

US007236811B2

(12) United States Patent Schmitt

(10) Patent No.: US 7,236,811 B2

(45) **Date of Patent: Jun. 26, 2007**

(54) DEVICE AND METHOD FOR MONITORING BODY FLUID AND ELECTROLYTE DISORDERS

(75) Inventor: Joseph M. Schmitt, Andover, MA (US)

(73) Assignee: Nellcor Puritan Bennett Incorporated,

Pleasanton, CA (US)

(*) Notice: Subject to any disclaimer, the term of this

patent is extended or adjusted under 35

U.S.C. 154(b) by 74 days.

(21) Appl. No.: 10/441,943

(22) Filed: May 20, 2003

(65) Prior Publication Data

US 2003/0220548 A1 Nov. 27, 2003

Related U.S. Application Data

- (63) Continuation of application No. 09/810,918, filed on Mar. 16, 2001, now Pat. No. 6,591,122.
- (51) **Int. Cl.** *A61B 5/1455* (2006.01)
- (52) **U.S. Cl.** 600/310; 600/322

(56) References Cited

U.S. PATENT DOCUMENTS

3,998,550 A 12/1976 Konishi et al.

(Continued)

FOREIGN PATENT DOCUMENTS

CA 2353007 A1 6/2000

(Continued)

OTHER PUBLICATIONS

Attas et al., *Vibrational Spectroscopy, 28(1)*:37–43 (2002). Edwardson et al., *J. Pharm. & Biomed. Analysis, 9(10–12)*:1089–1094 (1991).

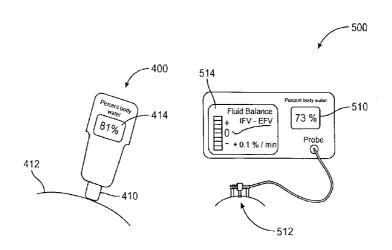
(Continued)

Primary Examiner—Teresa J. Walberg (74) Attorney, Agent, or Firm—Fletcher Yoder PC

(57) ABSTRACT

A device and a method for measuring body fluid-related metrics using spectrophotometry to facilitate therapeutic interventions aimed at restoring body fluid balance. The specific body fluid-related metrics include the absolute volume fraction of water in the extravascular and intravascular tissue compartments, as well as the shifts of water between these two compartments. The absolute volume fraction of water is determined using algorithms where received radiation measured at two or more wavelengths are combined to form either a single ratio, a sum of ratios or ratio of ratios of the form $\log[R(\lambda_1)/R(\lambda_2)]$ in which the received radiation in the numerator depends primarily on the absorbance of water and the received radiation in the denominator depends primarily on the absorbance of water and the sum of the absorbances of non-heme proteins, lipids and water in tissue. The difference between the fraction of water in the intravascular fluid volume ("IFV") and extravascular fluid volume ("EFV") compartments are also determined using a differential method that takes advantage of the observation that pulsations caused by expansion of blood vessels in the skin as the heart beats produce changes in the received radiation at a particular wavelength that are proportional to the difference between the effective absorption of light in the blood and the surrounding tissue. This difference, integrated over time, provides a measure of the quantity of the fluid that shifts into and out of the capillaries. A mechanism for mechanically inducing a pulse is built into the device to improve the reliability of measurements of 1FV-EFV under weak-pulse conditions.

61 Claims, 4 Drawing Sheets



U.S. PATENT	DOCUMENTS	WO WO 01/16577 A1 3/2001	
4,066,068 A 1/1978	Nilsson et al.	OTHER PUBLICATIONS	
	Jacques		
	Kuzara	Johnson et al., Kidney and Blood Pressure Research,	
	Oman et al.	<i>19</i> :94–99 (1996).	
	Jobsis	Lucassen et al., Proc. SPIE, 4162:39-45 (2000).	
	Rosenthal	Martin, Applied Spectroscopy, 52(7):1001–1007 (1998).	
	Amerena	Takeo et al., Applied Opliocs, 33(12):4267–4272 (1994).	
	Merrick	Thompson et al., <i>Physiol. Meas.</i> , 14:455–461 (1993).	
4,907,594 A 3/1990		Arimoto et al., Applied Spectroscopy, 58(12):1439–1445	
	Hirao et al. Bookspan	(2004), "Non–Contact Skin Moisture".	
	Clark et al.	Grandjean et al., <i>Nutrition Reviews</i> , 61(8):261–271 (2003,	
	Knudson	"Hydration Issues".	
	Sakai et al.	·	
	Mendelson et al.	Kotler, Am. J. Clin. Nutr. 64(suppl):489S–97S (1998),	
	Martens et al.	"Near-Infrared Spectroscopic".	
5,282,467 A 2/1994	Piantadosi et al.	Sowa et al., Journal of Surgical Research, 86:62–69 (1999),	
	Benaron	"Near-Infrared Spectroscopic".	
	Remiszewski et al.	Martin, "In vivo Measurements" Applied Spectroscopy,	
, ,	Hollub	v.52, No. 7, 1998.	
The state of the s	Thomas et al.	Wheeler, Owen H., "Near Infrared Spectra of Organic	
	Kuestner	Compounds," Department of Chemistry, College of Argri-	
	Steurer et al. Kotler	culture and Mechanic Arts, University of Puerto Rico (Mar.	
5,687,721 A 11/1997		1929).	
	Vari et al.	Pace, Nello, et al., "Studies on Body Composition: 111. The	
	Aoyagi et al.	body water and Chemically combined nitrogen content in	
	Tsoglin et al.	relation to fat content," Naval Medical Research Institute,	
	Godik	Bethesda, Maryland (Jan. 11, 1945).	
5,755,672 A 5/1998	Arai et al.	Mitchell, H. M., et al., The Chemical Composition of the	
5,788,643 A 8/1998	Feldman	Adult Human Body and its bearing on the Biochemistry of	
	Steuer et al.	Growth), Division of Animal Nutrition, Departments of	
	Dias et al.	Physiology and Animal Husbandry, University of Illinois,	
· ·	Osemwota	pp. 625–637 (Feb. 1945).	
	Baker, Jr. et al.	Schloerb, Paul R., et al., "The Measurement of Total Body	
	Kiani-Azarbayjany et al.	Water in the Human Subject by Deuterium Oxide Ditution,"	
	Kondo et al. Aldrich	Surgical Research Laboratories of the Peter Bent Brigham	
	Siconolfi	Hospital, and the Department of Surgery and the Biophysi-	
	Henderson et al.	cal Laboratory of the Harvard Medical School, pp.	
· · · · · · · · · · · · · · · · · · ·	Borgos et al.	1296–1310 (Mar. 20, 1950).	
	Misner et al.		
6,280,396 B1 8/2001	Clark	Forbes, R.M., et al., "The Composition of the Adult Human	
	Ghiassi et al.	Body as Determined by Chemical Analysis," Division of	
	Campbell et al.	Animal Nutrition, and the Department of Anatomy, Univer-	
	Finarov et al.	sity of Illinois, Jan. 19, 1953.	
	Rhee et al.	Bujis, K., et al., "Near–Infrared Studies of the Structure of	
	Wenzel et al. Monfre et al.	Water. 1. Pure Water," The Journal of Chemical Physics, vol.	
	Shimmick et al.	39, No. 8, pp. 2035–2041 (Oct. 15, 1963).	
6,600,946 B1 7/2003		Choppin, G.R., et al., "Near-Infrared Studies of the Struc-	
	Khalil et al.	ture of Water. 11. Ionic Soluation," The Journal of Chemical	
	Monfre et al.	Physics, vol. 39, No. 8, pp. 2042–2050 (Oct. 15, 1963).	
	Steuer et al.	Goldstein, R., et al., "The Near-Infrared Absorption of	
	Hazen et al.	Liquid Water at Temperatures Between 27 and 209°C," J.	
2001/0020122 A1 9/2001	Steuer et al.	Quant. Spectrosc. Radiat. Transfer., vol. 4, pp. 441-451	
		(1964).	
FOREIGN PATENT DOCUMENTS		Ben-Gera, l., et al., "Influence of Fat Concentration on the	
DE 19855521 A1	6/2000	Absorption Spectrum of Milk in the Near–Infrared Region,"	
EP 1135184 A1	6/2000	Isreal J. Agric. Res., vol. 18, No. 3, pp. 117–124 (Jul. 1968).	
EP 1184663 A2	3/2002	Houseman, R.A., et al., "The measurement of total body	
FR 2710517 A1	4/1995	water in living pigs by deuterium oxide dilution and its	
JP 4-40940	2/1992	relation to body composition," Br. J. Nutr., vol. 30, pp.	
JP 5-329163	12/1993	149–156 (1973).	
JP 11-244266	9/1999		
WO 98/34097 A1	8/1998	Krikorian, S. Edward, et al., "The Identification and orgin of	
WO WO 00/32262 A1	6/2000	N-H overtone and combination bands in the near-Infrared	
WO 00/71025 A1	11/2000	spectra of simple primary and secondary amides," Spectro-	
WO WO 93/13706 A2	1/2001	chimica Acta, vol. 29A, pp. 1233–1246 (1973).	

Lesser, G.T., et al., "Body water compartments with human aging using fat–free mass as the reference standard," *Am J. Physiol Regul Integr Comp Physiol.*, vol. 236, pp. 215–220 (1979).

Sheng, Hwai–Ping, et al., "A review of body composition studies with emphasis on total body water and fat," *The American Journal of Clinical Nutrition*, vol. 32., pp. 630–647 (Mar. 1979).

Martens, H., et al., "Unscrambling Multivariate Data from Mixtures: l: Fat, water and protein determination in meat by near–infrared reflectance spectroscopy, ll: soy protein and collagen determination in meat products from amino data," *Meat Res. Workers, Proc. European Meeting*, pp. 146–149 (1980).

Fomon, Samuel J., et al., "Body composition of reference children from birth to age 10 years," The American Journal of clinical Nutrition, vol. 35, pp. 1169–1175, (May 1982).

Lanza, Elaine, "Determination of Moisture, Protein, Fat, and Calories in Raw Pork and Beef by near Infrared Spectroscopy," *Journal of Food Science*, vol. 48, pp. 471–474 (1983).

Shields, R. G., Jr., et al., "Efficacy of Deuterium Oxide to Estimate Body Composition of Growing Swine", *Journal of Animal Science*, vol. 57, No. 1, pp. 66–73, (1983).

Wolfgang, Ameth, "Multivariate Infrared and near-infrared Spectroscopy: rapid analysis of protein, fat and water in meat," Food Res and Date Analysis, Proc from IUoST Symp, Oslo, Norway, pp. 239–251 (1983).

Cohn, S.H., et al., "Assessment of cellular mass and lean body mass by noninvasive nuclear techniques," *J. Lab Clin Med.*, vol. 105, pp. 305–311 (1985).

Hannon, John P., et al., Splenic red cell sequestration and Blood vol. 248, pp. R293–R301 (1985).

Potts, R.O., et al., "A Noninvasive, In Vivo Technique to Quantitatively measure Water Concentration of the Stratum Comeum Using Attenuated Total–Reflectance Infrared Spectroscopy," *Arch. Dermatol Res.*, vol. 277, pp. 489–495 (1985).

Cox, Patrick, et al., "Variations in Lipids in Different Layers of Porcine Epidermis," *J. Invest Dermatol.*, vol. 87, pp. 741–744 (1986).

Valdes, E. V., et al., "Determination of Crude Protein and Fat in Carcass and Breast Muscle Samples of Poultry by Near Infrared Reflectance Spectroscopy," *Poultry Science*, vol. 65, pp. 485–490 (1986).

Hedberg, Chrisopher L., et al., "The Time Course of Lipid Biosynthesis in Pig Epidermis," *J. Invest Dermatol.*, vol. 91, pp. 169–174 (1988).

Hedberg, Christopher L., et al., "The nonpolar Lipids of Pig Epidermis," *J. Invest Dermatol.*, vol. 90, pp. 225–229 (1988).

Trapp, Scott A., et al., "An improved spectrophotometric bromide assay for the estimation of extracellular water vol.," *Clinica Chimica Chimica Acta.*, vol. 181, pp. 207–212, (1989).

Bommannan, D., et al., "Examination of Stratum Comeum Barrier Function In Vivo by Infrared Spectroscopy," *J. Invest Dermatol*, vol. 95, pp. 403–408 (1990).

Hannon, John P., et al., "Normal pHysiological Values for Conscious Pigs Used in Biomedical Research," *Laboratory Animal Science*, vol. 40, No. 3, May 1990.

Mak, Vivien H.W., et al., "Oleic Acid Concentration and Effect in Human Stratum Commeum: Non–Invasive determination By Attenuated Total Reflectance Infrared Spectroscopy In Vivo," *Journal of Controlled Release*,vol. 12, pp. 67–75 (1990).

Drummer, C., et al., "Effects of an acute saline infusion on fluid and electrolyte metabolism in humans," *Am J. Physiol.*, vol. 262, pp. F744–F754 (1992).

Horber, F. F., et al., "Impact of hydration status on body composition as measured by dual energy X-ray absorptiometry in normal volunteers and patients on haemodialysis," *The British Journal of Radiology*, vol. 65, pp. 895–900 (1992).

Schmitt et al., *Proc. SPIE*, "Measurement of blood hematocrit by dual—wavelength near—IP photoplethysmography," 1641:150–161 (1992).

Diaz–Carrillo, E., et al., "Near infrared calibrations for goat's milk components; protein, total casein, α_3 -, β -and k-caseins, fat and lactose," *J. near Infrared Spectrosc.*, vol. 1, pp. 141–146 (1993).

Martin, K., "Direct Measurement of Moisture in Skin by NIR spectroscopy," *J. Soc. Cosmet. Chem.*, 44:249–261 (1993).

Richard, Stéphanie, et al., "Characterization of the Skin In Vivo by High Resolution Magnetic Resonance Imaging: Water Behavior and Age–Related Effects," *The Journal of Investigative Dermatology*, vol. 100, No. 5, pp. 705–709 (May 1993).

Bewig, Karen M., et al., "Discriminant Analysis of Vegatable Oils by Near–Infrared Reflectance Spectroscopy, "*JAOCS*, vol. 71, No. 2, pp. 195–200 (Feb. 1994).

Kamishikiryo-Yamashita, Hiromi, et al., "Protein Content in Milk by Near-Infrared Spectroscopy," *Journal of Food Science*, vol. 59, No. 2, pp. 313–315 (1994).

Matcher, S. J., et al., "Absolute quantification of deoxyhae-moglobin concentration in tissue near infrared spectroscopy," *Phys. Med. Biol.*, vol. 39, pp. 1295–1312 (1992).

Simanonok, Karl E., et al., "A Comprehensive Guyton Model Analysis of Physiologic Responses to Preadapting the Blood vol. as a Countermeasure to Fluid Shifts," *J. Clin Pharmacol*, vol. 34, pp. 440–453 (1994).

Steven, Alasdair C., et al., "Protein composition of comified cell envelopes of epidermal keratinocytes," *Journal of Cell Science*, vol. 107, pp. 693–700 (1994).

Warren, Joan L., et al., "The burden and Outcomes Associates with Dehydration among US Elderly, 1991," *American Journal of Public Health*, vol. 84, No. 8, pp. 1265–1269 (Aug. 1994).

Aneman, Anders, et al., "Splanchnic and Renal Sympathetic Activity in Relation to Hemodynamics During Isoflurane Administration in Pigs," *Anesth Analg.*, vol. 80, pp. 135–142, (1995).

Kisch, Hille, et al., "Accuracy and reproducibility of the measurement of actively circulating blood vol. with an integrated fiberoptic monitoring system," *Critical Care Medicine*, vol. 23, No. 5, pp. 885–893 (1995).

lsaksson, Tomas, et al., "Non-Destructive Determination of Fat, Moisture and Protein in Salmon Fillets by Use of Near-Infrared Diffuse Spectroscopy," *J. Sci Food Agric.*, vol. 69, pp. 95–100 (1995).

Quiniou, N., et al., "Predication of Tissular Body Composition from Protein and Deposition in Growing Pigs," *J. Anim. Sci.*, vol. 73, pp. 1567–1575 (1995).

Avis, N.J., et al.; "In vitro multifrequency electrical Impedance measurements and modeling of the cervix in late pregnancy", *Physiological Measurement*, vol. 17, pp. A97–103, 1996.

Gniadecka, M., et al., "Assessment of dermal water by high–Frequency ultrasound: comparative studies with nuclear magnetic resonance," *British Journal of Dermatology*, vol. 135, pp. 218–224, (1996).

Finn, Patrick J., et al., "Progressive celluari dehydration and proteolysis in critically ill patients," The Lancet, vol. 347, pp. 654–646 (Mar. 9, 1996).

Kumar, Gitesh, et al., "Non-Invasive Optical Assessment of Tissue Hydration," *International conference on Biomedical Engineering*, Jun. 3–5 1996, Hong Kong, pp. C2–5.

Schmitt et al., *Proc. SPIE*, "Optimum wavelengths for measurement of blood hemoglobin content and tissue hydration by NIR spectrophotometry," 2678:442–453 (1996).

De Fljter, W.M., et al., "Assessment of total body water ad lean body mass from anthropometry, Watson formula, creatinine kinetics, and body electrical impedance compared with antipyrine kinetics and peritoneal dialysis patients," *Nephrol Dial Transplant*, vol. 12, pp. 151–156 (1997).

Johansen, Lars Bo, et al., "Hemodilution central blood vol., and renal responses after an isotonic saline infusion in humans,", *Am J. Physiol.*, vol. 272, pp. R549–R556 (1997).

Visser, Marjolein, et al., "Density of fat-free body mass: relationship with race, age, and level of body fatness," *Am J. Physiol.*, vol. 272, pp. E781–E787, (1997).

Alanen, Esko, et al., "Measurement of dielectric properties of subcutaneous fat with open—ended coaxial sensors," *Phys. Med. Biol.*, vol. 43, pp. 475–485 (1998).

Bonadonna, Riccardo C., et al., "Tole of Tissue–Specific Blood Flow and Tissue Recruitment in Insulin–Mediated Glucose Uptake of Human Skeletal Musci," *Circulation*, vol. 98, pp. 234–241, (1998).

Bracco, David, et al., "Bedside determination of fluid accumulation after cardiac surgery using segmental bioelectrical impedance," *Crit Care Med*, vol. 26, No. 6, pp. 1065–1070 (1998).

Gniadecka, Monika, et al., "Water and Protein Structure in Photoaged and Chronically Aged Skin," *J. Invest Dermatol*, vol. 111, pp. 1129–1133 (1998).

Gniadecka, Monika, et al., "Structure of Water, Proteins, and Lipids in Intact Human Skin, Hair, and Nail," *J. Invest Dermatol*, vol. 110, pp. 393–398 (1998).

Gow, Kenneth W., et al., "Effect of crystalloid administration on oxygen extraction in endotoxemic pigs,", *J. Appl. Physiol.*, vol. 85, No. 5, pp. 1667–1675 (1998).

Husby, P., et al., "Midazolam–fentanyl–isoflurane anaesthesia is suitable for haemodynamic and fluid balance studies in pigs," *Laboratory Animals*, vol. 32, pp. 316–323 (1998).

Mitchell, A. D., et al., "Composition Analysis of Pork Carcasses by Dual–Energy X–Ray Absorptiometry," *J. Anim. Sci.*, vol. 76, pp. 2104–2114 (1998).

Mahan, D.C., et al., "Essential and Nonessential Amino Acid Composition of Pigs from Birth to 145 Kilograms of Body Weight, and Comparison to Other Studies," *J. Anim. Sci.*, vol. 76, pp. 513–521, (1998).

Martin, Kathleen, "In Vivo Measurements of Water in Skin by Near-Infrared Reflectance," *Applied Spectroscopy*, vol. 52, No. 7, 1998, pp. 1001–7.

Schou, Henning, et al., "Uncompensated Blood Los in not Tolerated During Acute Normovolemic Hemodilution in Anesthetized Pigs," *Anesth Analg.*, vol. 87, pp. 786–794 (1998)

Stranc, M.F., et al., "Assessment of tissue viability using near–infrared spectroscopy," *British Journal of Plastic Surgery*, vol. 51, pp. 210–217, (1998).

Thomas, B. J., et al., "Bioimpedance Spectrometry in the Determination of Body Water Compartments: Accuracy and Clinical Significance," *Appl. Radiat. Isot.*, vol. 49, No. 5/6, pp. 447–455, (1998).

Wilhelm, K.P., "Possible Pitfalls in Hydration Measurements," *Skin Bioengineering Techniques and Applications in Dermatology and Cosmetology*, vol. 26, pp. 223–234 (1998).

Vrhovski, Bernadette, et al., "Biochemistry of tropoelastin," Eur. J. Biochem., vol. 258, pp. 1–18 (1998).

Alanen, Esko, et al., "Penetration of electromagnetic fields of an open–ended coaxial probe between 1 MHz and 1 GHz in dielectric skin measurements," *Phys. Med. Biol.*, vol. 44 pp. N169–N176 (1999).

Dickens, Brian, et al., "Estimation of Concentration and Bonding Environment of Water Dissolved in Common Solvents Using Near Infrared Absorptivity," *J. Res. Natl. Inst. Stand. Technol.*, vol. 104, No. 2, pp. 173–183 (Mar.–Apr. 1999).

Formetti, Willa C., et al., "Reliability and validity of body composition measures in female athletes," Journal of Applied Physiology, vol. 87, pp. 1114–1122, (1999).

Fusch, Christoph, et al., "Neonatal Body COmponsition: Dual–Energy X–Ray Absorptiometry, Magnetic Resonance Imaging, and Three–Dimensional Chemical Shift Imaging versus Chemical Analysis in Piglets," *Pediatric Research*, vol. 46, No. 4, pp. 465–473 (1999).

Gudivaka, R., et al., "Single-and multifrequency models for bioelectrical impedance analysis of body water compartments," *J. Appl. Physiol.*, vol. 87, No. 3, pp. 1087–1096 (1999).

Jennings, Graham, et al., "The Use of Infrared Spectrophotometry for Measuring Body Water Spaces," vol. 45, No. 7, pp. 1077–1081 (1999).

Kalantar–Zadeh, Kamyar, et al., "Near infra–red interactance for nutritional assessment of dialysis patients," *Nephrol Dial Transplant*, vol. 14, pp. 169–175 (1999).

Kayser–Jones, Jeanie, et al., "Factors Contributing to Dehydration in Nursing Homes: lnadequate Staffing and Lack of Professional Supervision," *J. Am Geriatr. Soc.*, vol. 47, pp. 1187–1194 (1999).

Lange, Neale R., et al., "The measurement of lung water," *Critical Care*, vol. 3, pp. R19–R24 (1999).

Marken Lichtenbelt, Wouter D. Van, et al., Increased extracellular water compartment, relative to intracellular water compartment, after weight reduction, *Journal of Applied Physiology*, vol. 87, pp. 294–298 (1999).

Rennie, Michael J., "Perspectives —Teasing out the truth about collagen," *Journal of Physiology*, vol. 521, p. 1 (1999).

Wagner, J.R., et al., "Analysis of Body Composition Changes of Swine During Growth and Development," *J. Anim. Sci.*, vol. 77, pp. 1442–1466 (1999).

Wang, Zimian, et al., "Hydration of fat–free body mass: new physiological modeling approach," *Am. J. Physiol.*, vol. 276, pp. E995–E1003 (1999).

Wang, Zimian, et al., "Hydration of fat-free body mass: review and critique of a classic body-composition constant," *Am J. Clin. Nutr.*, vol. 69, pp. 833–841 (1999).

Ward, L., et al., "Multiple frequency bioelectrical impedance analysis: a cross-validation study of the inductor circuit and Cole models," *Physiol. Meas.*, vol. 20, pp. 333–347 (1999). Well, Jonathan CK, et al., "Four-component model of body composition in children: density and hydration of fat-free mass and comparison with simpler models," *Am J. Clin. Nutr.*, vol. 69, pp. 904–912 (1999).

Butte, Nancy F., et al., "Body Composition during the first 2 Years of Life; An Updated Reference," *Pediatric Research*, vol. 47, No. 5, pp. 578–585 (2000).

Feigenbaum, Matthew S., et al., "Contracted Plasma and Blood vol. in Chronic Heart Failure," *J Am Coll. Cardiol.*, vol. 35, No. 1, pp. 51–55 (Jan. 2000).

Kays, Sandra E., et al., "Predicting protein content by near infrared reflectance spectroscopy in diverse cereal food products," *J. Near Infrared Spectrosc.*, vol. 8, pp. 35–43 (2000).

Well, Jonathan CK, et al., "Four-component model of body composition in children: density and hydration of fat-free mass and comparsion with simpler models," *Am J. Clin. Nutr.*, vol. 69, pp. 904–912 (1999).

Plank, L. D., et al., "Similarity of Changes in Body Composition in Intensive Care Patients following Severe Sepsis or Major Blunt Injury," *Annals New York Academy of Sciences*, pp. 592–602 (2000).

Ritz, P., et al., "Body Water Spaces and Cellular Hydration during Healthy Aging," *Annals New York Academy of Sciences*, pp. 474–483 (2000).

Schoeller, Dale, "Bioelectrical Impedance Analysis —What does it measures?" Annals New York Academy of Sciences, pp. 159–162 (2000).

Stracher, Barry C., "Lung Elastin and Matrix," *Chest*, vol. 117, No. 5, pp. 2298–234S, May 2000 Supplement.

Young, A.E.R., et al., "Behaviour of near-infrared light in the adult human head: implications of clinical near-infrared spectroscopy," *British Journal of Anaesthesia*, vol. 84, No. 1, pp. 38–42 (2000).

Zembrzuski, Cora, "Nutrition and Hydration," Best Practices in Nursing Care to Older Adults, The Hartford Institute for Geriatric Nursing, vol. 2, No. 2, Sep. 2000, 2 pages.

for Geriatric Nursing, vol. 2, No. 2, Sep. 2000, 2 pages. Altas, Michael, et al., "Visualization of cutaneous hemoglobin oxygenation and skin hydration using near–infrared spectroscopic imaging," *Skin Research and Technology*, vol. 7, pp. 238–245, (2001).

Bray, George A., et al., "Evaluation of body fat in fatter and leaner 10-y-old African American and white children: The Baton Rouge Children's Study," *Am J. Clin Nutr*, vol. 73, pp. 687–702 (2001).

Campbell, Wayne W., et al., "The Recommended Dietary Allowance for Protein May Not Be Adequate for Older People to Maintain Skeletal Muscle," *Journal of Gerontology*, vol. 56A, No. 6, pp.M373–M–380 (2001).

Divert, Victor E., "Body Thermal State Influence on Local Skin Thermosensitivity," *International Journal of Circumpolar Health*, vol. 60, pp. 305–311 (2001).

Du, Y., et al., "Optical properties of porcine skin dermis between 900 nm and 1500 nm," *Phys. Med. Biol.*, vol. 46, pp. 167–181 (2001).

Endo, Yutaka, et al., "Water drinking causes a biphasic change in blood composition in humans," *Pflugers Arch—Eur J. Physiol*, vol. 442, pp. 362–368 (2001).

Garaulet, Marta, et al., "Site–specific differences in the fatty acid composition of abdominal adipose tissue in an obese population from a Mediterranean area: relation with dietary fatty acids, plasma lipid profile, Serum insulin, and central obesity," *Am J. Clin. Nutr.*, vol. 74, pp. 585–591 (2001).

Haga, Henning A., et al., "Electroencephalographic and cardiovascular indicators of nociception during isoflurane anaesthesia in pigs," *Veterinary Anaesthesis and Analgesia*, vol. 28, pp. 126–131 (2001).

Kalantar–Zadeh, Kamyar, et al., "Near infra–red interactactance for Longitudinal Assessment of Nutrition in Dialysis Patients," *Journal of Renal Nutrition*, vol. 11, No. 1, pp. 23–31 (Jan. 2001).

Kamba, Masayuki, et al., "Proton magnetic resonance spectroscopy for assessment of human body composition," *Am J. Clin. Nutr.*, vol. 73, pp. 172–176 (2001).

Lever, M., et al., "Some ways of looking at compensatory kosmotropes and different water environments," *Comparative Biochemistry and Physiolog.*, vol. 130, Part A, pp. 471–486, (2001).

Mingrone, G. et al., "Unreliable use of standard muscle hydration value in obesity," *Am J. Physiol Endocrinal Metab.*, vol. 280, pp. E365–371, (2001).

Šašic, Slobodan, et al., "Short–Wave Near–Infrared Spectroscopy of Biological Fluids. 1. Quantitative Analysis of Fat, Protein, and Lactose in Raw Milk by Partial Least–Squares Regression and Band Assignment," *Anal. Chem.*, vol. 73, pp. 64–71 (2001).

Schnickel, A.P., et al., "Evaluation of alternative measures of pork carcass composition," *J. Anim. Sci.*, vol. 79, pp. 1093–1119, (2001).

Sowa et al., "Near Infrared spectroscopic assessment of hemodynamic changes in the early post–burn period," *Burns*, 27(3):241–9 (2001).

Tsukahara, K., et al., "Dermal fluid translocation is an important determinant of the diurnal variation in human skin thickness," *British Journal of Dermatology*, vol. 145, pp. 590–596 (2001).

Vescovi, Jason D., et al., "Evalution of the BOD POD for estimating percentage body fat in a heterogeneous group of adult humans," *Eur J. Appl. Physiol.*, vol. 85, pp. 326–332 (2001).

Wang, Zimian, et al., "Magnitude and variation of ratio of total body potassium to fat–free mass: a cellular level modeling study," *Am J. Physiol. Endocrinal. Metab*, vol. 281, pp. E1–E7, (2001).

Watson, Walter, "Hydration of fat-free body mass: new physiological modeling approach," *Am J. Physiol. Endocrinol. Metab.*, Letters to the Editor, vol. 278, pp. E752–E753 (2001).

Ritz, Patrick, "Chronic Cellular Dehydration in the Aged Patient," Journal of Gerontology, vol. 56A, No. 6, pp. M349–M352 (2001).

Attas, E. Michael, et al., "Near–IR Spectroscopic Imaging for Skin Hydration: The Long and the short of it," *Biopolymers*, vol. 67, No. 2, pp. 96–106 (2002).

Garcia–Olmo, J., et al., "Advantages and disadvantages of multiple linear regression and partial least squares regression equations for the prediction of fatty acids," pp. 253–258 (undated).

Wang, Zimian, et al., "Cellular–Level Body Composition Model —A New Approach to Studying Fat–free Mass Hydration," *Annals New York Academy of Sciencei*, pp. 306–311 (undated).

Pig study: Hemorrhagic shock and fluid resuscitation

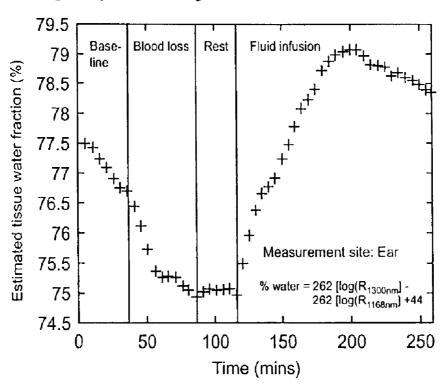


FIG. 1

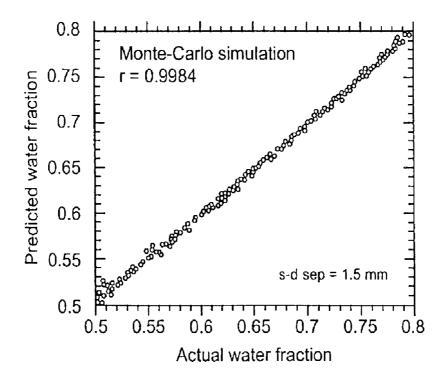


FIG. 2

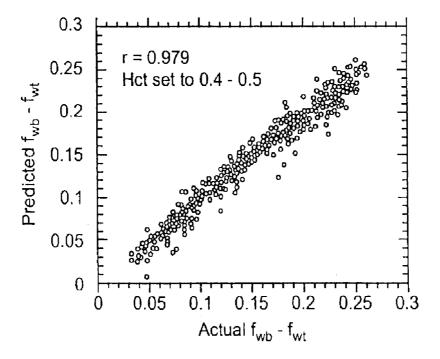


FIG. 3

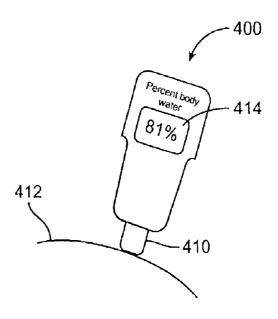


FIG. 4

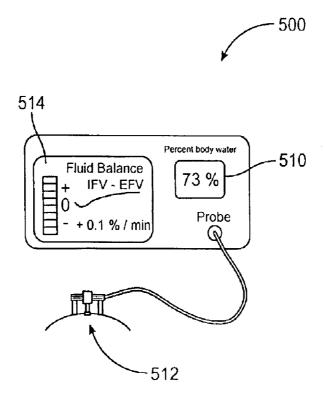


FIG. 5

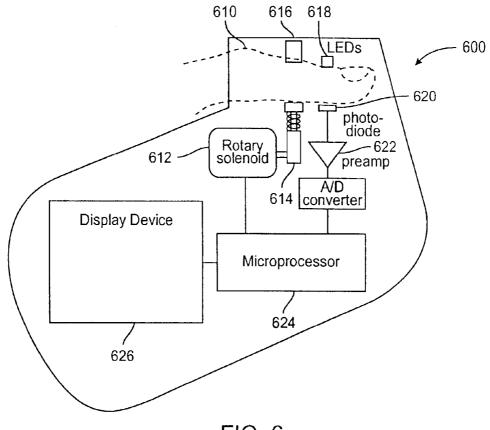


FIG. 6

DEVICE AND METHOD FOR MONITORING BODY FLUID AND ELECTROLYTE DISORDERS

CROSS REFERENCE TO RELATED APPLICATIONS

The application is a continuation application Ser. No. 09/810,918, filed on Mar. 16, 2001, which issued as U.S. Pat. No. 6,591,122, on Jul. 8, 2003.

BACKGROUND OF THE INVENTION

The maintenance of body fluid balance is of foremost concern in the care and treatment of critically ill patients, yet physicians have access to few diagnostic tools to assist them in this vital task. Patients with congestive heart failure, for example, frequently suffer from chronic systemic edema, which must be controlled within tight limits to ensure adequate tissue perfusion and prevent dangerous electrolyte disturbances. Dehydration of infants and children suffering from diarrhea can be life-threatening if not recognized and treated promptly.

The most common method for judging the severity of edema or dehydration is based on the interpretation of subjective clinical signs (e.g., swelling of limbs, dry mucous membranes), with additional information provided by measurements of the frequency of urination, heart rate, serum urea nitrogen SUN/creatinine ratios, and blood electrolyte levels. None of these variables alone, however, is a direct and quantitative measure of water retention or loss.

The indicator-dilution technique, which provides the most accurate direct measure of water in body tissues, is the present de facto standard for assessment of body fluid distribution. It is, however, an invasive technique that requires blood sampling. Additionally, a number of patents have disclosed designs of electrical impedance monitors for measurement of total body water. The electrical-impedance technique is based on measuring changes in the high-frequency (typically 10 KHz–1 MHz) electrical impedance of a portion of the body. Mixed results have been obtained with the electrical-impedance technique in clinical studies of body fluid disturbances as reported by various investigators. The rather poor accuracy of the technique seen in many studies point to unresolved deficiencies of these designs when applied in a clinical setting.

Therefore, there exists a need for methods and devices for monitoring total body water fractions which do not suffer from problems due to their being invasive, subjective and inaccurate.

SUMMARY OF THE INVENTION

Embodiments of the present invention provide devices and methods that measure body fluid-related metrics using spectrophotometry to facilitate therapeutic interventions aimed at restoring body fluid balance. The specific body 55 fluid-related metrics include the absolute volume fraction of water in the extravascular and intravascular tissue compartments, as well as the shifts of water between these two compartments. The absolute volume fraction of water is determined using algorithms where received radiation measured at two or more wavelengths are combined to form either a single ratio, a sum of ratios or ratio of ratios of the form $\log[R(\lambda_1)/R(\lambda_2)]$ in which the received radiation in the numerator depends primarily on the absorbance of water and the received radiation in the denominator depends primarily 65 on the absorbance of water and the sum of the absorbances of non-heme proteins and lipids in tissue.

2

The difference between the fraction of water in the intravascular fluid volume ("IFV") and extravascular fluid volume ("EFV") compartments are also determined using a differential method that takes advantage of the observation that pulsations caused by expansion of blood vessels in the skin, as the heart beats, produce changes in the received radiation at a particular wavelength that are proportional to the difference between the effective absorption of light in the blood and the surrounding tissue. This difference, integrated over time, provides a measure of the quantity of the fluid that shifts into and out of the capillaries. A mechanism for mechanically inducing a pulse is built into the device to improve the reliability of measurements of IFV–EFV under weak-pulse conditions.

For a fuller understanding of the nature and advantages of the embodiments of the present invention, reference should be made to the following detailed description taken in conjunction with the accompanying drawings.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 is a graph showing tissue water fraction measured on the ear of a pig during an experiment using reflectance measurements at two wavelengths.

FIG. 2 is a graph showing an example regression for prediction of water from reflectances measured at three wavelengths.

FIG. 3 is a graph showing an example regression of a two-wavelength algorithm for determination of the differ-30 ence between the intravascular and extravascular water fraction from pulsatile reflectances measured two wavelengths.

FIG. **4** is a diagram of an intermittent-mode version of a fluid monitor.

FIG. 5 is a diagram of a continuous-mode version of a fluid monitor.

FIG. 6 is a block diagram of a handheld apparatus for noninvasive measurement and display of tissue water.

DESCRIPTION OF THE SPECIFIC EMBODIMENTS

Embodiments of the present invention overcome the problems of invasiveness, subjectivity, and inaccuracy from which previous methods for body fluid assessment have suffered. The method of diffuse reflectance near-infrared ("NIR") spectroscopy is employed to measure the absolute fraction of water in skin. An increase or decrease in the free (non protein-bound) water content of the skin produces 50 unique alterations of its NIR reflectance spectrum in three primary bands of wavelengths (1100-1350 nm, 1500-1800 nm, and 2000-2300 nm) in which none-heme proteins (primarily collagen and elastin), lipids, and water absorb. According to the results of numerical simulations and experimental studies carried out by the inventor, the tissue water fraction f_{w} , defined spectroscopically as the ratio of the absorbance of water and the sum of the absorbances of none-heme proteins, lipids, and water in the tissue, can be measured accurately in the presence of nonspecific scattering variation, temperature, and other interfering variables.

In embodiments of this invention, the apparatus and its associated measurement algorithm are designed according to the following guidelines:

1. To avoid the shunting of light through the superficial layers of the epidermis, the light source and detector in optical reflectance probe have low numerical apertures, typically less than 0.3.

2. The spacing between the source and detector in the probe is in the range of 1–5 mm to confine the light primarily to the dermis.

- The reflectances are measured at wavelengths greater than 1150 nm to reduce the influence of hemoglobin 5 absorption.
- 4. To ensure that the expression that relates the measured reflectances and f_{w} yields estimates of water fraction that are insensitive to scattering variations, the lengths of the optical paths through the dermis at the wavelengths at which the reflectances are measured are matched as closely as possible. This matching is achieved by judicious selection of wavelength sets that have similar water absorption characteristics. Such wavelength sets may be selected from any one of the three primary wavelength bands (1100-1350 nm, 1500-1800 nm, and 2000-2300 nm) discussed above. Wavelength pairs or sets are chosen from within one of these three primary bands, and not from across the bands. More particularly the wavelength pair of 1180 and 1300 nm are one such wavelength set where the lengths of the optical paths through the dermis at these wavelengths are matched as closely as possible.
- 5. To ensure that the expression that relates the measured reflectances and f_{w} yields estimates of water fraction that are insensitive to temperature variations, the wavelengths at which the reflectances are measured are chosen to be either close to temperature isosbestic wavelengths in the water absorption spectrum or the reflectances are combined in a way that cancels the temperature dependencies of the individual reflectances. Typically, absorption peaks of various biological tissue components may shift with variations in temperature. Here, wavelengths are selected at points in the absorption spectrum where no significant temperature shift occurs. Alternately, by knowing the value of this temperature shift, wavelength sets may be chosen such that any temperature shift is mathematically canceled out when optical measurements are combined to compute the value of a tissue water metric. Such wavelength sets may be selected from any one of the three primary wavelength bands (1100-1350 nm, 1500-1800 nm, and 2000-2300 nm) discussed above. Wavelength pairs or sets are chosen from within one of these three primary bands, and not from across the bands. More particularly the wavelength pair of 1180 and 1300 nm are one such pair of temperature isosbestic wavelengths in the water absorption spectrum.
- 6. The reflectances measured at two or more wavelengths are combined to form either a single ratio, a sum of ratios or ratio of ratios of the form $\log[R(\lambda_1)/R(\lambda_2)]$ in which the reflectance in the numerator depends primarily on the absorbance of water and the reflectance in the denominator is nearly independent of the fraction of solids (lipids and proteins) in the tissue.

Thus, in one embodiment of the present invention the water fraction, f_w is estimated according to the following equation, based on the measurement of reflectances, $R(\lambda)$ at two wavelengths and the empirically chosen calibration constants c_0 and c_1 :

$$f_{w}=c_{1}\log[R(\lambda_{1})/R(\lambda_{2})]+c_{0} \tag{1}$$

Numerical simulations and in vitro experiments indicate that f_w can be estimated with an accuracy of approximately +/-2% over a range of water contents between 50 and 80% using Equation (1), with reflectances $R(\lambda)$ measured at two

4

wavelengths and the calibration constants c_0 and c_1 chosen empirically. Examples of suitable wavelength pairs are λ_1 =1300 nm, λ_2 =1168 nm, and λ_1 =1230 nm, λ_2 =1168 nm.

The ability to measure changes in the water content in the ear of a pig using two-wavelength NIR reflectometry was demonstrated experimentally in a study in which a massive hemorrhage was induced in a pig and the lost blood was replaced with lactated Ringer's solution over a period of several hours. Ringer's solution is a well-known solution of salts in boiled and purified water. FIG. 1 shows the water fraction in the skin of the ear of a pig, measured using Equation (1) with λ_1 =1300 nm and λ_2 =1168 nm. Referring to FIG. 1, it should be noted that experimental observations of concern to this embodiment commence when the lactated Ringer's solution was infused 120 minutes after the start of the experiment. It should also be noted that the drift in the water fraction from approximately 77.5% to 75% before the infusion is not related to this infusion experiment, but is related to the base-line hemorrhage portion of the experiment. The results show that the method of the present embodiment correctly reflects the effect of the infusion by showing an increase in tissue water fraction from approximately 75% to 79% while the infusion is continuing. These data suggest that the disclosed embodiment has a clear value as a monitor of rehydration therapy in a critical care setting.

In another embodiment of the present invention the water fraction, f_w is estimated according to Equation (2) below, based on the measurement of reflectances, $R(\lambda)$ at three wavelengths and the empirically chosen calibration constants c_0 , c_1 and c_2 :

$$f_{w} = c_{2} \log[R(\lambda_{1})/R(\lambda_{2})] + c_{1} \log[R(\lambda_{2})/R(\lambda_{3})] + c_{0}$$
 (2)

Better absolute accuracy can be attained using Equation (2) which incorporates reflectance measurements at an additional wavelength. The results of in vitro experiments on excised skin indicate that the wavelength triple (λ_1 =1190 nm, λ_2 =1170 nm, λ_3 =1274 nm) yields accurate estimates of skin water content based on Equation (2).

In yet another embodiment of the present invention the water fraction, f_w is estimated according to Equation (3) below, based on the measurement of reflectances, $R(\lambda)$ at three wavelengths and the empirically chosen calibration constants c_0 and c_1 :

$$f_w = c_1 \frac{\log[R(\lambda_1) / R(\lambda_2)]}{\log[R(\lambda_3) / R(\lambda_2)]} + c_0$$
(3)

Better absolute accuracy can be attained using Equations 50 (3), as is attained using Equations (2), which also incorporates reflectance measurements at an additional wavelength. Numerical simulations as shown in FIG. 2 indicate that an accuracy better than +/-0.5% can be achieved using Equation (3), with reflectances measured at three closely spaced wavelengths: λ_1 =1710 nm, λ_2 =1730 nm, and λ_3 =1740 nm.

Individuals skilled in the art of near-infrared spectroscopy would recognize that, provided that the aforementioned guidelines are followed, additional terms can be added to Equations (1)–(3) to incorporate reflectance measurements made at more than three wavelengths and thus improve accuracy further.

An additional embodiment of the disclosed invention provides the ability to quantify shifts of fluid into and out of the bloodstream through a novel application of pulse spectrophotometry. This additional embodiment takes advantage of the observation that pulsations caused by expansion of blood vessels in the skin as the heart beats produce changes

in the reflectance at a particular wavelength that are proportional to the difference between the effective absorption of light in the blood and the surrounding interstitial tissues. Numerical simulation indicate that, if wavelengths are chosen at which water absorption is sufficiently strong, the difference between the fractions of water in the blood, f_w^{blood} and surrounding tissue, f_w^{tissue} is proportional to the ratio of the denormalized reflectance changes ($\Delta R/R$) measured at two wavelengths, according to Equation (4) below:

$$f_w^{blood} - f_w^{fissue} = c_1 \left(\frac{\Delta R}{R}\right)_{\lambda_1} / \left(\frac{\Delta R}{R}\right)_{\lambda_2} + c_0,$$
 (4)

where c_0 and c_1 are empirically determined calibration constants. This difference, integrated over time, provides a measure of the quantity of fluid that shifts into and out of the capillaries. FIG. 3 shows the prediction accuracy expected for the wavelength pair λ_1 =1320 nm and λ_2 =1160 nm.

FIGS. 4 and 5 show diagrams of two different versions of an instrument for measuring the amount of water in body 20 tissues. The simplest version of the instrument 400 shown in FIG. 4 is designed for handheld operation and functions as a spot checker. Pressing the spring-loaded probe head 410 against the skin 412 automatically activates the display of percent tissue water 414. The use of the spring-loaded probe 25 head provides the advantages of automatically activating the display device when needed and turning the device off when not in use, thereby extending device and battery life. Moreover, this unique use of a spring-loaded probe also provides the force needed to improve the reliability of 30 measurements. Percent tissue water represents the absolute percentage of water in the skin beneath the probe (typically in the range 0.6-0.9). The force exerted by a spring or hydraulic mechanism (not shown) inside the probe head 410 pushes out most of the blood in the skin below the probe to 35 reduce the error caused by averaging the intravascular and extravascular fluid fractions. A pressure transducer (not shown) within the probe head 410 measures the compressibility of the skin for deriving an index of the fraction of free

The more advanced version of the fluid monitor 500 shown in FIG. 5 is designed for use as a critical-care monitor. In addition to providing a continuous display of the absolute volume fraction of water 510 at the site of measurement 512, it also provides a trend display of the time- 45 averaged difference between the intravascular fluid volume ("IFV") and extravascular fluid volume ("EFV") fractions 514, updated every few seconds. This latter feature would give the physician immediate feedback on the net movement of water into or out of the blood and permit rapid evaluation 50 of the effectiveness of diuretic or rehydration therapy. To measure the IFV-EFV difference, the monitor records blood pulses in a manner similar to a pulse oximeter. Therefore, placement of the probe on the finger or other well-perfused area of the body would be required. In cases in which 55 perfusion is too poor to obtain reliable pulse signals, the IFV-EFV display would be blanked, but the extravascular water fraction would continue to be displayed. A mechanism for mechanically inducing the pulse is built into the probe to improve the reliability of the measurement of IFV-EFV 60 under weak-pulse conditions.

FIG. 6. is a block diagram of a handheld device 600 for measuring tissue water fraction within the IFV and the EFV, as well as shifts in water between these two compartments with a pulse inducing mechanism. Using this device 600, 65 patient places his/her finger 610 in the probe housing. Rotary solenoid 612 acting through linkage 614 and collar 616

6

induces a mechanical pulse to improve the reliability of the measurement of IFV-EFV. LEDs 618 emit light at selected wavelengths and photodiode 620 measure the transmitted light. Alternately, the photodiode 620 can be placed adjacent to the LEDs to allow for the measurement of the reflectance of the emitted light. Preamplifier 622 magnifies the detected signal for processing by the microprocessor 624. Microprocessor 624, using algorithms described above, determines the tissue water fraction within the IFV and the EFV, as well as shifts in water between these two compartments, and prepares this information for display on display device 626. Microprocessor 624 is also programmed to handle the appropriate timing between the rotary solenoid's operation and the signal acquisition and processing. The design of the device and the microprocessor integrates the method and apparatus for reducing the effect of noise on measuring physiological parameters as described in U.S. Pat. No. 5,853,364, assigned to Nellcor Puritan Bennett, Inc., now a division of the assignee of the present invention, the entire disclosure of which is hereby incorporated herein by reference. Additionally, the design of the device and the microprocessor also integrates the electronic processor as described in U.S. Pat. No. 5,348,004, assigned to Nellcor Incorporated, now a division of the assignee of the present invention, the entire disclosure of which is hereby incorporated herein by reference.

As will be understood by those skilled in the art, other equivalent or alternative methods for the measurement of tissue water fraction within the IFV and the EFV, as well as shifts in water between these two compartments according to the embodiments of the present invention can be envisioned without departing from the essential characteristics thereof. For example, the device can be operated in either a handheld or a tabletop mode, and it can be operated intermittently or continuously. Moreover, individuals skilled in the art of near-infrared spectroscopy would recognize that additional terms can be added to the algorithms used herein to incorporate reflectance measurements made at more than three wavelengths and thus improve accuracy further. Also, light sources or light emission optics other then LED's including and not limited to incandescent light and narrowband light sources appropriately tuned to the desired wavelengths and associated light detection optics may be placed within the probe housing which is placed near the tissue location or may be positioned within a remote unit; and which deliver light to and receive light from the probe location via optical fibers. Additionally, although the specification describes embodiments functioning in a backscattering or a reflection mode to make optical measurements of reflectances, other embodiments can be working in a forward-scattering or a transmission mode to make these measurements. These equivalents and alternatives along with obvious changes and modifications are intended to be included within the scope of the present invention. Accordingly, the foregoing disclosure is intended to be illustrative, but not limiting, of the scope of the invention which is set forth in the following claims.

What is claimed is:

- 1. A device for measuring body fluid-related metrics using optical spectrophotometry comprising:
 - a probe housing configured to be placed proximal to a tissue location which is being monitored;

light emission optics connected to said housing and configured to direct radiation at said tissue location;

light detection optics connected to said housing and configured to receive radiation from said tissue location; and

- a processing device configured to process radiation from said light emission optics and said light detection optics to compute said body fluid-related metrics, wherein said light emission optics emit radiation at a plurality of spectral wavelengths chosen to be preferentially absorbed by tissue water, non-heme proteins and lipids, where preferentially absorbed wavelengths are wavelengths whose absorption is substantially independent of the individual concentrations of non-heme proteins and lipids, and is substantially dependent on the sum of the individual concentrations of non-heme proteins and lipids.
- 2. The device of claim 1, further comprising a display device connected to said probe housing and configured to display said body fluid-related metrics.
- 3. The device of claim 1, wherein said body-fluid metrics are monitored intermittently.
- **4**. The device of claim **1**, wherein said body-fluid metrics are monitored continuously.
- **5**. The probe housing of the device of claim **1** further 20 comprising a pressure transducer to measure the compressibility of tissue for deriving an index of a fraction of free water within said tissue.
- **6**. The device of claim **1**, wherein said wavelengths are chosen to ensure that measured received radiation are substantially insensitive to scattering variations and such that the optical path lengths through the dermis at said wavelengths are substantially equal.
- 7. The device of claim 1, wherein said wavelengths are chosen to ensure that measured received radiation from said 30 tissue location are insensitive to temperature variations, where said wavelengths are temperature isosbestic in the water absorption spectrum or said received radiation are combined in a way that substantially cancel temperature dependencies of said individual received radiation when 35 computing tissue water fractions.
- **8**. The device of claim **1**, wherein said wavelengths are chosen from one of three primary bands of wavelengths of approximately 1100–1350 nm, approximately 1500–1800 nm and approximately 2000–2300 nm.
- **9**. The device of claim **1**, wherein said light emission optics and said light detection optics are mounted within said probe housing and positioned with appropriate alignment to enable detection in a transmissive mode.
- 10. The device of claim 1, wherein said light emission 45 optics and said light detection optics are mounted within said probe housing and positioned with appropriate alignment to enable detection in a reflective mode.
- 11. The device of claim 1, wherein said light emission optics and said light detection optics are placed within a 50 remote unit and which deliver light to and receive light from said probe housing via optical fibers.
- 12. The device of claim 1, wherein said light emission optics comprise at least one of a (a) incandescent light source, (b) white light source, and (c) light emitting diode 55 ("LED").
- 13. The device of claim 1, wherein said processing device receives and compares at least two sets of optical measurements, where the at least first set of optical measurements corresponds to the detection of light whose 60 absorption is primarily due to water, lipids and non-heme proteins, and where the at least second set of optical measurements corresponds to the detection of light whose absorption is primarily due to water, and where a comparison of said at least two optical measurements provides a 65 measure of the absolute water fraction within said tissue location.

8

- 14. The device of claim 1, wherein said processing device receives and compares at least two sets of optical measurements, where said at least two sets of optical measurements are based on received radiation from at least two wavelengths and which are combined to form either a single ratio of said received radiation, a sum of ratios of said received radiation.
- 15. The device of claim 1, wherein said processing device receives and compares at least two sets of optical measurements from at least two different wavelengths, where absorption of light at said at least two different wavelengths is primarily due to water which is in the vascular blood and in the extravascular tissue, and where a ratio of said at least two measurements provides a measure of a difference between the fractions of water in the blood and surrounding tissue location.
- **16**. The device of claim **1**, wherein said body fluid-related metrics comprise tissue water fraction, and where said tissue water fraction, f_w is determined such that $f_w = c_1 \log[R(\lambda_1)/R(\lambda_2)] + c_0$, and where:

calibration constants c_0 and c_1 are chosen empirically; $R(\lambda_1)$ is a received radiation at a first wavelength; and $R(\lambda_2)$ is a received radiation at a second wavelength.

- 17. The device of claim 16 wherein said first and second wavelengths are approximately 1300 nm and approximately 1168 nm respectively.
- **18**. The device of claim **16** wherein said first and second wavelengths are approximately 1230 nm and approximately 1168 nm respectively.
- 19. The device of claim 1, wherein said body fluid-related metrics comprise tissue water fraction, and where said tissue water fraction, f_w is determined such that $f_w = c_2 \log[R(\lambda_1)/R(\lambda_2)] + c_1 \log[R(\lambda_2)/R(\lambda_3)] + c_0$, and where:

calibration constants c_0 , c_1 and c_2 are chosen empirically; $R(\lambda_1)$ is a received radiation at a first wavelength;

- $R(\lambda_2)$ is a received radiation at a second wavelength; and $R(\lambda_3)$ is a received radiation at a third wavelength.
- **20**. The device of claim **19** wherein said first, second and third wavelengths are approximately 1190 nm, approximately 1170 nm and approximately 1274 nm respectively.
- 21. The device of claim 1, wherein said body fluid-related metrics comprises tissue water fraction, and where said tissue water fraction, f_w is determined such that

$$f_w = c_1 \frac{\log[R(\lambda_1)/R(\lambda_2)]}{\log[R(\lambda_3)/R(\lambda_2)]} + c_0,$$

and where:

calibration constants c_0 and c_1 are chosen empirically;

 $R(\lambda_1)$ is a received radiation at a first wavelength;

 $R(\lambda_2)$ is a received radiation at a second wavelength; and $R(\lambda_3)$ is a received radiation at a third wavelength.

- 22. The device of claim 21, wherein said first, second and third wavelengths are approximately 1710 nm, approximately 1730 nm and approximately 1740 nm.
- 23. The device of claim 1, wherein said body fluid-related metrics comprises a quantified measure of a difference between the water fraction in the blood and the water fraction in the extravascular tissue, where said difference is determined such that

$$f_{w}^{blood} - f_{w}^{tissue} = c_{1} \left(\frac{\Delta R}{R}\right)_{\lambda_{1}} / \left(\frac{\Delta R}{R}\right)_{\lambda_{2}} + c_{0},$$

and where:

 $f_w^{\ blood}$ is the water fraction in the blood; $f_w^{\ tissue}$ is the water fraction in the extravascular tissue; calibration constants c_0 and c_1 are chosen empirically; and

$$\left(\frac{\Delta R}{R}\right)_{\lambda_1} / \left(\frac{\Delta R}{R}\right)_{\lambda_2}$$

is the ratio of dc-normalized received radiation changes at a first wavelength, λ_1 and a second wavelength, λ_2 respectively, where said received radiation changes are caused by a pulsation caused by expansion of blood vessels in tissue.

- 24. The device of claim 23 wherein the body fluid-metric further comprises an integral of said difference between the water fraction in the blood and the water fraction in the extravascular tissue to provide a measure of the water that shifts into and out of the capillaries.
- **25**. The device of claim **24** wherein said first and second wavelengths are approximately 1320 nm and approximately 1160 nm respectively.
- 26. A device for measuring body fluid-related metrics using optical spectrophotometry comprising:
 - a probe housing configured to be placed proximal to a $_{30}$ tissue location which is being monitored;

light emission optics connected to said housing and configured to direct radiation at said tissue location;

- light detection optics connected to said housing and configured to receive radiation from said tissue loca- 35 tion; and
- a processing device configured to process radiation from said light emission optics and said light detection optics to compute said body fluid-related metrics, wherein said light emission optics emit radiation at a plurality of 40 spectral wavelengths falling within the range of 1100 to 2300 nm and the separation between said light emission optics and said light detection optics falls within the range of 1 and 5 mm.
- 27. The device of claim 26, wherein said wavelengths are 45 chosen to ensure that measured received radiation are substantially insensitive to scattering variations and such that the optical path lengths through the dermis at said wavelengths are substantially equal.
- 28. The device of claim 26, wherein said wavelengths are 50 chosen to ensure that measured received radiation from said tissue location are insensitive to temperature variations, where said wavelengths are temperature isosbestic in the water absorption spectrum or said received radiation are combined in a way that substantially cancel temperature 55 dependencies of said individual received radiation when computing tissue water fractions.
- 29. The device of claim 26, wherein said wavelengths are chosen from one of three primary bands of wavelengths of approximately 1100–1350 nm, approximately 1500–1800 nm and approximately 2000–2300 nm.
- 30. The device of claim 26, wherein said processing device receives and compares at least two sets of optical measurements, where the at least first set of optical measurements corresponds to the detection of light whose absorption is primarily due to water, lipids and non-heme proteins, and where the at least second set of optical mea-

surements corresponds to the detection of light whose absorption is primarily due to water, and where a comparison of said at least two optical measurements provides a measure of the absolute water fraction within said tissue location.

- 31. The device of claim 26, wherein said processing device receives and compares at least two sets of optical measurements, where said at least two sets of optical measurements are based on received radiation from at least two wavelengths and which are combined to form either a single ratio of said received radiation, a sum of ratios of said received radiation or ratios of ratios of said received radiation.
- 32. The device of claim 26, wherein said body fluid-related metrics comprise tissue water fraction, and where said tissue water fraction, f_w is determined such that f_w = c_1 log[$R(\lambda_1)/R(\lambda_2)$]+ c_0 , and where:

calibration constants c_0 and c_1 are chosen empirically;

- $R(\lambda_1)$ is a received radiation at a first wavelength; and $R(\lambda_2)$ is a received radiation at a second wavelength.
- 33. The tissue water fraction as determined in claim 32, wherein said first and second wavelengths are approximately 1300 nm and approximately 1168 nm respectively.
- **34**. The tissue water fraction as determined in claim **32**, wherein said first and second wavelengths are approximately 1230 nm and approximately 1168 nm respectively.
- **35**. The device of claim **26**, wherein said body fluid-related metrics comprise tissue water fraction, and where said tissue water fraction, f_w is determined such that f_w = c_2 log[$R(\lambda_1)/R(\lambda_2)$]+ c_1 log[$R(\lambda_2)/R(\lambda_3)$]+ c_0 , and where:

calibration constants c_0 , c_1 and c_2 are chosen empirically; $R(\lambda_1)$ is a received radiation at a first wavelength;

 $R(\lambda_2)$ is a received radiation at a second wavelength; and $R(\lambda_3)$ is a received radiation at a third wavelength.

- **36**. The tissue water fraction as determined in claim **35**, wherein said first, second and third wavelengths are approximately 1190 nm, approximately 1170 nm and approximately 1274 nm respectively.
- 37. A device for measuring body fluid-related metrics using optical spectrophotometry comprising:
 - a probe housing configured to be placed proximal to a tissue location which is being monitored;

light emission optics connected to said housing and configured to direct radiation at said tissue location;

- light detection optics connected to said housing and configured to receive radiation from said tissue location; and
- a processing device configured to process radiation from said light emission optics and said light detection optics to compute said body fluid-related metrics, wherein said body fluid-related metrics comprises tissue water fraction, and where said tissue water fraction, f_w is determined such that

$$f_w = c_1 \frac{\log[R(\lambda_1)/R(\lambda_2)]}{\log[R(\lambda_3)/R(\lambda_2)]} + c_0$$

and where:

calibration constants \mathbf{c}_0 and \mathbf{c}_1 are chosen empirically;

 $R(\lambda_{\scriptscriptstyle 1})$ is a received radiation at a first wavelength;

 $R(\lambda_2)$ is a received radiation at a second wavelength; and $R(\lambda_3)$ is a received radiation at a third wavelength.

38. The device of claim 37, wherein said wavelengths are chosen to ensure that measured received radiation are sub-

stantially insensitive to scattering variations and such that the optical path lengths through the dermis at said wavelengths are substantially equal.

11

- 39. The device of claim 37, wherein said wavelengths are chosen to ensure that measured received radiation from said tissue location are insensitive to temperature variations, where said wavelengths are temperature isosbestic in the water absorption spectrum or said received radiation are combined in a way that substantially cancel temperature dependencies of said individual received radiation when computing tissue water fractions.
- **40**. The device of claim **37**, wherein said wavelengths are chosen from one of three primary bands of wavelengths of approximately 1100–1350 nm, approximately 1500–1800 nm and approximately 2000–2300 nm.
- 41. The device of claim 37, wherein said processing device receives and compares at least two sets of optical measurements, where the at least first set of optical measurements corresponds to the detection of light whose absorption is primarily due to water, lipids and non-heme proteins, and where the at least second set of optical measurements corresponds to the detection of light whose absorption is primarily due to water, and where a comparison of said at least two optical measurements provides a measure of the absolute water fraction within said tissue location.
- **42**. The device of claim **37**, wherein said first, second and third wavelengths are approximately 1710 nm, approximately 1730 nm and approximately 1740 nm.
- **43**. A device for measuring body fluid-related metrics using optical spectrophotometry comprising:
 - a probe housing configured to be placed proximal to a tissue location which is being monitored;
 - light emission optics connected to said housing and configured to direct radiation at said tissue location;
 - light detection optics connected to said housing and configured to receive radiation from said tissue location; and
 - a processing device configured to process radiation from said light emission optics and said light detection optics 40 to compute said body fluid-related metrics, wherein said light emission optics emit radiation at a plurality of spectral wavelengths chosen to ensure that said computed body fluid-related metrics are substantially insensitive to scattering variations and such that the optical 45 path lengths through said tissue location at said wavelengths are substantially equal.
- 44. The device of claim 43, wherein said wavelengths are chosen to ensure that measured received radiation from said tissue location are insensitive to temperature variations, 50 where said wavelengths are temperature isosbestic in the water absorption spectrum or said received radiation are combined in a way that substantially cancel temperature dependencies of said individual received radiation when computing tissue water fractions.
- **45**. The device of claim **43**, wherein said wavelengths are chosen from one of three primary bands of wavelengths of approximately 1100–1350 nm, approximately 1500–1800 nm and approximately 2000–2300 nm.
- **46**. The device of claim **43**, wherein said processing 60 device receives and compares at least two sets of optical measurements, where the at least first set of optical measurements corresponds to the detection of light whose absorption is primarily due to water, lipids and non-heme proteins, and where the at least second set of optical measurements corresponds to the detection of light whose absorption is primarily due to water, and where a compari-

son of said at least two optical measurements provides a measure of the absolute water fraction within said tissue location.

12

- 47. The device of claim 43, wherein said processing device receives and compares at least two sets of optical measurements, where said at least two sets of optical measurements are based on received radiation from at least two wavelengths and which are combined to form either a single ratio of said received radiation, a sum of ratios of said received radiation.
- **48**. The device of claim **43**, wherein said body fluid-related metrics comprise tissue water fraction, and where said tissue water fraction, f_w is determined such that f_w = c_1 15 $log[R(\lambda_1)/R(\lambda_2)]+c_0$, and where:

calibration constants c_0 and c_1 are chosen empirically; $R(\lambda_1)$ is a received radiation at a first wavelength; and $R(\lambda_2)$ is a received radiation at a second wavelength.

- **49**. The tissue water fraction as determined in claim **48**, wherein said first and second wavelengths are approximately 1300 nm and approximately 1168 nm respectively.
- **50**. The tissue water fraction as determined in claim **48**, wherein said first and second wavelengths are approximately 1230 nm and approximately 1168 nm respectively.
- **51**. The device of claim **43**, wherein said body fluid-related metrics comprise tissue water fraction, and where said tissue water fraction, f_w is determined such that f_w = c_2 log[$R(\lambda_1)/R(\lambda_2)$]+ c_1 log[$R(\lambda_2)/R(\lambda_3)$]+ c_0 , and where:

calibration constants c_0 , c_1 and c_2 are chosen empirically; $R(\lambda_1)$ is a received radiation at a first wavelength;

- $R(\lambda_2)$ is a received radiation at a second wavelength; and $R(\lambda_3)$ is a received radiation at a third wavelength.
- **52**. The tissue water fraction as determined in claim **51**, wherein said first, second and third wavelengths are approximately 1190 nm, approximately 1170 nm and approximately 1274 nm respectively.
- **53**. A device for measuring body fluid-related metrics using optical spectrophotometry comprising:
 - a probe housing configured to be placed proximal to a tissue location which is being monitored;
 - light emission optics connected to said housing and configured to direct radiation at said tissue location;
 - light detection optics connected to said housing and configured to receive radiation from said tissue location; and
 - a processing device configured to process radiation from said light emission optics and said light detection optics to compute said body fluid-related metrics, wherein said light emission optics emit radiation at a plurality of spectral wavelengths chosen to ensure that measured received radiation from said tissue location are insensitive to temperature variations, wherein said wavelengths are temperature isosbestic in the water absorption spectrum or the received radiation signals are combined in a way that substantially cancel temperature dependencies of said individual received radiation signals when computing tissue water fractions.
- **54**. The device of claim **53**, wherein said wavelengths are chosen from one of three primary bands of wavelengths of approximately 1100–1350 nm, approximately 1500–1800 nm and approximately 2000–2300 nm.
- 55. The device of claim 53, wherein said processing device receives and compares at least two sets of optical measurements, where the at least first set of optical measurements corresponds to the detection of light whose

absorption is primarily due to water, lipids and non-heme proteins, and where the at least second set of optical measurements corresponds to the detection of light whose absorption is primarily due to water, and where a comparison of said at least two optical measurements provides a 5 measure of the absolute water fraction within said tissue location.

- **56**. The device of claim **53**, wherein said processing device receives and compares at least two sets of optical measurements, where said at least two sets of optical measurements are based on received radiation from at least two wavelengths and which are combined to form either a single ratio of said received radiation, a sum of ratios of said received radiation.
- **57**. The device of claim **53**, wherein said body fluid-related metrics comprise tissue water fraction, and where said tissue water fraction, f_w is determined such that f_w = c_1 log[$R(\lambda_1)/R(\lambda_2)$]+ c_0 , and where:

calibration constants c_0 and c_1 are chosen empirically; $R(\lambda_1)$ is a received radiation at a first wavelength; and

14

 $R(\lambda_2)$ is a received radiation at a second wavelength.

- **58**. The tissue water fraction as determined in claim **57**, wherein said first and second wavelengths are approximately 1300 nm and approximately 1168 nm respectively.
- **59**. The tissue water fraction as determined in claim **57**, wherein said first and second wavelengths are approximately 1230 nm and approximately 1168 nm respectively.
- **60**. The device of claim **53**, wherein said body fluid-related metrics comprise tissue water fraction, and where said tissue water fraction, f_w is determined such that f_w = c_2 log[$R(\lambda_1)/R(\lambda_2)$]+ c_1 log[$R(\lambda_2)/R(\lambda_3)$]+ c_0 , and where:

calibration constants c_0 , c_1 and c_2 are chosen empirically; $R(\lambda_1)$ is a received radiation at a first wavelength;

- $R(\lambda_2)$ is a received radiation at a second wavelength; and $R(\lambda_3)$ is a received radiation at a third wavelength.
- **61**. The tissue water fraction as determined in claim **60**, wherein said first, second and third wavelengths are approximately 1190 nm, approximately 1170 nm and approximately 1274 nm respectively.

* * * * *



专利名称(译)	用于监测体液和电解质紊乱的装置和方法			
公开(公告)号	<u>US7236811</u>	公开(公告)日	2007-06-26	
申请号	US10/441943	申请日	2003-05-20	
[标]申请(专利权)人(译)	马林克罗特公司			
申请(专利权)人(译)	马林克罗特INC.			
当前申请(专利权)人(译)	COVIDIEN LP			
[标]发明人	SCHMITT JOSEPH M			
发明人	SCHMITT, JOSEPH M.			
IPC分类号	A61B5/1455 A61B5/00 A61B5/024 A61B5/145 G01N21/35			
CPC分类号	A61B5/0048 A61B5/0059 A61B5/14546 A61B5/4878 G01N21/359 A61B5/02438 A61B5/0053			
其他公开文献	US20030220548A1			
外部链接	Espacenet USPTO			

摘要(译)

一种用于使用分光光度法测量体液相关度量的装置和方法,以促进旨在恢复体液平衡的治疗干预。特定的体液相关指标包括血管外和血管内组织隔室中水的绝对体积分数,以及这两个隔室之间的水位变化。使用算法确定水的绝对体积分数,其中在两个或更多个波长处测量的接收辐射被组合以形成单个比率,比率之和或比率之比log [R (λ1)/R(λ2)]其中分子中接收的辐射主要取决于水的吸光度,分母中接收的辐射主要取决于水的吸光度和组织中非血红素蛋白质,脂质和水的吸光度之和。血管内液体体积("IFV")和血管外液体积("EFV")隔室中的水分数之间的差异也使用差分方法确定,该方法利用观察到由血管扩张引起的脉动。作为心脏跳动的皮肤在特定波长处产生接收辐射的变化,该变化与血液中的光和周围组织的有效吸收之间的差异成比例。随时间积分的这种差异提供了进出毛细管的流体量的量度。设备内置有机械诱导脉冲的机制,以提高弱脉冲条件下IFV-EFV测量的可靠性。

