Ile Lys Gly Leu Val Arg Ala Pro Gln Val Tyr Ile Leu Pro
 130 135 140

Pro Pro Ala Glu Gln Leu Ser Arg Lys Asp Val Ser Leu Thr Cys Leu
 145 150 155 160

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Val Val Gly Phe Asn Pro Gly Asp Ile Ser Val Glu Trp Thr Ser Asn
 165 170 175

5 Gly His Thr Glu Glu Asn Tyr Lys Asp Thr Ala Pro Val Leu Asp Ser
 180 185 190

10 Asp Gly Ser Tyr Phe Ile Tyr Ser Lys Leu Asn Met Lys Thr Ser Lys
 195 200 205

15 Trp Glu Lys Thr Asp Ser Phe Ser Cys Asn Val Arg His Glu Gly Leu
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Lys Asn Tyr Tyr Leu Lys Lys Thr Ile Ser Arg Ser Pro Gly Gly Gly
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20 Gly Ser Gly Gly Gly Gly Ser Gln Val Lys Leu Glu Glu Ser Gly Gly
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25 Gly Leu Val Gln Ala Gly Gly Ser Leu Arg Leu Ser Cys Ala Ala Ser
 260 265 270

30 Gly Arg Thr Ile Asp Asn Tyr Ala Met Ala Trp Ser Arg Gln Ala Pro
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Gly Lys Asp Arg Glu Phe Val Ala Thr Ile Asp Trp Gly Asp Gly Gly
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35 Ala Arg Tyr Ala Asn Ser Val Lys Gly Arg Phe Thr Ile Ser Arg Asp
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40 Asn Ala Lys Gly Thr Met Tyr Leu Gln Met Asn Asn Leu Glu Pro Glu
 325 330 335

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50 Val Ser Ser
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5 Ala Gly Val Cys Val Pro Ala Cys Pro Pro Asn Thr Tyr Arg Phe Glu
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10 Gly Trp Arg Cys Val Asp Arg Asp Phe Cys Ala Asn Ile Leu Ser Ala
275 280 285

Glu Ser Ser Asp Ser Glu Gly Phe Val Ile His Asp Gly Glu Cys Met
290 295 300

15 Gln Glu Cys Pro Ser Gly Phe Ile Arg Asn Gly Ser Gln Ser Met Tyr
305 310 315 320

20 Cys Ile Pro Cys Glu Gly Pro Cys Pro Lys Val Cys Glu Glu Glu Lys
325 330 335

Lys Thr Lys Thr Ile Asp Ser Val Thr Ser Ala Gln Met Leu Gln Gly
340 345 350

25 Cys Thr Ile Phe Lys Gly Asn Leu Leu Ile Asn Ile Arg Arg Gly Asn
355 360 365

30 Asn Ile Ala Ser Glu Leu Glu Asn Phe Met Gly Leu Ile Glu Val Val
370 375 380

35 Thr Gly Tyr Val Lys Ile Arg His Ser His Ala Leu Val Ser Leu Ser
385 390 395 400

Phe Leu Lys Asn Leu Arg Leu Ile Leu Gly Glu Glu Gln Leu Glu Gly
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40 Asn Tyr Ser Phe Tyr Val Leu Asp Asn Gln Asn Leu Gln Gln Leu Trp
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45 Asp Trp Asp His Arg Asn Leu Thr Ile Lys Ala Gly Lys Met Tyr Phe
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Ala Phe Asn Pro Lys Leu Cys Val Ser Glu Ile Tyr Arg Met Glu Glu
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465 470 475 480

55 Asn Asn Gly Glu Arg Ala Ser Cys Glu Ser Asp Val Leu His Phe Thr
485 490 495

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Ser Thr Thr Thr Ser Lys Asn Arg Ile Ile Ile Thr Trp His Arg Tyr
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5 Arg Pro Pro Asp Tyr Arg Asp Leu Ile Ser Phe Thr Val Tyr Tyr Lys
 515 520 525

10 Glu Ala Pro Phe Lys Asn Val Thr Glu Tyr Asp Gly Gln Asp Ala Cys
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15 Gly Ser Asn Ser Trp Asn Met Val Asp Val Asp Leu Pro Pro Asn Lys
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25 Tyr Ala Val Tyr Val Lys Ala Val Thr Leu Thr Met Val Glu Asn Asp
 580 585 590

30 His Ile Arg Gly Ala Lys Ser Glu Ile Leu Tyr Ile Arg Thr Asn Ala
 595 600 605

35 Ser Val Pro Ser Ile Pro Leu Asp Val Leu Ser Ala Ser Asn Ser Ser
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40 Ser Gln Leu Ile Val Lys Trp Asn Pro Pro Ser Leu Pro Asn Gly Asn
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45 Leu Ser Tyr Tyr Ile Val Arg Trp Gln Arg Gln Pro Gln Asp Gly Tyr
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50 Leu Tyr Arg His Asn Tyr Cys Ser Lys Asp Lys Ile Pro Ile Arg Lys
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Arg Ser Arg Asn Thr Thr Ala Ala Asp Thr Tyr Asn Ile Thr Asp Pro
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5 Glu Glu Leu Glu Thr Glu Tyr Pro Phe Phe Glu Ser Arg Val Asp Asn
770 775 780

10 Lys Glu Arg Thr Val Ile Ser Asn Leu Arg Pro Phe Thr Leu Tyr Arg
785 790 795 800

15 Ile Asp Ile His Ser Cys Asn His Glu Ala Glu Lys Leu Gly Cys Ser
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Ala Ser Asn Phe Val Phe Ala Arg Thr Met Pro Ala Glu Gly Ala Asp
820 825 830

20 Asp Ile Pro Gly Pro Val Thr Trp Glu Pro Arg Pro Glu Asn Ser Ile
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25 Phe Leu Lys Trp Pro Glu Pro Glu Asn Pro Asn Gly Leu Ile Leu Met
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Tyr Glu Ile Lys Tyr Gly Ser Gln Val Glu Asp Gln Arg Glu Cys Val
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40 Asn Gly Ser Trp Thr Asp Pro Val Phe Phe Tyr Val Gln Ala Lys Thr
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Gly Tyr Glu Asn Phe Ile His Leu Ile Ile Ala Leu Pro Val Ala Val
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50 Lys Arg Asn Asn Ser Arg Leu Gly Asn Gly Val Leu Tyr Ala Ser Val
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Asn Pro Glu Tyr Phe Ser Ala Ala Asp Val Tyr Val Pro Asp Glu Trp
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55 Glu Val Ala Arg Glu Lys Ile Thr Met Ser Arg Glu Leu Gly Gln Gly

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75	Glu	Gln	Val	Leu	Arg	Phe	Val	Met	Glu	Gly	Gly	Leu	Leu	Asp	Lys
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Pro Asp Asn Cys Pro Asp Met Leu Phe Glu Leu Met Arg Met Cys
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5 Trp Gln Tyr Asn Pro Lys Met Arg Pro Ser Phe Leu Glu Ile Ile
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10 Ser Ser Ile Lys Glu Glu Met Glu Pro Gly Phe Arg Glu Val Ser
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15 Phe Tyr Tyr Ser Glu Glu Asn Lys Leu Pro Glu Pro Glu Glu Leu
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Asp Leu Glu Pro Glu Asn Met Glu Ser Val Pro Leu Asp Pro Ser
 1295 1300 1305

20 Ala Ser Ser Ser Ser Leu Pro Leu Pro Asp Arg His Ser Gly His
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25 Lys Ala Glu Asn Gly Pro Gly Pro Gly Val Leu Val Leu Arg Ala
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Ser Phe Asp Glu Arg Gln Pro Tyr Ala His Met Asn Gly Gly Arg
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	65					70					75					80
15	Leu	Gln	Met	Asn	Xaa	Leu	Xaa	Xaa	Glu	Asp	Thr	Ala	Val	Tyr	Xaa	Cys
					85					90					95	
20	Ala	Met	Ala	Arg	Gln	Ser	Arg	Val	Asn	Leu	Asp	Val	Ala	Arg	Tyr	Asp
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25	Tyr	Trp	Gly	Gln	Gly	Thr	Xaa	Val	Thr	Val	Ser	Ser	Met	Thr	Val	Asp
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40	Pro	Ser	Val	Phe	Ile	Phe	Pro	Pro	Asn	Ile	Lys	Asp	Val	Leu	Met	Ile
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45	Ser	Leu	Thr	Pro	Lys	Val	Thr	Cys	Val	Val	Val	Asp	Val	Ser	Glu	Asp
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60	Val	Val	Ser	Thr	Leu	Pro	Ile	Gln	His	Gln	Asp	Trp	Met	Ser	Gly	Lys
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65	Glu	Phe	Lys	Cys	Lys	Val	Asn	Asn	Lys	Asp	Leu	Pro	Ser	Pro	Ile	Glu
					245					250					255	
70	Arg	Thr	Ile	Ser	Lys	Ile	Lys	Gly	Leu	Val	Arg	Ala	Pro	Gln	Val	Tyr
				260					265					270		
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			275					280					285			

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10	Leu	Asp	Ser	Asp	Gly	Ser	Tyr	Phe	Ile	Tyr	Ser	Lys	Leu	Asn	Met	Lys
					325					330					335	
15	Thr	Ser	Lys	Trp	Glu	Lys	Thr	Asp	Ser	Phe	Ser	Cys	Asn	Val	Arg	His
				340					345					350		
20	Glu	Gly	Leu	Lys	Asn	Tyr	Tyr	Leu	Lys	Lys	Thr	Ile	Ser	Arg	Ser	Pro
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5 Gly Leu Val Gln Xaa Gly Gly Ser Leu Arg Leu Ser Cys Ala Ala Ser
 260 265 270

10 Gly Arg Thr Ile Asp Asn Tyr Ala Met Ala Trp Xaa Arg Gln Ala Pro
 275 280 285

Gly Lys Xaa Xaa Glu Xaa Val Xaa Thr Ile Asp Trp Gly Asp Gly Gly
 290 295 300

15 Xaa Arg Tyr Ala Asn Ser Val Lys Gly Arg Phe Thr Ile Ser Arg Asp
 305 310 315 320

20 Asn Xaa Lys Xaa Thr Xaa Tyr Leu Gln Met Asn Xaa Leu Xaa Xaa Glu
 325 330 335

25 Asp Thr Ala Val Tyr Xaa Cys Ala Met Ala Arg Gln Ser Arg Val Asn
 340 345 350

Leu Asp Val Ala Arg Tyr Asp Tyr Trp Gly Gln Gly Thr Xaa Val Thr
 355 360 365

30 Val Ser Ser
 370

35 **Claims**

1. An isolated or purified antigen binding antibody fragment, comprising
 a complementarity determining region (CDR) 1 sequence of GRTIDNYA (SEQ ID NO:1);
 a CDR2 sequence of IDWGDGGX (SEQ ID NO:2); and
 a CDR3 sequence of AMARQSRVNLDVARYDY (SEQ ID NO:3),
 wherein the antibody fragment is specific for Insulin-Like Growth Factor 1 Receptor (IGF1R).
2. The isolated or purified antigen binding antibody fragment of any one of claims 1, comprising the sequence
 X₁VX₂LX₃ESGGGLVQX₄GGSLRLSCAASGRTIDNYAMAWX₅RQAPGKX₆X₇EX₈VX₉TID
 WGDGGX₁₀RYANSVKGRFTISRDNX₁₁KX₁₂TX₁₃YLQMNX₁₄LX₁₅X₁₆EDTAVYX₁₇CAMAR
 QSRVNLDVARYDYWGQGTX₁₈VTVSS (SEQ ID NO:4), where X₁ is E or Q; X₂ is K or Q; X₃ is V or E; X₄ is A or
 P; X₅ is V or S; X₆ is D or G; X₇ is L or R; X₈ is F or W; X₉ is A or S; X₁₀ is A or T; X₁₁ is A or S; X₁₂ is G or N; X₁₃
 is M or L; X₁₄ is N or R; X₁₅ is E or R; X₁₆ is P or A; X₁₇ is S or Y; and X₁₈ is Q or L,
 or a sequence substantially identical thereto.
3. The isolated or purified antigen binding antibody fragment of any one of claims 1 to 2, comprising a sequence
 selected from the group consisting of:

55 QVKLEESGGGLVQAGGSLRLSCAASGRTIDNYAMAWSRQAPGK DREFVATIDWGDG
 GARYANSVKGRFTISRDNKGTMYLQMNPLEPDTAVYSCAMARQSRVNLDVARYD
 YWGQGTQVTVSS (SEQ ID NO:5);

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EVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWVRQAPGKGLEWVSTIDWGD
GGTRYANSVKGRFTISRDN SKNTLYLQMNSLRAEDTAVYYCAMARQSRVNL DVARYD
5 YWGQGLTVTVSS (SEQ ID NO:6);

QVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWVRQAPGKGLEWVATIDWGD
GGTRYANSVKGRFTISRDN SKNTMYLQMNSLRAEDTAVYYCAMARQSRVNL DVARY
10 DYWGQGLTVTVSS (SEQ ID NO:7);

QVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWSRQAPGKGLEFVATIDWGDG
GTRYANSVKGRFTISRDN SKNTMYLQMNSLRAEDTAVYYCAMARQSRVNL DVARYD
15 YWGQGLTVTVSS (SEQ ID NO:8);

QVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWSRQAPGKDREFVATIDWGDG
GTRYANSVKGRFTISRDN SKGTMYLQMNSLRAEDTAVYSCAMARQSRVNL DVARYD
20 YWGQGLTVTVSS (SEQ ID NO:9); and

25 EVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWSRQAPGKDREFVSTIDWGDG
GTRYANSVKGRFTISRDN SKNTLYLQMNSLRAEDTAVYYCAMARQSRVNL DVARYDY WGQGLTVTVSS (SEQ
ID NO:10), or a sequence substantially identical thereto.

- 30 **4.** The isolated or purified antigen binding antibody fragment of any one of claims 1 to 3, wherein the antibody fragment is a single-domain antibody (sdAb).
- 5.** The isolated or purified antigen binding antibody fragment, wherein the antigen binding antibody fragment binds to an epitope comprising the sequence FENFLHNSIFVPR (SEQ ID NO:11).
- 35 **6.** The isolated or purified antibody or fragment thereof of any one of claims 1 to 4, wherein the antigen binding antibody fragment is in a multivalent display format.
- 7.** The isolated or purified antibody or fragment thereof of claim 6, wherein the antigen binding antibody fragment is linked to a Fc fragment.
- 40 **8.** The isolated or purified antibody fragment of any one of claims 2 to 7, wherein the isolated or purified antigen binding antibody fragment transmigrates the blood-brain barrier.
- 9.** A nucleic acid molecule encoding the isolated or purified antigen binding antibody fragment of any one of claims 1 to 8.
- 45 **10.** A vector comprising the nucleic acid molecule of claim 9.
- 11.** The isolated or purified antigen binding antibody fragment of any one of claims 1 to 8, wherein the antigen binding antibody fragment is linked to a cargo molecule.
- 50 **12.** The isolated or purified antigen binding antibody fragment of claim 11, wherein the cargo molecule has a molecular weight in the range of about 1kD to about 200 kDa and/or selected from the group consisting of a detectable agent, a therapeutic, a drug, a peptide, a growth factor, a cytokine, a receptor trap, a chemical compound, a carbohydrate moiety, an enzyme, an antibody or fragment thereof, a DNA-based molecule, a viral vector, or a cytotoxic agent;
- 55 one or more liposomes or nanocarriers loaded with a detectable agent, a therapeutic, a drug, a peptide, an enzyme, an antibody or fragment thereof, a DNA-based molecule, a viral vector, or a cytotoxic agent; or one or more nanoparticle, nanowire, nanotube, or quantum dots.

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13. A composition comprising one or more than one isolated or purified antigen binding antibody fragment of any one of claims 1 to 8 and 11 to 12 and a pharmaceutically-acceptable carrier, diluent, or excipient.

14. The isolated or purified antigen binding antibody fragment of any one of claims 1 to 8 and 11 to 12 for use in therapy.

15. The isolated or purified antigen binding antibody fragment of any one of claims 1 to 8 for use in a blood brain barrier model by detecting Insulin-Like Growth Factor 1 Receptor (IGF1R) or IGF1R expression, comprising

- a) contacting a tissue sample with one or more than one isolated or purified antigen binding antibody fragment of any one of claims 1 to 8 linked to a detectable agent; and
- b) detecting the detectable agent linked to the antigen binding antibody fragment bound to IGF1R in the tissue sample.

16. The isolated or purified antigen binding antibody fragment for the use according to claim 14 comprising the transportation of a cargo molecule across the blood-brain barrier (BBB), by:

- a) administering to a subject one or more than one isolated or purified antigen binding antibody fragment of claim 11 to 12 linked to the cargo molecule, wherein the one or more than one antibody or fragment thereof ferries the cargo molecule across the BBB.

Patentansprüche

1. Isoliertes oder gereinigtes Antigen bindendes Antikörperfragment, umfassend eine Komplementarität-bestimmende-Region- (CDR-) 1-Sequenz aus GRTIDNYA (SEQ ID NO:1); eine CDR2-Sequenz aus IDWGDGGX (SEQ ID NO:2) und eine CDR3-Sequenz aus AMARQSRVNLDVARYDY (SEQ ID NO:3), wobei das Antikörperfragment für Insulinartiger-Wachstumsfaktor-1- (IGF1R-) Rezeptor spezifisch ist.

2. Isoliertes oder gereinigtes Antigen bindendes Antikörperfragment nach einem der Ansprüche 1, umfassend die Sequenz
X₁VX₂LX₃ESGGGLVQX₄GGSLRLSCAASGRTIDNYAMAWX₅RQAPGX₆X₇EX₈VX₉TIDWGDGGX₁₀RYA
NSVKGRFTISRDNX₁₁KX₁₂TX₁₃YLQMNX₁₄LX₁₅X₁₆EDTAVYX₁₇CAMARQSRVNLDVARYDYWGQGT₁₈VTVSS
(SEQ ID NO:4), wobei X₁ E oder Q ist; X₂ K oder Q ist; X₃ V oder E ist; X₄ A oder P ist; X₅ V oder S ist; X₆ D oder G ist; X₇ L oder R ist; X₈ F oder W ist; X₉ A oder S ist; X₁₀ A oder T ist; X₁₁ A oder S ist; X₁₂ G oder N ist; X₁₃ M oder L ist; X₁₄ N oder R ist; X₁₅ E oder R ist; X₁₆ P oder A ist; X₁₇ S oder Y ist und X₁₈ Q oder L ist, oder eine damit im Wesentlichen identische Sequenz.

3. Isoliertes oder gereinigtes Antigen bindendes Antikörperfragment nach einem der Ansprüche 1 bis 2, umfassend eine Sequenz, die aus der Gruppe ausgewählt ist:

QVKLEESGGGLVQAGGSLRLSCAASGRTIDNYAMAWSRQAPGKDREFVATIDWGDGGARYANSVKG
RFTISRDNAGKTMYLQMNLEPEDTAVYSCAMARQSRVNLDVARYDYWGQGTQVTVSS (SEQ ID
NO:5);

EVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWVRQAPGKLEWVSTIDWGDGGTRYANSVK
GRFTISRDNKNTLYLQMNSLRAEDTAVYVCAMARQSRVNLDVARYDYWGQGLTVTVSS (SEQ ID
NO:6);

QVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWVRQAPGKLEWVATIDWGDGGTRYANSVK
GRFTISRDNKNTMYLQMNSLRAEDTAVYVCAMARQSRVNLDVARYDYWGQGLTVTVSS (SEQ ID
NO:7);

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QVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWSRQAPGKGLFVATIDWGDGGTRYANSVKG
RFTISRDN SKNTMYLQMNSLRAEDTAVYYCAMARQSRVNL DVARYDYWGQGLTVTVSS (SEQ ID
5 NO:8);

QVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWSRQAPGKDREFVATIDWGDGGTRYANSVK
GRFTISRDN SKGTYLQMNSLRAEDTAVYSCAMARQSRVNL DVARYDYWGQGLTVTVSS (SEQ ID
10 NO:9) und

EVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWSRQAPGKDREFVSTIDWGDGGTRYANSVKG
RFTISRDN SKNTLYLQMNSLRAEDTAVYYCAMARQSRVNL DVARYDYWGQGLTVTVSS (SEQ ID NO:10), oder ei-
15 ne damit im Wesentlichen identische Sequenz.

4. Isoliertes oder gereinigtes Antigen bindendes Antikörperfragment nach einem der Ansprüche 1 bis 3, wobei das Antikörperfragment ein Einzeldomänenantikörper (sdAb) ist.
- 20 5. Isoliertes oder gereinigtes Antigen bindendes Antikörperfragment, wobei das Antigen bindende Antikörperfragment an ein Epitop bindet, das die Sequenz FENFLHNSIFVPR (SEQ ID NO:11) umfasst.
6. Isolierter oder gereinigter Antikörper oder Fragment desselben nach einem der Ansprüche 1 bis 4, wobei das Antigen bindende Antikörperfragment in einem multivalenten Displayformat ist.
- 25 7. Isolierter oder gereinigter Antikörper oder Fragment desselben nach Anspruch 6, wobei das Antigen bindende Antikörperfragment mit einem Fc-Fragment verknüpft ist.
8. Isoliertes oder gereinigtes Antikörperfragment nach einem der Ansprüche 2 bis 7, wobei das isolierte oder gereinigte Antigen bindende Antikörperfragment durch die Blut-Hirn-Schranke transmigriert.
- 30 9. Nukleinsäuremolekül, das das isolierte oder gereinigte Antigen bindende Antikörperfragment nach einem der Ansprüche 1 bis 8 codiert.
- 35 10. Vektor, der das Nukleinsäuremolekül nach Anspruch 9 umfasst.
11. Isoliertes oder gereinigtes Antigen bindendes Antikörperfragment nach einem der Ansprüche 1 bis 8, wobei das Antigen bindende Antikörperfragment mit einem Frachtmolekül verknüpft ist.
- 40 12. Isoliertes oder gereinigtes Antigen bindendes Antikörperfragment nach Anspruch 11, wobei das Frachtmolekül ein Molekulargewicht im Bereich von circa 1kD bis circa 200 kDa hat und/oder ausgewählt ist aus der Gruppe bestehend aus einem detektierbaren Wirkstoff, einem Therapeutikum, einem Arzneistoff, einem Peptid, einem Wachstumsfaktor, einem Zytokin, einer Rezeptorfalle, einer chemischen Verbindung, einem Kohlenhydratanteil, einem Enzym, einem Antikörper oder Fragment desselben, einem DNA-basierten Molekül, einem viralen Vektor oder einem zyto-
45 toxischen Wirkstoff; einem oder mehreren Liposomen oder Nanoträgern, die mit einem detektierbaren Wirkstoff, einem Therapeutikum, einem Arzneistoff, einem Peptid, einem Enzym, einem Antikörper oder Fragment desselben, einem DNA-basierten Molekül, einem viralen Vektor oder einem zytotoxischen Wirkstoff geladen sind; oder einem oder mehreren Nanopartikeln, Nanodraht, Nanoröhrchen oder Quantenpunkten.
- 50 13. Zusammensetzung, die ein oder mehr als ein isoliertes oder gereinigtes Antigen bindendes Antikörperfragment nach einem der Ansprüche 1 bis 8 und 11 bis 12 und einen pharmazeutisch akzeptablen Träger, Verdünnungsstoff oder Hilfsstoff umfasst.
14. Isoliertes oder gereinigtes Antigen bindendes Antikörperfragment nach einem der Ansprüche 1 bis 8 und 11 bis 12
55 zur Verwendung bei der Therapie.
15. Isoliertes oder gereinigtes Antigen bindendes Antikörperfragment nach einem der Ansprüche 1 bis 8 zur Verwendung in einem Blut-Hirn-Schranken-Modell durch Detektieren von Insulinartiger-Wachstumsfaktor-1- (IGF1R-) Rezeptor

oder IGF1R-Expression, umfassend

- 5 a) Kontaktieren einer Gewebeprobe mit einem oder mehr als einem isolierten oder gereinigten Antigen bindenden Antikörperfragment nach einem der Ansprüche 1 bis 8, das mit einem detektierbaren Wirkstoff verknüpft ist; und
 b) Detektieren des detektierbaren Wirkstoffs, der mit dem Antigen bindenden Antikörperfragment verknüpft ist, das an IGF1R in der Gewebeprobe gebunden ist.

10 **16.** Isoliertes oder gereinigtes Antigen bindendes Antikörperfragment zur Verwendung gemäß Anspruch 14, umfassend den Transport eines Frachtmoleküls über die Blut-Hirn-Schranke (BBB), durch:

- 15 a) Verabreichen von einem oder mehr als einem von dem isolierten oder gereinigten Antigen bindenden Antikörperfragment nach Anspruch 11 bis 12, das mit dem Frachtmolekül verknüpft ist, an einen Patienten, wobei der eine oder mehr als ein Antikörper oder das Fragment desselben das Frachtmolekül über die BBB trägt.

Revendications

20 **1.** Fragment d'anticorps isolé ou purifié liant un antigène, comprenant :

25 une séquence de région déterminante de complémentarité (CDR) 1 de GRTIDNYA (N° ID SEQ : 1) ;
 une séquence CDR2 de IDWGDGGX (N° ID SEQ : 2) ; et
 une séquence CDR3 de AMARQSRVNLDVARYDY (N° ID SEQ : 3),
 dans lequel le fragment d'anticorps est spécifique pour le récepteur de facteur 1 de croissance analogue à l'insuline (IGF1R).

2. Fragment d'anticorps isolé ou purifié liant un antigène selon la revendication 1, comprenant la séquence :

30 X₁VX₂LX₃ESGGGLVQX₄GGSLRLSCAASGRTIDNYAMAWX₅RQAPGKX₆X₇EX₈VX₉TID
 WGDGGX₁₀RYANSVKGRFTISRDNX₁₁KX₁₂TX₁₃YLQMNX₁₄LX₁₅X₁₆EDTAVYX₁₇CAMAR
 QSRVNLDVARYDYWGQGTX₁₈VTVSS (N° ID SEQ : 4), où X₁ est E ou Q ; X₂ est K ou Q ; X₃ est V ou E ; X₄
 est A ou P ; X₅ est V ou S ; X₆ est D ou G ; X₇ est L ou R ; X₈ est F ou W ; X₉ est A ou S ; X₁₀ est A ou T ; X₁₁
 est A ou S ; X₁₂ est G ou N ; X₁₃ est M ou L ; X₁₄ est N ou R ; X₁₅ est E ou R ; X₁₆ est P ou A ; X₁₇ est S ou Y ;
 et X₁₈ est Q ou L,
 35 ou une séquence sensiblement identique à celle-ci.

3. Fragment d'anticorps isolé ou purifié liant un antigène selon une quelconque des revendications 1 à 2, comprenant une séquence sélectionnée à partir du groupe se composant de :

40 QVKLEESGGGLVQAGGSLRLSCAASGRTIDNYAMAWSRQAPGKDREFVATIDWGDG
 GARYANSVKGRFTISRDNKGTMYLQMNNLEPEDTAVYSCAMARQSRVNLDVARYD
 YWGQGTQVTVSS (N° ID SEQ : 5) ;

45 EVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWVRQAPGKGLEWVSTIDWGD
 GGTRYANSVKGRFTISRDNKNTLYLQMNSLRAEDTAVYYCAMARQSRVNLDVARYD
 YWGQGTTLTVSS (N° ID SEQ : 6) ;

50 QVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWVRQAPGKGLEWVATIDWGD
 GGTRYANSVKGRFTISRDNKNTMYLQMNSLRAEDTAVYYCAMARQSRVNLDVARY
 55 DYWGQGTTLTVSS (N° ID SEQ : 7) ;

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QVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWSRQAPGKGLEFVATIDWGDG
GTRYANSVKGRFTISRDN SKNTMYLQMNSLRAEDTAVYYCAMARQSRVNL DVARYD
5 YWQGGLTVTVSS (N° ID SEQ : 8) ;

QVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWSRQAPGKDREFVATIDWGDG
10 GTRYANSVKGRFTISRDN SKGTMYLQMNSLRAEDTAVYSCAMARQSRVNL DVARYD
YWQGGLTVTVSS (N° ID SEQ : 9) ; et

EVQLVESGGGLVQPGGSLRLSCAASGRTIDNYAMAWSRQAPGKDREFVSTIDWGDG
15 GTRYANSVKGRFTISRDN SKNTLYLQMNSLRAEDTAVYYCAMARQSRVNL DVARYDY WQGGLTVTVSS (N° ID
SEQ : 10), ou une séquence sensiblement identique à celle-ci.

4. Fragment d'anticorps isolé ou purifié liant un antigène selon une quelconque des revendications 1 à 3, dans lequel le fragment d'anticorps est un anticorps à domaine individuel (sdAb).
- 20 5. Fragment d'anticorps isolé ou purifié liant un antigène dans lequel le fragment d'anticorps liant un antigène se lie à un épitope comprenant la séquence FENFLHNSIFVPR (N° ID SEQ : 11).
6. Anticorps isolé ou purifié, ou fragment de celui-ci selon une quelconque des revendications 1 à 4, dans lequel le fragment d'anticorps liant un antigène est dans un format de manifestation polyvalente.
- 25 7. Anticorps isolé ou purifié, ou fragment de celui-ci selon la revendication 6, dans lequel le fragment d'anticorps liant un antigène est relié à un fragment Fc.
8. Anticorps isolé ou purifié, ou fragment de celui-ci selon une quelconque des revendications 2 à 7, dans lequel le fragment d'anticorps isolé ou purifié liant un antigène transmigre la barrière hémato-encéphalique.
- 30 9. Molécule d'acide nucléique codant le fragment d'anticorps isolé ou purifié liant un antigène selon une quelconque des revendications 1 à 8.
- 35 10. Vecteur comprenant la molécule d'acide nucléique selon la revendication 9.
11. Fragment d'anticorps isolé ou purifié liant un antigène selon une quelconque des revendications 1 à 8, dans lequel le fragment d'anticorps liant un antigène est relié à une molécule cargo.
- 40 12. Fragment d'anticorps isolé ou purifié liant un antigène selon la revendication 11, dans lequel la molécule cargo possède un poids moléculaire dans la fourchette d'environ 1kD à environ 200 kDa et/ou sélectionné à partir du groupe se composant d'un agent détectable, d'un thérapeutique, d'un médicament, d'un peptide, d'un facteur de croissance, d'une cytokine, d'un récepteur piège, d'un composé chimique, d'une moitié de carbohydrate, d'une enzyme, d'un anticorps ou d'un fragment de celui-ci, d'une molécule à base d'ADN, d'un vecteur viral, ou d'un agent cytotoxique ; d'un ou plusieurs liposomes ou nano-porteurs chargés d'un agent détectable, d'un thérapeutique, d'un médicament, d'un peptide, d'une enzyme, d'un anticorps ou d'un fragment de celui-ci, d'une molécule à base d'ADN, d'un vecteur viral, ou d'un agent cytotoxique ; ou d'une ou plusieurs nanoparticules, d'un nanofil, d'un nanotube, ou de points quantiques.
- 45 13. Composition comprenant un ou plus d'un fragment d'anticorps isolé ou purifié liant un antigène selon une quelconque des revendications 1 à 8 et 11 à 12 et un support, un diluant ou un excipient pharmaceutiquement acceptables.
- 50 14. Fragment d'anticorps isolé ou purifié liant un antigène selon une quelconque des revendications 1 à 8 et 11 à 12 en vue de l'utilisation en thérapie.
- 55 15. Fragment d'anticorps isolé ou purifié liant un antigène selon une quelconque des revendications 1 à 8 en vue de l'utilisation dans un modèle de barrière hémato-encéphalique par détection du récepteur de facteur 1 de croissance analogue à l'insuline (IGF1R) ou d'une expression IGF1R, comprenant

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- a) la mise en contact d'un échantillon de tissu avec un ou plus d'un fragment d'anticorps isolé ou purifié liant un antigène selon une quelconque des revendications 1 à 8 relié à un agent détectable ; et
b) la détection de l'agent détectable relié au fragment d'anticorps liant un antigène lié à IGF1R dans l'échantillon de tissu.

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- 16.** Fragment d'anticorps isolé ou purifié liant un antigène en vue de l'utilisation selon la revendication 14, comprenant le transport d'une molécule cargo à travers la barrière hémato-encéphalique (BBB), par:

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- a) l'administration à un sujet d'un ou plus d'un fragment d'anticorps isolé ou purifié liant un antigène selon une quelconque des revendications 11 à 12 relié à la molécule cargo, dans lequel l'unique ou les plus d'un anticorps ou fragment de celui-ci transportent la molécule cargo à travers la barrière hémato-encéphalique (BBB).

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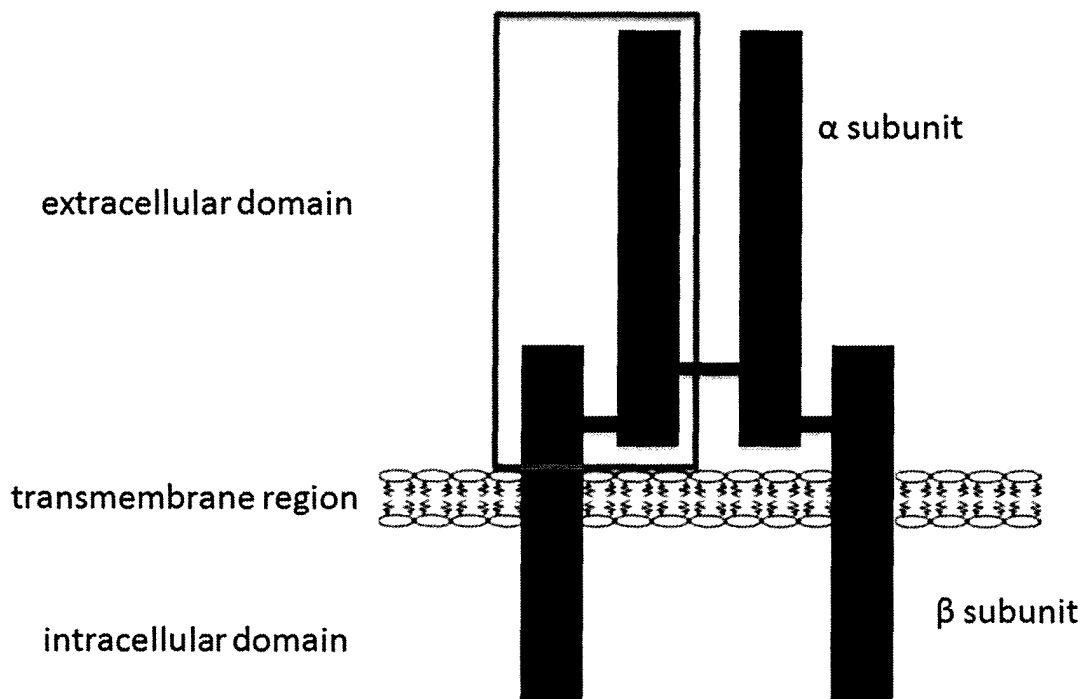


FIG. 1

<i>MKSGSGGGSP</i>	<i>TSLWGLLFLS</i>	<i>AALSLWPTSG</i>	<i>EICGPGIDIR</i>	<i>NDYQQLKRLE</i>	<i>NCTVIEGYLH</i>	60
<i>ILLISKAEDY</i>	<i>RSYRFPKLTIV</i>	<i>ITEYLLLFVRV</i>	<i>AGLES LGDLF</i>	<i>PNLTVIRGWK</i>	<i>LFYNYALVIF</i>	120
<i>EMTNLKDIGL</i>	<i>YNLRNITRGA</i>	<i>IRIEKNADLC</i>	<i>YLSTVDWSLI</i>	<i>LDAVSNNYIV</i>	<i>GNKPPKECGD</i>	180
<i>LCPGTMEEKP</i>	<i>MCEKTTINNE</i>	<i>YNYRCWTTNR</i>	<i>CQK MCPSTCG</i>	<i>KRACTENNEC</i>	<i>CHPECLGSCS</i>	240
<i>APDNDTACVA</i>	<i>CRHYYYAGVC</i>	<i>VPACPPNTYR</i>	<i>FEGWRCVDRD</i>	<i>FCANILSAES</i>	<i>SDSEGFVIHD</i>	300
<i>GECMQECPSG</i>	<i>FIRNGSQSMY</i>	<i>CIPCEGPCPK</i>	<i>VCEEEKTKTK</i>	<i>IDSV TSAQML</i>	<i>QGCTIFKGNL</i>	360
<i>LINIRRGNNI</i>	<i>ASELENFMGL</i>	<i>IEVVTGYVKI</i>	<i>RHSHALVSL</i>	<i>FLKNLRLILG</i>	<i>EEQLEGNYSF</i>	420
<i>YVLDNQNLQQ</i>	<i>LWDWDHRNLT</i>	<i>IKAGKMYFAF</i>	<i>NPKLCVSEIY</i>	<i>RMEEVTGTRG</i>	<i>RQSKGDINTR</i>	480
<i>NNGERASCES</i>	<i>DVLHFTSTTT</i>	<i>SKNRI IITWH</i>	<i>RYPDPYRDL</i>	<i>ISFTVYYKEA</i>	<i>PFKNVTEYDG</i>	540
<i>QDACGSNSWN</i>	<i>MVDVDLPPNK</i>	<i>DVEPGILLHG</i>	<i>LKPWTQYAVY</i>	<i>VKAVTLMVE</i>	<i>NDHIRGAKSE</i>	600
<i>ILYIRTNASV</i>	<i>PSIPLDVLSA</i>	<i>SNSSSQLIVK</i>	<i>WNPPSLPNGN</i>	<i>LSYYIVRWQR</i>	<i>QPQDGYLYRH</i>	660
<i>NYCSKDKIPI</i>	<i>RKYADGTIDI</i>	<i>EEVTENPKTE</i>	<i>VCGGEGKPPC</i>	<i>ACPKTEAEKQ</i>	<i>AEKEEAEYRK</i>	720
<i><u>VFENFLHNSI</u></i>	<i><u>FVPRPE</u><i>krkr</i></i>	<i>DVMQVANTTM</i>	<i>SSRSRNTTAA</i>	<i>DTYNITDPEE</i>	<i>LETEYPPFES</i>	780
<i>RVDNKERTVI</i>	<i>SNLRPFTLYR</i>	<i>IDIHSCNHEA</i>	<i>EKLGCSASNF</i>	<i>VFARTMPAEG</i>	<i>ADDIPGPVTW</i>	840
<i>EPRPENSIFL</i>	<i>KWPEPENPNG</i>	<i>LILMYEIKYG</i>	<i>SQVEDQRECV</i>	<i>SRQEYRKYGG</i>	<i>AKLNRLNPGN</i>	900
<i>YTARIQATSL</i>	<i>SGNGSWTDPV</i>	<i>FFYVQAKTGY</i>	<i>ENFIHLIIAL</i>	<i>PVAVLLIVGG</i>	<i>LVIMLYVFHR</i>	960
<i>KRNNSRLGNG</i>	<i>VLYASVNPEY</i>	<i>FSAADVVPD</i>	<i>EWEVAREKIT</i>	<i>MSRELQGGSF</i>	<i>GMVYEGVAKG</i>	1020
<i>VVKDEPETRV</i>	<i>AIKTVNEAAS</i>	<i>MRERIEFLNE</i>	<i>ASVMKEFNCH</i>	<i>HVVRLLGVVS</i>	<i>QGQPTLVIME</i>	1080
<i>LMTRGDLKSY</i>	<i>LRSLRPEMEN</i>	<i>NPVLAPPSLS</i>	<i>KMIQMAGEIA</i>	<i>DGMAYLNANK</i>	<i>FVHRDLAARN</i>	1140
<i>CMVAEDFTVK</i>	<i>IGDFGMTRDI</i>	<i>YETDYRKGK</i>	<i>KGLLPVRWMS</i>	<i>PESLKDGVFT</i>	<i>TYSDVWSFGV</i>	1200
<i>VLWEIATLAE</i>	<i>QPYQGLSNEQ</i>	<i>VLRFVMEGGL</i>	<i>LDKPDNCPDM</i>	<i>LFELMRMCWQ</i>	<i>YNPKMRPSFL</i>	1260
<i>EIISSIKEEM</i>	<i>EPGFREVSFY</i>	<i>YSEENKLPEP</i>	<i>EELDLEPENM</i>	<i>ESVPLDPSAS</i>	<i>SSSLPLDRH</i>	1320
<i>SGHKAENPG</i>	<i>PGVLVLRASF</i>	<i>DERQPYAHMN</i>	<i>GGRKNERALP</i>	<i>LPQSSTC</i>		1367

FIG. 2

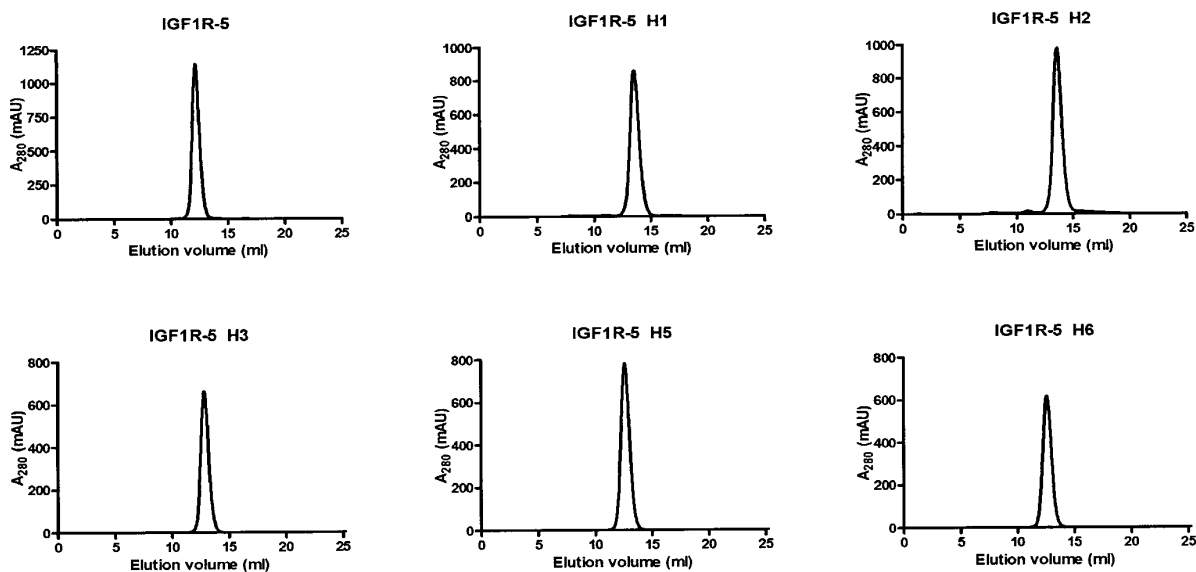


FIG. 3A

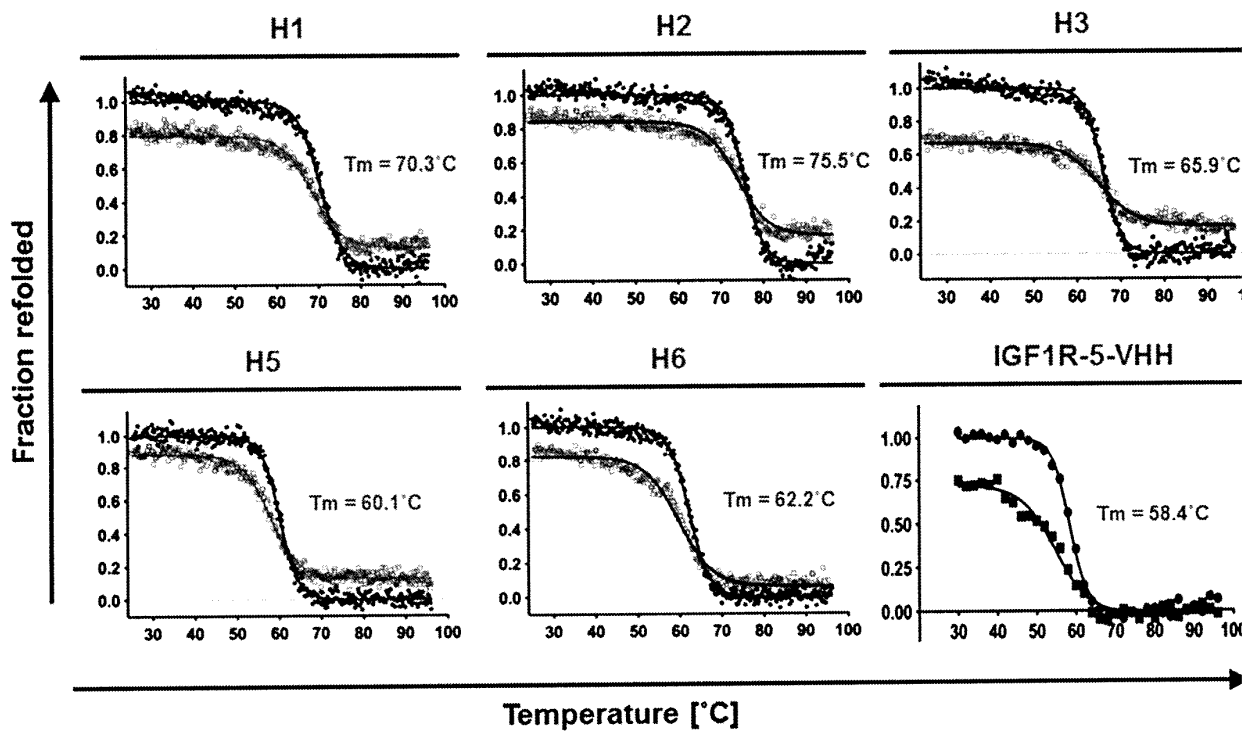


FIG. 3B

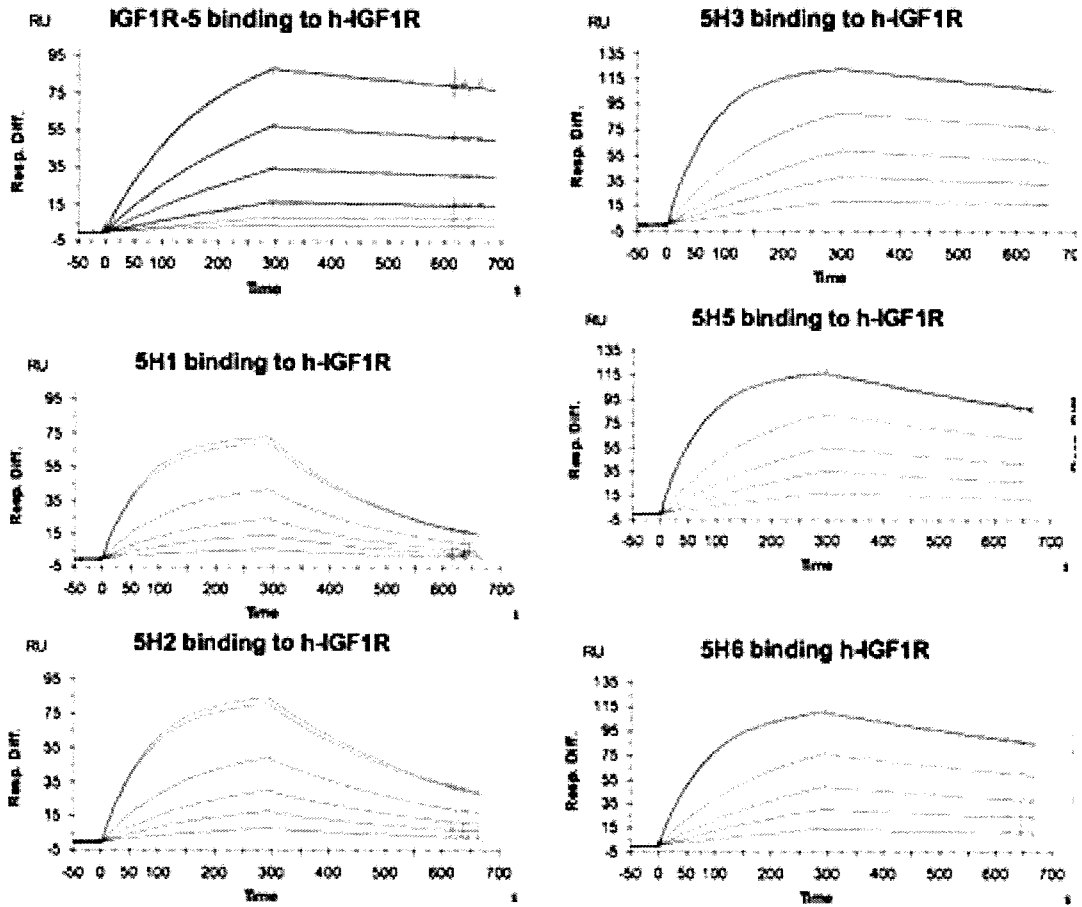


FIG. 3C

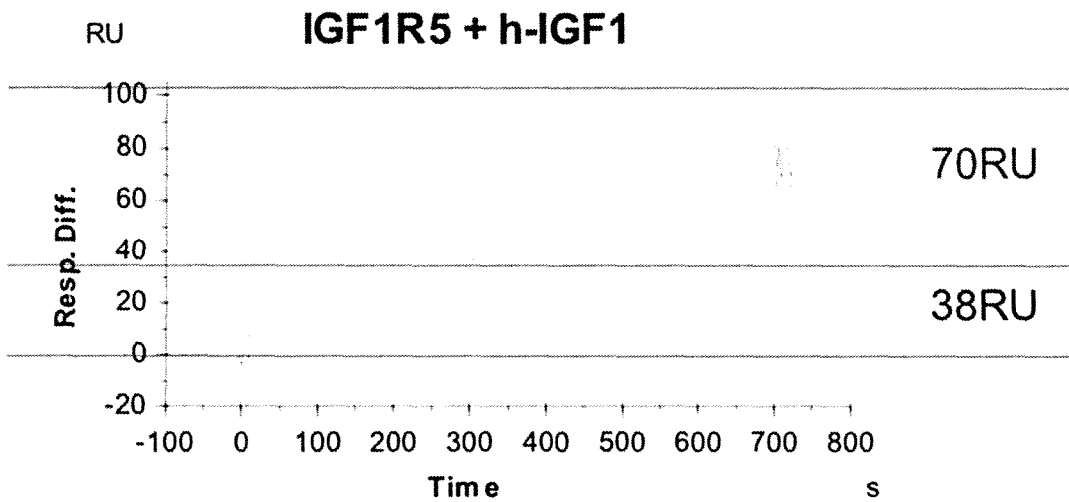


FIG. 3D

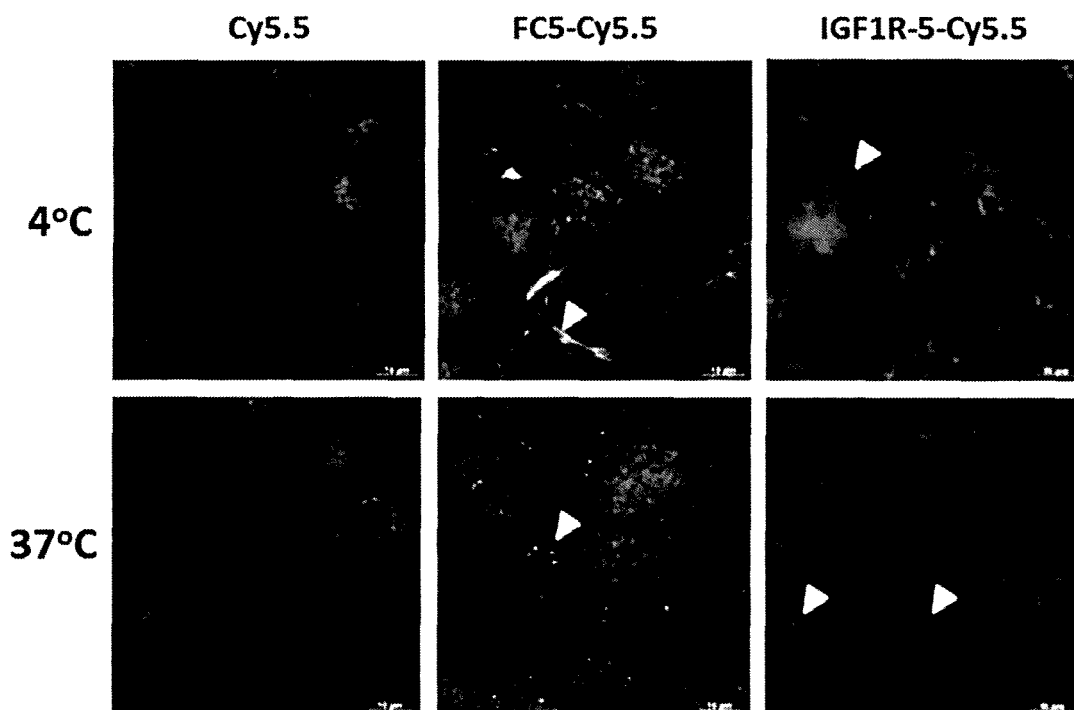


FIG. 4

QVKLEESGGGLVQAGGSLRLSCAASGRTIDNYAMAWSRQAPGKDREFVATIDWGDGGARYANS
 VKGRFTISRDNAGTMYLQMNNLEPEDTAVYSCAMARQSRVNLDVARYDYWGQGTQVTVSS
 MTVDKKLEPSGPISTINPCPPCKECHKCPAPNLEGGPSVFIFPPNIKDVLMISLTPKVTVCVVVDVSE
 DDPDVQISWVFNNEVHTAQTQTHREDYNSTIRVVSTLPIQHQDWMSGKEFKCKVNNKDLPSPI
 IERTISKIKGLVRAPQVYILPPPAEQLSRKDVSLTCLVVGFNPGDISVEWTSNGHTEENYKDTAPVLD
 SDGSYFIYSKLNMKTSKWEKTDSFSCNVRHEGLKNYYLKKTISRSPGK

FIG. 5A

EPSPGPISTINPCPPCKECHKCPAPNLEGGPSVFIFPPNIKDVLMISLTPKVTVCVVVDVSEDDPDVQI
 SWVFNNEVHTAQTQTHREDYNSTIRVVSTLPIQHQDWMSGKEFKCKVNNKDLPSPIERTISKI
 KGLVRAPQVYILPPPAEQLSRKDVSLTCLVVGFNPGDISVEWTSNGHTEENYKDTAPVLDSDGSY
 FIYSKLNMKTSKWEKTDSFSCNVRHEGLKNYYLKKTISRSPGGGGSGGGGSQVKLEESGGGLVQ
 AGGSLRLSCAASGRTIDNYAMAWSRQAPGKDREFVATIDWGDGGARYANSVKGRFTISRDNAG
 TMYLQMNNLEPEDTAVYSCAMARQSRVNLDVARYDYWGQGTQVTVSS

FIG 5B

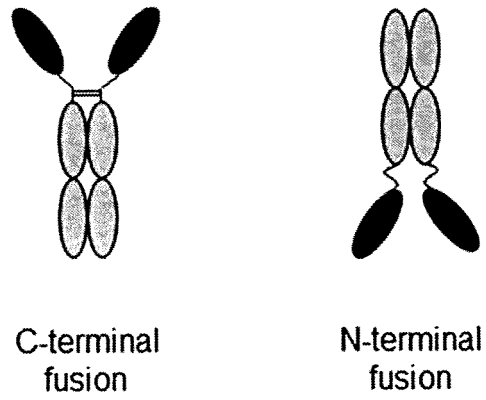


FIG. 5C

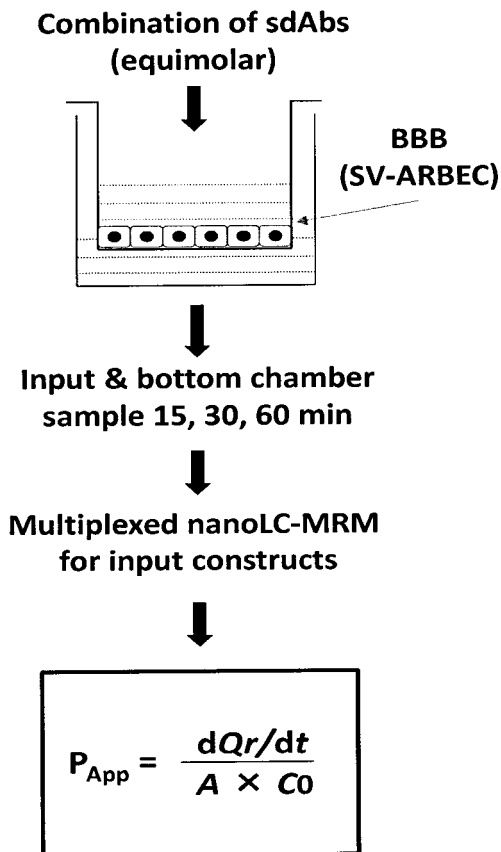


FIG. 6A

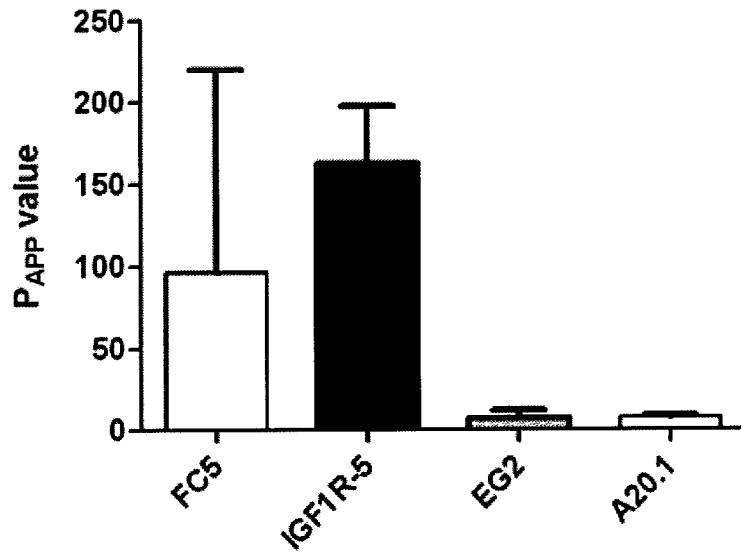


FIG. 6B

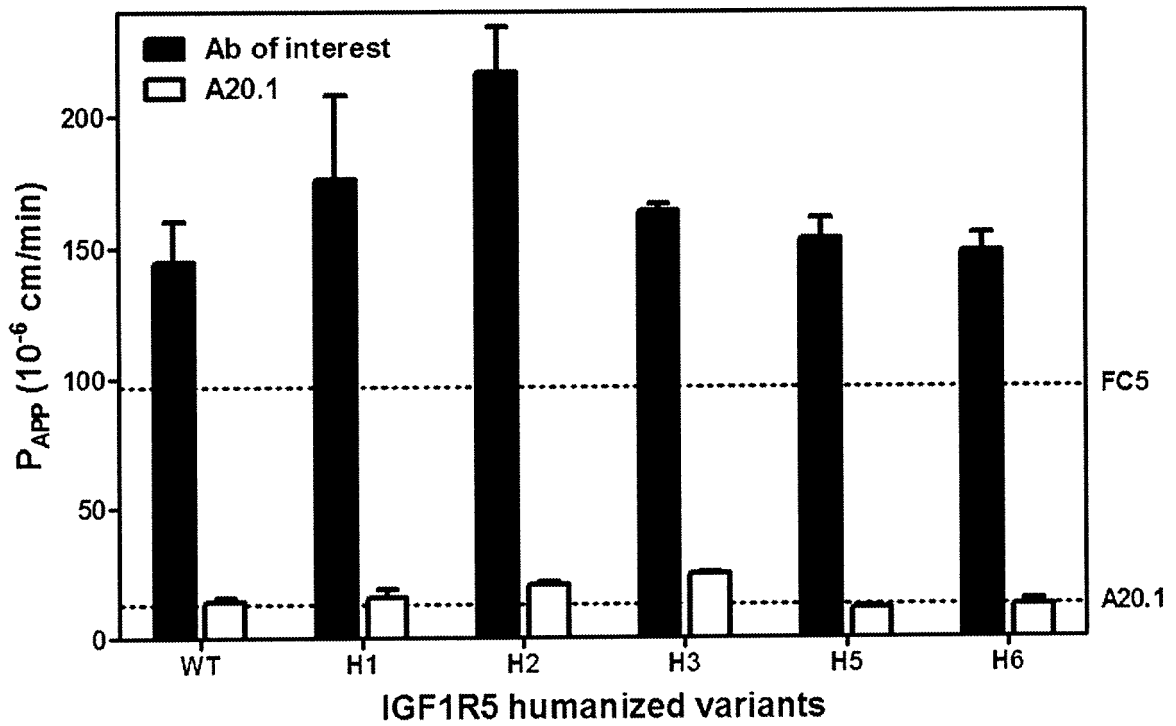


FIG. 6C

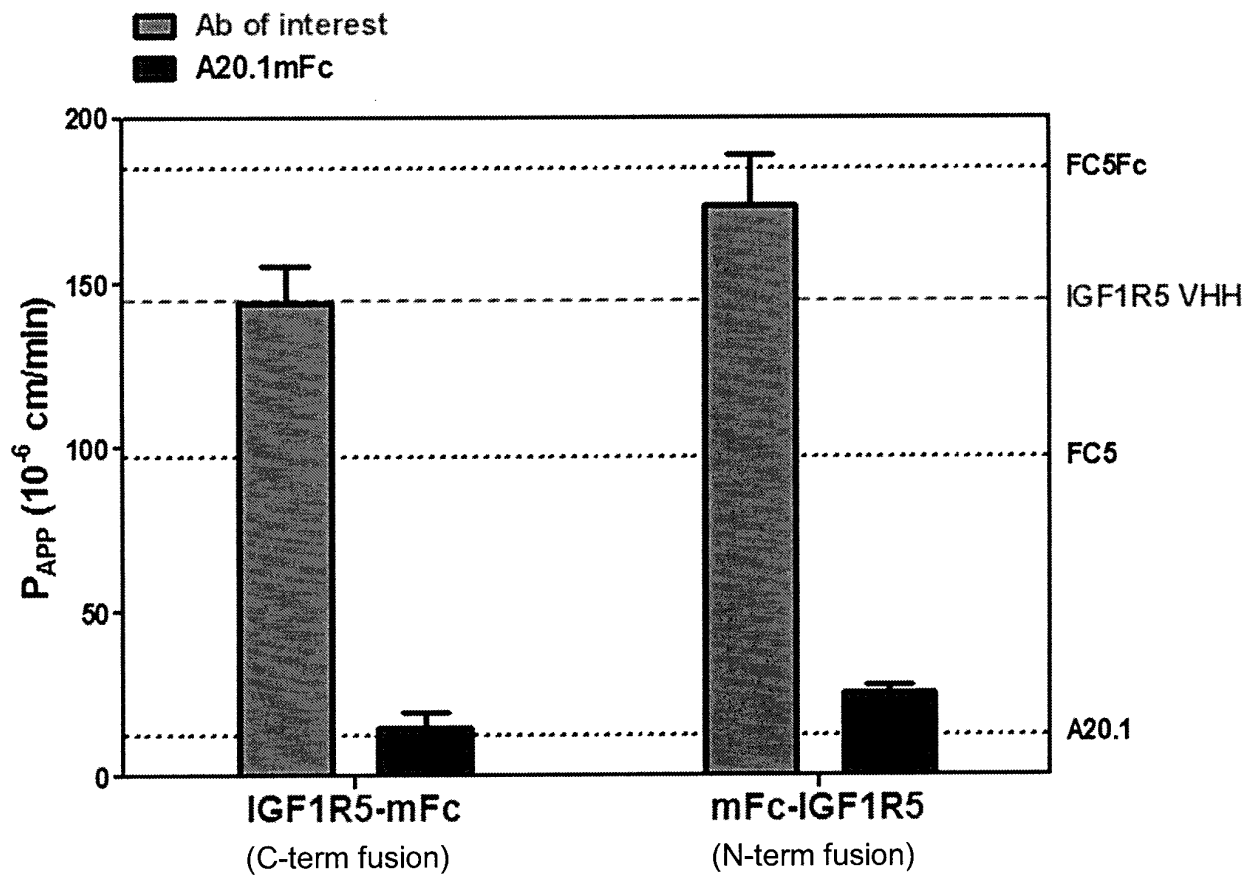


FIG. 6D

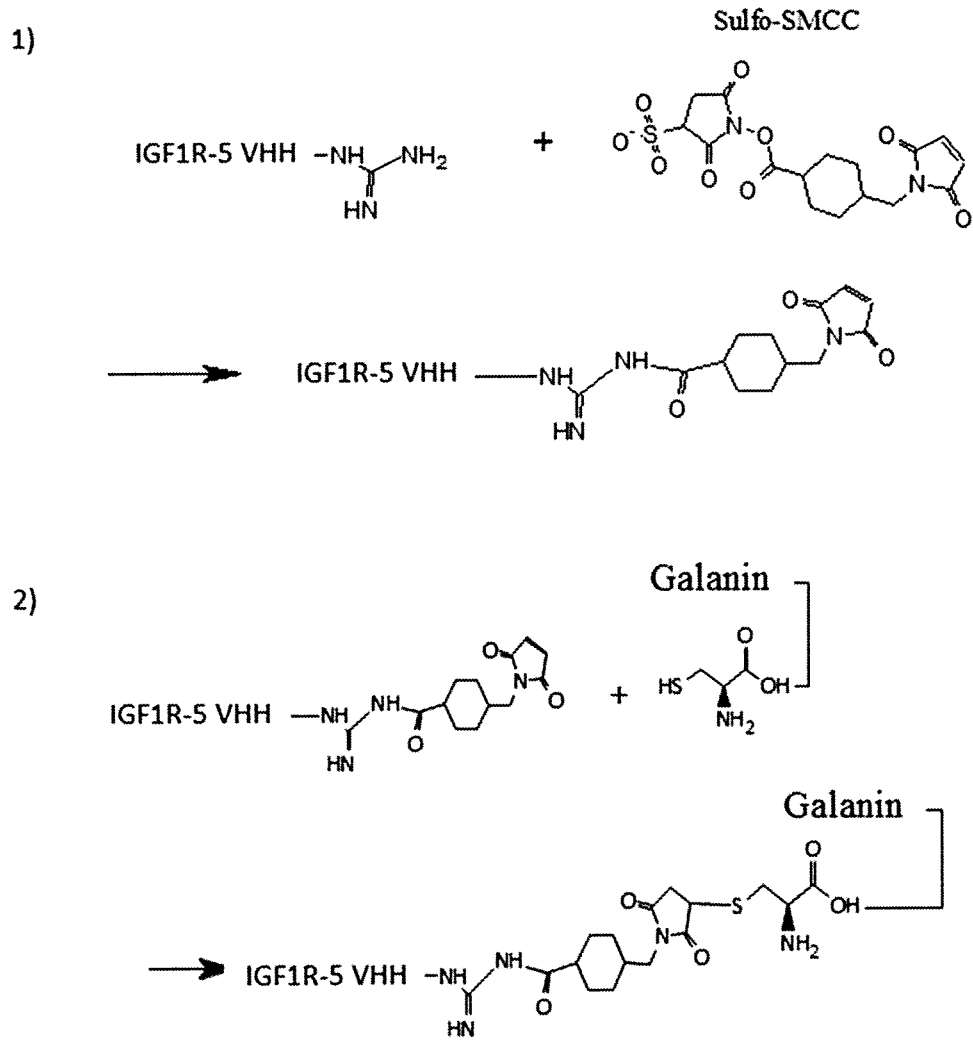


FIG. 7

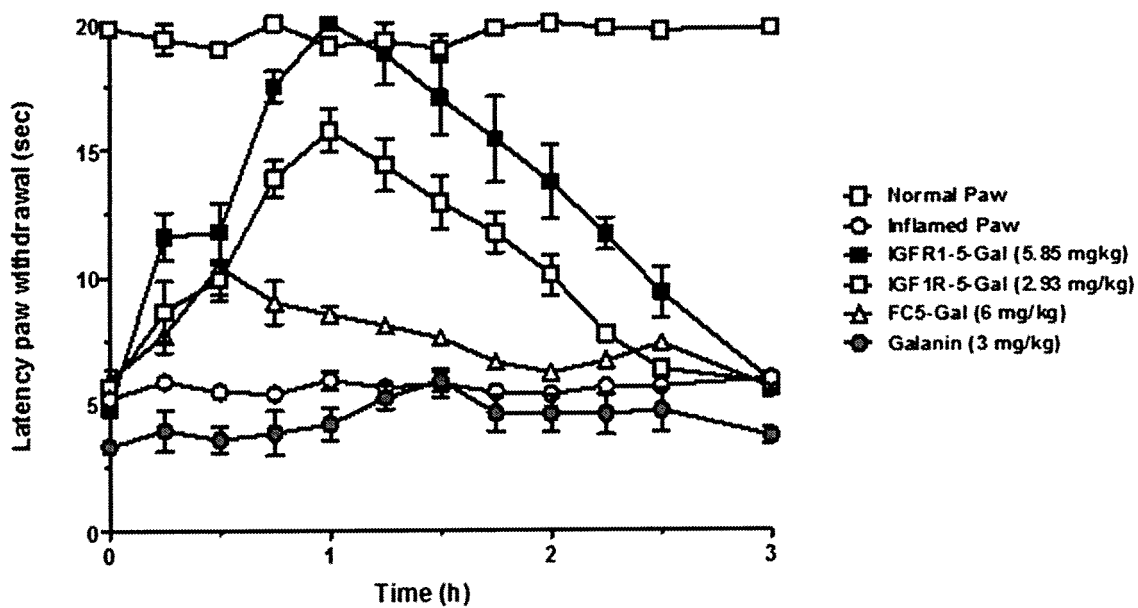


FIG. 8A

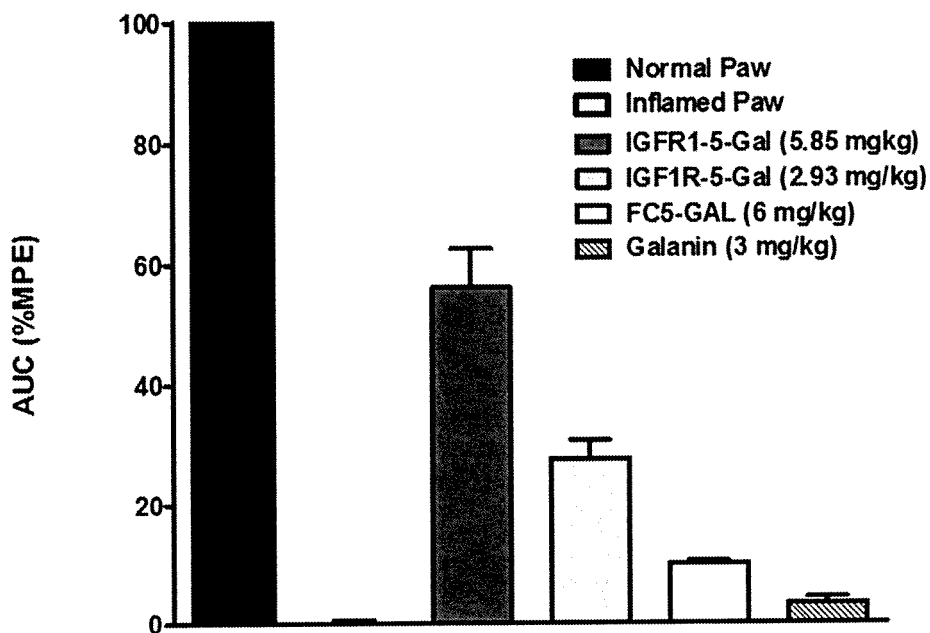


FIG. 8B

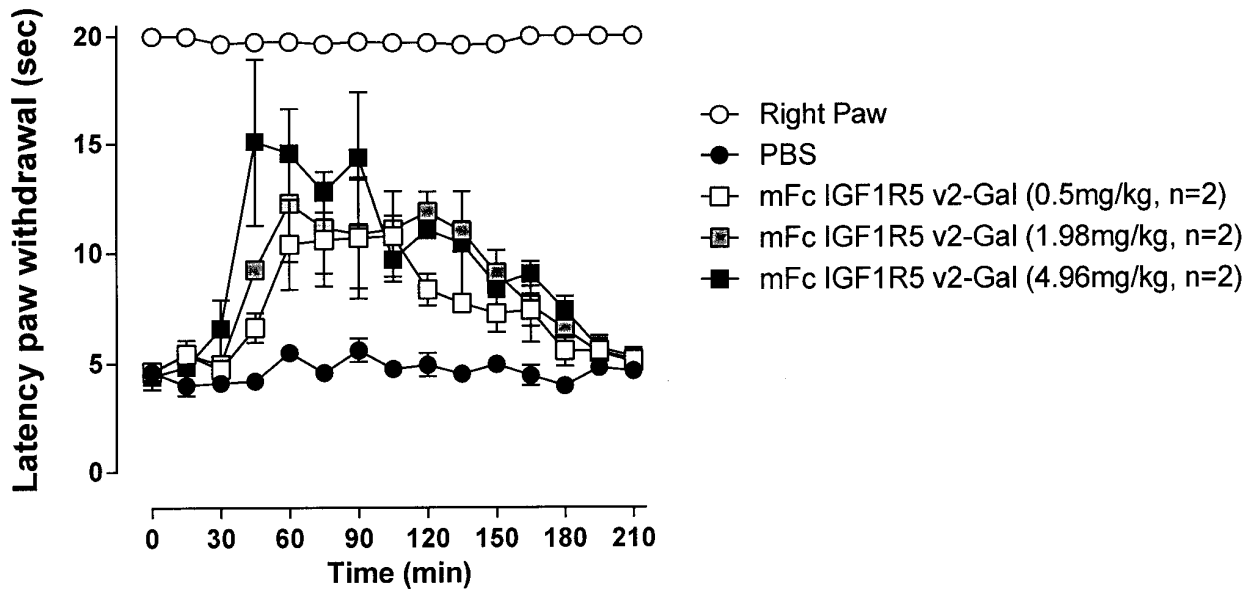


FIG. 8C

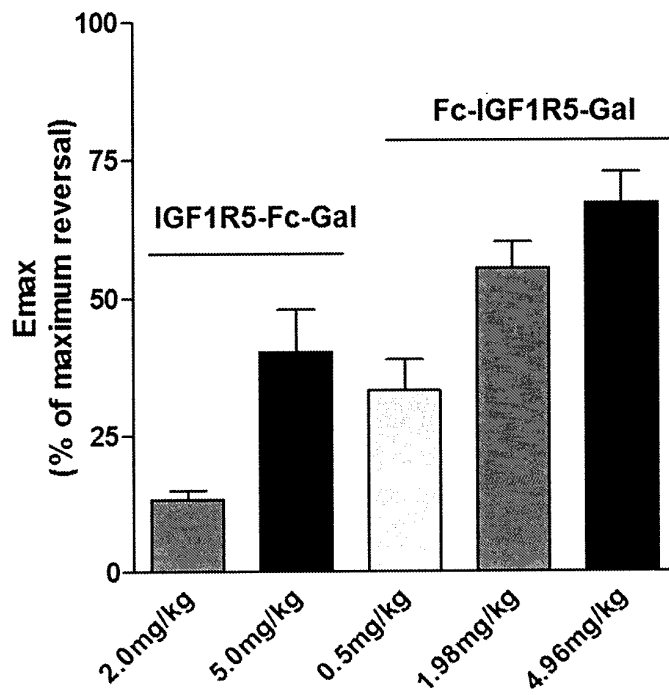


FIG. 8D

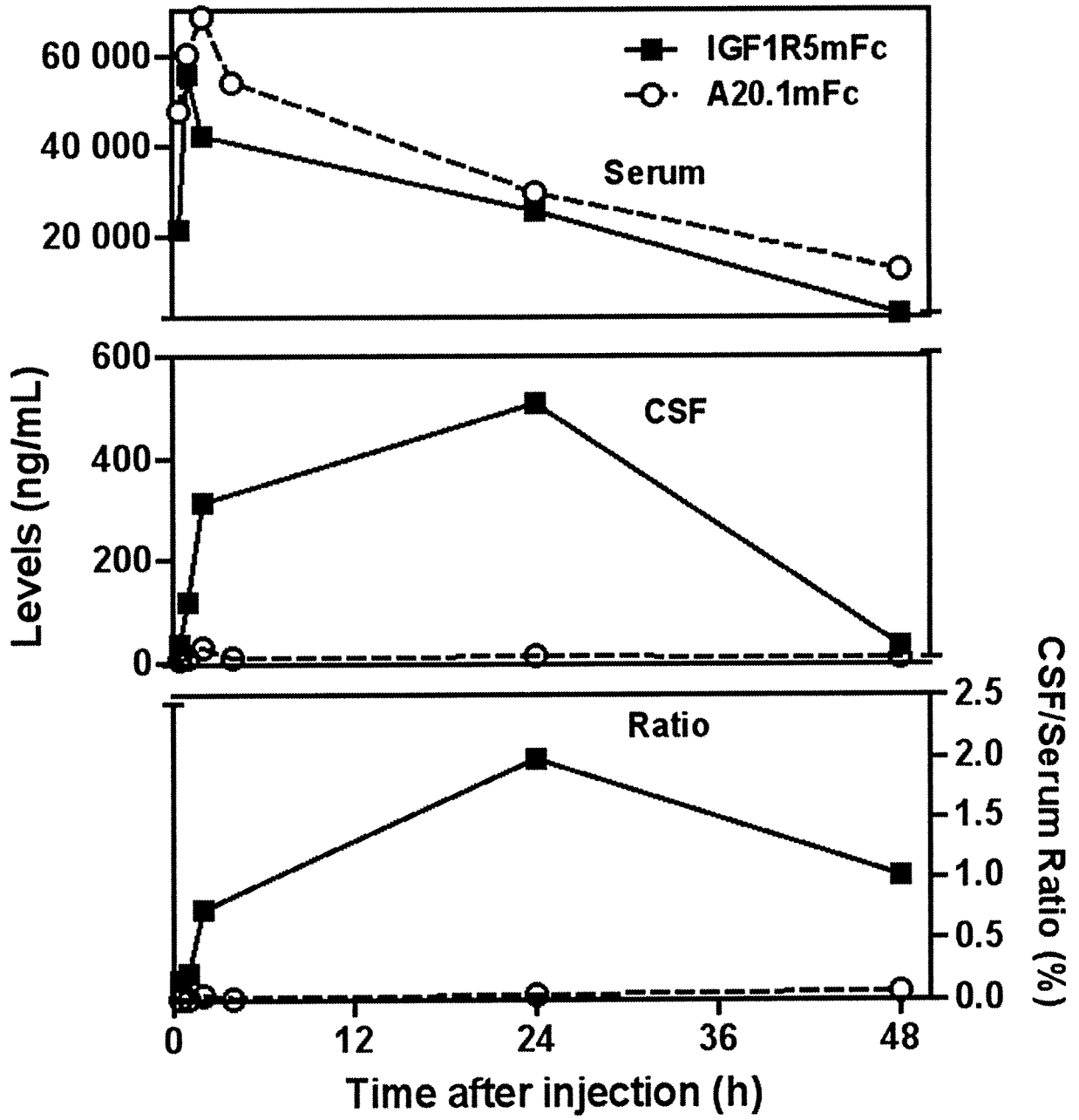


FIG. 9

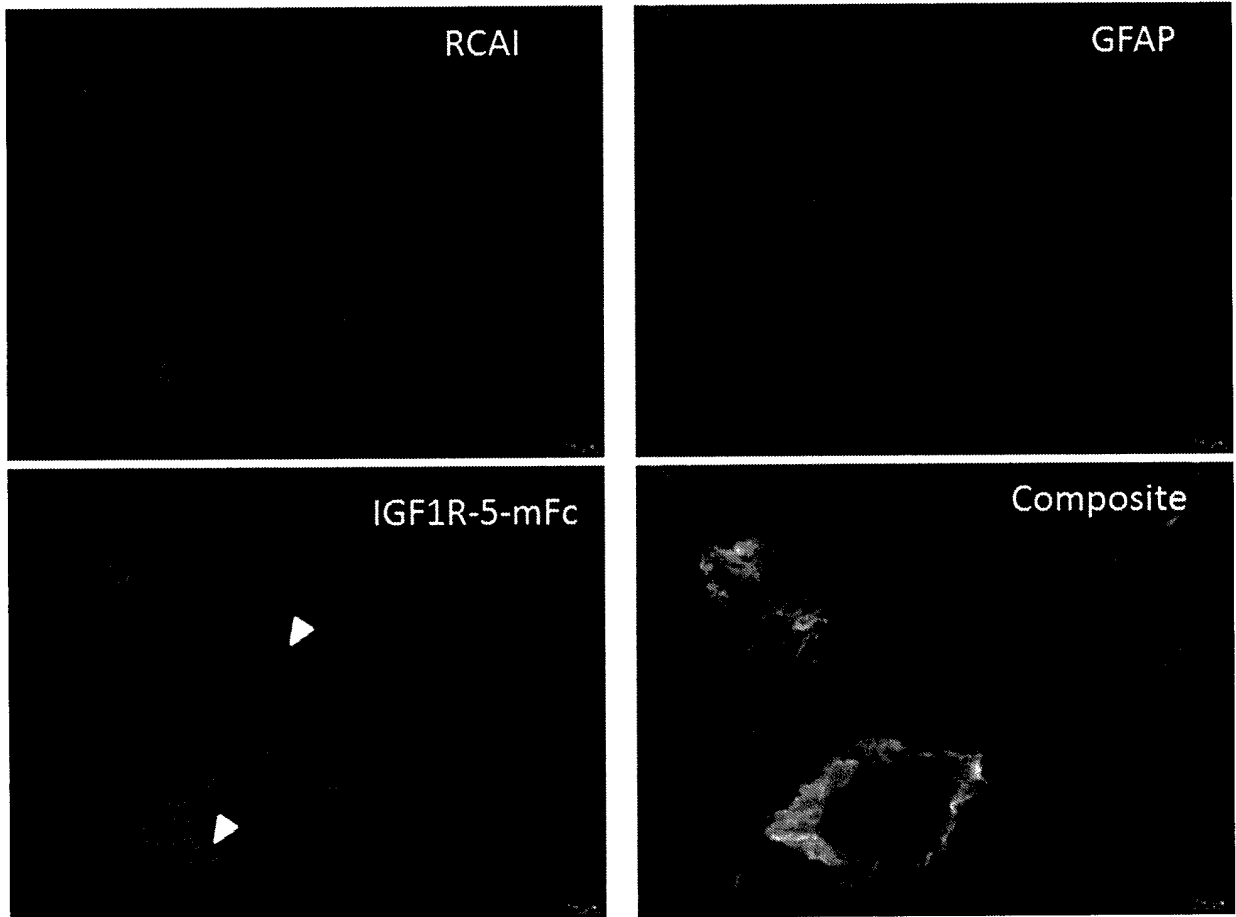


FIG. 10

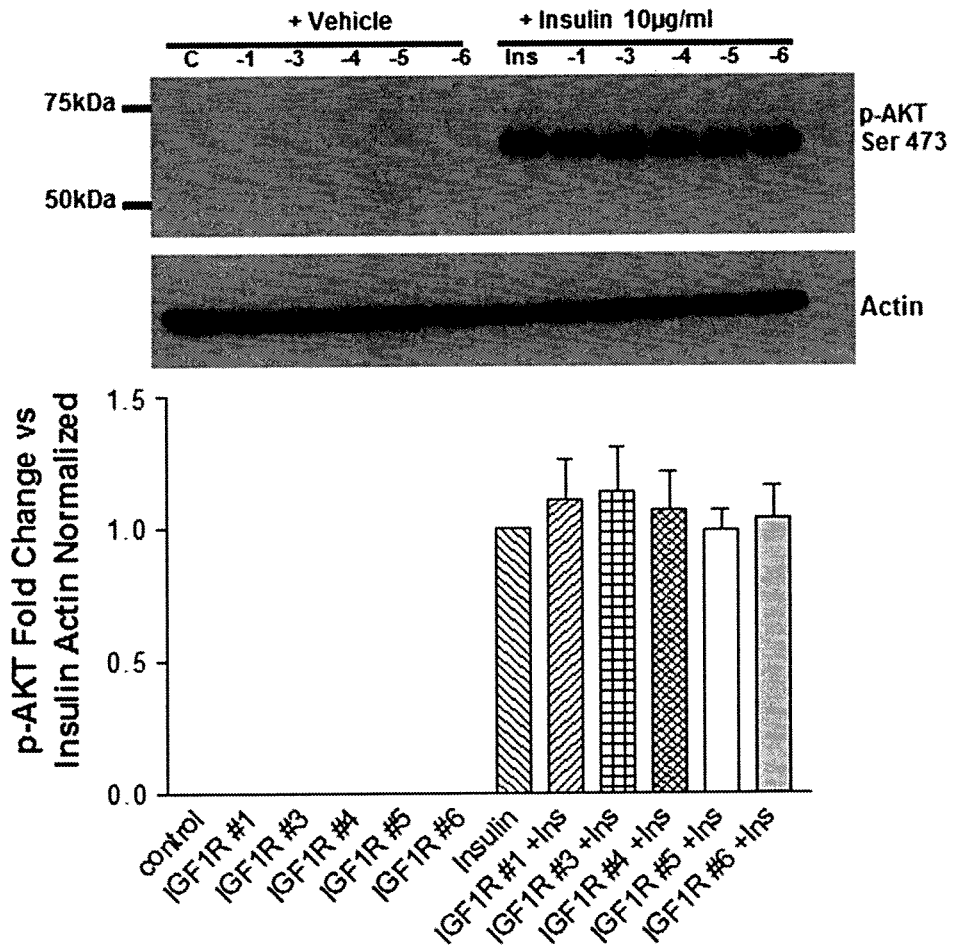


FIG. 11A

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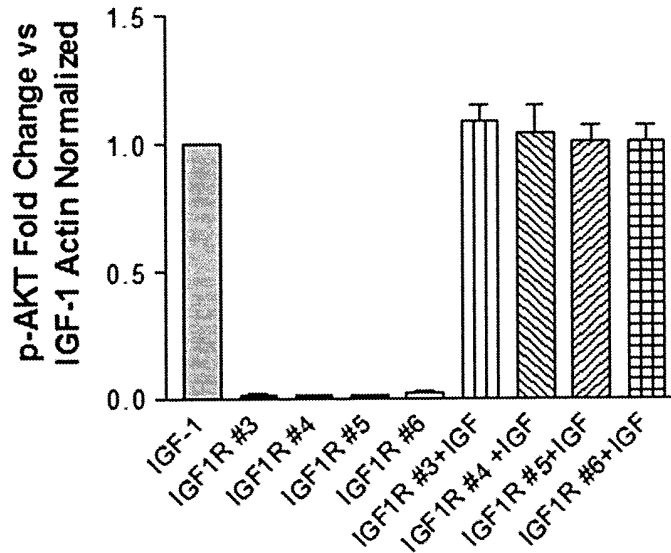
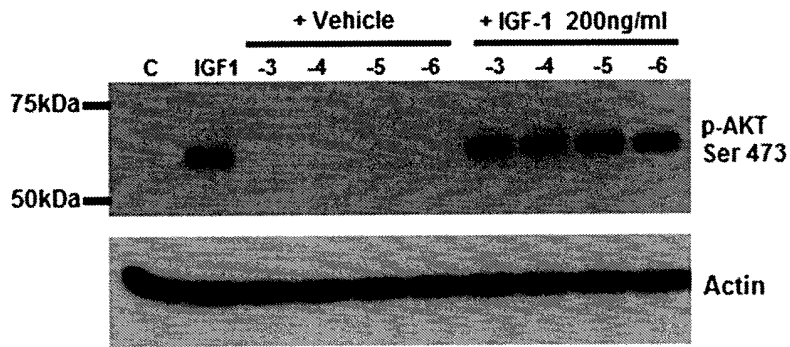


FIG. 11B

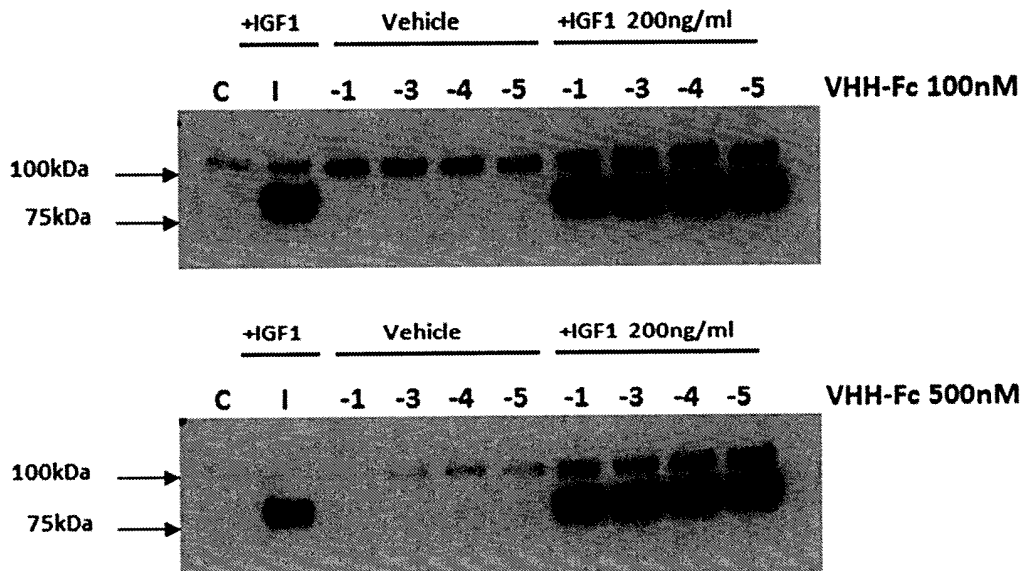


FIG. 11C

REFERENCES CITED IN THE DESCRIPTION

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专利名称(译)	胰岛素样生长因子1受体特异性抗体及其用途		
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[标]发明人	STANIMIROVIC DANICA KEMMERICH KRISTIN HAQQANI ARSALAN S SULEA TRAIAN ARBABI GHAHROUDI MEHDI MASSIE BERNARD GILBERT RENALD		
发明人	STANIMIROVIC DANICA KEMMERICH KRISTIN HAQQANI, ARSALAN S. SULEA, TRAIAN ARBABI-GHAHROUDI, MEHDI MASSIE BERNARD GILBERT RÉNALD		
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摘要(译)

血脑屏障 (BBB) 可防止大于500道尔加仑的分子从血液运输到大脑。受体介导的转胞吞作用 (RMT) 促进跨特定分子的BBB的转运，所述特定分子结合形成BBB的脑内皮细胞上的受体。鉴定了通过RMT转移BBB的胰岛素样生长因子1受体 (IGF1R) 结合抗体或其片段。抗体或片段用于将货物分子递送穿过BBB，其中货物分子可以是治疗剂或可检测剂。抗体是骆驼科动物VHH，通过用933个氨基酸的IGF1R多肽免疫美洲驼制备。还产生骆驼科动物VHH的人源化形式。